

Appendix: Codes and extra material

1 Credit Card Data

1.1 data cleaning and extraction of small subset

```
set.seed(123)

file_path_credit <- "default-credit-card.csv"
file_path_credit_names <- "names-default-credit-card.csv"
data_credit <- read.csv(file_path_credit)
data_credit_names <- read.csv(file_path_credit_names)

credit_short_colnames <- colnames(data_credit)
credit_long_colnames <- colnames(data_credit_names)
colnames(data_credit) <- credit_long_colnames

data_credit_unclean=data_credit

data_credit <- data_credit %>%
  filter(PAY_0 != -2) %>% filter(PAY_0 != 0) %>%
  filter(PAY_2 != -2) %>% filter(PAY_2 != 0) %>%
  filter(PAY_3 != -2) %>% filter(PAY_3 != 0) %>%
  filter(PAY_4 != -2) %>% filter(PAY_4 != 0) %>%
  filter(PAY_5 != -2) %>% filter(PAY_5 != 0) %>%
  filter(PAY_6 != -2) %>% filter(PAY_6 != 0) %>%
  filter(EDUCATION > 0) %>% filter(EDUCATION < 4)

data_credit_edu <-function(edu,def){data_credit %>% filter(EDUCATION == edu) %>% filter(default == c

combined_df <- rbind(data_credit_edu(1,0), data_credit_edu(1,1),
                     data_credit_edu(2,0), data_credit_edu(2,1),
                     data_credit_edu(3,0), data_credit_edu(3,1))

write.csv(combined_df, file = "credit_data_education_short.csv")

data_credit_full = data_credit
data_credit = combined_df
```

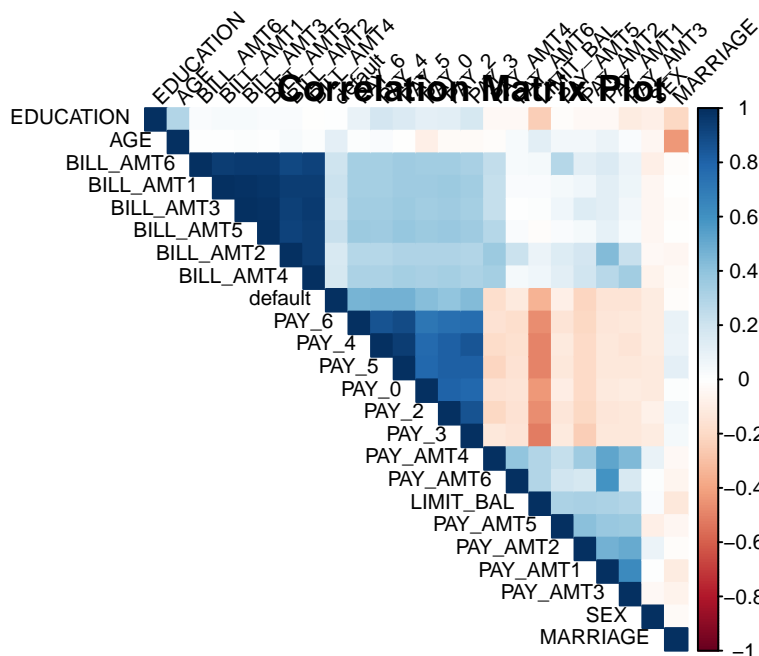
1.2 correlations in subset

```
colnames(data_credit) <- credit_long_colnames
df = data_credit
correlations <- sapply(df[, -which(names(df) == "default")], function(x) biserial.cor(x, df$default))
list_cor = sort(round(abs(correlations),3))
correlations_df <- data.frame(
  Variable = names(list_cor),
  Correlation = as.vector(list_cor)
)
```

```
#correlations_df %>% filter(Correlation > 0)
```

```
# Calculate the correlation matrix
cor_matrix <- cor(data_credit[, -1])
```

```
# Create a correlation matrix plot
corrplot(cor_matrix, method = "color", type = "upper", order = "hclust", tl.col = "black", tl.srt = 45,
title("Correlation Matrix Plot"))
```



```
correlations <- cor(data_credit[, 'default'], data_credit[, -which(names(data_credit) == 'default')])
round(correlations,3)
```

```

ID LIMIT_BAL SEX EDUCATION MARRIAGE AGE PAY_0 PAY_2 PAY_3 PAY_4
[1,] -0.035 -0.344 -0.104 0 -0.015 0.12 0.428 0.393 0.432 0.47
PAY_5 PAY_6 BILL_AMT1 BILL_AMT2 BILL_AMT3 BILL_AMT4 BILL_AMT5 BILL_AMT6
[1,] 0.475 0.466 0.213 0.164 0.204 0.18 0.223 0.204
PAY_AMT1 PAY_AMT2 PAY_AMT3 PAY_AMT4 PAY_AMT5 PAY_AMT6
[1,] -0.14 -0.218 -0.147 -0.179 -0.086 -0.118
```

1.3 Stan model PAY_0 + PAY_2

```
#Time 1.36 mins
start.time <- Sys.time()
fit1 <- brm(default ~ PAY_0 + PAY_2,
  data = data_credit,
  refresh = 0,
  prior=c(
    prior(normal(0,100), class="Intercept")
    , prior(normal(0, 100), class = b)
  ),
  family = bernoulli(),
  file = "pooled",
  backend = "cmdstanr",
  seed = 123
)
end.time <- Sys.time()
time.taken <- round(end.time - start.time,2)
time.taken
```

Time difference of 0.01 secs

```
predictions <- predict(fit1, newdata = data_credit, type = "response")
binary_predictions <- ifelse(predictions[,1] > 0.5, 1, 0)
conf_matrix <- confusionMatrix(factor(binary_predictions), factor(data_credit$default))
conf_matrix
```

Confusion Matrix and Statistics

	Reference	
Prediction	0	1
0	94	36
1	26	84

Accuracy : 0.7417
95% CI : (0.6814, 0.7958)
No Information Rate : 0.5
P-Value [Acc > NIR] : 1.785e-14

Kappa : 0.4833

Mcnemar's Test P-Value : 0.253

Sensitivity : 0.7833
Specificity : 0.7000
Pos Pred Value : 0.7231
Neg Pred Value : 0.7636
Prevalence : 0.5000
Detection Rate : 0.3917
Detection Prevalence : 0.5417
Balanced Accuracy : 0.7417

'Positive' Class : 0

```
start.time <- Sys.time()
batch_size = 1000;
start = 1;
binary_predictions_unclean = c();
predictions_batch_list = c();
for (i in 1:30) {
  if (i%%10 == 0) {
    print(i);
  }
  end = start + batch_size - 1;
  predictions_batch <- predict(fit1, newdata = data_credit_unclean[start:end,],
                             type = "response", allow_new_levels = TRUE)
  predictions_batch_list = c(predictions_batch_list, predictions_batch)
  start = end + 1
  binary_predictions_batch <- ifelse(predictions_batch[,1] > 0.5, 1, 0)
  binary_predictions_unclean = c(binary_predictions_unclean, binary_predictions_batch);
}
```

```
[1] 10
[1] 20
[1] 30
```

```
end.time <- Sys.time()
time.taken <- round(end.time - start.time,2)
time.taken
```

Time difference of 45.19 secs

```
conf_matrix <- confusionMatrix(factor(binary_predictions_unclean), reference = factor(data_credit_un
conf_matrix
```

Confusion Matrix and Statistics

	Reference	
Prediction	0	1
0	20637	3452
1	2727	3184

Accuracy : 0.794
95% CI : (0.7894, 0.7986)
No Information Rate : 0.7788
P-Value [Acc > NIR] : 7.634e-11

Kappa : 0.3779

McNemar's Test P-Value : < 2.2e-16

Sensitivity : 0.8833
 Specificity : 0.4798
 Pos Pred Value : 0.8567
 Neg Pred Value : 0.5387
 Prevalence : 0.7788
 Detection Rate : 0.6879
 Detection Prevalence : 0.8030
 Balanced Accuracy : 0.6815

'Positive' Class : 0

1.4 Stan model EDUCATION Hierarchical

```

start.time <- Sys.time()
fit2 <- brm(default ~ PAY_0 + PAY_2 + (1| EDUCATION), # short dataset
  data = data_credit,
  refresh = 0,
  prior=c(
    prior(normal(0,100), class="Intercept"),
    prior(normal(0,100), class="b"),
    prior(exponential(.02), class="sd")
  ),
  family = bernoulli(),
  file = "model2_education_small_data_simple",
  backend = "cmdstanr",
  iter = 5000,
  warmup = 2500,
  seed = 123
)
end.time <- Sys.time()
time.taken <- round(end.time - start.time,2)
time.taken

```

Time difference of 0.02 secs

```

predictions <- predict(fit2, newdata = data_credit, type = "response", allow_new_levels = TRUE)
binary_predictions <- ifelse(predictions[,1] > 0.5, 1, 0)
conf_matrix <- confusionMatrix(factor(binary_predictions), factor(data_credit$default))
conf_matrix

```

Confusion Matrix and Statistics

	Reference	
Prediction	0	1
0	95	37
1	25	83

Accuracy : 0.7417
 95% CI : (0.6814, 0.7958)
 No Information Rate : 0.5

P-Value [Acc > NIR] : 1.785e-14

Kappa : 0.4833

Mcnemar's Test P-Value : 0.1624

Sensitivity : 0.7917

Specificity : 0.6917

Pos Pred Value : 0.7197

Neg Pred Value : 0.7685

Prevalence : 0.5000

Detection Rate : 0.3958

Detection Prevalence : 0.5500

Balanced Accuracy : 0.7417

'Positive' Class : 0

```
start.time <- Sys.time()
batch_size = 1000;
start = 1;
binary_predictions_unclean = c();
predictions_batch_list = c();
for (i in 1:30) {
  if (i%%10 == 0) {
    print(i);
  }
  end = start + batch_size - 1;
  predictions_batch <- predict(fit2, newdata = data_credit_unclean[start:end,],
                              type = "response", allow_new_levels = TRUE)
  predictions_batch_list = c(predictions_batch_list, predictions_batch)
  start = end + 1
  binary_predictions_batch <- ifelse(predictions_batch[,1] > 0.5, 1, 0)
  binary_predictions_unclean = c(binary_predictions_unclean, binary_predictions_batch);
}
```

[1] 10

[1] 20

[1] 30

```
end.time <- Sys.time()
time.taken <- round(end.time - start.time,2)
time.taken
```

Time difference of 1.28 mins

```
confusionMatrix(factor(binary_predictions_unclean), reference = factor(data_credit_unclean$default))
```

Confusion Matrix and Statistics

	Reference	
Prediction	0	1
0	20409	3372
1	2955	3264

Accuracy : 0.7891
 95% CI : (0.7844, 0.7937)
 No Information Rate : 0.7788
 P-Value [Acc > NIR] : 8.003e-06

Kappa : 0.3738

Mcnemar's Test P-Value : 1.696e-07

Sensitivity : 0.8735
 Specificity : 0.4919
 Pos Pred Value : 0.8582
 Neg Pred Value : 0.5248
 Prevalence : 0.7788
 Detection Rate : 0.6803
 Detection Prevalence : 0.7927
 Balanced Accuracy : 0.6827

'Positive' Class : 0

1.5 Model comparison (e.g. with LOO-CV).

```

l1 = loo(fit1)
l2 = loo(fit2)
loo_compare(l1,l2)

```

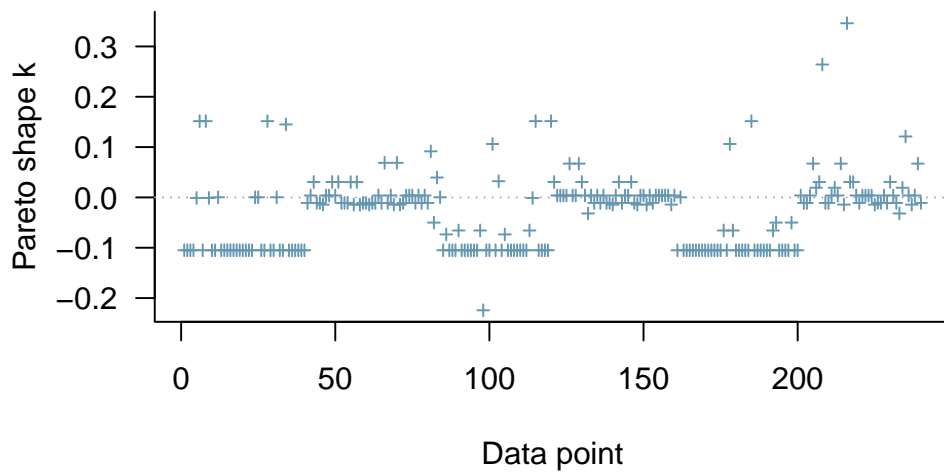
	elpd_diff	se_diff
fit1	0.0	0.0
fit2	-0.4	1.0

```

plot(l1, label_points = TRUE, main = 'PSIS diagnostic plot for pooled model')

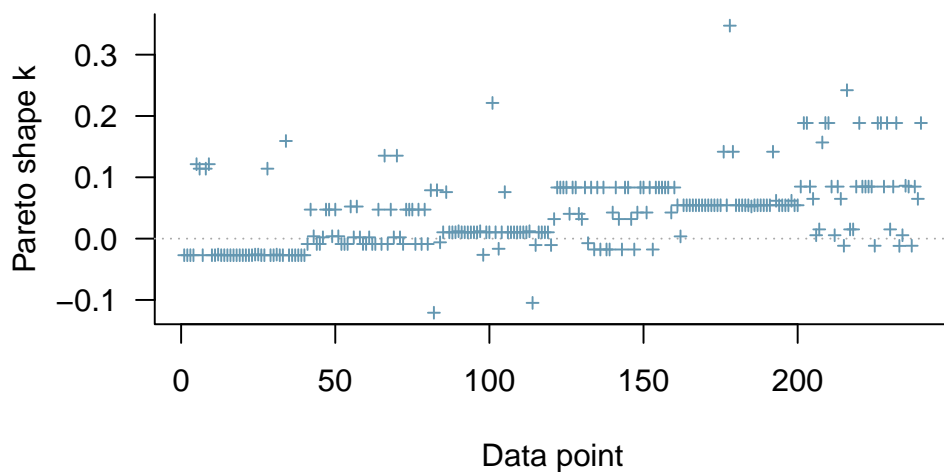
```

PSIS diagnostic plot for pooled model



```
plot(l2, label_points = TRUE, main = 'PSIS diagnostic plot for hierarchical model')
```

PSIS diagnostic plot for hierarchical model



1.6 Convergence diagnostic

```
summarize_draws(fit1)
```

A tibble: 5 x 10

	variable	mean	median	sd	mad	q5	q95	rhat	ess_bulk
	<chr>	<dbl>	<dbl>	<dbl>	<dbl>	<dbl>	<dbl>	<dbl>	<dbl>
1	b_Intercept	-0.213	-0.212	0.150	0.151	-4.58e-1	2.94e-2	1.00	2592.
2	b_PAY_0	0.521	0.518	0.154	0.157	2.79e-1	7.81e-1	1.00	2058.
3	b_PAY_2	0.165	0.165	0.131	0.131	-5.02e-2	3.85e-1	1.00	2075.

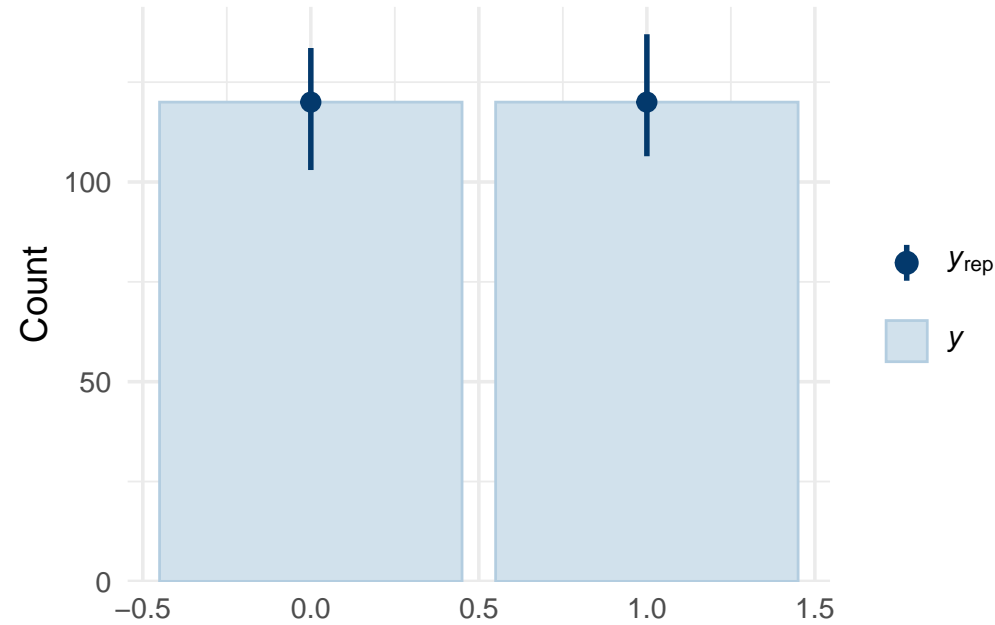

```
4 lprior      -16.6    -16.6    0.0000398 0      -1.66e+1 -1.66e+1  1.00    2418.
5 lp__        -158.    -158.     1.21      0.981 -1.61e+2 -1.57e+2  1.00    1876.
# i 1 more variable: ess_tail <dbl>
```

```
summarize_draws(fit2)
```

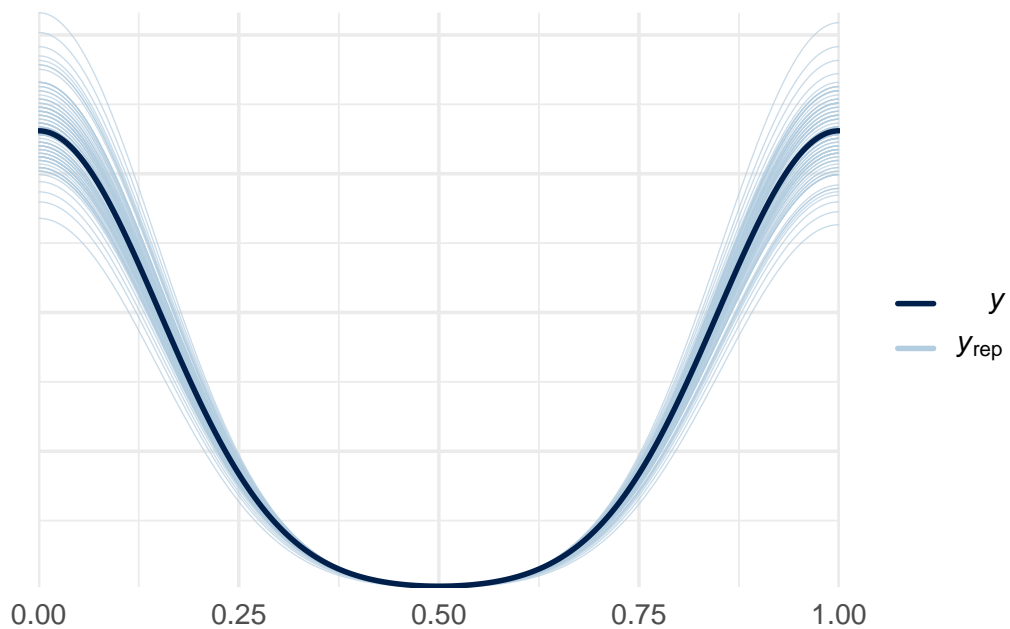
```
# A tibble: 9 x 10
  variable      mean  median    sd    mad    q5    q95  rhat ess_bulk
  <chr>        <dbl>   <dbl>  <dbl>  <dbl>  <dbl>  <dbl> <dbl>   <dbl>
1 b_Intercept -2.49e-1 -2.25e-1 0.364  0.269  -9.08e-1  0.293  1.00    1622.
2 b_PAY_0      5.51e-1  5.48e-1 0.157  0.156   2.95e-1  0.811  1.00    3162.
3 b_PAY_2      1.64e-1  1.65e-1 0.130  0.130  -4.99e-2  0.380  1.00    3102.
4 sd_EDUCATIO~ 5.73e-1  3.90e-1 0.605  0.368   4.01e-2  1.69   1.01    1170.
5 r_EDUCATION~ 2.08e-1  1.25e-1 0.392  0.258  -3.01e-1  0.982  1.01    1601.
6 r_EDUCATION~ -1.61e-1 -1.16e-1 0.384  0.256  -8.30e-1  0.407  1.00    1859.
7 r_EDUCATION~ 2.89e-2  5.26e-3 0.377  0.235  -5.62e-1  0.699  1.00    1774.
8 lprior      -2.05e+1 -2.05e+1 0.0121 0.00741 -2.05e+1 -20.5   1.01    1167.
9 lp__        -1.67e+2 -1.66e+2 2.33    2.26   -1.71e+2 -163.   1.01    1550.
# i 1 more variable: ess_tail <dbl>
```

#PPC

```
pp_check(fit1, type = "bars", ndraws = 150)
```



```
pp_check(fit1, type = "dens_overlay", ndraws = 100)
```



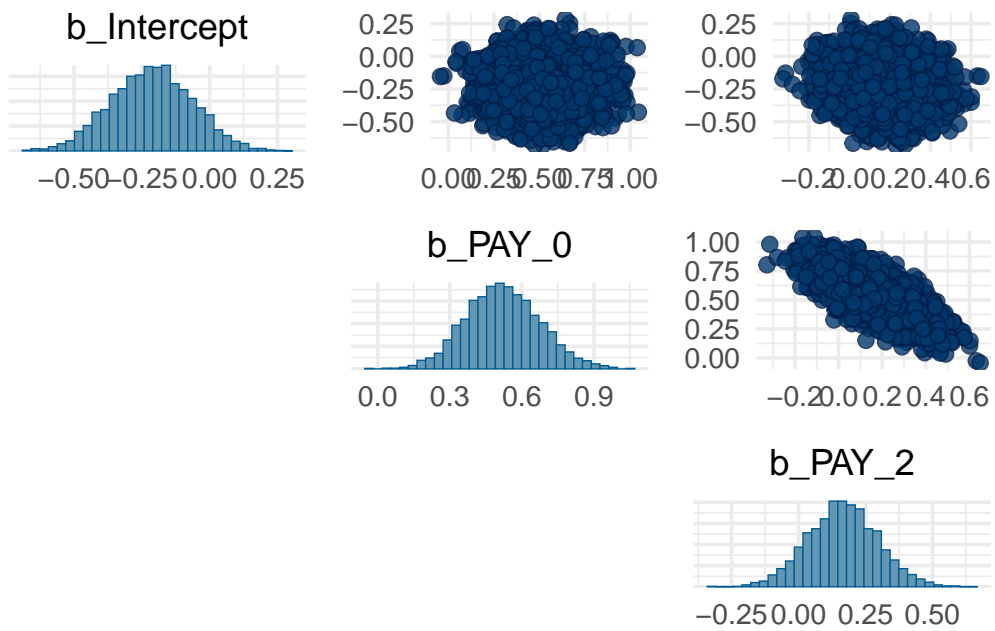
```
posterior_samples <- posterior_samples(fit1)
```

Warning: Method 'posterior_samples' is deprecated. Please see ?as_draws for recommended alternatives.

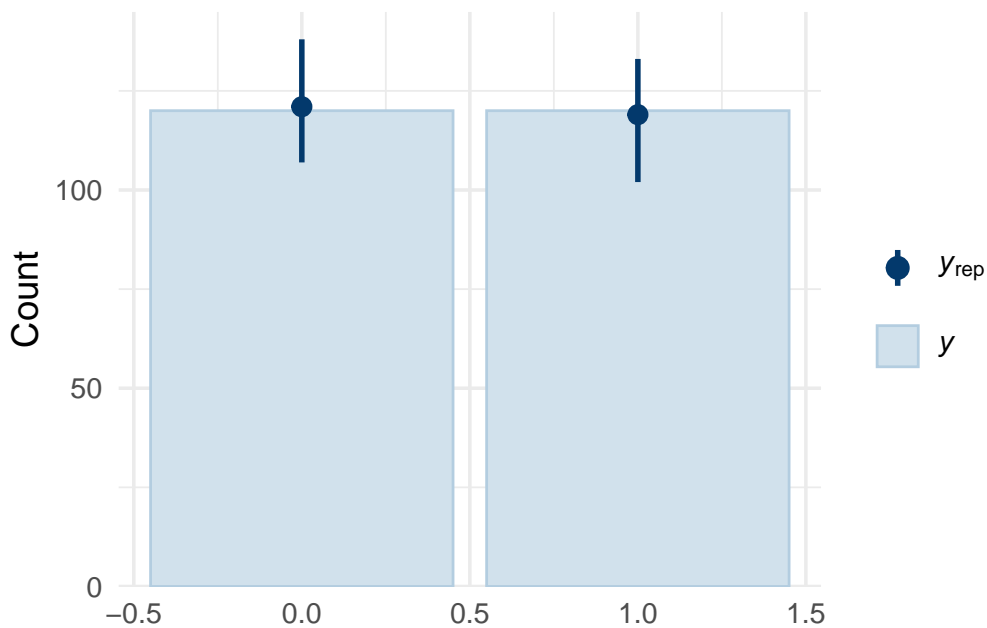
```
# Use mcmc_pairs to create a scatterplot matrix for selected parameters
# Select a subset of parameters if you have many;
selected_params <- posterior_samples[, c("b_Intercept", "b_PAY_0", "b_PAY_2")]
np <- nuts_params(fit1)

# Create the pairs plot
mcmc_pairs(
  selected_params,
  np = np,
  pars = c("b_Intercept", "b_PAY_0", "b_PAY_2")
)
```

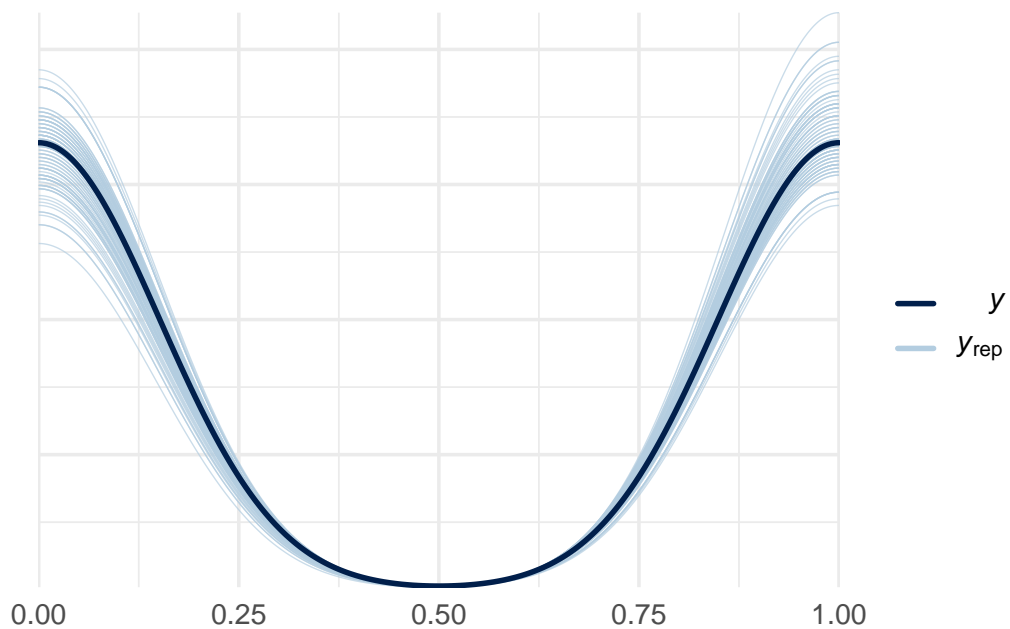
Warning: Only one chain in 'x'. This plot is more useful with multiple chains.



```
# Bar plot of the observed outcomes compared to the replicated datasets
pp_check(fit2, type = "bars", ndraws = 100)
```



```
# Density plot of the predicted probabilities
pp_check(fit2, type = "dens_overlay", ndraws = 100)
```



```
posterior_samples <- posterior_samples(fit2)
```

Warning: Method 'posterior_samples' is deprecated. Please see ?as_draws for recommended alternatives.

```
# Use mcmc_pairs to create a scatterplot matrix for selected parameters
# Select a subset of parameters if you have many;
selected_params <- posterior_samples[, c("b_Intercept", "b_PAY_0", "b_PAY_2")]
np <- nuts_params(fit2)

# Create the pairs plot
mcmc_pairs(
  selected_params,
  np = np,
  pars = c("b_Intercept", "b_PAY_0", "b_PAY_2")
)
```

Warning: Only one chain in 'x'. This plot is more useful with multiple chains.

