

# 1   **Multimodal Integration of Alzheimer's Plasma Biomarkers, MRI,** 2   **and Genetic Risk for Individual Prediction of Cerebral Amyloid** 3   **Burden**

4  
5   Yichen Wang<sup>1,2,3#</sup>, Lairai H.J. Chen<sup>1,2,3#</sup>, Yuxin Cheng<sup>4</sup>, Yuyan Cheng<sup>5</sup>, Shiyun Zhao<sup>5</sup>,  
6   Yidong Jiang<sup>1</sup>, Tianyu Bai<sup>1,2,3</sup>, Yanxi Huo<sup>1,2,3</sup>, Kexin Wang<sup>1,2,3</sup>, Mingkai Zhang<sup>6</sup>, Weijie  
7   Huang<sup>7</sup>, Guozheng Feng<sup>8</sup>, Ying Han<sup>6,9,10,11,12,13\*</sup>, Ni Shu<sup>1,2,3\*</sup>

8  
9   <sup>1</sup>State Key Laboratory of Cognitive Neuroscience and Learning & IDG/McGovern  
10   Institute for Brain Research, Beijing Normal University, Beijing, China

11   <sup>2</sup>BABRI Centre, Beijing Normal University, Beijing, China

12   <sup>3</sup>Beijing Key Laboratory of Brain Imaging and Connectomics, Beijing Normal University,  
13   Beijing, China

14   <sup>4</sup>State Key Laboratory of Oil and Gas Reservoir Geology and Exploitation, Southwest  
15   Petroleum University, Chengdu, 610500, Sichuan, P. R. China

16   <sup>5</sup>School of Mathematical Sciences, Beijing Normal University, Beijing, China

17   <sup>6</sup>Department of Neurology, XuanWu Hospital of Capital Medical University, Beijing,  
18   China

19   <sup>7</sup>College of Artificial Intelligence, Nanjing University of Aeronautics and Astronautics,  
20   MOE Key Laboratory of Brain Computer Intelligence Technology, Nanjing 211106,  
21   China

1 <sup>8</sup>Tri-Institutional Center for Translational Research in Neuroimaging and Data Science  
2 (TReNDS), Atlanta, GA, USA

3 <sup>9</sup>School of Biomedical Engineering, Hainan University, Haikou 570228, China

4 <sup>10</sup>Center of Alzheimer's Disease, Beijing Institute for Brain Disorders, Beijing 100053,  
5 China

6 <sup>11</sup>National Clinical Research Center for Geriatric Diseases, Beijing 100053, China

7 <sup>12</sup>The Central Hospital of Karamay, Xinjiang 834000, China

8 <sup>13</sup>Institute of Biomedical Engineering, Shenzhen Bay Laboratory, Shenzhen 518132,  
9 China

10 Yichen Wang and Lairai H.J. Chen contributed equally to this work.

# 11 **Corresponding author**

12 \*Ni Shu, Email: [nshu@bnu.edu.cn](mailto:nshu@bnu.edu.cn), No. 19, Xijiekouwai St, Haidian District, Beijing,  
13 100875, P. R. China

14 \*Han Ying, Email: [hanying@xwh.ccmu.edu.cn](mailto:hanying@xwh.ccmu.edu.cn), XuanWu Hospital of Capital Medical  
15 32 University, Beijing, 100053, China

# 16 **ORCID**

17 Yichen Wang: <https://orcid.org/0009-0006-4283-7986>

18 Lairai H.J. Chen: <https://orcid.org/0000-0002-4362-1056>

19 Yuxin Cheng: <https://orcid.org/0009-0008-7684-0723>

20 Yuyan Cheng: <https://orcid.org/0009-0000-1355-7383>

21 Shiyun Zhao: <https://orcid.org/0009-0008-3816-1367>

- 1 Yidong Jiang: <https://orcid.org/0009-0005-4406-9549>
- 2 Tianyu Bai: <https://orcid.org/0009-0002-9357-5497>
- 3 Yanxi Huo: <https://orcid.org/0009-0002-8448-3275>
- 4 Mingkai Zhang: <https://orcid.org/0009-0001-9846-123X>
- 5 Weijie Huang: <https://orcid.org/0000-0002-2481-1188>
- 6 Guozheng Feng: <https://orcid.org/0000-0002-6937-8592>
- 7 Han Ying: <https://orcid.org/0000-0003-0377-7424>
- 8 Ni Shu: <https://orcid.org/0000-0003-2420-2910>
- 9

## Abstract

Alzheimer's disease (AD), the most prevalent neurodegenerative disorder, is marked by the accumulation of amyloid- $\beta$  (A $\beta$ ) plaques. Although cerebral A $\beta$  positron emission tomography (A $\beta$ -PET) remains the gold standard for assessing cerebral A $\beta$  burden, its clinical utility is hindered by cost, radiation exposure, and limited availability. Plasma biomarkers serve as promising non-invasive predictors of cerebral A $\beta$  burden, but reliance on a single marker often leads to suboptimal predictive performance. To address this, we proposed a multimodal machine learning strategy that integrates readily accessible and non-invasive features—such as plasma biomarkers, structural magnetic resonance imaging (sMRI)-derived atrophy measures, diffusion tensor imaging (DTI)-based structural connectomes (SCs), and genetic risk profiles—to predict cerebral A $\beta$  burden and evaluate the relative contribution of each modality to predictive performance. Specifically, a random forest regressor was trained using data from the Alzheimer's Disease Neuroimaging Initiative (ADNI;  $n = 150$ ) and evaluated with leave-one-out cross-validation. Our results showed that integrating multimodal features improves the predictive power on cerebral amyloid burden: while the baseline model using plasma and clinical variables alone achieved an  $R^2$  of 0.52, adding neuroimaging and apolipoprotein E (APOE) genotype features improved performance ( $R^2 = 0.617$ ), and replacing APOE with polygenic risk scores (PRS) further enhanced accuracy ( $R^2 = 0.637$ ). The predictive value of multimodal integration was also replicated in an independent cohort (SILCODE;  $n = 101$ ). Moreover, a multiclass classifier trained with the same multimodal features achieved high accuracy in distinguishing clinical stages of A $\beta$  burden—normal controls (NC), mild cognitive impairment (MCI), and Alzheimer's disease (AD)—with area under the curve (AUC) values of 0.86, 0.77, and 0.93, respectively. These findings highlight the value of combining plasma, imaging, and genetic data to non-invasively estimate cerebral A $\beta$  burden, offering a potential alternative to PET imaging for early AD risk assessment.

1    **Keyword:** Alzheimer’s disease; Amyloid- $\beta$  burden; Plasma biomarkers; Structural MRI;  
2    Structural connectome; Polygenic risk score; APOE  $\epsilon$ 4 genotype; Multimodal machine  
3    learning  
4

1

## 2 1 Introduction

3 Alzheimer's disease (AD) is the most prevalent neurodegenerative disorder, characterized  
 4 by the accumulation of amyloid- $\beta$  ( $A\beta$ ) plaques and neurofibrillary tangles, both of which  
 5 are closely linked to cognitive decline<sup>1,2</sup>.  $A\beta$  positron emission tomography ( $A\beta$ -PET)  
 6 remains the gold standard for in vivo quantification of cerebral  $A\beta$  burden, providing  
 7 region-specific estimates of amyloid deposition<sup>3</sup>. However, its high cost, invasive nature,  
 8 and limited accessibility hinder its applicability for large-scale or routine individual  
 9 screening.

10 Plasma biomarkers—including phosphorylated tau (reflecting tau hyperphosphorylation),  
 11  $\beta$ -amyloid 42/40 ratio ( $A\beta_{42}/A\beta_{40}$ ; indexing amyloid burden), neurofilament light chain  
 12 (NfL; indicating axonal injury), and glial fibrillary acidic protein (GFAP; reflecting  
 13 astrocytic activation)—have shown promise as accessible, non-invasive indicators for  
 14 estimating cerebral  $A\beta$  burden<sup>4</sup>. However, their standalone predictive performance  
 15 remains limited, particularly at the individual level, due to biological heterogeneity and  
 16 the indirect nature of peripheral measurements. Structural MRI (sMRI) and diffusion  
 17 tensor imaging (DTI) provide non-invasive insights into brain changes associated with  
 18 Alzheimer's disease<sup>5</sup>. sMRI characterizes regional atrophy, whereas DTI-derived  
 19 structural connectomes (SCs), capturing inter-regional white matter connectivity<sup>6,7</sup>.  
 20 Genetic factors also modulate the predictive capacity of plasma biomarkers. The

1    apolipoprotein E (APOE)  $\epsilon 4$  allele, the most established genetic risk factor for AD<sup>8</sup>, is  
 2    easily genotyped and widely used in clinical settings. Polygenic risk scores (PRS),  
 3    although theoretically more comprehensive by aggregating genome-wide variants, are  
 4    often limited by their cost and complexity<sup>9</sup>. Both may provide complementary information  
 5    beyond plasma biomarkers<sup>10,11</sup>. However, given the higher cost of PRS assessment, it is  
 6    necessary to consider the feasibility of using APOE genotype as a cost-effective proxy.

7    Together, plasma, neuroimaging, and genetic biomarkers capture distinct yet  
 8    complementary dimensions of Alzheimer's disease pathophysiology<sup>12</sup>. However,  
 9    integrating these heterogeneous data sources remains challenging due to their different  
 10    dimensionality and complex interdependencies. Among machine learning approaches,  
 11    random forest (RF) algorithms are particularly well-suited for modeling multimodal  
 12    biomedical data<sup>13</sup>. RFs can effectively accommodate nonlinear relationships, and provide  
 13    interpretable feature importance metrics, making them ideal for individualized prediction  
 14    of cerebral A $\beta$  burden.

15    Previous studies have explored A $\beta$  burden prediction using selected data modalities, but  
 16    few have integrated multiple complementary biological domains, such as plasma, imaging,  
 17    and genetics. For example, Ramanan et al. demonstrated that adding genetic risk scores to  
 18    plasma biomarkers improved diagnostic accuracy for brain amyloidosis<sup>10</sup>. Santos et al  
 19    validated the utility of plasma biomarkers for predicting dementia conversion in a  
 20    Brazilian cohort<sup>14</sup>. Other studies have utilized machine learning on imaging and cognitive

features to identify incipient dementia cases<sup>15</sup>, while some reviews have summarized available plasma assays and highlighted their promise for clinical translation<sup>16</sup>. However, these efforts typically focus on one or two modalities. A truly integrative multimodal framework—combining plasma, neuroimaging, and genetic data—is still lacking. Furthermore, no prior study has systematically evaluated the relative contribution of each modality within a unified predictive framework, leaving it unclear which data sources offer the most substantial incremental value.

To address these gaps, we proposed a multimodal machine learning framework based on ML to predict continuous cerebral A $\beta$  burden. Specifically:

(1) We evaluated the model using data from the ADNI, systematically examining the added predictive value of structural MRI, DTI-derived structural connectomes, and genetic features beyond plasma biomarkers; (2) We compared the predictive performance of APOE genotype and PRS within this framework to evaluate their respective utility. (3) We validated the model in an independent Chinese cohort (SILCODE) to assess cross-cohort generalizability

## 2 Methods

### 2.1 Participants and Study Design

This study conducted a secondary analysis using publicly available data from the ADNI, a longitudinal project aimed at identifying early biomarkers of AD through multimodal data



1 collection. ADNI participants were included if they were aged 55–90 years, generally in  
2 good health, fluent in English or Spanish, and had a Geriatric Depression Scale score  
3 below 6. Clinical diagnoses—cognitively unimpaired (CU), mild cognitive impairment  
4 (MCI), or AD—were determined according to standard ADNI criteria based on subjective  
5 memory complaints, objective neuropsychological assessments, and clinician-rated global  
6 functioning. To ensure multimodal data availability, participants with sMRI, DTI, genetic  
7 data, plasma biomarkers, and Aβ-PET imaging acquired within a one-year interval were  
8 selected from the ADNI-GO, ADNI-2, and ADNI-3 phases.

9 To assess the generalizability, we performed external validation using the SILCODE  
10 cohort—a prospective multicenter study of AD and cognitive impairment in the mainland  
11 Chinese population. SILCODE participants were included if they were aged 45–90 years,  
12 right-handed, native Mandarin speakers, and had complete multimodal data for sMRI, DTI,  
13 plasma biomarkers, APOE genotyping, and Aβ-PET imaging within a one-year window.  
14 Demographic characteristics of both cohorts, stratified by diagnostic group, are  
15 summarized in **Table 1**.

	ADNI			SILCODE		
Diagnose	CU	MCI	AD	CU	MCI	AD
N	97	43	10	57	24	20
Age	71.36±6.02	74.84±8.62	72.27±6.57	68.16±5.71	70.46±6.31	71.40±10.98
Female	36(37%)	26(60%)	7(70%)	40(61%)	14(42%)	12(60%)

<b>Pet_CL</b>	21.81±31.15	44.53±54.78	102.10±46.53	\	\	\
<b>SUVr</b>	\	\	\	1.03±0.11	1.09±0.16	1.3±0.17
<b>Pla_tau217</b>	0.21±0.27	0.34±0.34	0.73±0.38	\	\	\
<b>Pla_tau181</b>	\	\	\	2.04±0.94	2.82±1.04	3.79±1.17
<b>Pla_Aβ42/Aβ40</b>	0.09±0.01	0.13±0.2	0.08±0.01	0.06±0.02	0.06±0.01	0.05±0.02
<b>Pla_NfLQ</b>	19.56±8.7	23.24±12.84	25.58±8.55	17.64±7.1	22.09±7.71	31.62±22.82
<b>Pla_GFAP</b>	159.96±78.9	172.83±107.16	244.32±114.55	102.32±44.49	118.08±70.02	164.09±114.95
<b>APOE_e4</b>	0.41±0.63	0.65±0.65	1±0.67	0.23±0.46	0.5±0.59	0.55±0.51
<b>PRS_1e-6</b>	-0.16±0.94	0.12±0.89	0.25±0.62	\	\	\
<b>Education</b>	16.93±2.28	15.63±2.55	15.6±1.6	14.51±3.24	13.29±2.8	11.75±4.24
<b>MMSE</b>	29.24±1.17	26.63±2.9	23.4±2.17	28.42±1.7	24.54±2.38	18.35±6.39

Table 1 | Demographic, clinical, imaging, and biomarker characteristics of participants in the ADNI and SILCODE cohorts, stratified by diagnostic group: cognitively unimpaired (CU), mild cognitive impairment (MCI), and Alzheimer's disease (AD). Values are reported as mean ± standard deviation or N (%). PET\_CL: Centiloid values from amyloid PET; SUVr: standardised uptake value ratio from amyloid PET; Pla\_tau217 and Pla\_tau181: plasma phosphorylated tau at threonine 217 and 181, respectively; Pla\_Aβ42/Aβ40: plasma amyloid-β 42/40 ratio; Pla\_NfLQ: plasma neurofilament light chain; Pla\_GFAP: plasma glial fibrillary acidic protein; APOE\_e4: apolipoprotein E ε4 dosage (range: 0–2); PRS\_1e-6: polygenic risk score based on SNPs with  $p < 1 \times 10^{-6}$ ; Education: years of formal education; MMSE: Mini-Mental State Examination score. “\” indicates variables not available in the corresponding cohort or subgroup.

## 2.2 MRI Data Acquisition

sMRI and DTI data were collected from both the ADNI and SILCODE cohorts. In ADNI, MRI scans were acquired using 3.0 Tesla (3T) scanners from Philips, Siemens, or GE

1 Healthcare across the ADNI-GO, ADNI-2, and ADNI-3 phases. For sMRI, a standardized  
2 protocol was used, with typical parameters including a repetition time (TR) of 6.7 ms,  
3 echo time (TE) of 3.1 ms, flip angle of  $9^\circ$ , field of view (FOV) of  $256 \times 256 \times 170 \text{ mm}^3$ ,  
4 slice thickness of 1.2 mm, and voxel size of 1.0–1.11  $\text{mm}^3$ , covering 170 sagittal slices.  
5 DTI scans in ADNI-GO and ADNI-2 were acquired using GE 3T scanners with TR =  
6 9000 ms, TE = 60–70 ms, and voxel size =  $1.37 \times 1.37 \times 2.7 \text{ mm}^3$ . Each scan included 30  
7 diffusion-weighted directions ( $b = 1000 \text{ s/mm}^2$ ) and 5 non-diffusion-weighted ( $b = 0$ )  
8 images. For ADNI-3, an updated protocol was used with a resolution of  $2 \times 2 \times 2 \text{ mm}^3$ ,  
9 TR = 7200 ms, TE = 56 ms, 35 axial slices, and a total scan time of approximately 7.5  
10 minutes.

11 In the SILCODE cohort, imaging was performed using a 3T integrated PET/MR scanner  
12 (SIGNA PET/MR, GE Healthcare) at Xuanwu Hospital, Capital Medical University.  
13 sMRI was acquired using a gradient-recalled echo sequence with TR = 2300 ms, inversion  
14 time (TI) = 900 ms, TE = 2.26 ms, flip angle =  $8^\circ$ , FOV =  $256 \times 256 \text{ mm}^2$ , slice thickness  
15 = 1 mm, and isotropic voxel size of 1  $\text{mm}^3$ . DTI data were collected using a spin-echo  
16 echo-planar imaging (EPI) sequence with TR = 4500 ms, TE = 65 ms, voxel size =  $2 \times 2 \times$   
17  $2 \text{ mm}^3$ , and 64 slices. Diffusion gradients were applied in 128 directions, including 64 at  $b$   
18 =  $1000 \text{ s/mm}^2$  and 64 at  $b = 2000 \text{ s/mm}^2$ , along with one non-diffusion-weighted ( $b = 0$ )  
19 image.

## 2.3 MRI Data Processing

All imaging data were preprocessed using standardized protocols adapted from the UK Biobank pipeline<sup>17</sup>, implemented using *FSL*<sup>18</sup>, *FreeSurfer*<sup>19</sup>, and *MRtrix3*<sup>20</sup>. Raw DICOM images were converted to NIfTI format using *dcm2niix*<sup>21</sup>. For diffusion imaging, b-value distributions were inspected prior to preprocessing. sMRI preprocessing for both cohorts included skull stripping and bias field correction using FSL, followed by cortical reconstruction and volumetric segmentation using FreeSurfer v6.0. This yielded native-space parcellations based on the Desikan–Killiany atlas. These parcellations were further used to extract mean SUVR values within regions of interest (ROIs) for each participant. DTI data were corrected for head motion, eddy currents, and EPI distortions using FSL’s eddy<sup>22</sup>.

SCs were constructed using a unified tractography pipeline. Fiber orientation distributions were estimated using constrained spherical deconvolution, followed by probabilistic tractography<sup>23</sup>. Tractography was seeded from the gray–white matter interface using the *5ttgmwmi* algorithm<sup>24</sup>. Brain network nodes were defined using a hybrid atlas comprising 100 cortical regions from the Schaefer atlas<sup>25</sup> and 16 subcortical regions from the Melbourne Subcortex Atlas (MSA)<sup>26</sup>. The resulting SC matrices were used for feature extraction and predictive modeling. To further characterize topological changes at the network level, we grouped the 116 regions into 8 functional modules based on Yeo’s 7-network parcellation scheme and the MSA-defined subcortical network. The Yeo-7

1 system defines seven large-scale cortical functional networks: visual, somatomotor,  
2 default mode, dorsal attention, salience/ventral attention, executive control and limbic  
3 networks. The eighth module, subcortical, includes 16 subcortical nuclei defined by the  
4 MSA.

## 5 2.4 Quantification and Harmonization of Amyloid PET Measures

6 In the ADNI cohort, A $\beta$  burden was quantified using Centiloid (CL) values<sup>27,28</sup>, processed  
7 uniformly by the University of California, Berkeley, and publicly released alongside the  
8 official dataset. CL values were derived from cortex-wide *SUMMARY\_SUVR* measures,  
9 normalized to the whole cerebellum, and converted using tracer-specific formulas:

10 For [ $^{18}$ F]-florbetapir (FBP):

$$CL = 188.22 \times SUMMARY\_SUVR - 189.16$$

11 For [ $^{18}$ F]-florbetaben (FBB):

$$CL = 157.15 \times SUMMARY\_SUVR - 151.87$$

12 *SUMMARY\_SUVR* values were based on sMRI-guided segmentation of four cortical  
13 regions (frontal, cingulate, parietal, and temporal), with normalization to the whole  
14 cerebellum; regional definitions are detailed in the Supplementary Materials (Table S1). In  
15 ADNI cohort, we utilized the standardized CL values provided by ADNI rather than  
16 recalculating SUVRs. This decision was based on two considerations: (i) the CL  
17 framework enables cross-tracer and cross-scanner harmonization, improving

1 comparability and robustness; and (ii) the official CL pipeline has been extensively  
2 validated and widely adopted, ensuring high reproducibility.

3 In the SILCODE cohort, A $\beta$  PET imaging was conducted using the radiotracer  
4 [ $^{18}$ F]-florbetapir (AV-45). Participants received an intravenous injection of 7 to 10 mCi of  
5 [ $^{18}$ F]-florbetapir, followed by a rest period of approximately 40 min, after which a  
6 20-minute static PET scan was performed. PET data acquisition employed a time-of-flight  
7 ordered subset expectation maximization (TOF-OSEM) algorithm<sup>29,30</sup> with 8 iterations, a  
8 field of view of 350  $\times$  350 mm<sup>2</sup>, subset matrices of 32 at 192  $\times$  192, and a full width at  
9 half maximum (FWHM) of 3 mm. A $\beta$  burden was quantified using *SUMMARY\_SUVR*  
10 values, calculated with the same cortical and reference regions as in the ADNI processing  
11 pipeline.

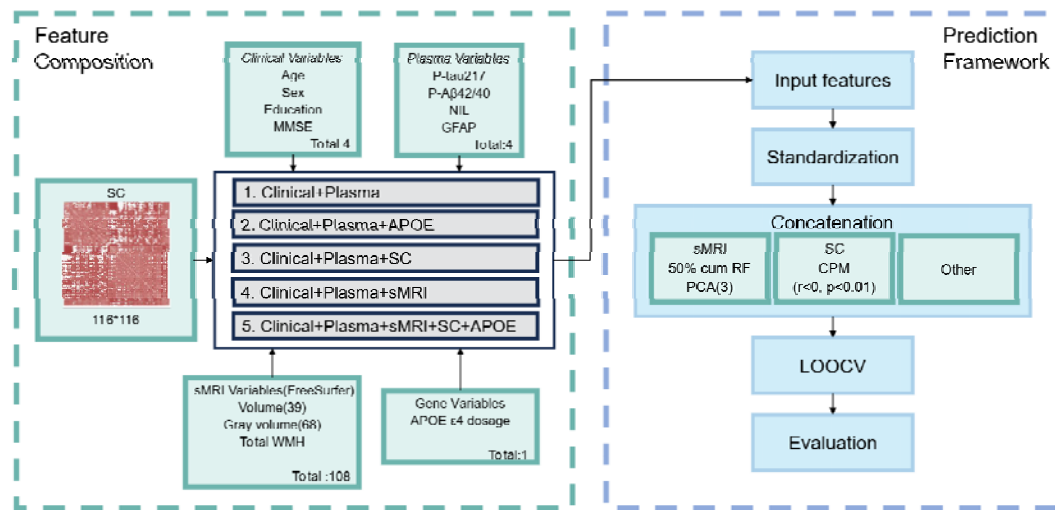
## 12 2.5 Genetic Risk Assessment

13 In ADNI, genotyping was performed using either the Illumina HumanOmniExpress or  
14 Infinium Global Screening Array v2. Each dataset underwent separate preprocessing,  
15 imputation, and quality control (QC) steps to ensure batch consistency. Preprocessing  
16 involved recoding chromosome labels, removing non-autosomal variants and duplicate  
17 SNPs, and correcting strand orientations using PLINK<sup>31</sup> and Bcftools<sup>32</sup>. Phasing used the  
18 1000 Genomes Phase 3 panel<sup>33</sup>, with imputation via Impute5<sup>34</sup>. Variants with imputation  
19 INFO scores < 0.8 were excluded. After merging batches, unified QC removed SNPs with  
20 minor allele frequency < 0.01, genotype missingness > 1%, or significant deviation from

1 Hardy–Weinberg equilibrium ( $p < 1 \times 10^{-6}$ ). Individuals were excluded for high genotype  
2 missingness, excessive heterozygosity (F-statistic  $\pm 3$  SD), cryptic relatedness  
3 (identity-by-descent  $> 0.125$ ), or discrepancies between genetically inferred and reported  
4 sex. PRS were computed using clumping and thresholding (C+T) based on AD GWAS  
5 summary statistics<sup>35</sup>, testing multiple p-value inclusion thresholds ( $p < 10^{-6}$ ) combined  
6 with LD-based clumping (window size=250 kb,  $r^2 < 0.01$ ). Resulting scores were  
7 standardized (Z-scores) for analysis. To control population stratification, the top five  
8 genetic principal components derived via PCA were included as covariates. In SILCODE,  
9 genome-wide genotyping was unavailable. Only APOE genotype data obtained through  
10 standardized laboratory protocols were included as categorical covariates ( $\epsilon 2$ ,  $\epsilon 3$ ,  $\epsilon 4$ ) in  
11 analyses.

## 12 2.6 Multimodal Feature Integration and Predictive Modeling

13 To build an accurate prediction model and assess the added predictive value of sMRI, SC,  
14 and genetic features beyond plasma biomarkers, we implemented a modular machine  
15 learning framework enabling multimodal integration and systematic comparison across  
16 feature combinations<sup>10,11,36</sup>. Taking plasma features as the baseline, we defined and  
17 compared five feature combinations: Clinical + Plasma, Clinical + Plasma + APOE,  
18 Clinical + Plasma + SC, Clinical + Plasma + MRI, and Clinical + Plasma + MRI + SC +  
19 APOE. Two regression algorithms, linear regression and random forest (RF) regression,  
20 were used for model benchmarking (Fig. 1).



1

2 Figure 1 | Multimodal machine learning framework for predicting cerebral amyloid-β burden.

3 An integrative pipeline was constructed to combine clinical, plasma, genetic, sMRI, and SC features. Five feature  
4 combinations were tested under a LOOCV framework. sMRI features and SC were dimensionally reduced or selected  
5 based on established criteria. Models were trained using both linear and random forest regressors.

6 Abbreviations: Aβ, amyloid-β; APOE, apolipoprotein E4 dosage; CPM, connectome-based predictive modeling; DTI,  
7 diffusion tensor imaging; LOOCV, leave-one-out cross-validation; PCA, principal component analysis; PRS, polygenic  
8 risk score; SC, structural connectome; sMRI, structural MRI; MMSE, Mini-Mental State Examination.

9 A total of 108 sMRI features were included, encompassing cortical volumes, surface areas,  
10 and white matter hyperintensities (WMHs), all normalized by intracranial volume (ICV)  
11 to control for inter-individual differences in brain size<sup>37</sup>. Within each training set, features  
12 were ranked by importance using RF regression, with the top 50% retained and further  
13 reduced to three orthogonal components using PCA. The SC features were extracted using  
14 the CPM (connectome-based predictive modeling) approach<sup>38,39</sup>. For each training fold,  
15 edges negatively correlated with CL values (Pearson's  $r < 0$ ,  $p < 0.01$ ) were identified.



1 The average strength of these negatively correlated edges was computed as a summary  
2 feature per subject.

3 Model training and validation were performed using leave-one-out cross-validation  
4 (LOOCV)<sup>40</sup>. All preprocessing, feature selection, and dimensionality reduction steps were  
5 strictly confined to the training data in each fold. The resulting model was then applied to  
6 the held-out subject to obtain unbiased performance estimates. The ML framework  
7 modelling was conducted separately within ADNI and SILCODE.

8 To further evaluate model performance in classifying clinical A $\beta$  burden status, we trained  
9 multi-class Random Forest classifiers using sing clinical diagnosis labels (NC, MCI, AD)  
10 as outcomes. Classification was conducted in ADNI cohort under the same LOOCV  
11 framework, and model performance was assessed using area under the receiver operating  
12 characteristic curve (AUC) for each class, as well as macro-average AUC to evaluate  
13 overall discriminative ability. The same feature extraction and selection pipeline was  
14 applied as in the regression setting, ensuring consistent input modalities across tasks.

## 15 2.7 Interpretability Analysis

16 To further elucidate the contributions of individual features to model predictions and  
17 enhance interpretability, we employed SHapley Additive exPlanations (SHAP)<sup>41</sup> to  
18 interpret the predictive power of each feature. SHAP quantifies the marginal contribution  
19 of each input feature to a given prediction while accounting for feature interactions by

1 constructing a local linear model to approximate and interpret complex nonlinear models

2  $f(x)$ , formulated as:

$$g(x') = \phi_0 + \sum_{i=1}^M \phi_i x'_i$$

3 where  $x'$  represents a simplified binary version of the input features (indicating feature

4 presence or absence),  $\phi_0$  denotes the expected output when all features are absent, and

5  $\phi_i$  represents the Shapley value of feature  $i$ , reflecting its marginal contribution to the

6 prediction.

## 7 2.8 Sensitivity Analysis

8 To evaluate the robustness of the proposed framework, we conducted sensitivity analyses

9 across three methodological dimensions: (1) PRS construction, and (2) SC parcellation

10 templates

11 First, PRSs were reconstructed using clumping and thresholding with a range of p-value

12 cutoffs ( $p < 10^{-5}$ ,  $10^{-6}$ ,  $10^{-7}$ ,  $10^{-8}$ ,  $10^{-9}$ , and  $10^{-10}$ ). Second, to evaluate the effect of brain

13 parcellation schemes, we derived SC features using an alternative atlas combining aParc

14 and MSA-16 (total 84 regions), replacing the default Schaefer-100 + MSA-16 template

15 (total 116 regions).

# 3 Results

## 3.1 Associations between features and cerebral amyloid- $\beta$ burden

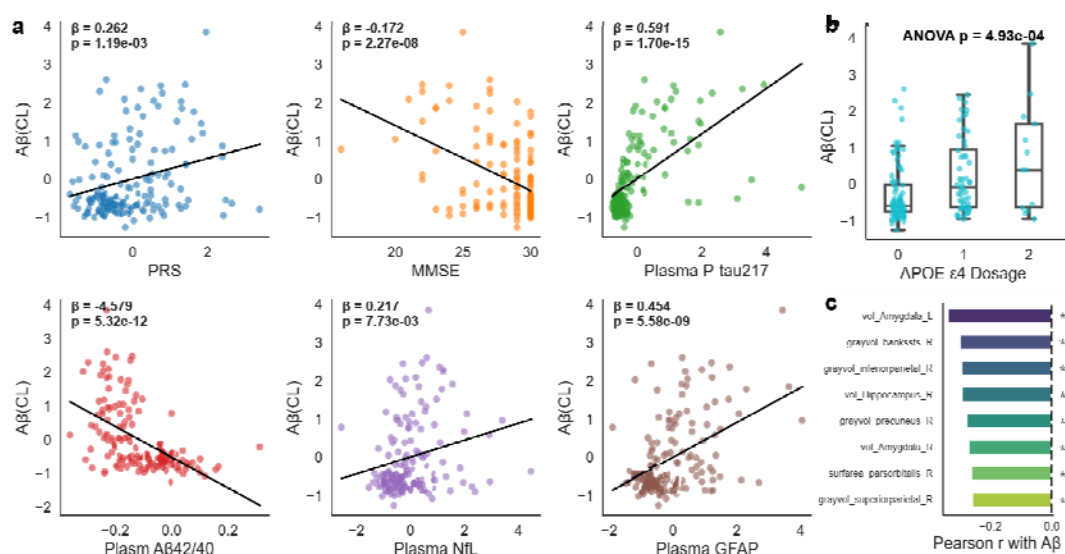


Figure 2 | Associations between plasma biomarkers, cognitive function, genetic risk, and brain structural features with standardized Aβ burden.

(a) Scatter plots showing the linear associations between Z-scored CL values and PRS, MMSE, p-tau<sub>217</sub>, Aβ<sub>42</sub>/Aβ<sub>40</sub> ratio, NFL, and GFAP. Standardized regression coefficients (β) and corresponding p-values are annotated in each panel.

(b) Boxplots illustrating the distribution of Z-scored CL values stratified by APOE ε4 allele dosage (0, 1, or 2 copies).

(c) The top eight ICV-normalized sMRI features (volume or surface area) most strongly correlated with Aβ burden, ranked by Pearson correlation coefficients. Asterisks (\*) indicate significant associations (p < 0.05, uncorrected).

Abbreviations: Aβ, amyloid-β; APOE, apolipoprotein E; CL, Centiloid; GFAP, glial fibrillary acidic protein; MMSE, Mini-Mental state Examination; NFL, neurofilament light chain; PRS, polygenic risk score; ICV, Intracranial Volume; ANOVA, analysis of variance.

In the ADNI cohort, among plasma biomarkers, p-tau<sub>217</sub> showed the strongest positive association with cerebral Aβ burden (β = 0.591, p = 1.70 × 10<sup>-15</sup>), followed by GFAP (β =

1    0.454,  $p = 5.58 \times 10^{-11}$ ) and NfL ( $\beta = 0.217$ ,  $p = 7.73 \times 10^{-3}$ ). In contrast, the  $A\beta_{42}/A\beta_{40}$   
2    ratio was strongly and negatively associated with  $A\beta$  burden ( $\beta = -4.579$ ,  $p = 5.32 \times$   
3     $10^{-12}$ ). Cognitive performance, as measured by MMSE, exhibited a modest negative  
4    association with  $A\beta$  burden ( $\beta = -0.172$ ,  $p = 2.27 \times 10^{-11}$ ), suggesting that amyloid  
5    deposition contributes only partially to cognitive decline. Compared to plasma markers,  
6    genetic features showed weaker associations: the PRS was positively correlated with  $A\beta$   
7    burden ( $\beta = 0.262$ ,  $p = 1.19 \times 10^{-3}$ ), while APOE  $\epsilon 4$  dosage revealed a clear  
8    dose-dependent increase in amyloid levels (Fig. 2a–b). Structural MRI analyses revealed  
9    multiple brain regions associated with  $A\beta$  burden. The eight regions with the strongest  
10    correlations, as shown in Figure 2c, were mainly located in the medial temporal lobe (e.g.,  
11    amygdala, hippocampus), parietal cortex (e.g., precuneus, inferior parietal lobule), and  
12    subcortical areas. These spatial patterns are consistent with regions known to be  
13    vulnerable in the early stages of Alzheimer’s disease, reinforcing the link between  
14    structural atrophy and amyloid deposition.

15

## 3.2 Multimodal feature integration enhances A $\beta$ burden prediction performance

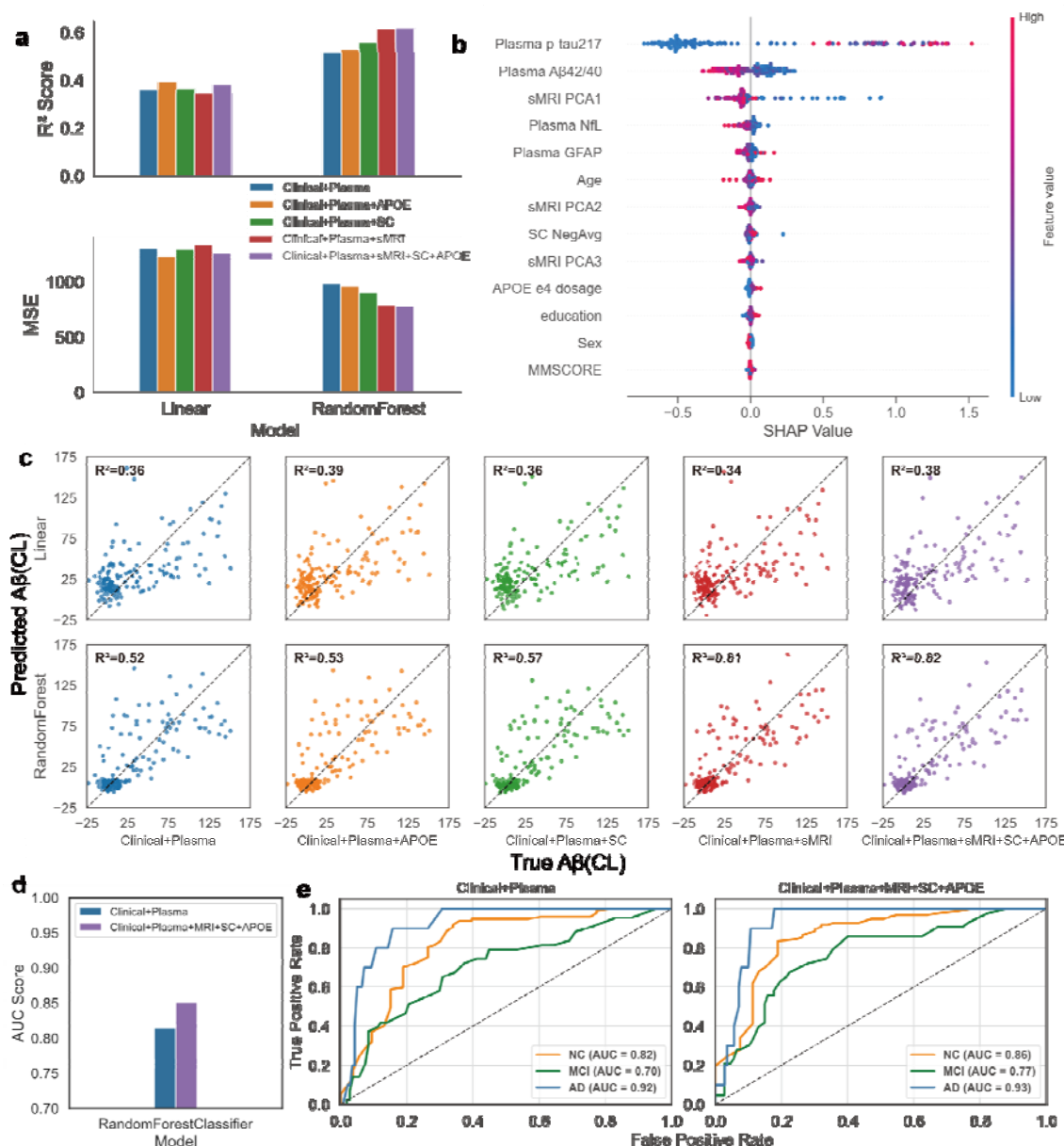


Figure 3 | Model performance and SHAP-based feature importance for A $\beta$  burden prediction in the ADNI cohort.

(a)  $R^2$  and mean squared error (MSE) for linear regression (LR) and random forest (RF) models across different feature combinations under leave-one-out cross-validation (LOOCV).

(b) SHAP summary plot showing the top features in the best-performing RF model (Clinical + Plasma + sMRI + SC + APOE), ranked by mean absolute SHAP value. Each point represents a subject, with colour indicating feature value (red = high, blue = low).

(c) Scatter plots of observed versus predicted A $\beta$  burden (Centiloid values) for LR and RF models across all feature combinations.  $R^2$  values are shown for each model.

(d) Macro-average area under the receiver operating characteristic curve (AUC) for multi-class classification (NC, MCI, AD) across different feature combinations using RF classifier.

(e) Class-specific AUC values for NC, MCI, and AD groups based on RF models trained with various combinations of features.

Abbreviations: A $\beta$ , amyloid- $\beta$ ; APOE, apolipoprotein E; GFAP, glial fibrillary acidic protein; MMSE, Mini-Mental State Examination; NfL, neurofilament light chain; PCA, principal component analysis; SC, structural connectivity; SHAP, SHapley Additive exPlanations; RF, random forest; LR, linear regression; MSE, mean squared error; ROC, receiver operating characteristic; AUC, area under the curve; NC, cognitively normal; MCI, mild cognitive impairment; AD, Alzheimer's disease; Clinical, demographic and cognitive measures.

We systematically evaluated the predictive performance of different feature combinations under a LOOCV framework using data from the ADNI cohort. Linear regression showed limited ability to model cerebral A $\beta$  burden across all feature sets ( $R^2 = 0.38$  for the Clinical + Plasma + sMRI + SC + APOE model), indicating its inability to capture the complex, nonlinear interactions inherent to multimodal data. In contrast, the RF model consistently outperformed linear regression, with performance improving as more modalities were integrated (Fig. 3a–b). Notably, the full model incorporating clinical features, plasma biomarkers, sMRI, SC, and APOE genotype achieved the best predictive accuracy ( $R^2 = 0.617$ ) and lowest mean squared error ( $MSE = 770.1$ ) (Fig. 3c), highlighting the synergistic value of combining diverse biological information. Specifically, the RF model using only clinical and plasma features achieved an  $R^2$  of 0.515. Incremental gains were observed with the addition of APOE genotype ( $R^2 = 0.527$ ), SC features ( $R^2 = 0.556$ ), and sMRI measures ( $R^2 = 0.615$ ), with maximal performance attained when all modalities were combined ( $R^2 = 0.617$ ). Detailed  $R^2$  and MSE metrics for all feature sets are provided in Supplementary Table S2.

To interpret the model, we applied SHAP analysis to the best-performing RF model. The most influential predictors were plasma biomarkers, particularly p-tau<sub>217</sub> and the A $\beta$ <sub>42</sub>/A $\beta$ <sub>40</sub> ratio, followed by principal components from sMRI, SC-derived connectivity metrics and genetic feature. These results suggest that while plasma biomarkers offer strong individual predictive power, imaging and genetic feature provide valuable complementary information.

To assess classification performance across clinical stages, we further trained a multi-class RF classifier using clinical diagnosis labels (NC, MCI, AD). Compared to the baseline model using clinical and plasma features (AUC = 0.82 for NC, 0.70 for MCI, and 0.92 for AD), the full multimodal model improved classification performance across all subgroups (AUC = 0.86 for NC, 0.77 for MCI, and 0.93 for AD) (Fig. 3e). Macro-average AUC also improved (Fig. 3d), reinforcing that multimodal integration enhances both continuous and categorical prediction of cerebral amyloid pathology.

### 3.3 Comparing the predictability of APOE and PRS

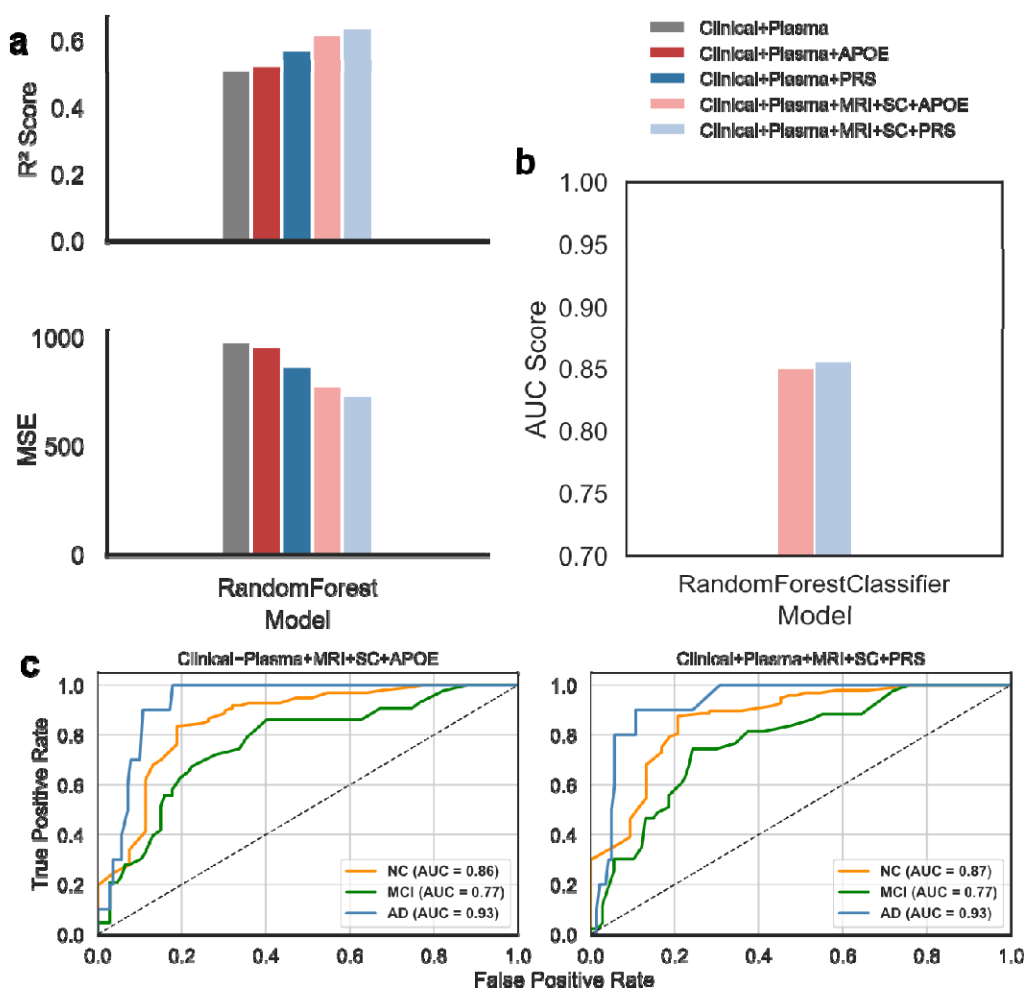


Figure 4 | Model performance with APOE or PRS for predicting A $\beta$  burden in the ADNI cohort.

Bar plots display R<sup>2</sup> (top) and MSE (bottom) for models using different combinations of clinical, plasma, genetic, and imaging features.

(b) AUC scores from multi-class Random Forest classifiers trained using CL-based A $\beta$  labels (NC, MCI, AD) with either APOE or PRS added to the baseline Clinical + Plasma + sMRI + SC model.

(c) ROC curves for each class (NC, MCI, AD) derived from models using APOE (left) or PRS (right) in the full feature combination.

To evaluate the additive predictive value of genetic information, we compared the performance of APOE genotype and PRS in predicting cerebral A $\beta$  burden within the ADNI cohort (Fig. 4). In the baseline model comprising clinical and plasma biomarkers, the inclusion of APOE  $\epsilon$ 4 status led to a modest improvement in prediction accuracy ( $\Delta R^2 = 0.012$ ). In contrast, substituting APOE with PRS yielded a more substantial gain ( $\Delta R^2 = 0.056$ ), suggesting a broader capture of genetic risk beyond the APOE locus. A detailed comparison is provided in Supplementary Materials (Table S3).

This advantage of PRS persisted when more complex models incorporating structural MRI and structural connectivity features were used. Although the performance gap between APOE and PRS narrowed in the full multimodal model ( $R^2_{\text{APOE}} = 0.617$  vs.  $R^2_{\text{PRS}} = 0.637$ ), PRS consistently demonstrated higher predictive accuracy across all combinations.

This trend was also reflected in classification performance. As shown in Fig. 4b–c, both APOE- and PRS-based models achieved similarly high AUCs in distinguishing between NC, MCI, and AD groups based on CL-derived A $\beta$  burden. The PRS-based model achieved AUCs of 0.87 (NC), 0.77 (MCI), and 0.93 (AD), closely matching the APOE-based model (0.86, 0.77, and 0.93, respectively). These results indicate that while PRS provides superior continuous-level prediction of A $\beta$  burden, its classification performance is comparable to APOE when stratifying disease stages.



# 3.4 Distributed Network Disruption Associated with A $\beta$ Burden

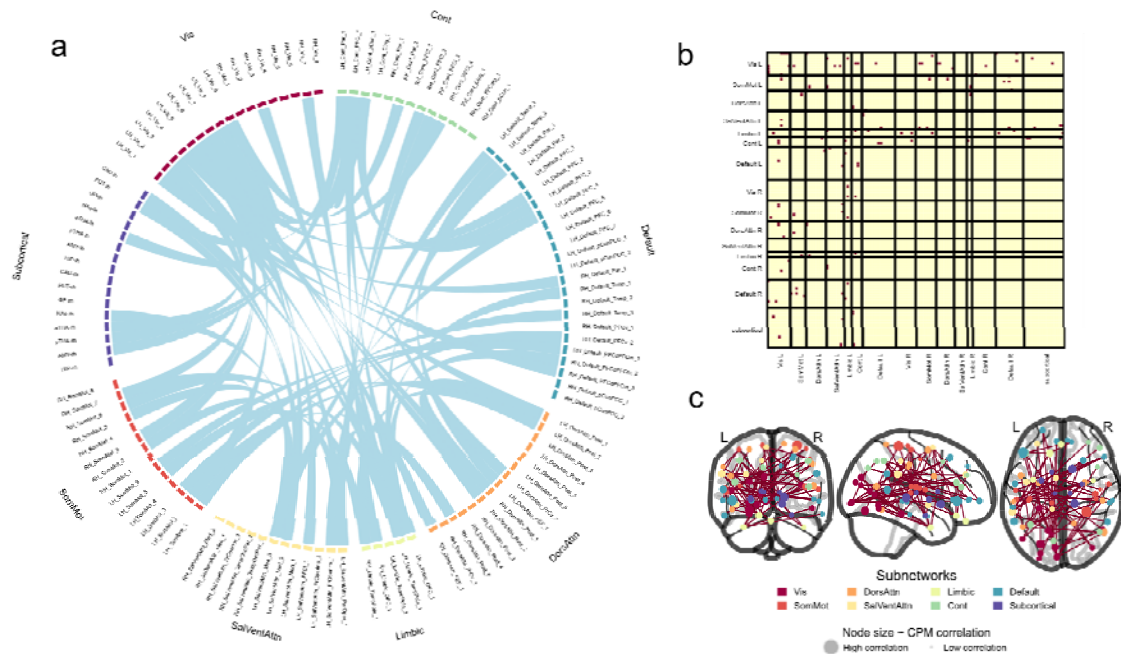


Figure 5 | Structural connections with common associations with CL value across all folds in ADNI.

(a) Chord diagram illustrating the intersection of significantly positive connections identified by CPM across all training sets under the LOOCV framework. Nodes are grouped by functional subnetworks.

(b) Inter-subnetwork connectivity matrix displaying the distribution of stable connections among major functional modules. Red dots indicate the presence of consistently selected edges; rows and columns represent different subnetworks.

(c) Cortical surface projection of stable structural connections. Node size indicates CPM correlation strength, and both nodes and edges are colour-coded by functional subnetwork.

Abbreviations: CPM, connectome-based predictive modeling; LOOCV, leave-one-out cross-validation; Vis, visual network; SomMot, somatomotor network; DorsAttn, dorsal attention network; SalVentAttn, salience/ventral attention network; Limbic, limbic network; Default, default mode network; Cont, control network; Subcortical, subcortical structures.

To identify SC consistently weakened by A $\beta$  burden, we applied CPM under a LOOCV framework. In each fold, connections negatively associated with CL values ( $p < 0.01$ ) were selected, and those recurring across all folds were retained, yielding 82 stable disconnections (Fig. 5). To identify SC consistently weakened by A $\beta$  burden, we applied CPM under a LOOCV framework. In each fold, connections negatively associated with

1 CL values ( $p < 0.01$ ) were selected, and those recurring across all folds were retained,  
2 yielding 82 stable disconnections (Fig. 5).

3 Among all functional subnetworks, the visual network exhibited the highest number of  
4 disconnections, both within itself ( $n = 10$ ) and in its connections with other  
5 networks—particularly the control ( $n = 5$ ), somatomotor ( $n = 4$ ), and default mode ( $n = 4$ )  
6 networks. This dominant pattern of disconnection suggests early vulnerability of  
7 perceptual and attentional integration systems during amyloid accumulation.

8 In addition to the visual network, spatial mapping of affected regions revealed consistent  
9 involvement of the medial prefrontal cortex and lateral parietal cortex—key hubs of the  
10 default mode and frontoparietal control networks. These findings indicate that A $\beta$ -related  
11 disruptions extend beyond primary sensory systems and impact higher-order cognitive  
12 networks implicated in early-stage AD.

13 Collectively, the observed disconnection patterns support a network-level framework for  
14 understanding AD, wherein early amyloid deposition perturbs distributed integrative  
15 systems. Structural connectome features derived from robust CPM analysis may thus  
16 serve as sensitive and interpretable biomarkers for early disease characterization and risk  
17 stratification.

18

### 3.5 External validation in the SILCODE cohort

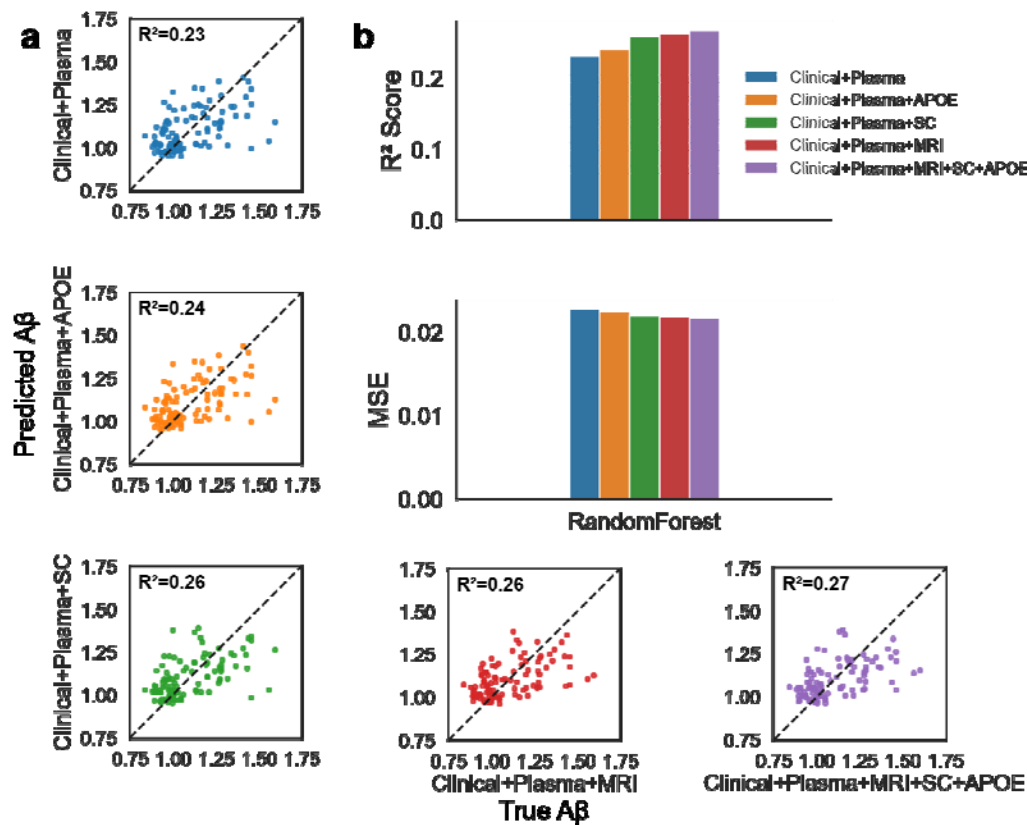


Figure 6 | Model performance for Aβ burden prediction in the SILCODE cohort.

(a) Scatter plots of observed vs. predicted Aβ SUVR values across five feature combinations using RF models. R² values indicate model fit. The x-axis denotes observed SUVR values; the y-axis shows model-predicted values.

(b) Bar plots summarize model performance under a LOOCV framework. The upper panel shows R² values, and the lower panel displays MSE across feature combinations.

Abbreviations: Aβ, amyloid-β; APOE, apolipoprotein E; GFAP, glial fibrillary acidic protein; MMSE, Mini-Mental State Examination; NfL, neurofilament light chain; PCA, principal component analysis; SC, structural connectivity; RF, random forest; MSE, mean squared error. Clinical, demographic and cognitive measures.

To assess generalizability, we performed external validation using the SILCODE cohort, employing five feature combinations with RF models to predict Aβ PET SUVR (Fig. 6).

Relative to the baseline model (Clinical+Plasma; R²=0.232), incremental performance

improvements were observed upon adding APOE genotype ( $\Delta R^2=+0.009$ ), SC features ( $\Delta R^2=+0.027$ ), and sMRI features ( $\Delta R^2=+0.032$ ). The full model (Clinical+Plasma+MRI+SC+APOE) achieved the highest  $R^2$  of 0.267, a detailed comparison of  $R^2$  and MSE across models is provided in Supplementary Materials (Table S4). Although overall predictive accuracy was lower than in ADNI, the incremental benefit pattern remained consistent, highlighting multimodal robustness. Lower performance in SILCODE primarily reflected plasma biomarker differences: ADNI utilized plasma p-tau<sub>217</sub>, known for superior amyloid sensitivity, whereas SILCODE employed the less sensitive but widely used p-tau<sub>181</sub>. Prior studies have confirmed that p-tau<sub>217</sub> better discriminates amyloid-positive individuals, partly explaining the observed performance gap<sup>42</sup>.

### 3.6 Sensitivity Analysis

First, we tested multiple p-value thresholds ( $p < 10^{-5}$  to  $10^{-10}$ ) for PRS construction using the clumping-and-thresholding approach. All thresholds except  $p < 10^{-5}$  improved prediction over the plasma-only baseline, with the best performance observed at  $p < 10^{-6}$ . These results underscore the importance of optimal thresholding for leveraging genome-wide polygenic data (Table S5 and Fig S1). Second, we evaluated the impact of different parcellation schemes on SC-based prediction. Alternative atlases with 84 and 116 regions both outperformed the baseline, with minor differences (Table S6 and Fig S2). These findings suggest that SC features provide stable predictive value across anatomical resolutions, with higher-resolution schemes offering more refined disease-relevant

connectivity patterns. Together, these analyses show that predictive gains from integrating imaging and genetic features are robust to key modeling and design choices.

## 4 Discussion

We proposed a multimodal machine learning framework that integrates plasma biomarkers, sMRI, DTI-derived structural connectivity (SC), and genetic risk to non-invasively predict cerebral A $\beta$  burden. In the ADNI cohort, adding sMRI, SC features, and APOE genotype to clinical and plasma measures substantially improved model performance ( $R^2 = 0.617$  vs.  $R^2 = 0.515$  for clinical + plasma only). Replacing APOE with a genome-wide polygenic risk score (PRS) further enhanced predictive accuracy ( $R^2 = 0.637$ ), highlighting the added value of capturing distributed genetic risk beyond the APOE locus<sup>10,35</sup>. These results underscore the complementary strengths of different modalities in characterizing early Alzheimer's disease pathology. External validation in the SILCODE cohort supported the generalizability of our findings. Despite differences in demographics, imaging protocols, and biomarker assays, consistent multimodal predictive improvements across cohorts underscore the framework's generalizability and translational potential.

Beyond continuous prediction, our multimodal model also showed improved classification of individuals along the AD spectrum. Compared to the clinical + plasma model, the full model achieved higher AUC scores across all diagnostic stages (NC: 0.86; MCI: 0.77; AD:

0.93), reinforcing its utility for both quantitative and categorical assessments of A $\beta$  burden.

Structurally, sMRI and SC features offered complementary insights into disease mechanisms. While sMRI captured diffuse cortical atrophy, SC analysis revealed spatially organized disconnection patterns. Notably, the visual network exhibited the highest number of disrupted connections, both internally and with the control, somatomotor, and default mode networks—suggesting early vulnerability of perceptual and integrative attention systems in preclinical AD<sup>25</sup>. In addition, disconnections frequently involved the medial prefrontal and lateral parietal cortices—core hubs of the default mode and frontoparietal networks—coinciding with known regions of early A $\beta$  deposition<sup>43</sup>. These findings support a network-level interpretation of amyloid pathology and underscore the utility of CPM-derived SC features as sensitive and interpretable markers of early brain changes.

From a genetic perspective, APOE  $\epsilon$ 4 consistently demonstrated predictive utility across cohorts. However, PRS outperformed APOE in the ADNI dataset, likely due to its capacity to aggregate genome-wide susceptibility variants<sup>10</sup>. Despite its superior predictive power, PRS remains less accessible and more costly, making APOE genotyping a more pragmatic option in resource-limited clinical settings<sup>8</sup>.

1 Sensitivity analyses confirmed that our findings were robust across different PRS p-value  
2 thresholds and SC parcellation schemes. Nevertheless, several limitations warrant  
3 consideration. First, our sample sizes were modest, which may restrict the generalizability  
4 of the model to broader populations<sup>44</sup>. Second, the use of the Schaefer-100/MSA-16 atlas  
5 prioritized interpretability but may have limited our ability to detect finer-grained network  
6 effects<sup>25</sup>. Future work should explore higher-resolution or individualized parcellations to  
7 refine SC modeling.

8 Implications. Collectively, our study demonstrates that integrating multimodal  
9 information—including plasma biomarkers, neuroimaging, and genetic risk—can  
10 significantly enhance non-invasive prediction of cerebral amyloid pathology. This scalable  
11 framework holds potential for improving early AD risk stratification and may inform  
12 future screening strategies in both clinical and population-based settings.

13

## 14 References

- 15 1. Selkoe, D. J. & Hardy, J. The amyloid hypothesis of Alzheimer's disease at 25 years. *EMBO*  
16 *Mol Med* **8**, 595–608 (2016).
- 17 2. DeTure, M. A. & Dickson, D. W. The neuropathological diagnosis of Alzheimer's disease.  
18 *Molecular Neurodegeneration* **14**, 32 (2019).
- 19 3. Jack, C. R. *et al.* NIA-AA Research Framework: Toward a biological definition of  
20 Alzheimer's disease. *Alzheimers Dement* **14**, 535–562 (2018).
- 21 4. Nakamura, A. *et al.* High performance plasma amyloid- $\beta$  biomarkers for Alzheimer's disease.  
22 *Nature* **554**, 249–254 (2018).

- 1 5. Zhang, M. *et al.* Relationship between topological efficiency of white matter structural  
2 connectome and plasma biomarkers across the Alzheimer's disease continuum. *Human Brain*  
3 *Mapping* **45**, e26566 (2024).
- 4 6. Racine, A. M. *et al.* Associations between white matter microstructure and amyloid burden in  
5 preclinical Alzheimer's disease: A multimodal imaging investigation. *NeuroImage: Clinical* **4**,  
6 604–614 (2014).
- 7 7. Huang, W. *et al.* Individual Variability in the Structural Connectivity Architecture of the  
8 Human Brain. *J Neurosci* **45**, e2139232024 (2025).
- 9 8. Liu, C.-C., Liu, C.-C., Kanekiyo, T., Xu, H. & Bu, G. Apolipoprotein E and Alzheimer  
10 disease: risk, mechanisms and therapy. *Nat Rev Neurol* **9**, 106–118 (2013).
- 11 9. Leonenko, G. *et al.* Identifying individuals with high risk of Alzheimer's disease using  
12 polygenic risk scores. *Nat Commun* **12**, 4506 (2021).
- 13 10. Ramanan, V. K. *et al.* Genetic risk scores enhance the diagnostic value of plasma biomarkers  
14 of brain amyloidosis. *Brain* **146**, 4508–4519 (2023).
- 15 11. Karlsson, L. *et al.* Machine learning prediction of tau  $\square$  PET in Alzheimer's disease using  
16 plasma, MRI, and clinical data.
- 17 12. Ying, Q. *et al.* Multi-Modal Data Analysis for Alzheimer's Disease Diagnosis: An Ensemble  
18 Model Using Imagery and Genetic Features. *Annu Int Conf IEEE Eng Med Biol Soc* **2021**,  
19 3586–3591 (2021).
- 20 13. Sarica, A., Cerasa, A. & Quattrone, A. Random Forest Algorithm for the Classification of  
21 Neuroimaging Data in Alzheimer's Disease: A Systematic Review. *Front Aging Neurosci* **9**, 329  
22 (2017).
- 23 14. Santos, L. E. *et al.* Performance of plasma biomarkers for diagnosis and prediction of  
24 dementia in a Brazilian cohort. *Nat Commun* **16**, 2911 (2025).
- 25 15. Identifying incipient dementia individuals using machine learning and amyloid imaging.  
26 *Neurobiology of Aging* **59**, 80–90 (2017).
- 27 16. Pais, M. V., Forlenza, O. V. & Diniz, B. S. Plasma Biomarkers of Alzheimer's Disease: A  
28 Review of Available Assays, Recent Developments, and Implications for Clinical Practice. *J*  
29 *Alzheimers Dis Rep* **7**, 355–380.
- 30 17. Alfaro-Almagro, F. *et al.* Image processing and Quality Control for the first 10,000 brain  
31 imaging datasets from UK Biobank. *NeuroImage* **166**, 400–424 (2018).
- 32 18. Jenkinson, M., Beckmann, C. F., Behrens, T. E., Woolrich, M. W. & Smith, S. M. FSL.  
33 *NeuroImage* **62**, 782–790 (2012).
- 34 19. Fischl, B. FreeSurfer. *NeuroImage* **62**, 774–781 (2012).
- 35 20. Tournier, J.-D. *et al.* MRtrix3: A fast, flexible and open software framework for medical  
36 image processing and visualization. *NeuroImage* **202**, 116137 (2019).
- 37 21. Li, X., Morgan, P. S., Ashburner, J., Smith, J. & Rorden, C. The first step for neuroimaging  
38 data analysis: DICOM to NIfTI conversion. *Journal of Neuroscience Methods* **264**, 47–56 (2016).
- 39 22. Andersson, J. L. R. & Sotiropoulos, S. N. An integrated approach to correction for  
40 off-resonance effects and subject movement in diffusion MR imaging. *Neuroimage* **125**,  
41 1063–1078 (2016).



23. Tournier, J.-D., Calamante, F. & Connelly, A. Robust determination of the fibre orientation distribution in diffusion MRI: Non-negativity constrained super-resolved spherical deconvolution. *NeuroImage* **35**, 1459–1472 (2007).
24. Smith, R. E., Tournier, J.-D., Calamante, F. & Connelly, A. Anatomically-constrained tractography: Improved diffusion MRI streamlines tractography through effective use of anatomical information. *NeuroImage* **62**, 1924–1938 (2012).
25. Schaefer, A. *et al.* Local-global parcellation of the human cerebral cortex from intrinsic functional connectivity MRI. *Cerebral Cortex* **28**, 3095–3114 (2018).
26. Tian, Y., Margulies, D. S., Breakspear, M. & Zalesky, A. Hierarchical organization of the human subcortex unveiled with functional connectivity gradients. *Nature Neuroscience* **23**, 1421–1432 (2020).
27. Landau, S. M. *et al.* Amyloid deposition, hypometabolism, and longitudinal cognitive decline. *Annals of Neurology* **72**, 578–586 (2012).
28. Klunk, W. E. *et al.* The Centiloid Project: standardizing quantitative amyloid plaque estimation by PET. *Alzheimers Dement* **11**, 1-15.e1–4 (2015).
29. Chen, H.-J. *et al.* Amyloid pathology related to aberrant structure-function coupling of brain networks in Alzheimer’s disease: insights from [18F]-florbetapir PET imaging. *Eur J Nucl Med Mol Imaging* (2025) doi:10.1007/s00259-025-07172-8.
30. Clark, C. M. *et al.* Use of florbetapir-PET for imaging beta-amyloid pathology. *JAMA* **305**, 275–283 (2011).
31. Purcell, S. *et al.* PLINK: a tool set for whole-genome association and population-based linkage analyses. *Am J Hum Genet* **81**, 559–575 (2007).
32. Danecek, P. *et al.* Twelve years of SAMtools and BCFtools. *Gigascience* **10**, giab008 (2021).
33. Auton, A. *et al.* A global reference for human genetic variation. *Nature* **526**, 68–74 (2015).
34. Rubinacci, S., Ribeiro, D. M., Hofmeister, R. J. & Delaneau, O. Efficient phasing and imputation of low-coverage sequencing data using large reference panels. *Nat Genet* **53**, 120–126 (2021).
35. Jansen, I. E. *et al.* Genome-wide meta-analysis identifies new loci and functional pathways influencing Alzheimer’s disease risk. *Nat Genet* **51**, 404–413 (2019).
36. Prakash, R. S. *et al.* A whole-brain functional connectivity model of Alzheimer’s disease pathology. *Alzheimers Dement* **21**, e14349 (2025).
37. Fjell, A. M. *et al.* One-year brain atrophy evident in healthy aging. *J Neurosci* **29**, 15223–15231 (2009).
38. Finn, E. S. Functional connectome fingerprinting: identifying individuals using patterns of brain connectivity. *nature NEUROSCIENCE* **18**, (2015).
39. Shen, X. *et al.* Using connectome-based predictive modeling to predict individual behavior from brain connectivity. *Nat Protoc* **12**, 506–518 (2017).
40. Varoquaux, G. Cross-validation failure: Small sample sizes lead to large error bars. *Neuroimage* **180**, 68–77 (2018).
41. Lundberg, S. M. & Lee, S.-I. A unified approach to interpreting model predictions. in *Advances in Neural Information Processing Systems* vol. 30 4765–4774 (2017).

- 1 42. Palmqvist, S. *et al.* Discriminative Accuracy of Plasma Phospho-tau217 for Alzheimer  
2 Disease vs Other Neurodegenerative Disorders. *JAMA* **324**, 772–781 (2020).
- 3 43. Thal, D. R., Rüb, U., Orantes, M. & Braak, H. Phases of A $\beta$ -deposition in the human brain  
4 and its relevance for the development of AD. *Neurology* **58**, 1791–1800 (2002).
- 5 44. Lockett, E. S. *et al.* Association of Alzheimer’s disease polygenic risk scores with amyloid  
6 accumulation in cognitively intact older adults. *Alzheimers Res Ther* **14**, 138 (2022).

7

## 8 Ethics declarations

9 All procedures performed in studies involving human participants were in accordan  
10 ce with the ethical standards of the institutional and/or national research committee  
11 s and with the 1964 Declaration of Helsinki and its later amendments or comparab  
12 le ethical standards.

13 For the ADNI cohort, ethical approval was obtained from the institutional review b  
14 oards of all participating institutions (e.g., University of California, San Diego). For  
15 the SILCODE cohort, the study was approved by the Ethics Committee of Xuanw  
16 u Hospital, Capital Medical University. Written informed consent was obtained fro  
17 m all participants in both cohorts.

## 18 Consent for publication

19 Not applicable. No individual person’s data is included in this study.

## 1 Data availability

2 The data used in this study are publicly available from the Alzheimer's Disease  
3 Neuroimaging Initiative (ADNI; <http://adni.loni.usc.edu>) and the SILCODE data are  
4 available upon reasonable request from the corresponding author.

## 5 Conflicts of Interest

6 The authors declare that they have no conflicts of interest regarding the publication of this  
7 manuscript.

## 9 Funding

10 This work was supported by the STI2030-Major Projects (2022ZD0213300,  
11 2022ZD0211600), National Natural Science Foundation of China (82301608, 32271145,  
12 81871425, 210510238), Open Research Fund of the State Key Laboratory of Cognitive  
13 Neuroscience and Learning (CNLZD2101, CNLZD2303), Fundamental Research Funds  
14 for the Central Universities (2017XTCX04).

## 15 Acknowledgments

16 We thank all study participants and their families for their generous contributions.  
17 We are grateful to the Alzheimer's Disease Neuroimaging Initiative (ADNI) investi  
18 gators and staff for participant recruitment and data acquisition, and to the SILCO  
19 DE research team for providing external validation data.

1

2

3

4

5

6