

Subs Paper Things to Do:

- ~~# of coding and non-coding sites~~
- ~~# of subs in each of  $\uparrow$~~
- ~~Look into *Streptomyces* non-coding issue~~
- ~~Look into *E. coli* coding issue~~
- ~~Look into pSymB coding/non-coding trend weirdness~~
- ~~Figure out why *Streptomyces* appears to have tons of coding data missing~~
- ~~Figure out what is going on with cod/non-cod code and why it is still not working!~~
- ~~write up methods for coding/non-coding~~
- ~~write methods and results for clustering~~
- ~~start code to split alignment into multiple alignments of each gene~~
- ~~figure out how to deal with overlapping genes~~
- ~~figure out how to deal with gaps in gene of ref taxa~~
- ~~split up the alignment into multiple alignments of each gene~~
- ~~check if each gene alignment is a multiple of 3 (proper codon alignment)~~
- ~~get dN/dS for coding/non-coding stuff per gene~~
- ~~Or get 1st, 2nd, 3rd codon pos log regs~~
- ~~write up coding/non-coding results~~
- ~~take out gene expression from this paper~~
- ~~write better intro/methods for distribution of subs graphs~~
- ~~write discussion for coding/non-coding~~
- ~~write coding/non-coding into conclusion~~
- ~~figured out pipeline for CODEML to calculate dN/dS for each gene~~
- ~~make a list of what should be in supplementary files for subs paper~~
- ~~put everything in list into supplementary file for subs paper~~
- ~~write dN/dS methods~~
- ~~write dN/dS results~~
- ~~write dN/dS discussion~~

- write dN/dS into conclusion
- ~~new bar graph with coding and non-coding sites separated~~
- mol clock for my analysis?
- GC content? COG? where do these fit?

#### Gene Expression Paper Things to Do:

- ~~look for more GEO expression data for *S. meliloti*~~
- ~~look for more GEO expression data for *Streptomyces*~~
- ~~look for more GEO expression data for *B. subtilis*~~
- format paper and put in stuff that is already written
- ~~look for more GEO expression data for *E. coli*~~
- ~~Get numbers for how many different strains and multiples of each strain I have for gene expression~~
- ~~re-do gene expression analysis for *B. subtilis*~~
- ~~re-do gene expression analysis for *E. coli*~~
- ~~find papers about what has been done with gene expression~~
- read papers ↑
- ~~put notes from ↑ papers into word doc~~
- write abstract
- ~~write intro~~
- add stuff from outline to Data section
- create graphs for expression distribution (no sub data)
- add # of genes to expression graphs (top)
- average gene expression
- ~~write discussion~~
- write conclusion
- add into methods: filters for Hiseq, RT PCR and growth phases for data collection
- update supplementary figures/file

#### Inversions and Gene Expression Letter Things to Do:

- ~~get as much GEO data as possible~~
- ~~find papers about inversions and expression~~
- ~~see how many inversions I can identify in these strains of *Escherichia coli* with gene expression data~~
- ~~read papers about inversions~~
- check if opposite strand in progressiveMauve means an inversions (check visual matches the xmfa)
- check if PARSNP and progressiveMauve both identify the same inversions (check xmfa file)
- create latex template for paper
- ~~put notes from papers into doc~~
- use large PARSNP alignment to identify inversions
- confirm inversions with dot plot
- write outline for letter
- write Abstract
- write intro
- write methods
- compile tables (supplementary)
- write results
- write discussion
- write conclusion
- do same ancestral/phylogenetic analysis that I did in the subs paper

## Last Week

- ✓ write results and discussion for gene expression paper
- ✓ write methods for gene expression paper
- ✓ get coding and non-coding substitution graphs for each replicon
- ✓ get proportions of coding and non-coding sections for each genome
- ✓ results and outline of slides for OE3C

I obtained a break down of the coding and non-coding proportions of the genome for each replicon. These can be found in Table 1. The proportion of coding sites that have substitutions, and non-coding sites that have substitutions are about the same (5% - 10% of sites). The only concerning thing is that because we are now only looking at sites in the alignment where all taxa are the same classification (codon position 1, 2, 3, non-coding...etc), we are now only looking at less than half of the total sites in the genome. I am slightly concerned that some of the regions in the genome may be underrepresented. What are your thoughts on this?

I realized when working through the dN/dS pipeline that there was still “stop” codons present and so I then realized that I forgot to account for the reverse complement of the sequence. So I had to add in this extra step into the pipeline, and re-run the coding/non-coding substitution analysis.

I worked on writing the methods, results and discussion for the gene expression paper/letter. I also have started working on my presentation for the OE3C, I have most of the intro, and results.

## This Week

I realized that I made a slight mistake in the genome positions that each block prints out, so I have fixed this and need to re-do the substitution analysis again. I began this process this past weekend but it takes some time to run so I am still working on this.

I would like to run the dN/dS pipeline on all of the replicons.

I would like to have a presentation complete for the OE3C so I can practice it in front of everyone next week.

## Next Week

I want to begin figuring out how to obtain all inversions from the Mauve or PARSNP alignment for the inversions analysis.

I will mostly be at the OE3C conference (May 2-4).

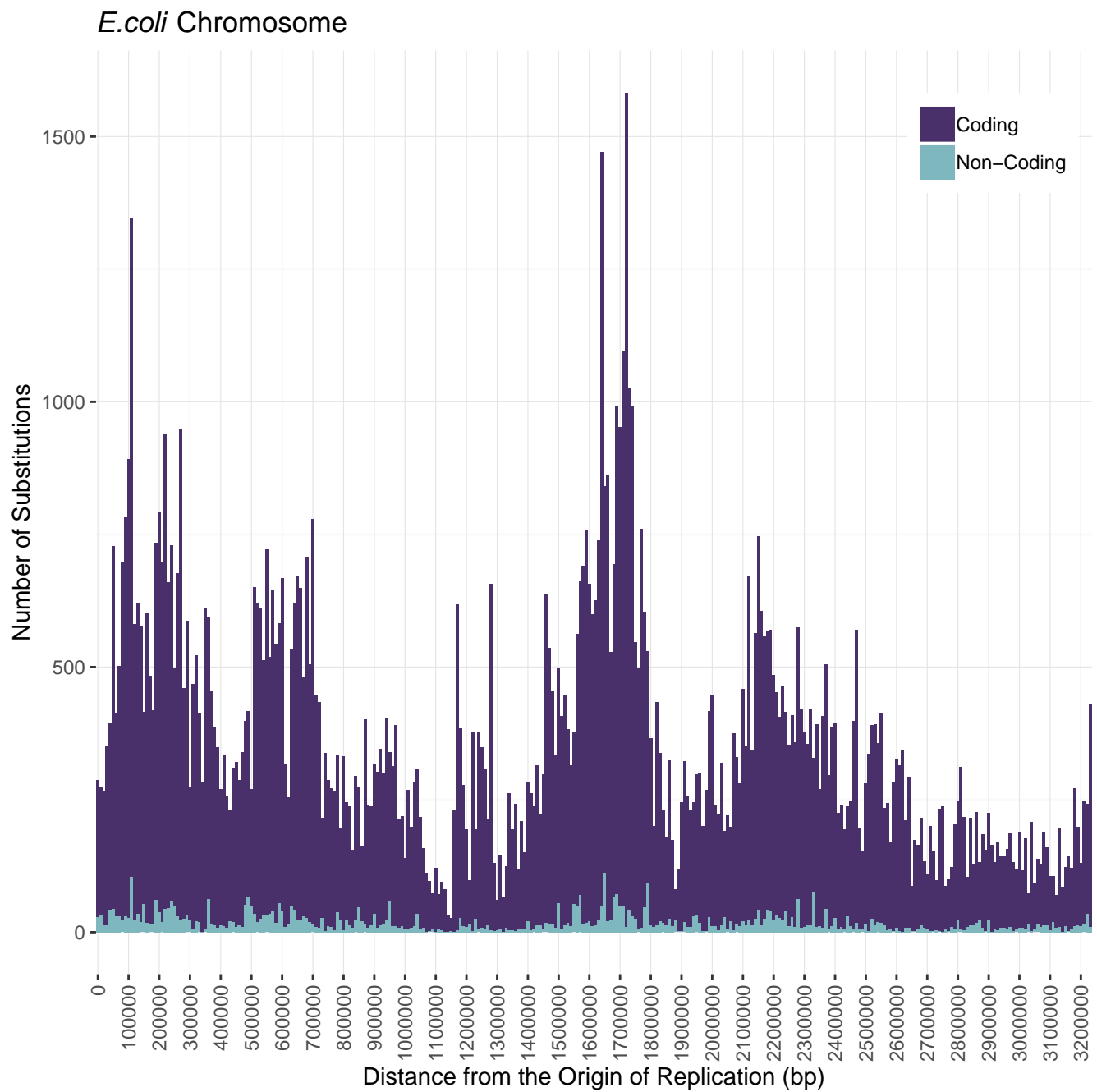
Bacteria and Replicon	Average Replicon Length	# of Coding Sites	# of Non-Coding Sites	# of Subs Coding	# of Subs Non-Coding
<i>E. coli</i> Chromosome	5082529	2243082	141885	129595	6721
<i>B. subtilis</i> Chromosome	4077077	2073824	102906	171510	6187
<i>Streptomyces</i> Chromosome	8497577	1996354	18584	246447	2808
<i>S. meliloti</i> Chromosome	3426881	1928634	199425	5153	842
<i>S. meliloti</i> pSymA	1455940	419531	68426	7709	943
<i>S. meliloti</i> pSymB	1664597	559311	22098	10039	645

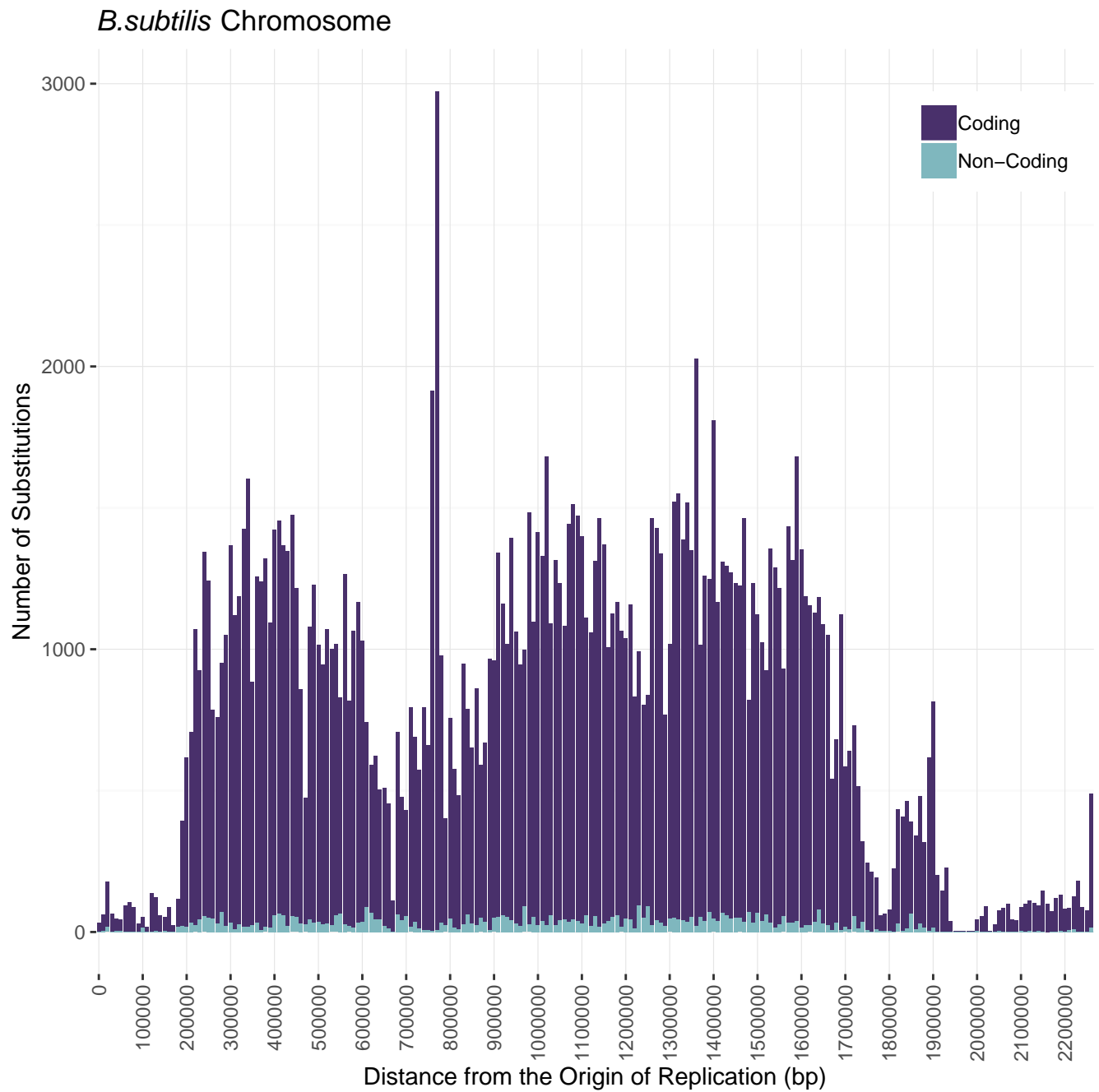
Table 1: Total proportion of coding and non-coding sites in each replicon and the percentage of those sites that have a substitution (multiple substitutions at one site are counted as two substitutions).

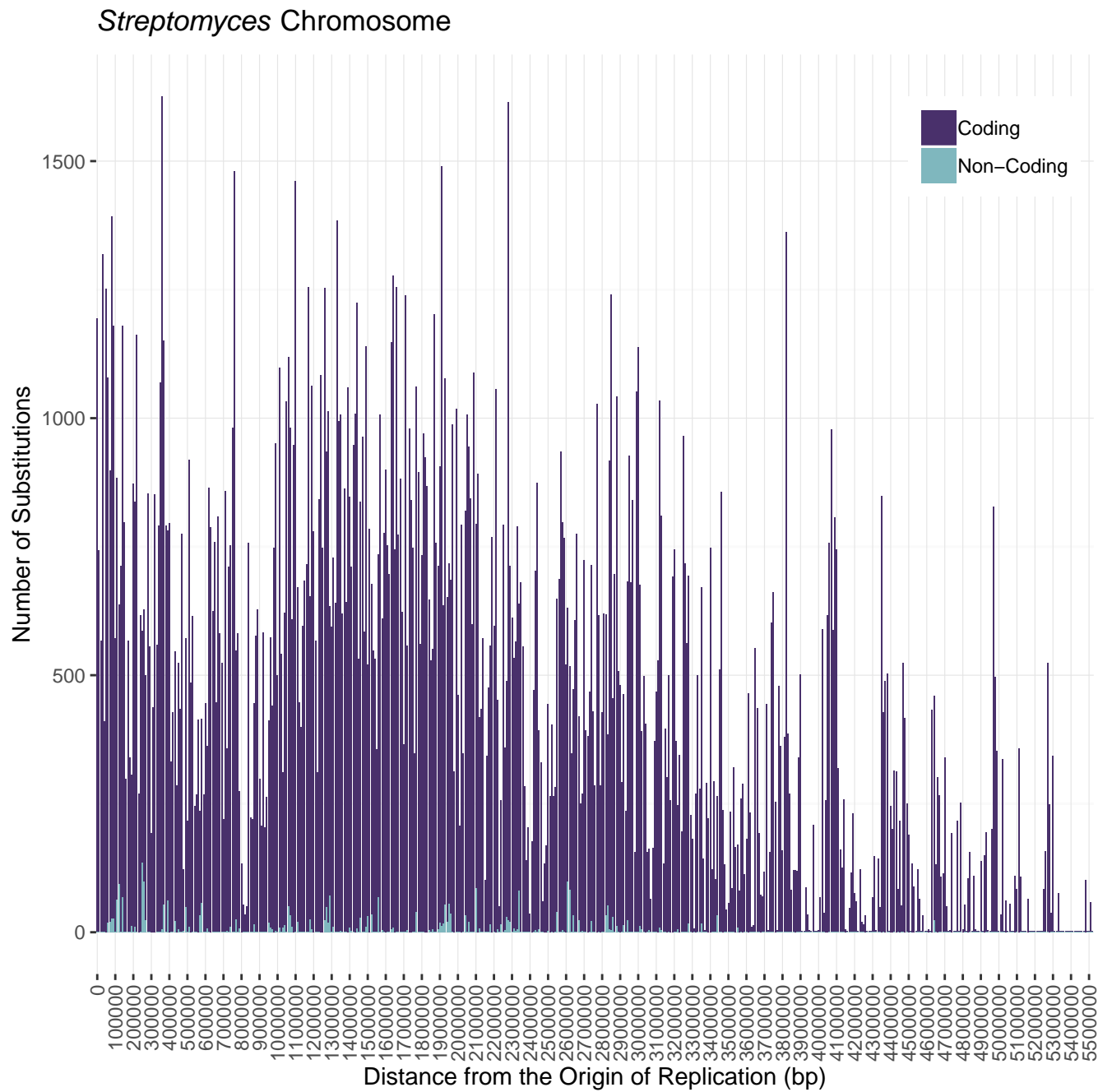
Bacteria and Replicon	Coding Sequences	Non-Coding Sequences
<i>E. coli</i> Chromosome	$-6.454 \times 10^{-8***}$	NS
<i>B. subtilis</i> Chromosome	$-1.159 \times 10^{-7***}$	$-9.861 \times 10^{-8***}$
<i>Streptomyces</i> Chromosome	$-8.464 \times 10^{-9***}$	$3.572 \times 10^{-7***}$
<i>S. meliloti</i> Chromosome	$-1.269 \times 10^{-7***}$	$-1.51 \times 10^{-7*}$
<i>S. meliloti</i> pSymA	$-2.02 \times 10^{-7***}$	NS
<i>S. meliloti</i> pSymB	$2.618 \times 10^{-7***}$	$8.591 \times 10^{-7***}$

Table 2: Logistic regression analysis of the number of substitutions along all positions of the genome of the respective bacteria replicons. These genomic positions were split up into the coding and non-coding regions of the genome. Grey coloured boxes indicate a negative logistic regression coefficient estimate. All results are statistically significant. Logistic regression was calculated after the origin of replication was moved to the beginning of the genome and all subsequent positions were scaled around the origin accounting for bidirectionality of replication. All results are marked with significance codes as followed:  $< 0.001 = '***'$ ,  $0.001 < 0.01 = '**'$ ,  $0.01 < 0.05 = '*'$ ,  $> 0.05 = 'NS'$ .

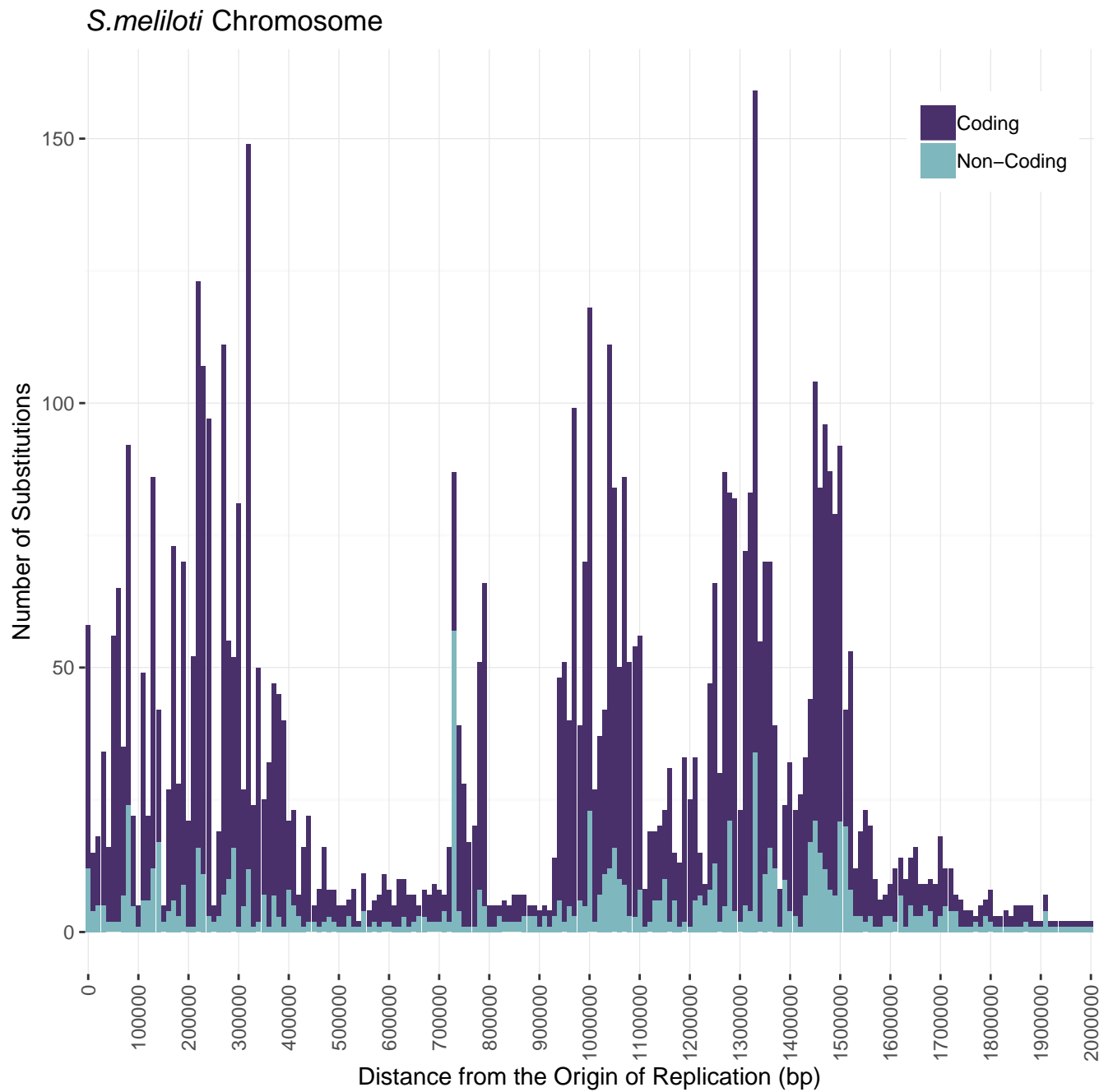
Sub density graphs with coding and non-coding information

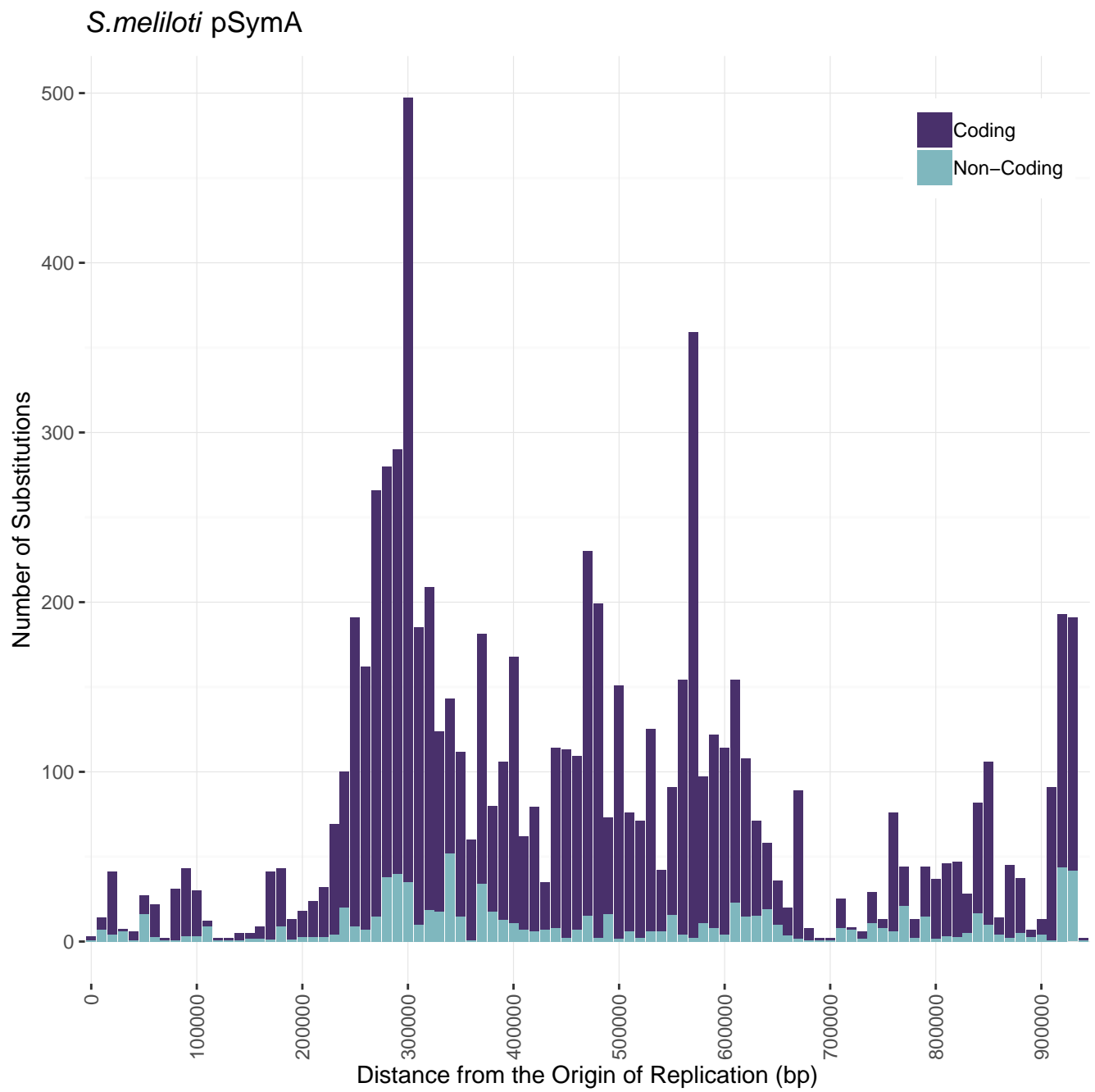


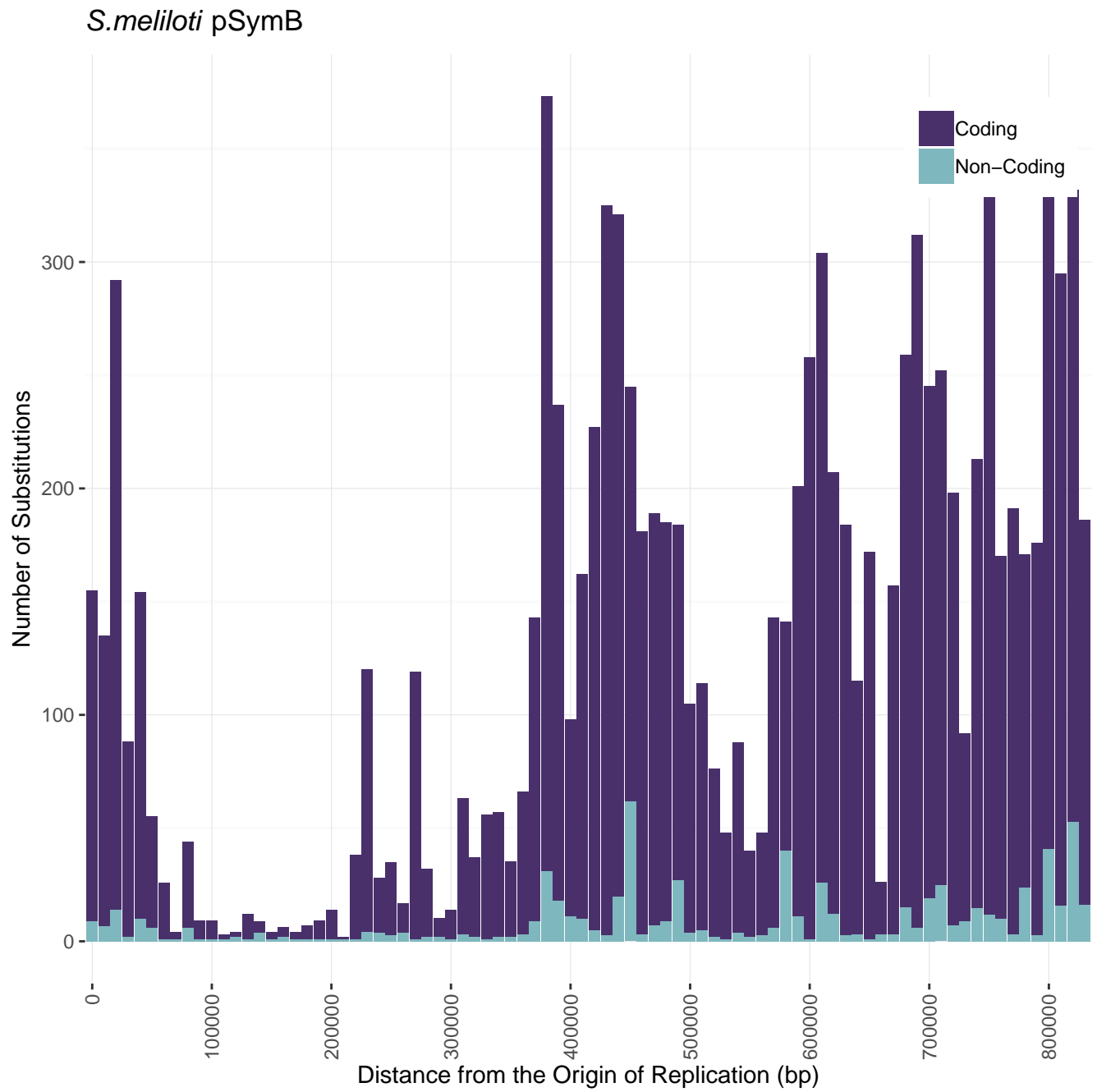




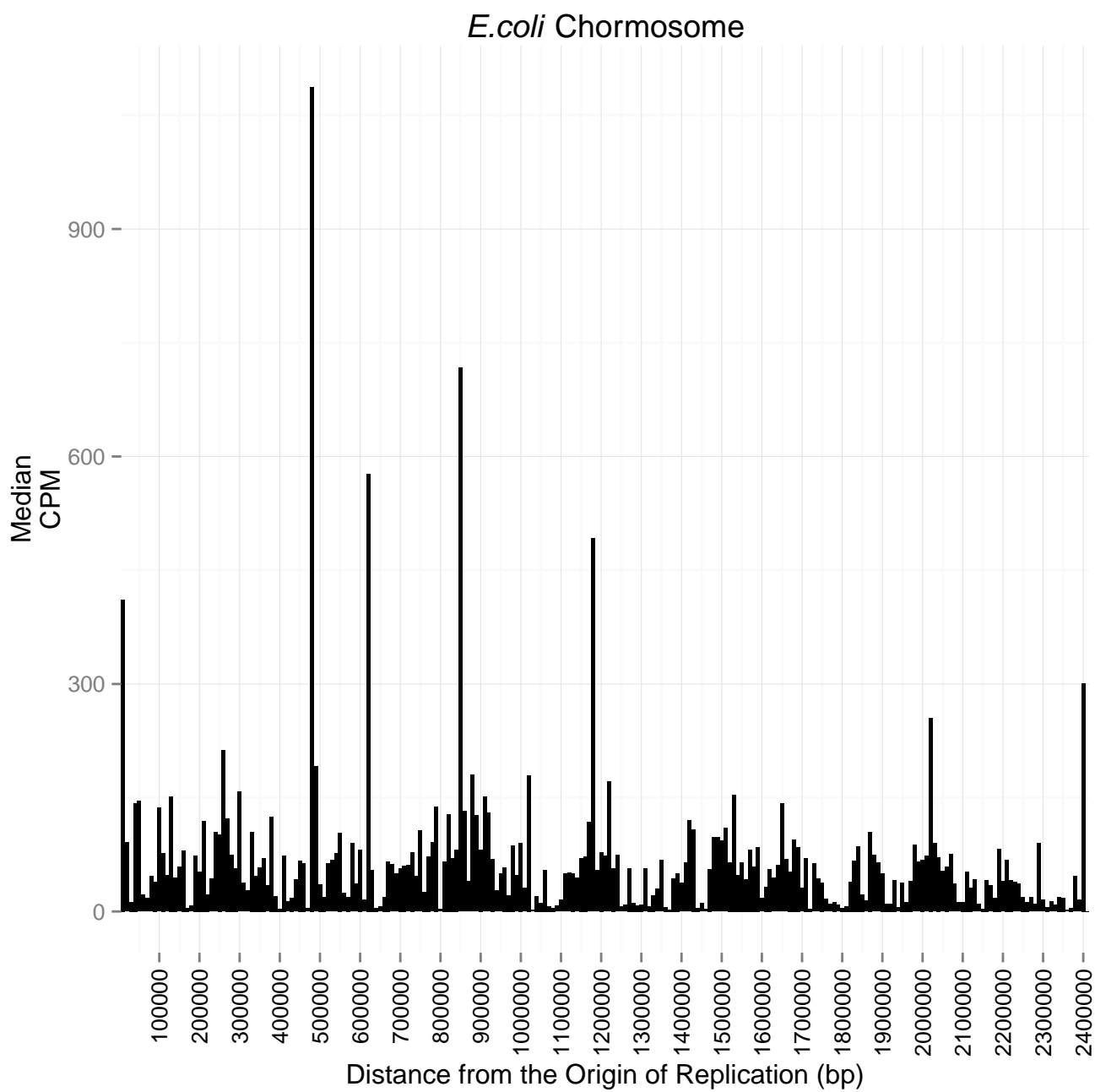


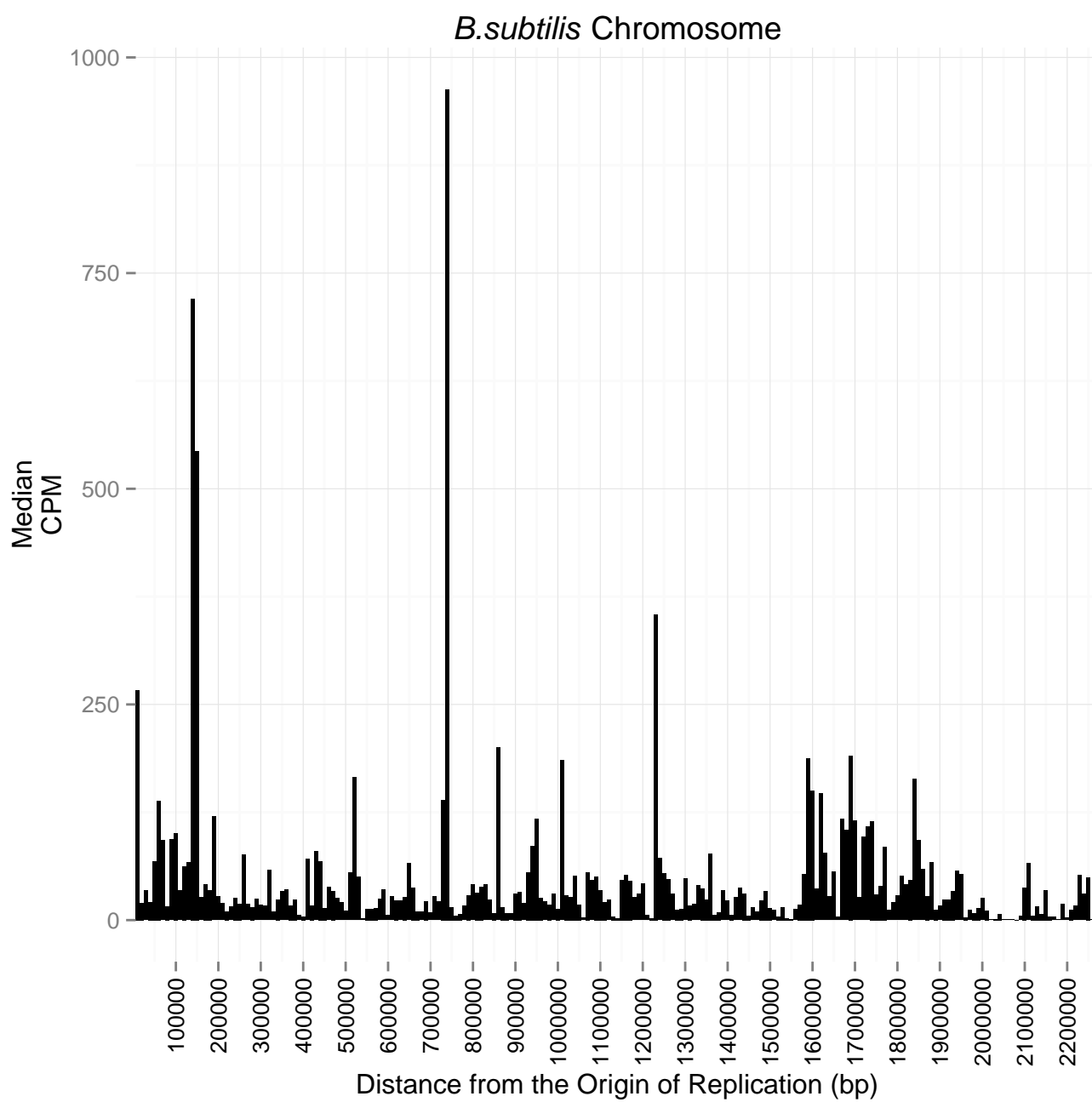


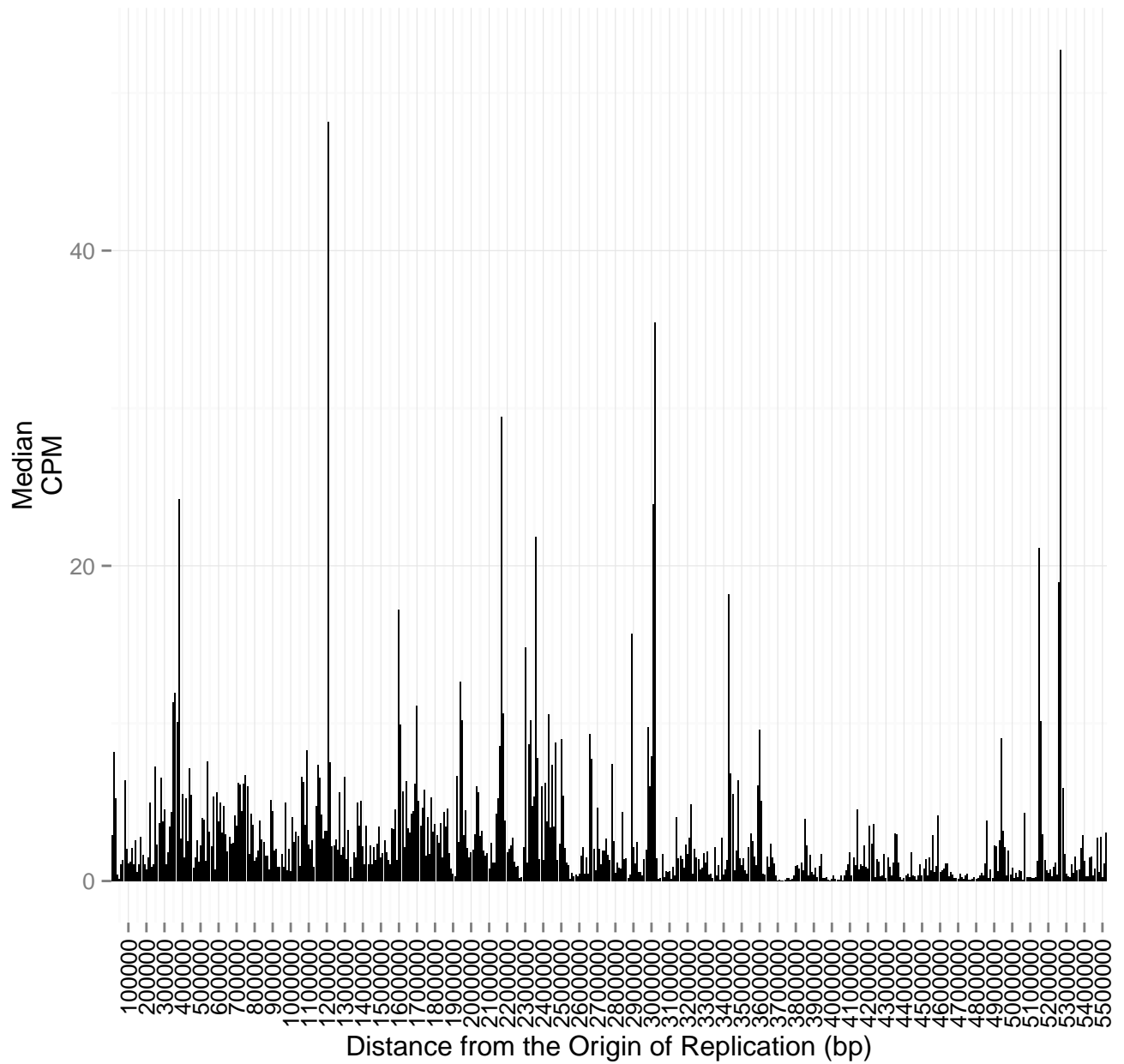


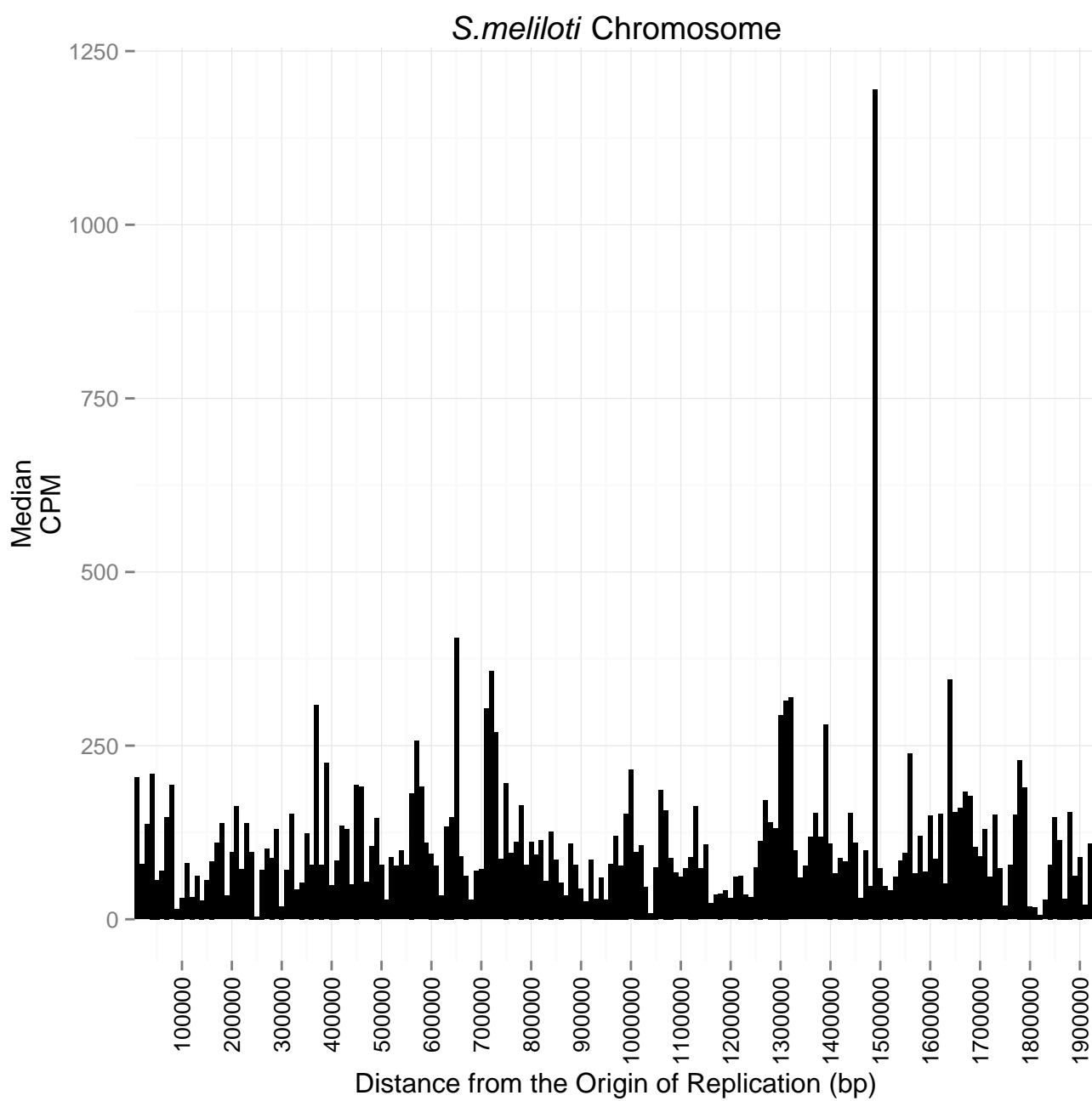


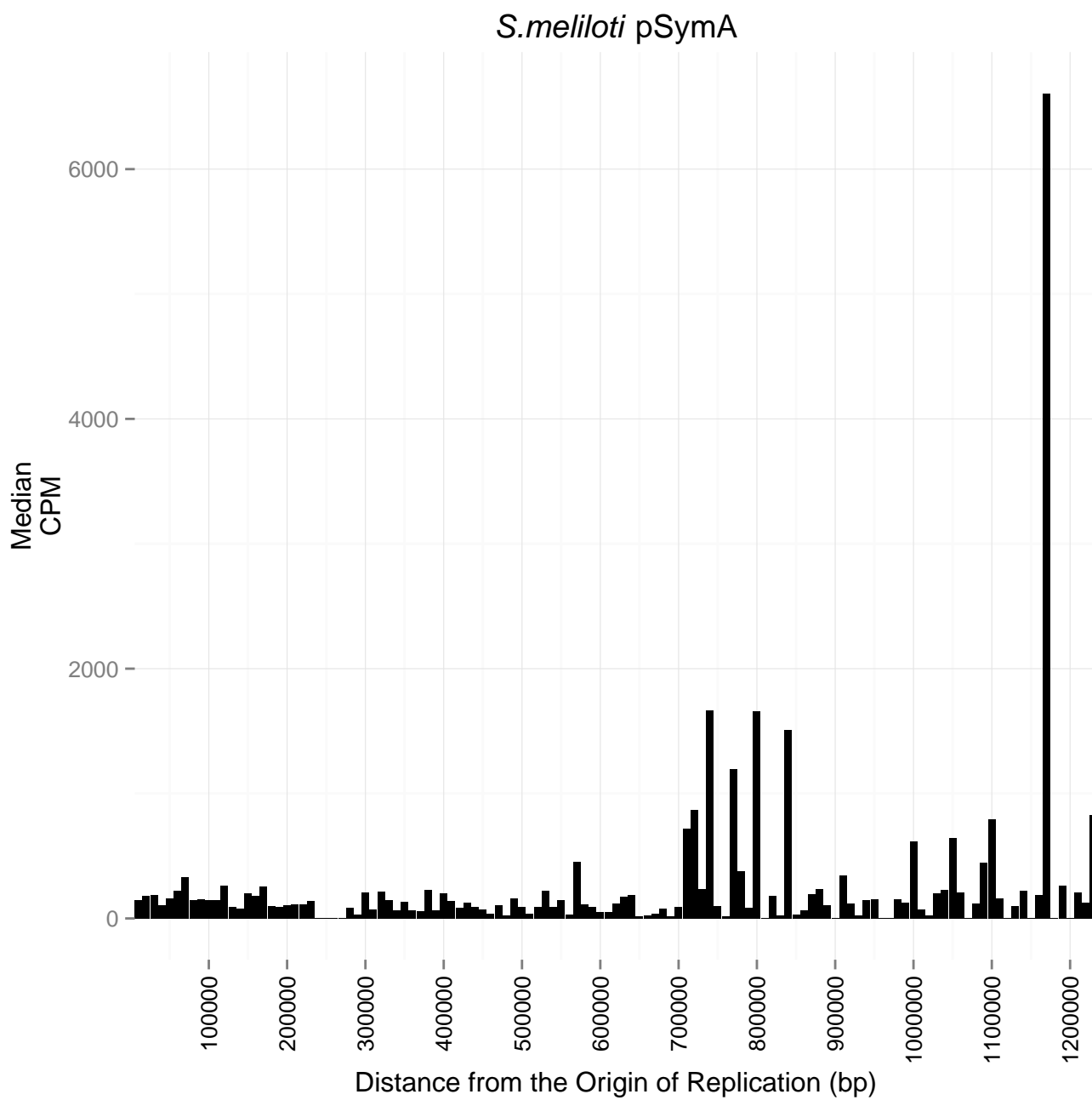
Gene expression graphs



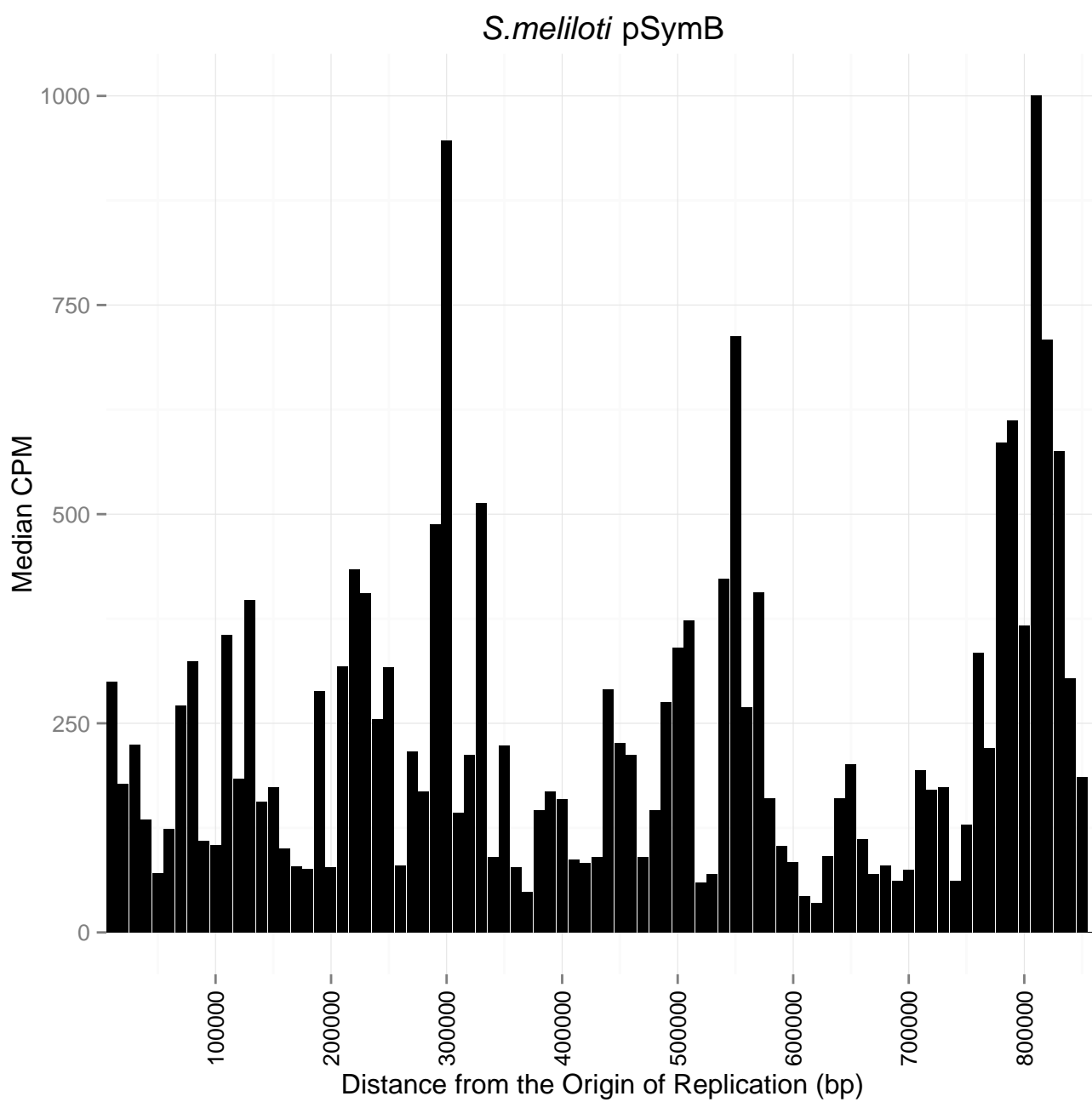


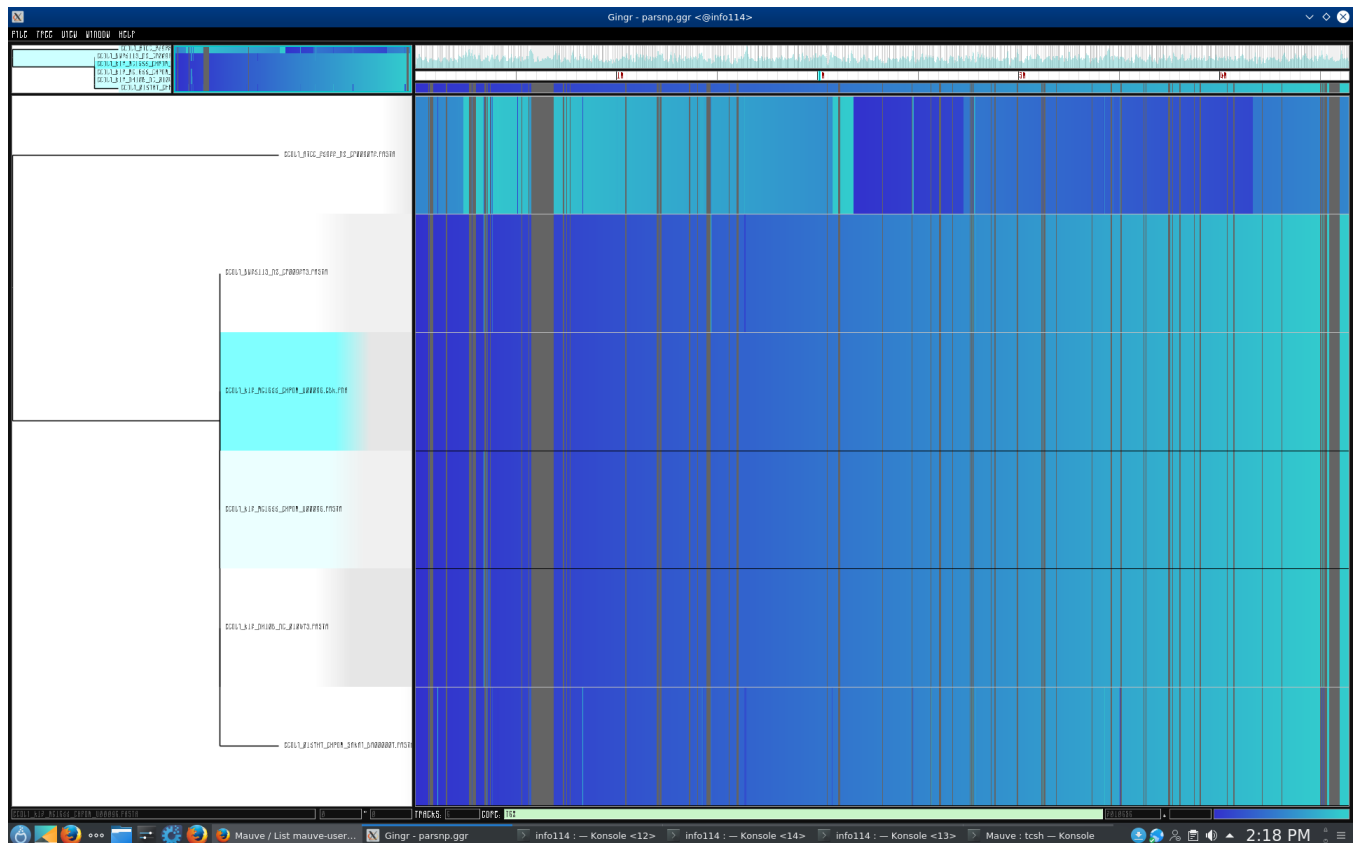
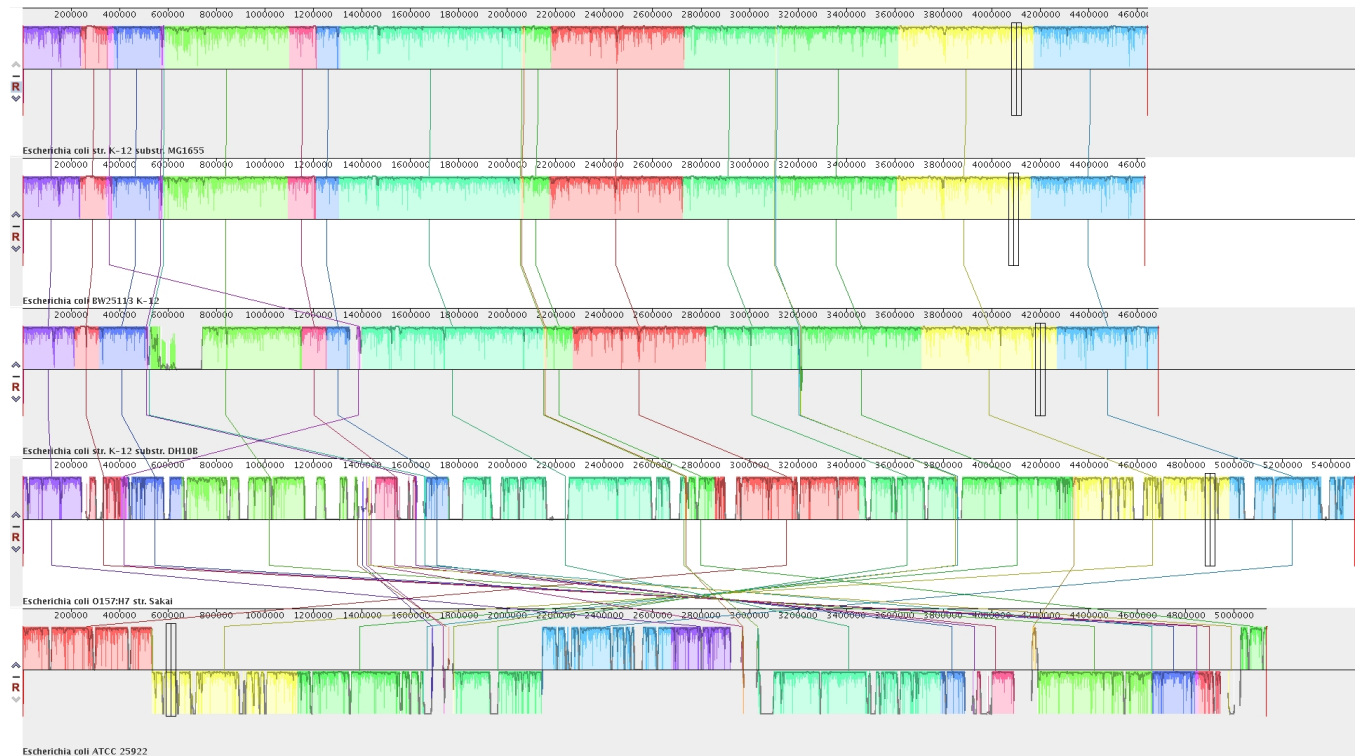
*Streptomyces* Chromosome











Bacteria Strain/Species	GEO Accession Number	Date Accessed
<i>E. coli</i> K12 MG1655	GSE60522	December 20, 2017
<i>E. coli</i> K12 MG1655	GSE73673	December 19, 2017
<i>E. coli</i> K12 MG1655	GSE85914	December 19, 2017
<i>E. coli</i> K12 MG1655	GSE40313	November 21, 2018
<i>E. coli</i> K12 MG1655	GSE114917	November 22, 2018
<i>E. coli</i> K12 MG1655	GSE54199	November 26, 2018
<i>E. coli</i> K12 DH10B	GSE98890	December 19, 2017
<i>E. coli</i> BW25113	GSE73673	December 19, 2017
<i>E. coli</i> BW25113	GSE85914	December 19, 2017
<i>E. coli</i> O157:H7	GSE46120	August 28, 2018
<i>E. coli</i> ATCC 25922	GSE94978	November 23, 2018
<i>B. subtilis</i> 168	GSE104816	December 14, 2017
<i>B. subtilis</i> 168	GSE67058	December 16, 2017
<i>B. subtilis</i> 168	GSE93894	December 15, 2017
<i>B. subtilis</i> 168	GSE80786	November 16, 2018
<i>S. coelicolor</i> A3	GSE57268	March 16, 2018
<i>S. natalensis</i> HW-2	GSE112559	November 15, 2018
<i>S. meliloti</i> 1021 Chromosome	GSE69880	December 12, 2017
<i>S. meliloti</i> 2011 pSymA	NC_020527 (Dr. Finan)	April 4, 2018
<i>S. meliloti</i> 1021 pSymA	GSE69880	November 15, 18
<i>S. meliloti</i> 2011 pSymB	NC_020560 (Dr. Finan)	April 4, 2018
<i>S. meliloti</i> 1021 pSymB	GSE69880	November 15, 18

Table 3: Summary of strains and species found for each gene expression analysis. Gene Expression Omnibus accession numbers and date accessed are provided.

Bacteria and Replicon	Coefficient Estimate	Standard Error	P-value
<i>E. coli</i> Chromosome	$-6.03 \times 10^{-5}$	$1.28 \times 10^{-5}$	$2.8 \times 10^{-6}$
<i>B. subtilis</i> Chromosome	$-9.7 \times 10^{-5}$	$2.0 \times 10^{-5}$	$1.2 \times 10^{-6}$
<i>Streptomyces</i> Chromosome	$-1.17 \times 10^{-6}$	$1.04 \times 10^{-7}$	$< 2 \times 10^{-16}$
<i>S. meliloti</i> Chromosome	$3.97 \times 10^{-5}$	$4.25 \times 10^{-5}$	NS ( $3.5 \times 10^{-1}$ )
<i>S. meliloti</i> pSymA	$1.39 \times 10^{-3}$	$2.53 \times 10^{-4}$	$4.9 \times 10^{-8}$
<i>S. meliloti</i> pSymB	$1.46 \times 10^{-4}$	$2.03 \times 10^{-4}$	NS ( $5.34.7 \times 10^{-1}$ )

Table 4: Linear regression analysis of the median counts per million expression data along the genome of the respective bacteria replicons. Grey coloured boxes indicate statistically significant results at the 0.5 significance level. Linear regression was calculated after the origin of replication was moved to the beginning of the genome and all subsequent positions were scaled around the origin accounting for bidirectionality of replication.