

This program is very similar to the original SOP program in terms of setup and running, there are just a few extra parameters to define.

1 - In the “def_param.h” file, you can set the fixed_beads and pulled_bead (which determines the pulling vector), as well as the integration time step (h), eH, deltax, R_limits and spring constants.

Lines 40 to 54 are the parameters that define the interaction between the protein and the nucleotide. Line 50 is the eH for this interaction (0.2 for Hsp70). On line 54, you need to define the number of residues in the protein, as the program will count the nucleotide as a second chain starting where the protein chain stops. On line 55 you need to define the type of nucleotide you want to model. 0 = rigid/static, 1 = normal, 2 = rigid but COM moves. 2 is the standard option for Hsp70.

2 - In the protein pdb file, after the protein chain atoms are listed, you need to include the nucleotide coordinates. Each atom needs to be listed as a “CA” type atom. Additionally, each atom needs to be listed as a valid amino acid type, “VAL” or similar. The chain label should be different than the protein chain label, and the atom and residue numbers need to go in order. See example below.

```

Atom Number (column 2)
|   Atom Type (3)
|   |   Amino Acid Type (4)
|   |   |   chain label (5)
|   |   |   |   Amino Acid Number (6)   X, Y, Z coordinates (columns 7-9)
|   |   |   |   |

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ATOM	2894	N	THR	A	383	86.461	93.710	98.574	1.00	78.54	N
ATOM	2895	CA	THR	A	383	85.604	94.293	97.553	1.00	81.73	C
ATOM	2896	C	THR	A	383	84.127	94.147	97.920	1.00	87.92	C
ATOM	2897	O	THR	A	383	83.321	95.042	97.667	1.00	93.10	O
ATOM	2898	CB	THR	A	383	85.868	93.675	96.165	1.00	79.19	C
ATOM	2899	OG1	THR	A	383	85.939	92.252	96.276	1.00	78.83	O
ATOM	2900	CG2	THR	A	383	87.176	94.194	95.589	1.00	72.24	C
ATOM	2901	N	GLY	A	384	83.789	93.014	98.524	1.00	86.36	N
ATOM	2902	CA	GLY	A	384	82.421	92.718	98.896	1.00	77.50	C
ATOM	2903	C	GLY	A	384	81.921	91.470	98.200	1.00	79.55	C
ATOM	2904	O	GLY	A	384	80.842	90.970	98.498	1.00	96.10	O
ATOM	2905	CA	VAL	D	385	107.345	78.748	100.059	1.00	29.94	P
ATOM	2906	CA	VAL	D	386	105.874	79.100	100.230	1.00	26.91	O
ATOM	2907	CA	VAL	D	387	107.966	79.197	98.771	1.00	21.55	O
ATOM	2908	CA	VAL	D	388	108.314	79.192	101.283	1.00	27.47	O
ATOM	2909	CA	VAL	D	389	106.758	76.040	99.541	1.00	36.86	P
ATOM	2910	CA	VAL	D	390	105.484	75.859	100.294	1.00	37.56	O
ATOM	2911	CA	VAL	D	391	106.688	76.306	98.061	1.00	42.90	O
ATOM	2912	CA	VAL	D	392	107.624	77.172	100.297	1.00	29.89	O
ATOM	2913	CA	VAL	D	393	107.794	74.840	99.809	1.00	33.13	O
ATOM	2914	CA	VAL	D	394	109.022	74.729	99.085	1.00	25.69	C
ATOM	2915	CA	VAL	D	395	109.990	73.959	99.974	1.00	25.89	C
ATOM	2916	CA	VAL	D	396	109.443	72.652	100.191	1.00	48.19	O
ATOM	2917	CA	VAL	D	397	111.342	73.754	99.346	1.00	46.64	C
ATOM	2918	CA	VAL	D	398	112.273	73.658	100.427	1.00	32.55	O
ATOM	2919	CA	VAL	D	399	111.189	72.397	98.690	1.00	49.75	C
ATOM	2920	CA	VAL	D	400	112.432	71.743	98.448	1.00	41.64	O
ATOM	2921	CA	VAL	D	401	110.353	71.666	99.718	1.00	53.18	C
ATOM	2922	CA	VAL	D	402	109.631	70.546	99.086	1.00	46.97	N

3 - You need to obtain a contact map of your pdb file to input to the program. The “get_contacts_PDB_nucl.c” gives the command to make the executable “run_contacts”. This produces four files, “Contact_map_” for the inter- and intra-chain contacts. There are two of each, one with backbone only contacts (labeled “b”) and one with backbone and side-chain contacts (labeled “sc_and_b”). To get the correct contacts for the program, you need to cat the inter and intra-chain backbone only contact files into one file. This is the file you will pass to the SOP program when running. If using a different version of the contact program, just be sure that your file contains the correct contacts (inter and intra-chain, backbone only), the output formats are pretty standard (exception: compare contacts program output has an extra column).

4 - The pbs script is set up the same as for a normal SOP run, with the compile and run commands the same, only using the SOP-nucleotide program files. The run command is:

```
./run_protein_Fene <pdb_file>.pdb <Contacts_file> <run_number>
```

5 - The output file you will want to use for plotting the force vs. extension curve is the “out<pdb_file>_<k_trans>_<deltax>_<h>.dat”. Column 3 is the extension in Å, column 4 is the force (*70 to convert to pN). The other file ends in “_d.dat”, which Ji Young programmed to print, not sure what’s in this file exactly.