ETV2 functions as a pioneer factor and regulates the endothelial lineage

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**Introduction**

Ischemic heart and vascular diseases are common and deadly and result in considerable morbidity and mortality. Progression of these diseases results in myocardial infarction requiring vascular bypass grafting or limb amputation and these therapeutic interventions have significant limitations. Therefore, new therapies are warranted. One strategy is to enhance or facilitate vascular development and requires the definition of the earliest mechanisms that govern endothelial and vascular fate determination.

During development, pioneer transcription factors have the essential and unique role of opening new regulatory chromatin landscapes on genomic DNA. Compared with the non-pioneer transcription factors, the pioneer factors can recognize their target DNA sequences in closed heterochromatin and trigger the remodeling of the adjacent chromatin landscape to provide accessibility to non-pioneer transcription factors1. Through the relaxation of the chromatin state, these pioneer factors enable other transcription factors to sequentially access their binding motifs through histone modifications and collectively activate gene expression resulting in reprogramming the cell fate. A limited number of pioneer factors have been defined and all of these factors have been shown to have important roles in various biological processes. Examples of pioneer factors include pluripotent factors OSK (Oct4, Sox2, and Klf4) for iPSC reprogramming, FoxA for hepatocyte reprogramming2, GATA3 for mesenchymal to epithelial transition (MET)3, PU.1 and C/EBPa for macrophage differentiation4, Pax7 for pituitary development5, Ebf1 for B cell lineage specification6 and Ascl1 for neurogenesis7.

Studies from our laboratory and others have identified ETV2 as an essential transcription factor for the development of cardiac, endothelial and hematopoietic lineages. Our laboratory has previously shown, using the 3.9kb *Etv2-Cre* and *Rosa-EYFP* reporter alleles, that *Etv2*-EYFP progenitors daughter endocardial/endothelial and hematopoietic lineages in the WT background. Furthermore, when crossed into the *Etv2* mutant background, the *Etv2-*EYFP progenitor cells daughtered cardiomyocytes in the absence of *Etv2*. *Etv2* has been shown to be responsive to BMP, WNT and NOTCH signaling pathways and synergizes with FOXC2 to regulate the endothelial program by directly targeting *Lmo2,*  *Cdh5*, *Tie2* and *Flk1.* Importantly, the ability of ETV2 to reprogram fibroblasts to endothelial cells points to its master regulatory role and supports the notion that ETV2 functions as a pioneer factor in endothelial development.

In the present study, we used two distinct biological systems: ES/EB differentiation and MEF reprogramming, to define the role for Etv2 as a pioneer factor that drives endothelial development. Even though these two model systems have very different global expression, chromatin accessibility and epigenetic profiles, we surprisingly found similar molecular programs and downstream genes that were regulated by Etv2 induction. We showed that Etv2 cooperates with Brg1, an ATPase of the SWI/SNF chromatin remodeling enzyme, and together they function to relax the closed chromatin and convert the nucleosome-enriched to the nucleosome-depleted region during the endothelial lineage development in both MEFs and EBs. To our knowledge, this is the first study that highlights the pioneer function of Etv2 in two distinct systems.

**Results**

**Etv2 reprograms fibroblasts into an endothelial cell population**

Using the *iHA-Etv2* ES cell line, we engineered a mouse that inducibly overexpresses ETV2. We isolated embryonic fibroblasts from this mouse line and demonstrated that these cell populations uniformly expressed fibroblast markers (Thy1.2, CD44 and CD29) and lacked HE expression by FACS (in the absence of Dox) (Supplementary Figure 1a). Using western blot analysis, we further demonstrated that ETV2 was robustly expressed within 3 hrs post-Dox treatment (Supplementary Figure 1b). ETV2 overexpression resulted in more than a 50-fold increase in cells expressing FLK1/TIE2 by FACS (Supplementary Figure 1c - 1f). At day 7 post-Etv2 induction, we found Ac-LDL uptake, positive sprouting assays and an induction of ETV2 downstream endothelial target expression (Supplementary Figure 1g - 1m).

To further investigate the molecular dynamics of Etv2 inducible reprogramming, we captured and sequenced 3,539, 2,936 and 7,202 high quality single cells from 24 hours, 48 hours and 7 days post-induction of Etv2 in mouse embryonic fibroblasts (MEFs). We also performed the scRNA-seq of 948 undifferentiated MEF cells as well as 827 sorted Flk1+ cells from day 7 reprogrammed cells. The dimension reduction analysis by scVI8, followed by uniform manifold approximation and projection (UMAP) and k-means clustering identified seven distinct cell clusters (Figure 1a, and Figure 1c-1d). We noted that during the first 48 hours of reprogramming, even though Etv2 was significantly up-regulated as early as 24 hours, endothelial markers such as Kdr, Lmo2, Emcn, Cdh5 and Sox18 were significantly activated only in a subpopulation of cells (cluster 1 for MEFs and day 1 cells and cluster 2 for day 2 cells) (Figure 1d, Supplementary Figure 2a and 2b). The pathway analysis suggested that the cell cycle process and chromatin organization related genes were significantly up-regulated in cluster 1 MEF cells, and may be required for the initiation of Etv2 induced reprogramming process (Supplementary Figure 2c). At day 7 of reprogramming, the FACS analysis showed that 17% of the cells were Flk1+ cells (Supplementary Figure 1c and 1d). The scRNA-seq showed that the Flk1+ cells at day 7 formed a unique cell population (cluster 7). The endothelial marker genes such as Lmo2 and Emcn were robustly expressed in the cluster 7 cells, while the fibroblast markers such as Cd44 and Fosl1 were down-regulated (Figure 1f). The pathway analysis confirmed that genes with functions related to vasculature development and blood vessel development were more abundantly expressed in cluster 7 compared with the remaining clusters (Figure 1g). In summary, the scRNA-seq analysis suggested that inducing Etv2 in MEFs activated the downstream endothelial marker genes and promoted the development of endothelial lineages.

The embryoid body (EB) formation from ES cells has been broadly used for studying the role of Etv2 in endothelial development9. To investigate the molecular programs that are commonly or differentially responsive to the Etv2 induction in EBs and MEFs, we induced Etv2 at day 2.5 developing EBs and performed bulk RNA-seq of Flk1+ cells at 3 hours post-induction (Figure 1b and Supplementary Figure 3). Compared with the gene expression profiles in D2.5 EBs without induction, we identified 2,324 and 2,328 genes that were up- and down-regulated, respectively. Among them, 554 and 1,507 genes were up- and down-regulated in both EBs and MEFs (Supplementary Figure 4a and 4b). As expected, the commonly up-regulated genes were closely related to vasculature development, including endothelial markers Myct1 and Sox18, while the commonly down-regulated genes were significantly associated with the translation and RNA processing (Supplementary Figure 4c - 4f).

To examine the chromatin accessibility changes in Etv2 induced cell differentiation, we also performed ATAC-seq (Assays for Transposase-Accessible Chromatin with high-throughput sequencing) at 24 hours, 48 hours and 7 days post induction in MEFs and 3 hours post induction in EB (Figure 1a and 1b). The analysis of transcription factors (TF) associated chromatin accessibility analysis suggested that although there existed significant batch effects between MEFs and EBs (PC1 in Figure 1h), the Flk1+ cells from day 7 MEFs and day 2.5 EBs post-Etv2 induction share a common global chromatin accessibility pattern (PC2 in Figure 1h), including 113 and 246 transcription factors whose chromatin accessibility that were commonly up-regulated and down-regulated in MEFs and EBs, respectively (Supplementary Figure 5a - 5d). By integrating the RNA-seq and ATAC-seq datasets from MEFs and EBs, we identified 13 TFs whose expression levels and TF associated accessibility consistently increased in Flk1+ cells in both MEFs and EBs, and 18 TFs decreased in Flk1+ cells (Figure 1i and Supplementary 5e and 5f). It is interesting that mesodermal factors such as Msx2, Eomes and Foxk2 were among the genes that were consistently down-regulated in both MEFs and EBs, suggesting the critical role of Etv2 as a suppressor of non-endothelial lineage development.

Collectively, these results demonstrated that induction of Etv2 was able to promote the cell differentiation toward the endothelial lineage and the suppression of non-endothelial lineages, in two very distinct cellular environments: MEFs and EBs, supporting the notion that Etv2 functioned as a master regulator to drive the endothelial lineage development.

**Etv2 targets nucleosomes during reprogramming**

To investigate how Etv2 binding reshapes the genomic accessibility landscape and drives the endothelial lineage differentiation in different cellular environments, we performed Etv2 ChIP-seq at 24 hours, 48 hours and 7 days post-Etv2 induction in reprogrammed MEFs, and Etv2 ChIP-seq at 3 hours and 12 hours post-Etv2 induction in developing EBs at day 2.5. The initial Etv2 binding events were captured by the Etv2 ChIP-seq at 24 hours in MEFs and 3 hours in EBs, resulting in 18,024 and 131,001 peaks, respectively (Figure 2a). The 11,751 common Etv2 peaks overlapped between MEFs and EBs and represented the majority (65.2%) of the ETV2 peaks in EBs, and were located predominately in the promoter region. In contrast, the EB and MEF specific Etv2 peaks were more likely distributed at the distal intergenic regions (Figure 2b). One of the key features of pioneer factors is their capability of targeting the nucleosomes. To test whether Etv2 was able to target the nucleosome, we first examined the nucleosome profiles of 24 hours Etv2 ChIP-seq peaks in undifferentiated MEFs using published histone H3 ChIP-seq and MNase-seq datasets10. We divided the Etv2 ChIP-seq peaks into four quartiles based on the mean MNase-seq signals of the central 200-bp region, and we used the first (lowest mean signal) and the fourth quartile (highest mean signal) to represent the nucleosome free region (NFR) and the nucleosome (Figure 2c and Supplementary Figure 6a). Thus, the nucleosome quartile represented the Etv2 peaks with their summits located at the nucleosome centers, suggesting that Etv2, like other reported pioneer factors, was also able to target the nucleosome during MEF reprogramming. Similarly, we divided the Etv2 ChIP-seq peaks at 3h post-induction in day 2.5 EB into NFR (5,291 peaks) and nucleosome (8,843 peaks) groups according to the local V-plot and fragment size profiles of ATAC-seq day 2.5 EB without Etv2 induction (Supplementary Figure 6b-6d). We found that similar to the MEF reprogramming, Etv2 also targets the nucleosome centers at day 2.5 EB differentiation (Figure 2d). Our results in MEFs and EBs suggested that Etv2 was intrinsically able to target and bind the nucleosomes, and this capability appeared to be independent of cellular context. Sequence motif analysis identified a common GGAAAT motif that was significantly more enriched in NFR regions compared with the nucleosomes in both MEFs and EBs (Fisher's adjusted p-value=6.0E-05 and 6.8E-5) (Figure 2e and Supplementary Figure 7a). This motif has an additional terminal "AT" nucleotides compared with the canonical motifs identified from the canonical Etv2 motifs. These findings suggested that Etv2, similar to other pioneer factors, such as Oct4, Sox2 and Klf4, were able to target nucleosome-enriched sites using partial or degenerate motifs, and targeted their full canonical motif in nucleosome-depleted sites1.

Moreover, we found that Etv2 targeted nucleosome regions were characterized by low Brg1 ChIP-seq signals at 24 hours post induction in MEFs and 3 hours post induction in EBs, suggesting that the initial recognition of the nucleosome may not require Brg1 (Figure 2c and 2d). Interestingly, the Etv2 targeted nucleosomes had a distinct H3K27ac surrounding pattern in MEFs and EBs: Etv2 targeted H3K27ac depleted nucleosomes in MEFs and H3K27ac enriched ones in EBs, suggesting that Etv2 recognized the nucleosomes regardless of H3K27ac enrichment.

**Brg1 maintains and stabilizes the binding of Etv2**

To further elaborate how Etv2 induction drives the endothelial lineage development during the reprogramming of MEFs and ES/EB differentiation, we performed Etv2 ChIP-seq at 48 hours and 7 days post Etv2 induction in reprogramming MEFs and Etv2 ChIP-seq at 12 hours post induction in EBs. We identified in total 154,468 and 19,651 non-overlapping Etv2 peaks in MEFs and EBs, respectively. Similar to the OSK-binding sites during the reprogramming toward iPS, the majority (more than 80%) of the unique Etv2 peaks were only present at early stages (day 1 for MEFs and 3 hours for EBs) but absent during the latter stages (day 7 for MEFs and 12 hours for EBs), suggesting that most of the initial Etv2 binding events were short-lived and were not sustained during differentiation (Supplementary Figure 8a and 8b). We divided the Etv2 peaks into "early", "late" and "sustained" groups according to whether Etv2 peaks were present in the early stage, late stage or both stages (Figure 3a and 3b). Although we previously found that the majority of initial Etv2 binding events were independent of Brg1, the "late" Etv2 peaks were associated with increasing levels of Brg1 in the late stage of EBs and MEFs, while the "sustained" Etv2 peaks were associated with the highest levels of Brg1 in the early stage. Note that the establishment of "late" Etv2 peaks were also coupled with the increasing H3K27ac enrichment, in contrast to the "sustained" Etv2 peaks that were initially located at a high H3K27ac environment. These results suggested a significant role for Brg1 on the maintainance and stabilization of the binding of Etv2 during the reprogramming period and facilitation of Etv2 activation of downstream endothelial programs. Indeed, we found there were significant overlap between the "late" Etv2 peaks, as well as the nearby genes (Figure 3c and 3d). Moreover, the "late" Etv2 peaks were located near a higher proportion of endothelial genes, in contrast to the "early" Etv2 peaks (Figure 3e). Similar to Ascl1 induced neural reprogramming, we also found the significant phasing events around the Etv2 binding motifs in the Flk1+ cells in both MEFs and EBs (Supplementary Figure 9)11. In summary, these results suggested that Brg1 maintained and stabilized Etv2 binding, promoted the local H3K27ac enrichment, and activated the downstream endothelial genes (Figure 3f).

**Etv2 requires Brg1 to activate downstream genes during reprogramming**

Brg1 is required for ATP-dependent pioneer factors such as GATAs to open the chromatin. To examine whether Brg1 is also required by Etv2 to perform its pioneer function, we knocked down Brg1 in iHA-Etv2 MEFs using shRNAs 48 hours prior to the doxycycline induction of Etv2, and continued inducing Etv2 at 48 hours, 4 days and 6 days post induction (Supplementary Figure 10a). We found that the Brg1 RNA levels and the Flk1+ cell populations were significantly reduced at 24 hours, 48 hours and 7 days post induction, compared with the control Etv2 induced MEF reprogramming (Supplementary Figure 10b and 10c). Moreover, we found that the expression levels of several downstream endothelial genes were also significantly reduced at 7 days post induction when Brg1 was knocked down, suggesting that Brg1 is required for Etv2 to play its pioneer role in open the chromatin and activate the downstream endothelial target genes (Supplementary Figure 10e-10n).

**Figure Legends**

**Figure 1. Etv2 promotes the endothelial program in both MEF and EBs. (a)** Schematic of the reprogramming strategy in iHA-Etv2 MEFs by overexpression of Etv2 with doxycycline (Dox). **(b)** Schematic of the differentiation of embryoid body (EB) and induction of Etv2 at day 2.5 of differentiation. **(c-d)** The UMAP plot shows the scRNA-seq of 948 undifferentiated MEFs, 3,539 reprogrammed cells at 24 hours, 2,936 cells at 48 hours and 7,202 cells at 7 days and 827 Flk1+ cells at 7 days post-induction of Etv2 in MEFs. The dimension reduction analysis by scVI, followed by uniform manifold approximation and projection (UMAP) and k-means clustering identified seven distinct cell clusters. The color represents **(c)** the cell sources and **(d)** cell clusters. **(e)** The expression profiles of Etv2 and Kdr (Flk1). **(f)** The volcano plot of genes differentially expressed between cluster 1 and cluster 7. The *p*-values were determined by Wilcoxon ran sum test of the normalized read counts. **(g)** The biological process that are significantly associated with the up-regulated in genes in cluster 7 (Flk1+ cells at day 7 of reprogramming) compared with cluster 1 (undifferentiated MEFs). **(h)** The PCA of the TF deviations of the ATAC-seq of MEF reprogramming (MEF, 24 hours, 48 hours and 7 days post induction) and EB differentiation (2.5 day and 3 hours post induction). The TF deviations were inferred by chromVAR. **(i)** The 31 transcription factors which expression levels and motif associated chromatin accessibility showed consistently directional change in both EB and MEF (13 up-regulated TFs and 18 down-regulated TFs).

**Figure 2. Etv2 targets nucleosomes during reprogramming. (a)** The Venn diagram shows 131,001 and 18,024 Etv2 ChIP-seq peaks at 24 hours post induction in MEF reprogramming and 3 hours post induction in day 2.5 EB, respectively. There are 11,751 common Etv2 peaks overlapped between MEF and EB. **(b)** The genomic distribution of EB specific, MEF specific and common Etv2 peaks. The EB and MEF specific Etv2 peaks were more likely distributed at the distal intergenic regions. **(c)** The heatmap shows the read density of MNase-seq, Brg1 ChIP-seq and H3K27ac ChIP-seq in MEFs, surrounding 131,001 Etv2 ChIP-seq peaks at 24 hours post induction in MEF reprogramming. The Etv2 peaks were divided into four quartiles based on the mean MNase-seq signals of the central 200-bp region. The first (lowest mean signal) and the fourth quartile (highest mean signal) were used to represent the nucleosome free region (NFR) and nucleosome. **(d)** The heatmap shows the ratio of NFR / nucleosome reads density, reads density of Brg1 ChIP-seq and H3K27ac ChIP-seq at EB day 2.5, surrounding 18,024 Etv2 ChIP-seq at 3 hours post induction. The Etv2 peaks were divided into NFR (5,291 peaks) and nucleosome (8,843 peaks) groups according to the local V-plot and fragment size profiles of ATAC-seq day 2.5 EB without Etv2 induction. **(e)** Sequence motif analysis by DREME and CentriMo identified a common GGAAAT motif that are significantly more enriched in NFR regions compared with the nucleosomes in both MEF and EB (Fisher's adjusted p-value=6.0E-05 and 6.8E-5).

**Figure 3.** **Brg1 maintains and stabilize the binding of Etv2. (a)** The heatmap shows the fold enrichment of Etv2 ChIP-seq, Brg1 ChIP-seq and H3K27ac ChIP-seq, at 24 hours, 48 hours and 7 days post-induction of Etv2. We also included the Brg1 and H3K27ac ChIP-seq at undifferentiated MEF. Each site is centered at the 154,468 non-overlapping Etv2 ChIP-seq summits in MEF reprogramming. The Etv2 peaks into "early", "late" and "sustained" groups according to whether Etv2 peaks were present in 24 hours post induction (early stage), 48 hours post induction (late stage) or both stages. **(b)** The heatmap shows the fold enrichment of Etv2 ChIP-seq, Brg1 ChIP-seq and H3K27ac ChIP-seq, at 3 hours and 12 hours post-induction of Etv2 in day 2.5 EB. We also included the Brg1 and H3K27ac ChIP-seq at day 2.5 EB without induction. Each site is centered at the 19,651 non-overlapping Etv2 ChIP-seq summits in EB differentiation. The Etv2 peaks into "early", "late" and "sustained" groups according to whether Etv2 peaks were present in 3 hours post induction (early stage), 12 hours post induction (late stage) or both stages. **(c-d)** The Venn diagram shows the overlap between late Etv2 peaks in MEF and EB, as well as the overlap between the genes near the late Etv2 peaks. **(e)** The percent of blood vessel development related genes near the early, late or sustainedEtv2 peaks in EB and MEF. (f) The model of Etv2 induced reprogramming. Etv2 targets the nucleosome during the early stage of reprogramming. Brg1 help to maintain the stabilize the Etv2 binding near the endothelial genes, coupled with the increasing of local H3K27ac levels.

**References**

1. Soufi, A. *et al.* Pioneer transcription factors target partial DNA motifs on nucleosomes to initiate reprogramming. *Cell* **161**, 555–568 (2015).   
  
2. Gualdi, R. *et al.* Hepatic specification of the gut endoderm in vitro: cell signaling and transcriptional control. *Gene Dev* **10**, 1670–1682 (1996).   
  
3. Takaku, M. *et al.* GATA3-dependent cellular reprogramming requires activation-domain dependent recruitment of a chromatin remodeler. *Genome Biol* **17**, 36 (2016).   
  
4. van Oevelen, C. *et al.* C/EBPα Activates Pre-existing and De Novo Macrophage Enhancers during Induced Pre-B Cell Transdifferentiation and Myelopoiesis. *Stem Cell Rep* **5**, 232–247 (2015).   
  
5. Mayran, A. *et al.* Pioneer and nonpioneer cooperation drives lineage specific chromatin opening. *bioRxiv* 472647 (2019) doi:10.1101/472647 .   
  
6. Györy, I. *et al.* Transcription factor Ebf1 regulates differentiation stage-specific signaling, proliferation, and survival of B cells. *Gene Dev* **26**, 668–82 (2012).   
  
7. Wapinski, O. L. *et al.* Hierarchical mechanisms for direct reprogramming of fibroblasts to neurons. *Cell* **155**, 621–635 (2013).   
  
8. Lopez, R., Regier, J., Cole, M. B., Jordan, M. I. & Yosef, N. Deep generative modeling for single-cell transcriptomics. *Nat Methods* **15**, 1053–1058 (2018).   
  
9. Rasmussen, T. L. *et al.* ER71 directs mesodermal fate decisions during embryogenesis. *Development (Cambridge, England)* **138**, 4801–4812 (2011).   
  
10. Chronis, C. *et al.* Cooperative Binding of Transcription Factors Orchestrates Reprogramming. *Cell* (2017) doi:10.1016/j.cell.2016.12.016 .   
  
11. Wapinski, O. L. *et al.* Rapid Chromatin Switch in the Direct Reprogramming of Fibroblasts to Neurons. *Cell reports* **20**, 3236–3247 (2017).