

NIH grant proposal

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Things to do

4: Finish

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6: Douglas add more background here?

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Specific aims

We propose a large-scale bioinformatics study to identify the effects of synonymous codon usage on protein structure. We intend to address causal relationships rather than statistical associations by developing a mathematically and statistically rigorous framework, which we will use to address a number of hypotheses.

Aim 1: Develop a comprehensive, verifiable, and rigorous framework to elucidate relationships between codon usage bias and protein structure.

We will develop a mathematical and computational framework that will allow the robust detection of relationships between codon usage and variables describing protein structure, even when the overall statistical correlation between the two variables is low.

Aim 2: Identify particular effects of synonymous mutations on protein structure.

We will use the framework established in aim 1 to develop a computational pipeline to detect proteins whose structures are affected by differential codon usage. We hypothesize that many of these structural changes can cause protein misfolding, which may be clinically important.

Aim 3: Elucidate general mechanisms that underlie differential codon usage.

We intend to use the same framework and pipeline described in the previous aims to investigate our hypotheses that codon usage bias is causally related to protein domain organization, secondary structure, knotting, folding environment, and structural complexity. Secondly, we wish to establish codon usage bias as an important biological mechanism.

Significance

Existing literature

Correlation with expression

It is known that codon usage bias correlates with expression levels [15, 19]. There are strong indications that abundance of isoaccepting tRNA molecules is a causal factor of this differential expression [21, 30, 32, 40]. Proteins with high expression levels contain greater levels of frequently used codons, and frequently used codons are associated with higher levels of corresponding tRNAs. Thus there is believed to be a positive selection for a small, constrained set of codon–tRNA combinations, and concentrations of codons and tRNAs are related in a positive feedback cycle, where bias in either causes a positive selection for bias in the other. Therefore, statistically significant violations of this general trend are interesting because they indicate the presence of other

selection biases. Such selection biases may be at the heart of important mechanisms of protein expression, some which are probably currently unknown.

Terminology

Because of strong correlation described above, we call infrequently used codons *slow*, and frequently used codons *fast*, except in cases where we address this correlation directly. Furthermore, we consider sequences containing a large proportion of fast codons to have high *codon usage bias*, and sequences containing either a moderate or low proportion to be *unbiased*, even though the bias of sequences with low proportion still deviate from the statistical mean.

Importance in translational dynamics

There is also substantial evidence that codon usage bias is also fundamentally linked to translation dynamics [4, 7, 8, 14, 26, 28]. Factors including mRNA secondary structure [3, 10, 39], mRNA stability [18], codon–tRNA affinity [21], and translational errors [13, 30, 47] have been suggested. In addition, studies have shown that codon usage bias is strongly correlated to ribosomal traffic [7, 9, 22]. Particularly, was demonstrated that, for efficient translation, overall codon usage should be skewed in favor of fast codons, and there should be a gradient in which slow codons are prevalent at the beginning of the transcript but rare toward the middle and end. Mitarai and Pedersen [28] used a simple computational model to show that the introduction of even a single slow codon near the end of the transcript can cause ribosomal traffic jams that drastically decrease translation rate, and presumably also expression. This is believed to be due to ribosomal queueing, a ribosome translating an mRNA transcript interacts physically with the translation process of another ribosome on the same transcript. This prevents both ribosomes from proceeding to subsequent codons.

Effects on protein structure

Because slow codons can cause pauses in translation, it has been suggested that synonymous codon usage can influence protein folding, leading to different folded states for transcripts with differing synonymous codon usage [7, 11, 23, 44]. Several studies have found that codon usage bias is correlated to protein structure [1, 5, 16, 34], including protein secondary structure [16, 31, 34, 41] and domain organization [17, 31].

The first known effect of synonymous mutations on protein structure came from Crombie et al. [11], who replaced 10 consecutive slow codons with fast codons in the *E. coli* TRP3 protein. Doing so reduced the native enzymatic activity by 1.5-fold. Still more surprising was the finding by [?] that a silent polymorphism in the mammalian multi-drug-resistant gene MDR1 altered its folded state and decreased its substrate specificity, without affecting its expression. A recent study by Zhou et al. [46] found a similar result for a natural mutation in the clinically important circadian rhythm protein FRQ. These studies hint to the existence of more examples of such effects. Furthermore, we argue that additional such cases are likely to be clinically important.

Limitations of existing approaches

In general, the bioinformatics studies by Adzhubei et al. [1], Biro [5], Gu et al. [16], Saunders and Deane [34] controlled for very few variables and were therefore able to identify only a few clear correlations (most notably with protein secondary structure). However, we hypothesize that these results were negative because the effect of codon usage bias on structure is a weak signal: the effects on most protein structure are minor or nonexistent for most proteins. However, the weakness of the signal does not belie the presence of general mechanisms behind the effects; this is evidence because, as shown by Crombie et al. [11], Zhou et al. [46], differential codon

usage can dramatically affect the folding of certain proteins. Therefore, we further hypothesize that general mechanisms exist even if few general correlations do, and that controlling for more variables will allow us to elucidate general mechanisms.

Further applications

In addition to clinically significant synonymous mutations and other differential codon usage, our data will have impact on additional applications. It has been recently shown that codon usage bias can be a pivotal factor in de novo protein design [20]. Although it is known that designed transcripts should contain mostly fast codons, and that there should be a gradient of codon bias along the transcript sequence, potential unwanted effects of synonymous codon usage on a protein product is not generally considered as part of protein design. We therefore note that the discovery of general mechanisms may be important to this application.

Assessing the impact of codon usage on protein structure has implications for protein folding analyses, especially if correlation is found between fast or slow codons and domain or interdomain regions. Prediction of protein structure given amino acid sequence is one of the foremost problems in biochemistry, however known determinants of structure are few [6] and the predictions made by current computational models frequently fall short of native conformations [12, 36]. Finding a relationship between codon bias and protein structure would provide considerable additional predictive power to such models.

Innovation

The previous studies by Adzhubei et al. [1], Biro [5], Gu et al. [16], Saunders and Deane [34] examining protein structure in the context of codon usage bias have been constrained to examinations of statistical correlations. Although they and many other studies [] have suggested that differential codon usage may influence protein structure, such effects have only been demonstrated in vivo by Zhou et al. [46?].

No studies attempting to investigate causal relationships between codon usage bias and protein structure have been thus far published in peer-reviewed journals, and, to our knowledge, no such investigations have been performed. Therefore, we conclude that our proposal is strictly unique in this endeavor.

In addition, the previous studies were partly unsuccessful in establishing even statistical correlations. The investigation by Saunders and Deane [34], which is the most recent, interrogated differences in codon usage within helices, strands, and coils by applying both Mantel–Haenszel statistics and a χ^2 -test. They found few significant differences between the three secondary structural types, but did find a significant decrease in codon usage bias near the transitions between secondary structural elements. They also investigated the hypothesis that slow codons are frequent around domain boundaries; the results in this case were negative. However, these studies relied on simple statistical techniques that lack the power necessary to find statistical associations for weaker signals.

Immense progress has been made in both the quality and quantity of information contained within bioinformatics databases within the past few years, making analyses of codon usage across thousands or more proteins possible. Examining larger sample sizes enables the study of family or activity based codon bias as well as allowing for a more robust statistical analysis when compared with previous studies where fewer than 300 proteins were examined.

Approach

Aim 1

We primarily aim to investigate the relationship between secondary structural elements (SSEs) and slow codons, with the aim of finding biases towards slow codon enrichment in certain elements. The null hypothesis for this situation is that there is no meaningful relationship between frequency of slow codons and domain features. However, we hypothesize that such a relationship does exist, as previous research indicates the introduction or removal of slow codons has a dramatic effect on protein expression.

Hypotheses

We are currently interested in several hypotheses and open questions:

1. **Secondary structure** Are particular secondary structural elements (SSEs) enriched for slow codons? Are the transitions between SSEs, in agreement with Saunders and Deane [34], enriched for slow codons?

To investigate this hypothesis, we will use Define Secondary Structure of Proteins (DSSP) [.] to identify putative SSEs. Although DSSP is currently outperformed in accuracy by sophisticated machine learning methods, the use of machine learning algorithms such as PROTEUS presents significant pitfalls in performing statistical analysis because decisions by the classifier depend on the training data, and individual classification decisions are unstable subject to arbitrary bounds. Because our methodology depends so heavily on such analysis to improve overall sensitivity, we consider this loss of statistical rigor unacceptable.

We will first attempt to verify the results of Saunders and Deane [34] by duplicating their approach. Specifically, we will calculate the distribution of codon usage bias within helices, strands, and coils. We will apply this method to 5–10 species-dependent data sets, including eukaryotes, invertebrates, and *Homo sapiens*. We will then apply a Mantel–Haenszel and a χ^2 -test to determine significance between the categories. Secondly, we select 2–5 large data sets (at least 2,000 genes each), and expand the categories to also include, at a minimum, 3_{10} -helices, β -helices, and π -helices. We will also distinguish between β -sheets and isolated β -strands. We will then determine significance using the same statistical techniques.

To address the second part of this question, we will calculate codon usage bias around the transitions between SSEs, using windows of 0, 1, 2, 4, and 6 residues.

Preliminary data: In addition to the work by Gu et al. [16], Oresic et al. [31], Saunders and Deane [34], Thanaraj and Argos [41], we investigated the hypothesis that strands are enriched for slow codons on a data set of 1,000 **TODO: fix this description of the data** genes, much larger than the data sets of any previous works. The results showed that **TODO: something**.

2. **Domain boundaries** Are domain boundaries enriched for slow codons? To investigate this question, [2][29]
3. **Structural complexity and knotting** [24, 43]
4. **Solvent accessibility and oligomeric assembly**
[6]

Determination of causation

Ultimately, we are interested in causation related to codon usage bias. The analyses described above are independently useful, both for novel findings and verification of previous results. While we argue that such

results are interesting, biologically relevant, and warrant further investigation, they may indicate only indirect correlations. It is entirely possible for one variable to account for another in whole or in part; for example, domain boundaries may be enriched for slow codons only because they contain more strands than helices, or that solvent accessibility around domain boundaries may be the more fundamental explanation.

The determination of causal relationships from correlation data consists of two tasks:

1. The removal of indirect correlations, and
2. The assignment of a direction to each relationship

To formulate these two tasks concretely, we will model the entire problem as a Markov Random Field.

[37][38]

Mathematical model

TODO: Douglas add more background here? Bias towards fast or slow codons in mRNA sequence will be established using the codon adaptation index (CAI) described by Sharp and Li [35]. Codons are assigned weights depending on the frequency of their corresponding tRNA in the appropriate organism:

$$w_{ij} = \frac{X_{ij}}{X_{i*}} \quad (1)$$

The codon usage bias in a particular region is then the geometric mean of the codons that comprise it:

$$CAI = (\prod_{k=1}^L w_k)^{\frac{1}{L}} \quad (2)$$

$$= \exp\left(\frac{1}{L} \sum_{k=1}^L \log w_k\right) \quad (3)$$

Benchmarking

Many proteins will be examined over a number of species. While our preliminary analysis has been performed in approximately 500 yeast proteins, we hope to analyze over 10,000 human proteins in hopes of maximizing sample size.

Data sets

Domain information will be obtained from the Structural Classification of Proteins (SCOP) database [29]. Coding mRNA sequences for the protein will be obtained from the NCBI Reference Sequence Database (RefSeq) [33]. Sequences will be correlated to structures by means of Structure Integration with Function, Taxonomy, and Sequence (SIFTS) database [42].

Analysis of accuracy

Codon usage bias will be examined on domain boundary regions and non-boundary regions, and the overall CAI value will be compared across these regions. If a significant difference in CAI across secondary structure features is found, the hypothesis will be supported. [25]

Aim 2

We also aim to examine existing proteins that have been demonstrated to be affected by the removal of slow codons, and characterize in what circumstances a protein could be reasonably assumed to require slow codons for normal function.

We will run our analysis on species-independent protein families that contain at least one protein shown experimentally to require slow codons. We will complement our structural analysis with a phylogenetic examination of conserved regions to see if the bias towards slow codons is strictly evolutionarily enforced or if it is an isolated development in some species.

Aim 3

Folding study

As an auxillary study, we will investigate the effects of synonymous mutations in detail for select cases using MD-based folding software. Although de novo folding is still largely unsolved, and de novo folding algorithms are still in their infancy, such an investigation may still reveal insight for some cases that is unavailable through other means. [45]

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