

Subs Paper Things to Do:

- why are the lin reg of dN , dS and ω NS but the subs graphs are...explain!
- mol clock for my analysis?
- GC content? COG? where do these fit?

Inversions and Gene Expression Letter Things to Do:

- create latex template for paper
- confirm inversions with dot plot
- make dot plot of just gene presence and absence matrix (instead of each site) to see if this will go better
- look up inversions and small RNA's paper Marie was talking about at Committee meeting
- write outline for letter
- write Abstract
- write intro
- write methods
- compile tables (supplementary)
- write results
- write discussion
- write conclusion
- do same ancestral/phylogenetic analysis that I did in the subs paper

General Things to Do:

- summarize references 40 and 56 from Committee meeting report (Brian was asking)

Last Week

- ✓Downloaded proteomes for Inversions blast analysis
- ✓detailed list of things for Queenie to do
- ✓(virtual) meeting with Queenie to update her on next steps for the inversions project

- ✓organize inversion paper notes into broad categories for intro
- ✓outline for inversions paper
- ✓wrote first rough draft of intro for inversions paper
- ✓Brian's edits to substitution paper

Most of last week I spent getting Queenie caught up on the inversions project. She is still excited to work on it and we will be meeting twice a week (virtually) to discuss her progress. So far she is working on combining the dataframes with information on gene expression and gene position.

I finished the edits you asked me to make on the substitutions paper and have attached the latest draft to this email. **If you could look over this again that would be great!**

This Week

- check on Queenie's progress and double check her normalization code
- look over requirements for GBE submission and create a folder/complete draft of the paper to be submitted
- write cover letter for substitutions paper submission
- work on code to grab gene info for inversions project

Next Week

- submit substitutions paper
- begin to look into how to perform blast to confirm inversions
- start writing methods for the inversions and gene expression paper
- help queenie with anything else she might need

Bacteria and Replicon	Genome Average		
	dS	dN	ω
<i>S. meliloti</i> Chrom + <i>A. tumefaciens</i>	12.5529	0.0553	0.0265
<i>E. coli</i> Chromosome	0.2387	0.0101	0.0441
<i>B. subtilis</i> Chromosome	0.4201	0.0243	0.0714
<i>Streptomyces</i> Chromosome	0.0458	0.0011	0.0335
<i>S. meliloti</i> Chromosome	0.0029	0	0
<i>S. meliloti</i> pSymA	0.0835	0.0099	0.1645
<i>S. meliloti</i> pSymB	0.0940	0.0084	0.1142

Table 1: Weighted averages calculated for each bacterial replicon on a per genome basis using the gene length as the weight. Arithmetic mean calculated for the per gene averages for each bacterial replicon.

Bacteria and Replicon	Protein Coding Sequences
<i>E. coli</i> Chromosome	$-1.43 \times 10^{-8}***$
<i>B. subtilis</i> Chromosome	$-5.55 \times 10^{-8}***$
<i>Streptomyces</i> Chromosome	$7.49 \times 10^{-8}***$
<i>S. meliloti</i> Chromosome	$-5.99 \times 10^{-7}***$
<i>S. meliloti</i> pSymA	$-5.18 \times 10^{-7}***$
<i>S. meliloti</i> pSymB	$1.67 \times 10^{-7}***$

Table 2: Logistic regression analysis of the number of substitutions along all protein coding positions of the genome of the respective bacteria replicons. Grey coloured boxes indicate a negative logistic regression coefficient estimate. All results are statistically significant. Logistic regression was calculated after the origin of replication was moved to the beginning of the genome and all subsequent positions were scaled around the origin accounting for bidirectional replication. All results are marked with significance codes as followed: $< 0.001 = '***'$, $0.001 < 0.01 = '**'$, $0.01 < 0.05 = '*'$, $> 0.05 = 'NS'$.

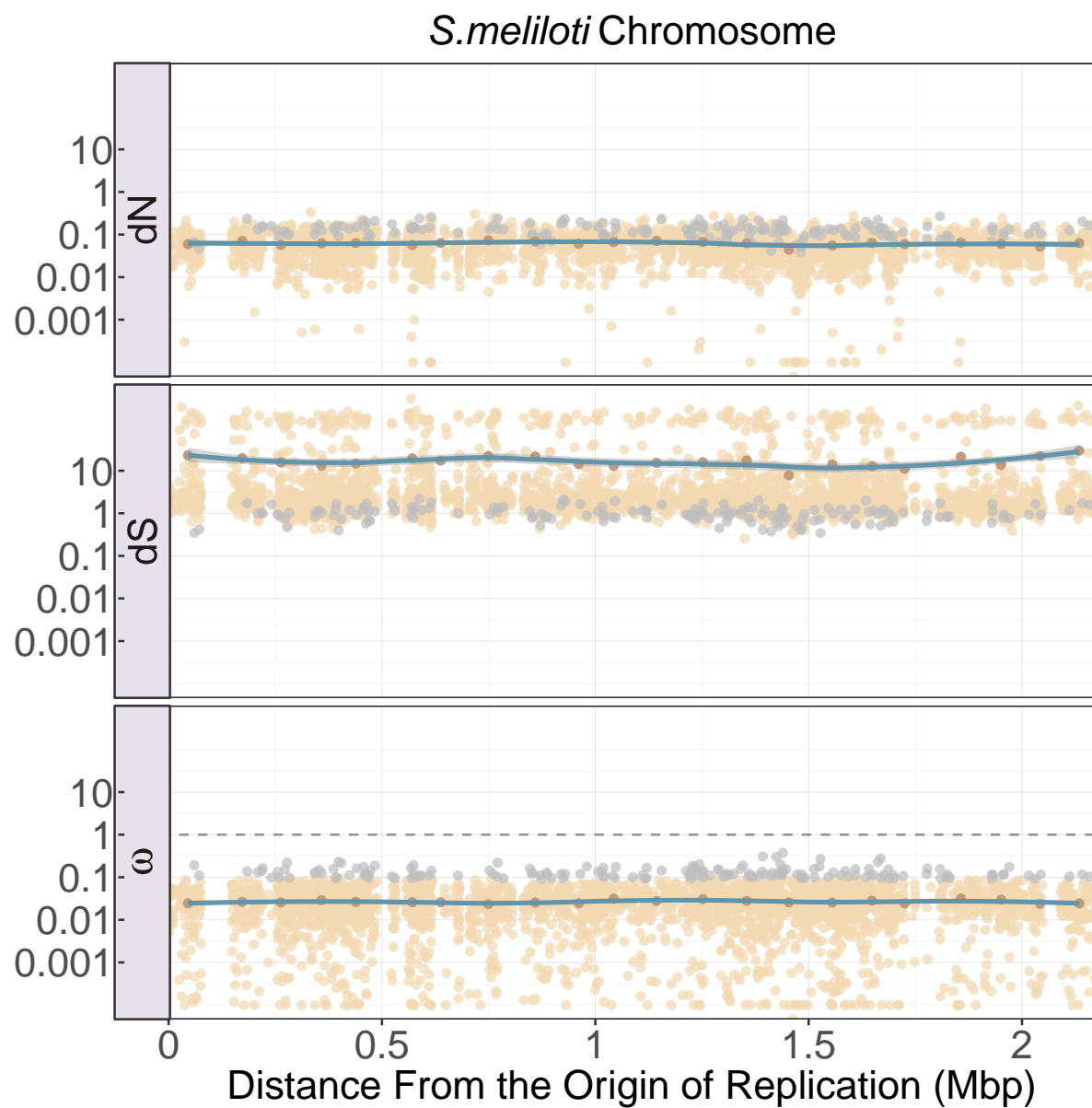
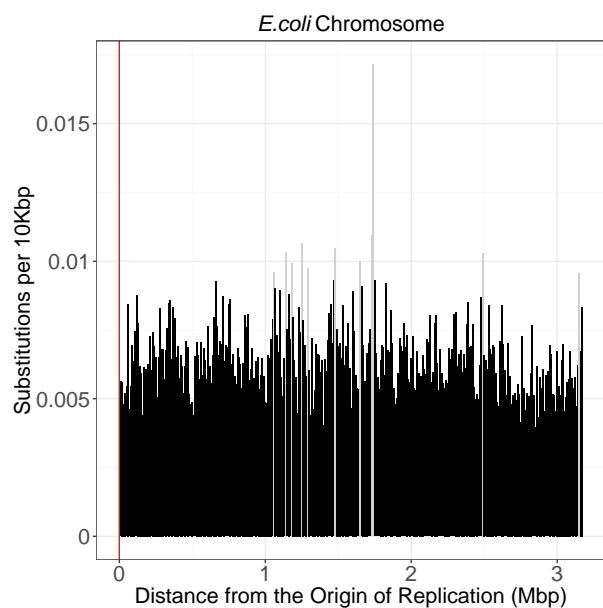
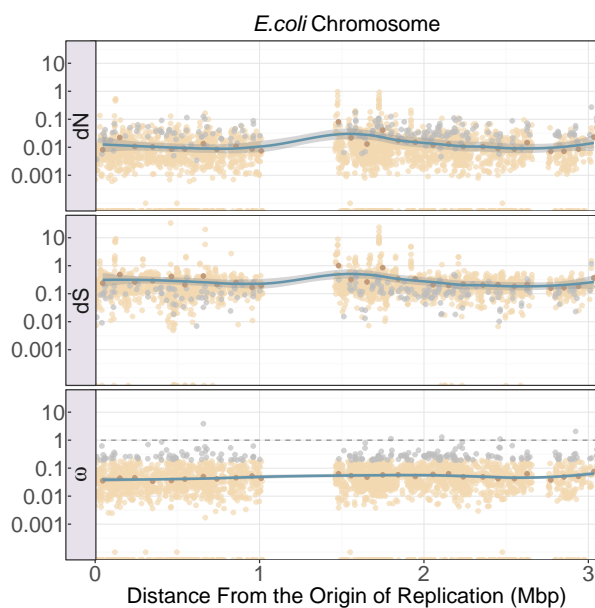


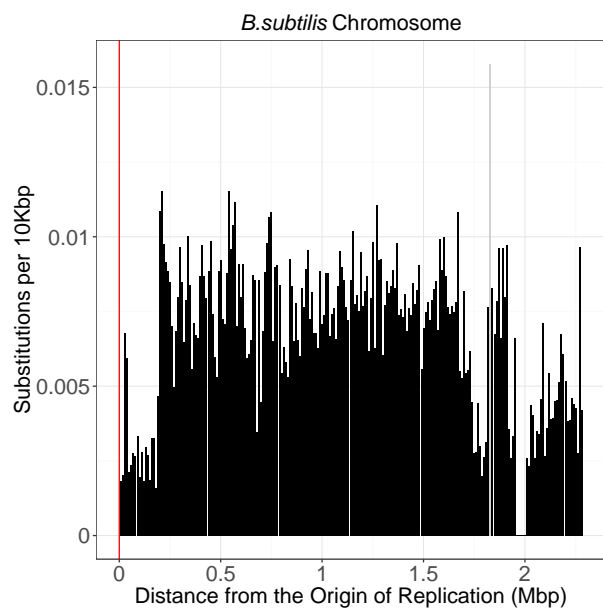
Figure 1: dN , dS , and ω values for *S. meliloti* chromosomes and *A. tumefaciens*.



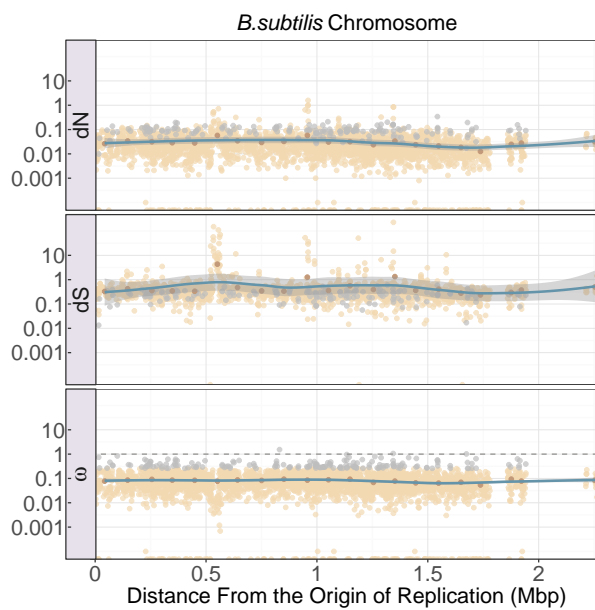
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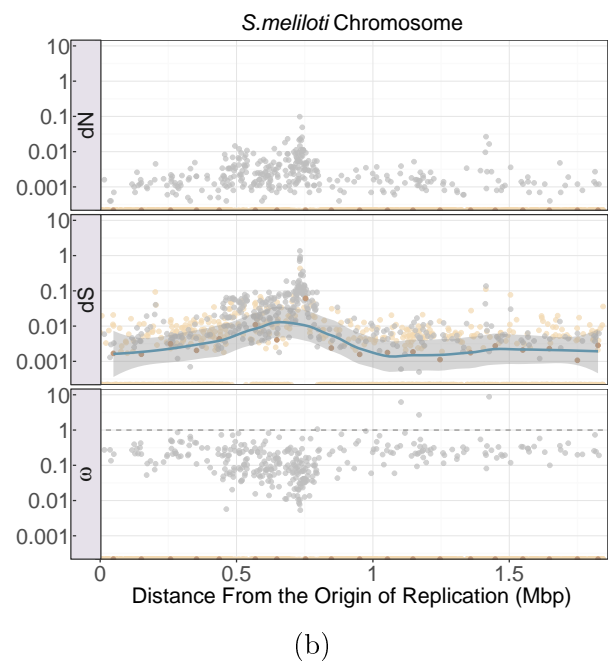
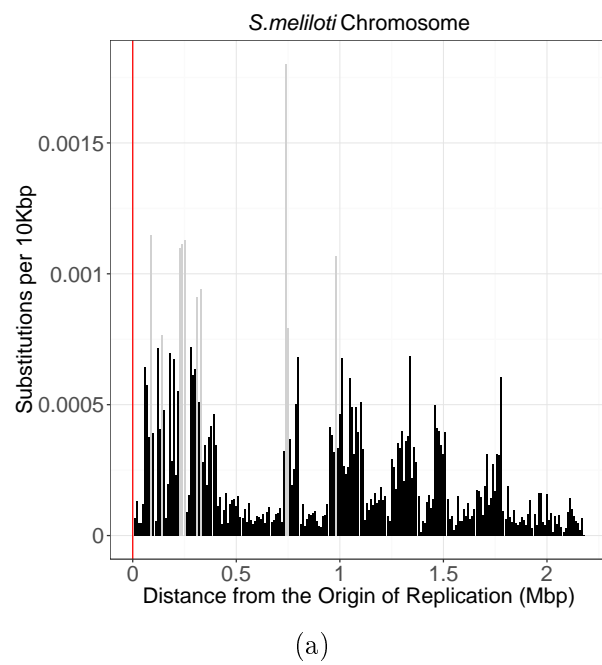
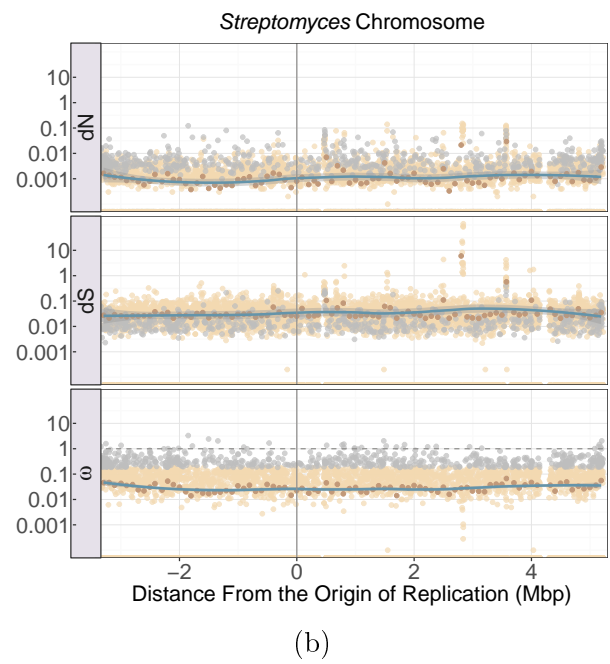
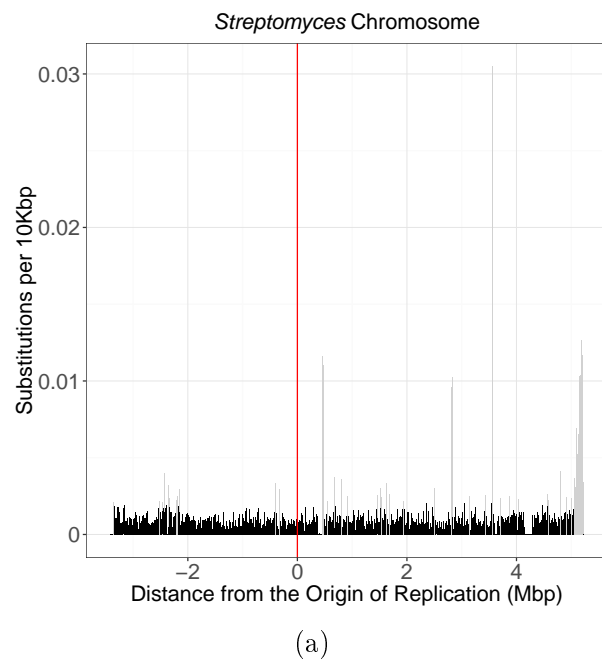
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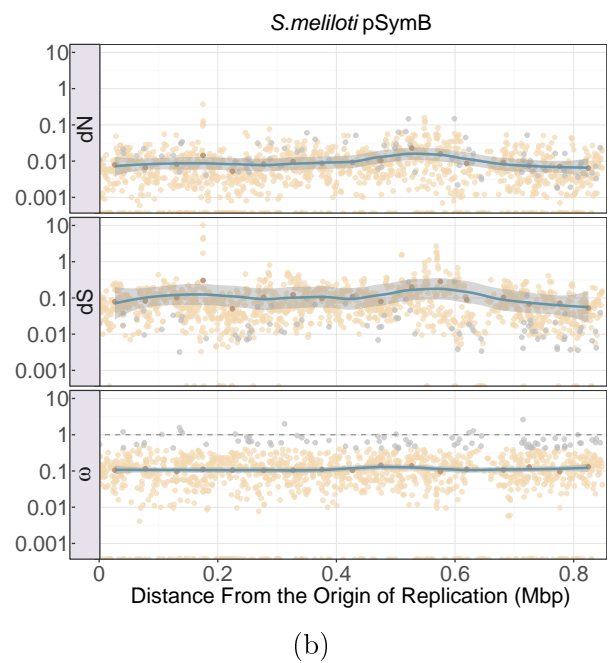
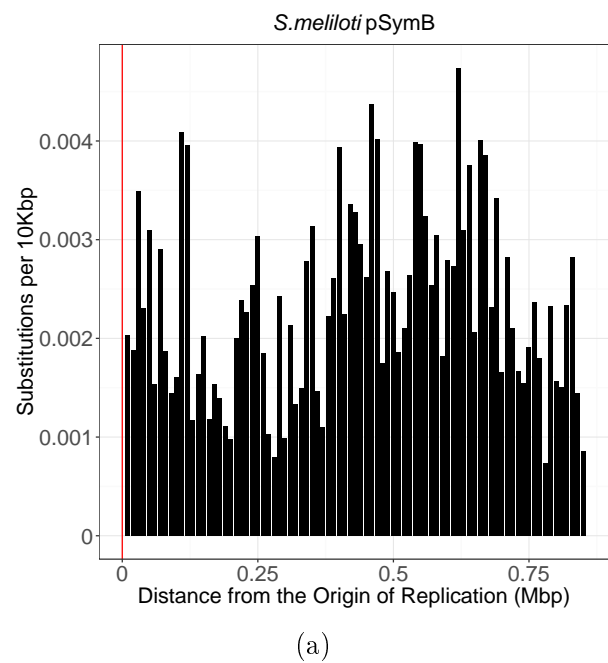
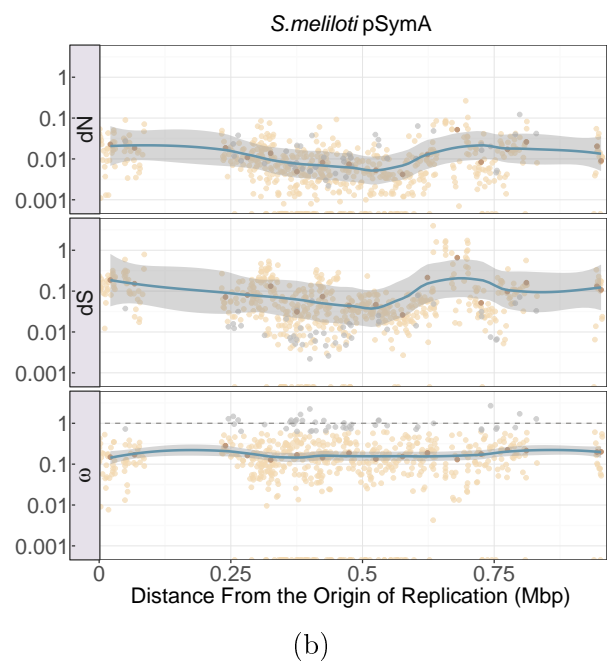
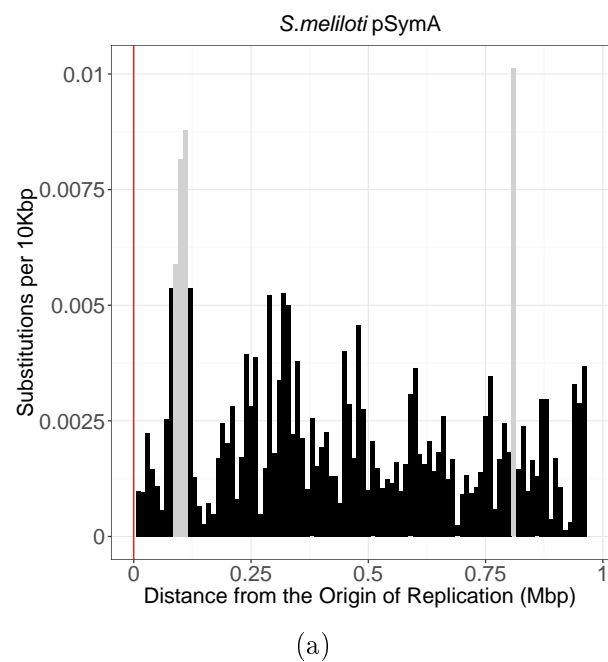


(a)



(b)





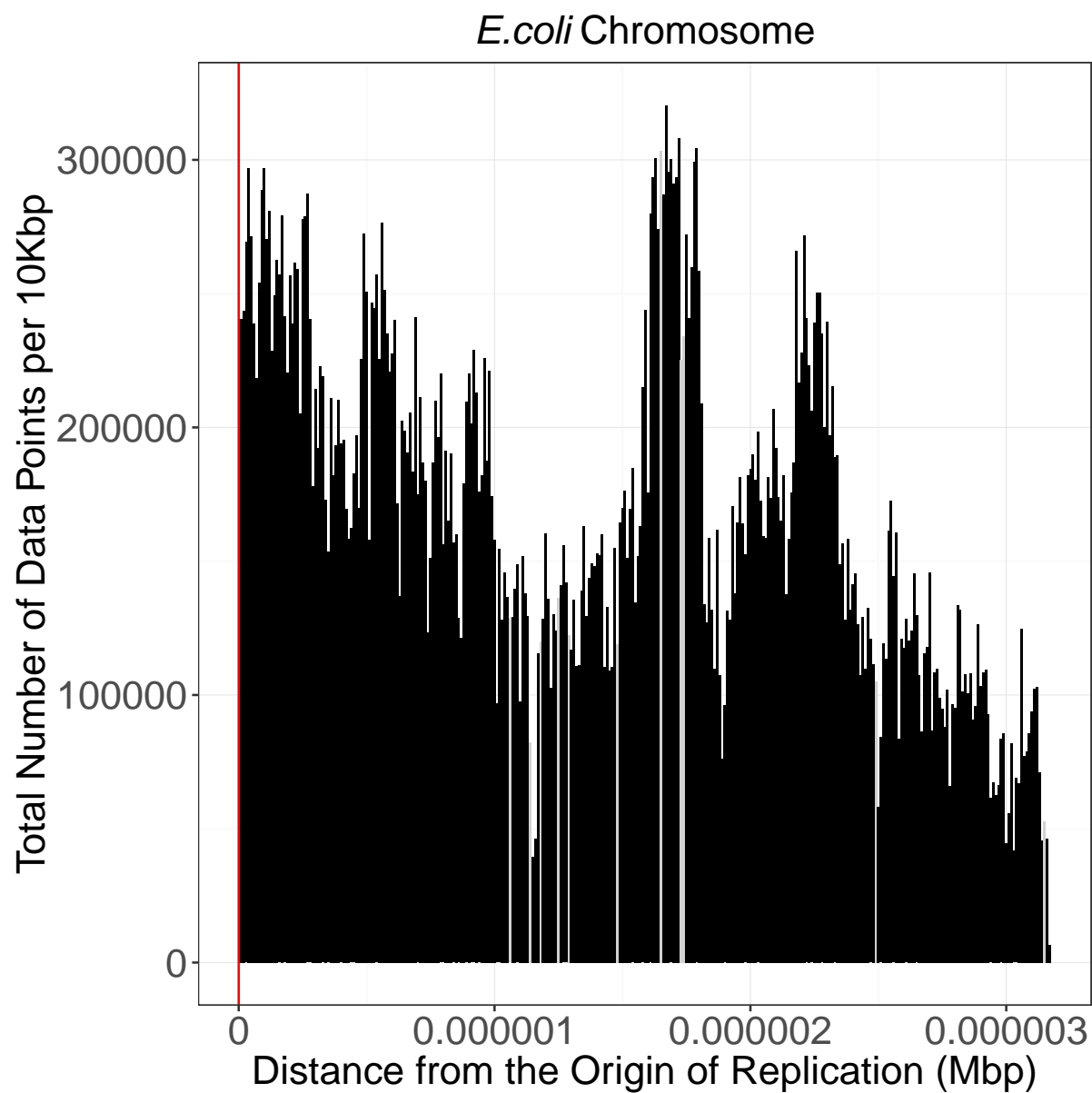


Figure 8: Distribution of total number of substitution data points per 10Kbp in genome.

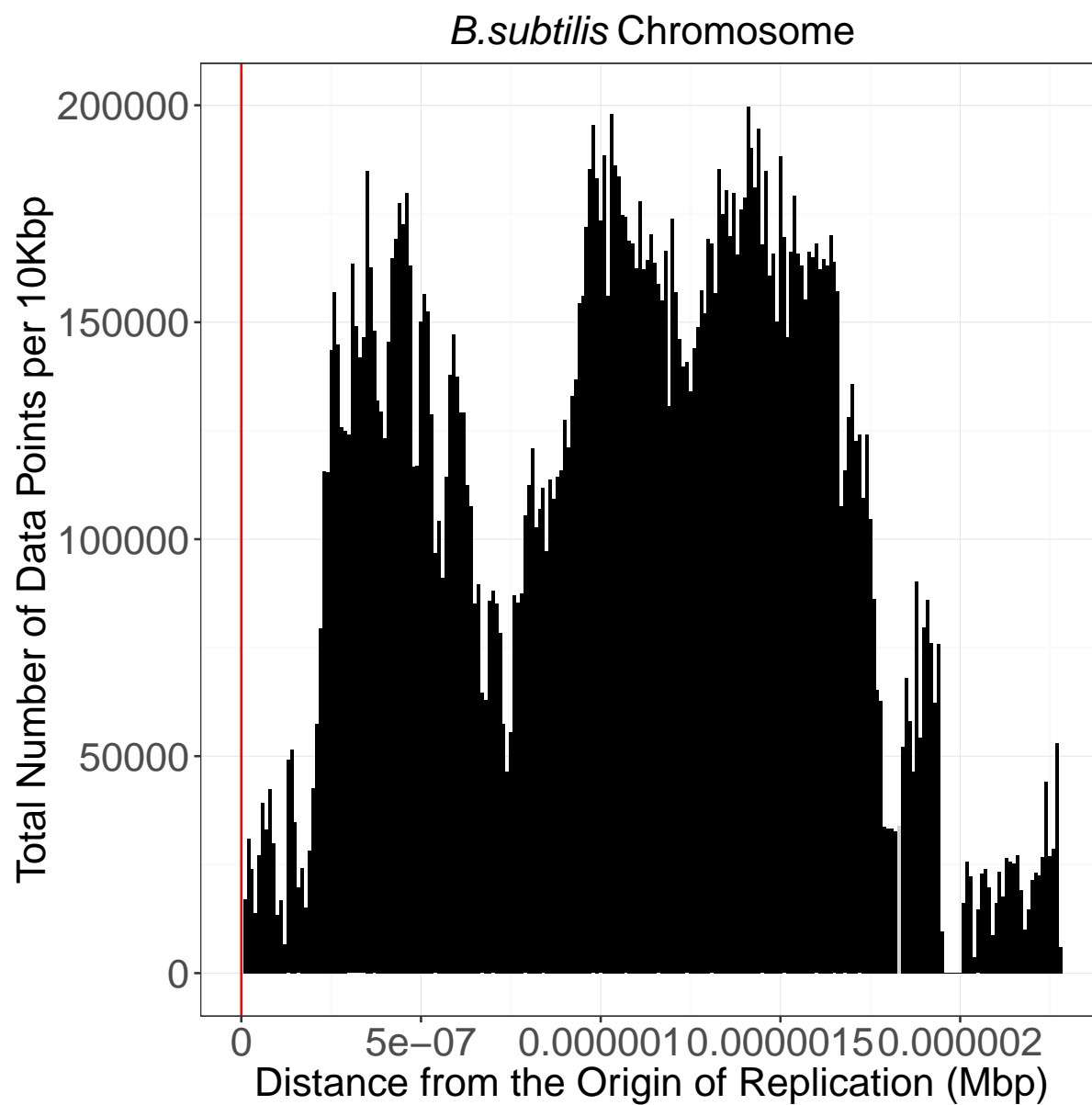


Figure 9: Distribution of total number of substitution data points per 10Kbp in genome.

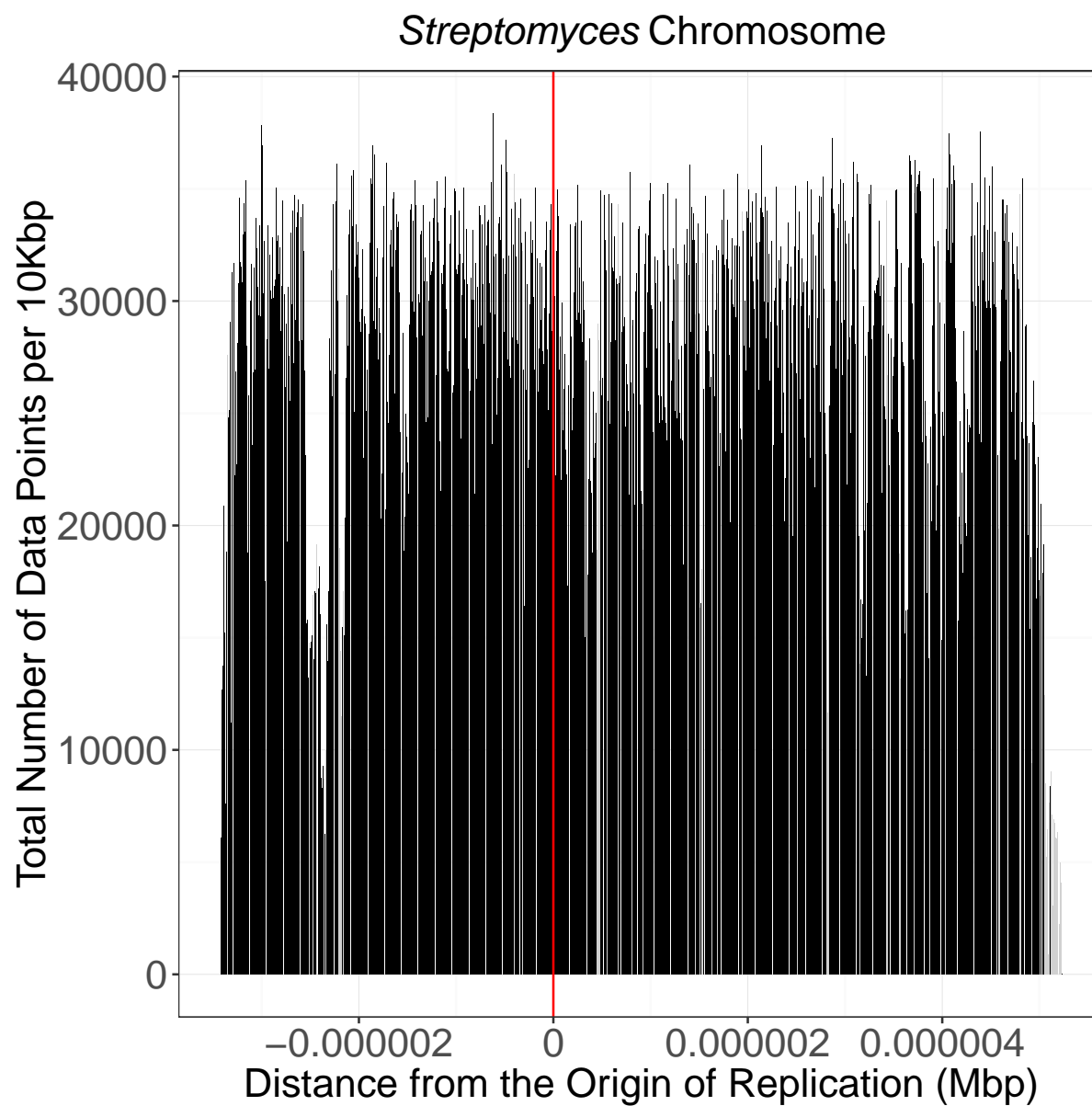


Figure 10: Distribution of total number of substitution data points per 10Kbp in genome.

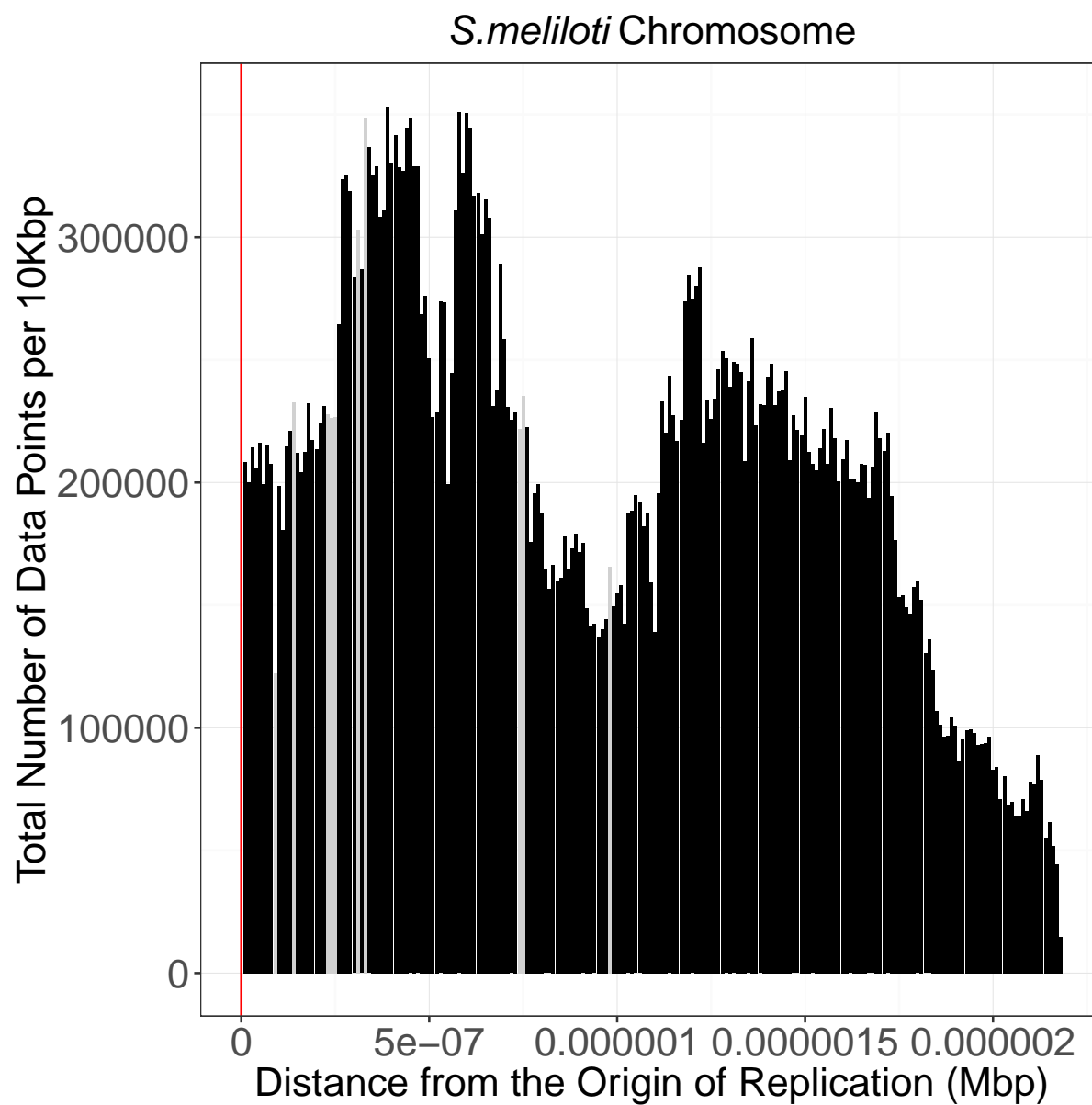


Figure 11: Distribution of total number of substitution data points per 10Kbp in genome.

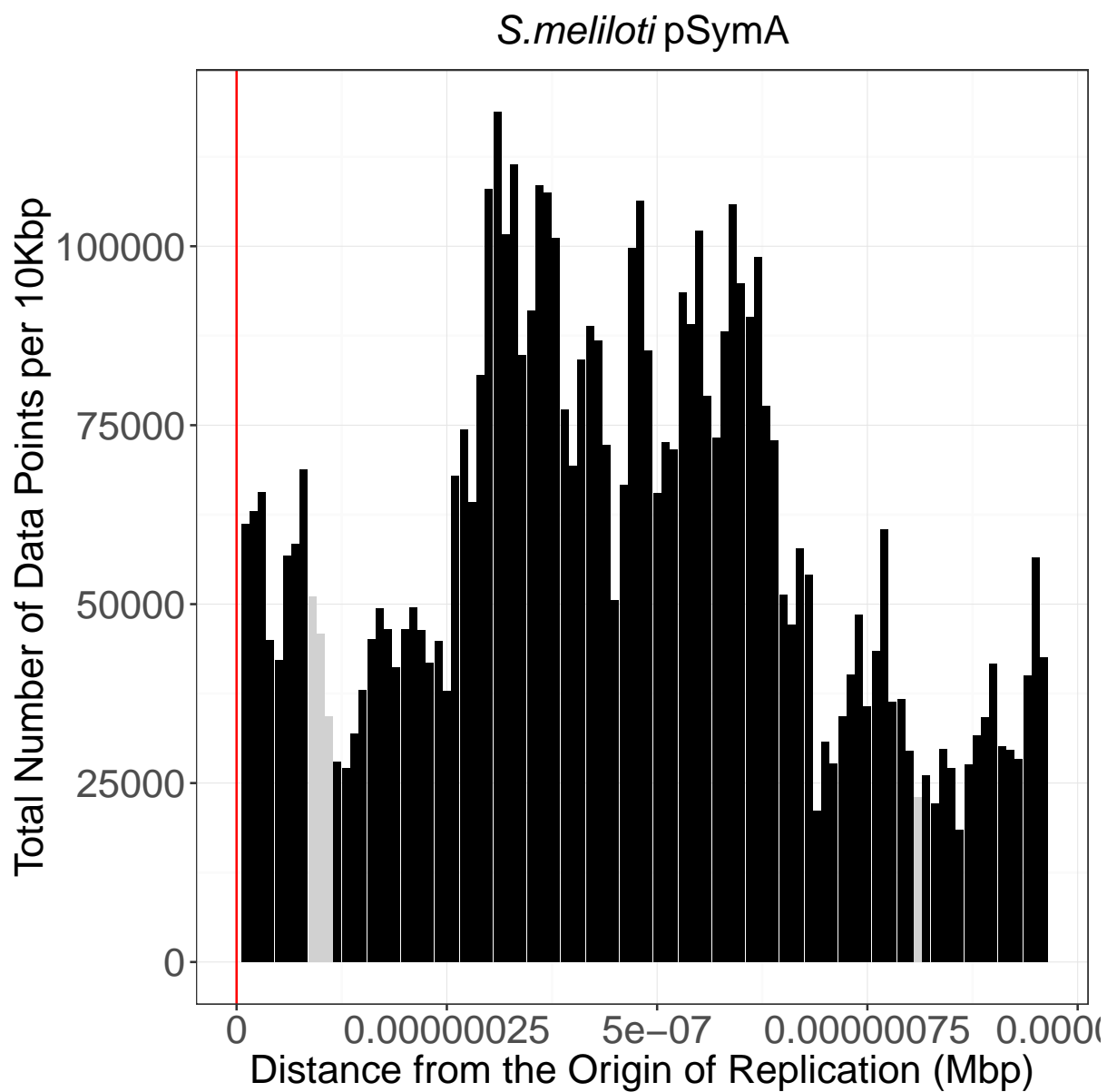


Figure 12: Distribution of total number of substitution data points per 10Kbp in genome.

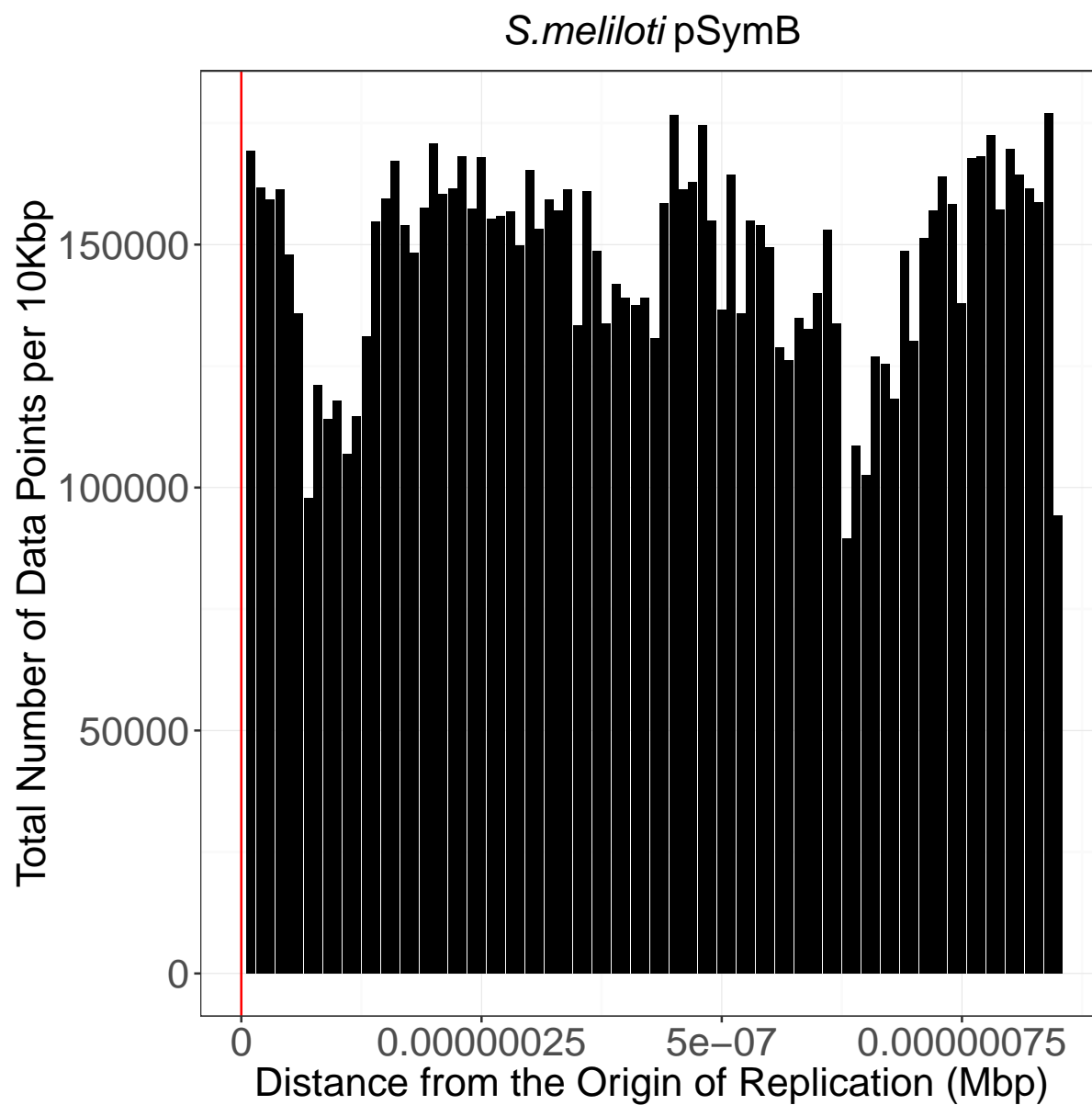


Figure 13: Distribution of total number of substitution data points per 10Kbp in genome.

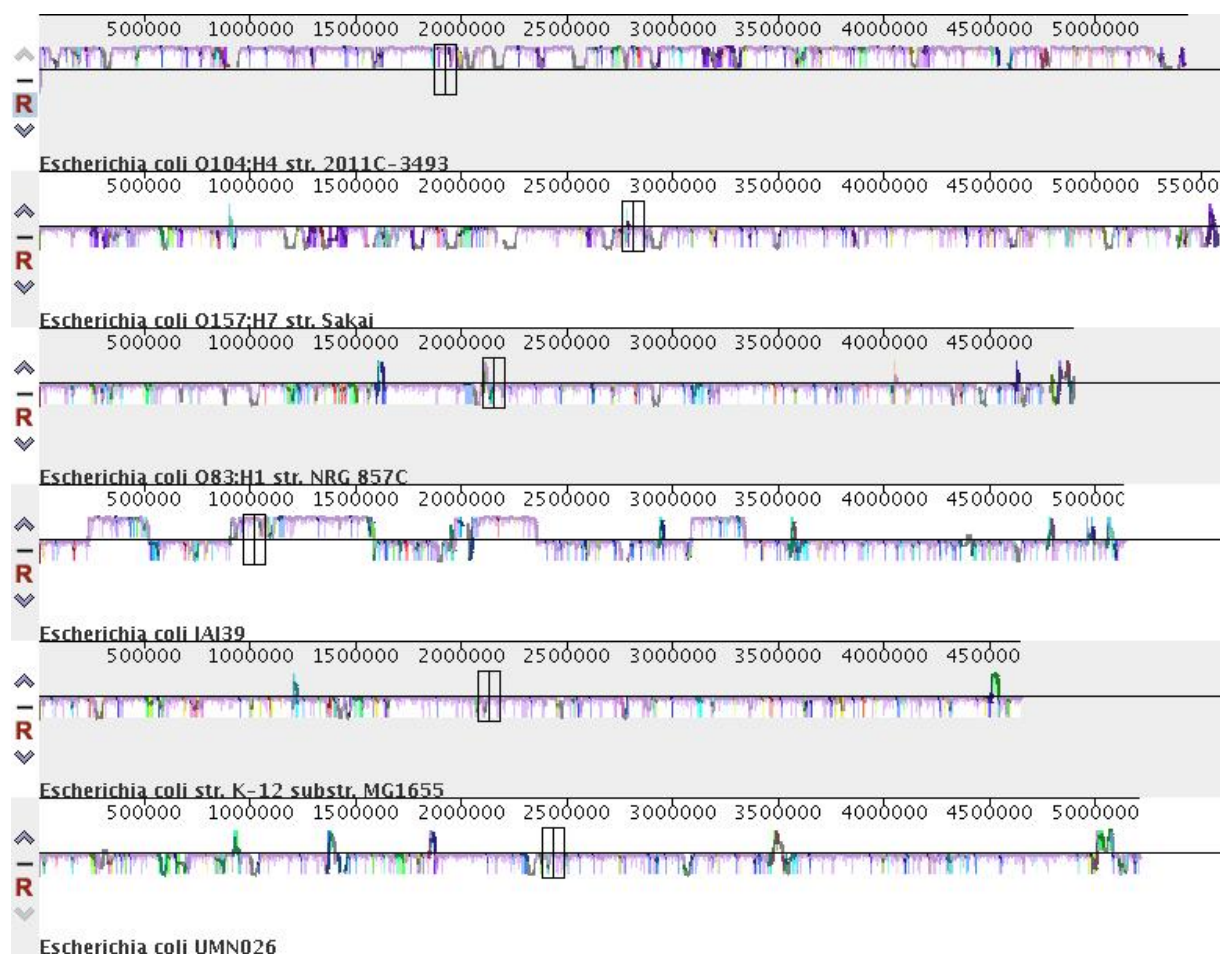


Figure 14: progressiveMauve alignment of *Escherichia coli* genomes highlighting the “backbone” of the alignment (matching regions).



Figure 15: progressiveMauve alignment of *S. meliloti* Chromosomes highlighting the “backbone” of the alignment (matching regions).

Bacteria and Replicon	Average Number of Substitutions per bp
<i>E. coli</i> Chromosome	1.97×10^{-4}
<i>B. subtilis</i> Chromosome	1.93×10^{-4}
<i>Streptomyces</i> Chromosome	2.74×10^{-6}
<i>S. meliloti</i> Chromosome	9.72×10^{-5}
<i>S. meliloti</i> pSymA	6.54×10^{-5}
<i>S. meliloti</i> pSymB	1.99×10^{-4}

Table 3: Average number of protein coding substitutions calculated per base across all bacterial replicons. Outliers and missing data was not included in the calculation.

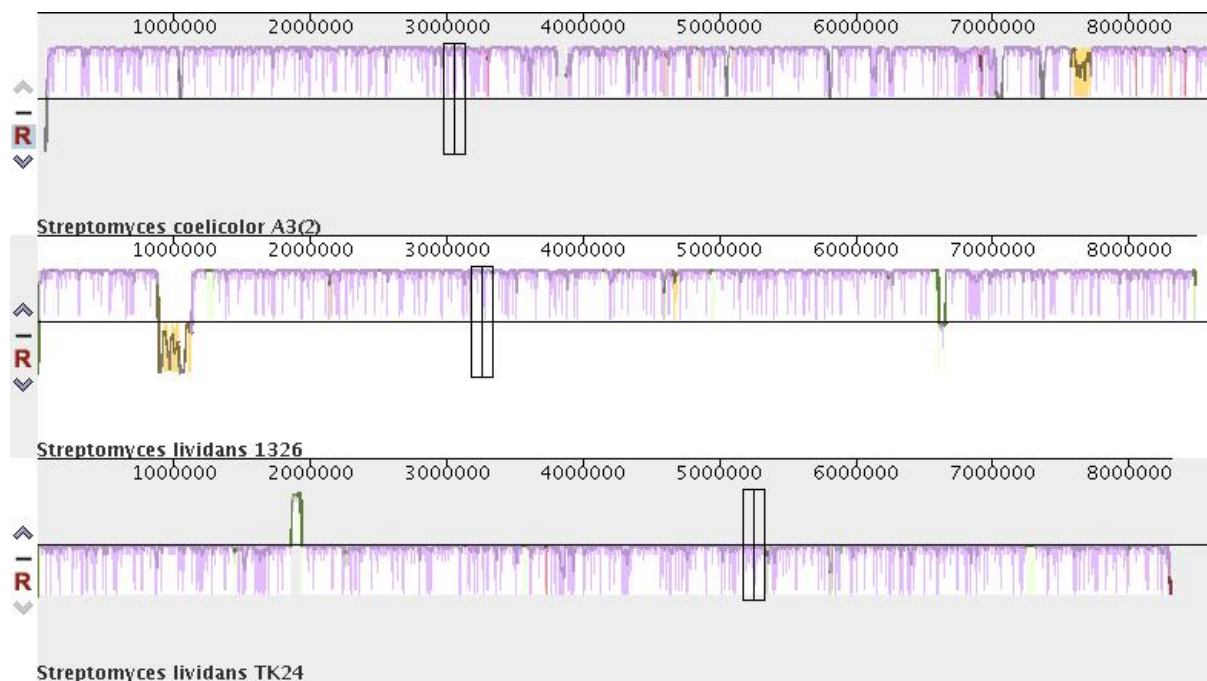


Figure 16: progressiveMauve alignment of *Streptomyces* genomes highlighting the “backbone” of the alignment (matching regions).