

# Solving SATISFIABILITY with Molecular Algorithms

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## **Abstract**

Molecular computation uses techniques from molecular biology and combinatorial chemistry to perform generalized computations. We explore, via simulation, three distinct molecular algorithms for solving SATISFIABILITY. The simulation measures the number of molecular operations for solving SATISFIABILITY. The test input consists of a set of random 3-SAT instances distributed over a range of clause-variable ratios ( $\alpha = [0.2, 14.0]$ ).

# Contents

<b>1</b>	<b>Introduction</b>	<b>1</b>
1.1	Introduction to molecular computation . . . . .	1
1.2	Simulation of molecular SATISFIABILITY solvers . . . . .	5
1.3	Report Overview . . . . .	6
<b>2</b>	<b>Background</b>	<b>8</b>
2.1	On nanotechnology and construction of molecules . . . . .	8
2.2	On microbiology and computation . . . . .	9
2.3	Adleman’s molecular toolbox for solving HAMITONIAN PATH . . . . .	11
2.3.1	Additional molecular operators . . . . .	14
2.4	Definition of SATISFIABILITY . . . . .	14
2.5	Evaluating SAT Solvers . . . . .	16
2.5.1	Input and output . . . . .	16
2.5.2	Metrics for classifying SATISFIABILITY . . . . .	17
2.5.3	SATISFIABILITY instances . . . . .	18
<b>3</b>	<b>Existing molecular algorithms for SATISFIABILITY</b>	<b>19</b>
3.1	Lipton’s algorithm for SATISFIABILITY . . . . .	19
3.1.1	Description of Lipton’s algorithm . . . . .	20

3.1.2	Pseudocode for Lipton’s algorithm . . . . .	21
3.2	Ogihara and Ray’s algorithm for SATISFIABILITY . . . . .	22
3.2.1	Description of Ogihara and Ray’s algorithm . . . . .	22
3.2.2	Pseudocode for Ogihara and Ray’s algorithm . . . . .	25
3.3	Implementations of molecular SATISFIABILITY solvers . . . . .	26
3.3.1	Physical implementations . . . . .	26
3.3.2	Simulations . . . . .	26
<b>4</b>	<b>A new molecular algorithm for SATISFIABILITY</b>	<b>27</b>
4.1	Distribution algorithm for SATISFIABILITY . . . . .	27
4.1.1	Description of the Distribution algorithm . . . . .	27
4.1.2	Pseudocode for Distribution algorithm . . . . .	29
<b>5</b>	<b>Molecular Simulation: A system for molecular computation</b>	<b>31</b>
5.1	Overview . . . . .	31
5.2	Download . . . . .	32
5.3	Requirements . . . . .	32
5.3.1	Hardware requirements . . . . .	32
5.3.2	Software requirements . . . . .	32
5.4	Documentation . . . . .	33
5.5	Tools . . . . .	33
5.5.1	Perl utilities . . . . .	33
5.5.2	Data Visualization . . . . .	33
5.6	Input . . . . .	35
5.7	Output . . . . .	35
5.8	Execution . . . . .	37

<b>6</b>	<b>Experimental Setup</b>	<b>39</b>
6.1	Setup . . . . .	39
6.2	Create dataset . . . . .	40
6.3	Import dataset . . . . .	40
6.4	Configure test . . . . .	41
6.5	Execution and collection of data . . . . .	42
6.5.1	Execution output . . . . .	42
<b>7</b>	<b>Results</b>	<b>43</b>
7.1	Algorithm metric comparison . . . . .	43
<b>8</b>	<b>Conclusions</b>	<b>52</b>
8.1	Contributions . . . . .	52
8.2	Future work . . . . .	52
	<b>Bibliography</b>	<b>54</b>
<b>A</b>	<b>Source</b>	<b>58</b>
A.1	Contributed . . . . .	58
A.2	External . . . . .	59
<b>B</b>	<b>Molecular algorithm trace</b>	<b>60</b>
B.1	Example SATISFIABILITY instance . . . . .	60
B.2	Lipton’s Algorithm . . . . .	60
B.3	Ogihara and Ray’s Algorithm . . . . .	63
B.4	Distribution Algorithm . . . . .	66

# Chapter 1

## Introduction

Molecular computing uses parallel interactions between genetic molecules, such as DNA or RNA, to perform computational tasks. We provide an experimental system for simulating three molecular algorithms. In this chapter we discuss the process of solving a combinatorial problem with both standard or molecular models of computation. This discussion includes an introduction to simulating molecular algorithms. We conclude the chapter with the contents of this report.

### 1.1 Introduction to molecular computation

NP-complete problems, such as SATISFIABILITY, may be verified in polynomial time with the aid of a short proof called a *witness*; NP-complete problems may be solved by checking the state for all possible *witness candidates*. In a standard computing environment, brute force search can check all  $2^n$  witness candidates in exponential time.

Molecular computing requires exponential space to represent all witness candidates. This combinatorial space of witness candidates can be filtered in polynomial time with parallel molecular operators.



Conjunctive normal form (CNF) can be used to represent SATISFIABILITY instances as a structured input format for Boolean formulas. A CNF expression  $\phi$  consists of a conjunctive set of  $m$  Boolean disjunctive clauses. The expression  $\phi$  consists of  $n$  independent Boolean variables. We have, e.g.,

$$\phi = C_1 \wedge C_2 \wedge \cdots \wedge C_m,$$

where each clause  $C_i$  contains  $k$  disjunctive Boolean variables

$$C_i = (v_1 \vee v_2 \vee \cdots \vee v_k).$$

A potentially satisfying witness for a SATISFIABILITY instance is a Boolean assignment to the variables that make the formula true. Such a witness can be represented as an  $n$ -bit Boolean vector. Validating a witness candidate for a SATISFIABILITY instance may be verified in polynomial time with  $\text{CHECKSAT}(\phi, B)$  (Algorithm 1.1.1 below).  $\text{CHECKSAT}(\phi, B)$  iterates over each of the clauses  $C$  in the expression  $\phi$ . The *test\_clause* variable gets set to *False*, assuming that the clause cannot be satisfied. If the clause can be satisfied with the input configuration  $B$ , then the algorithm continues. If each of the  $m$  clauses can be

satisfied, then  $\text{CHECKSAT}(\phi, B)$  returns *True*; otherwise the algorithm returns *False*.

**Algorithm 1.1.1:**  $\text{CHECKSAT}(\phi, B)$

```

for each clause  $C$  in  $\phi$ 
     $test\_clause \leftarrow False$ 
    for each variable  $v$  in  $C$ 
        do  $\left\{ \begin{array}{l} \text{if } v \in B \\ \text{then } test\_clause \leftarrow True \end{array} \right.$ 
    if  $test\_clause = False$ 
        then return (False)
return (True)

```

**Algorithm 1.1.2:**  $\text{BRUTESAT}(\phi)$

$n$  number of variables in  $\phi$

$t$  is bit vector representing a witness candidate

```

for  $t \leftarrow 0$  to  $2^n - 1$ 
    do  $\left\{ \begin{array}{l} \text{if } \text{CHECKSAT}(\phi, t) \\ \text{then return } (\text{SATISFIABLE}) \end{array} \right.$ 
return (UNSATISFIABLE)

```

$\text{CHECKSAT}(\phi, B)$  may be applied as a subroutine in a brute force SATISFIABILITY solver. Algorithm 1.1.2 provides pseudocode for a brute force SATISFIABILITY solver  $\text{BRUTESAT}(\phi)$ . The algorithm  $\text{BRUTESAT}(\phi)$  tests a maximum of  $2^n$  Boolean configurations, using the  $\text{CHECKSAT}(\phi, B)$  algorithm. If the test configuration  $t$  satisfies the input instance  $\phi$ , then the algorithm returns SATISFIABLE; otherwise the algorithm returns UNSATISFIABLE.

In this project, we consider molecular algorithms to solve SATISFIABILITY. Molecular algorithms permit many combinations to occur in parallel [1, 15]. This permits molecular operations, such as *append* or *extract*, to perform in parallel on all of the string contents of a test tube [1, 15, 12]. In Chapter 3, we explore techniques from combinatorial chemistry to generate combinatorial sets [15, 9, 12]. The function `COMBINATORIALGENERATE( $n$ )`, which we introduce in Chapter 3, constructs an exponential number of configurations in linear time.

Let us consider Algorithm 1.1.3 as a simplified version of Lipton’s algorithm [15, 12].

**Algorithm 1.1.3:** `EXTRACTSAT( $\phi$ )`

$n$  number of variables in  $\phi$

$T \leftarrow \text{COMBINATORIALGENERATE}(n)$

**for each** clause  $C$  in  $\phi$

**do**  $\left\{ \begin{array}{l} T_C \leftarrow \emptyset \\ \textbf{for each} \text{ variable } v \text{ in } C \\ \textbf{do} \left\{ \begin{array}{l} T_T \leftarrow \text{extract}(T, v) \\ T_C \leftarrow \text{mix}(T_C, T_T) \end{array} \right. \\ T \leftarrow T_C \end{array} \right.$

**if**  $T = \emptyset$

**then return** (UNSATISFIABLE)

**return** (SATISFIABLE)

`EXTRACTSAT( $\phi$ )` collects configurations satisfying Boolean variables from each clause in  $\phi$ . Initially, `EXTRACTSAT( $\phi$ )` constructs a combinatorial space  $T$  with the subroutine `COMBINATORIALGENERATE( $n$ )`. The initial space  $T$  contains configurations representing all potential witness candidates for  $\phi$ . The space  $T$  gets filtered for each clause to only

those configurations that satisfy any of the Boolean variables contained within a clause. These potential solutions are incrementally mixed into the tube  $T_C$  for each clause. Once extracting the contents for the current clause  $C$  the tube  $T_C$  of partial assignments gets stored as  $T$ . The set  $T$  now contains all witnesses that can be satisfied with the previous clauses. If  $T$  contains no string configurations after filtering variables for each clause, then  $\phi$  is **UNSATISFIABLE**; otherwise  $\phi$  is **SATISFIABLE**, and  $T$  contains configurations for all satisfying witnesses.

We consider Lipton’s algorithm in detail in Chapter 3. However, the  $\text{EXTRACTSAT}(\phi)$  function provides an introductory view of a molecular algorithm.  $\text{EXTRACTSAT}(\phi)$  differs from  $\text{BRUTESAT}(\phi)$  in the method of determining the state for a **SATISFIABILITY** instance. With the  $\text{BRUTESAT}(\phi)$  algorithm, exponential configurations get generated in exponential time; on the other hand, with  $\text{EXTRACTSAT}(\phi)$  exponential configurations get filtered from exponential space.

## 1.2 Simulation of molecular **SATISFIABILITY** solvers

We consider three molecular algorithms for solving **SATISFIABILITY**: Lipton’s algorithm [15], Ogihara and Ray’s algorithm [18, 19], and a new algorithm, introduced here, that we call the ‘Distribution’ algorithm. Lipton’s algorithm begins with a combinatorial space of all  $n$ -bit witness candidates and filters the combinatorial space so that only those that satisfy the input formula remain. Ogihara and Ray’s algorithm constructs a space of witness candidates with a heuristic search. The Distribution algorithm expands a set of witnesses with non-conflicting variables from each clause. Chapters 3 and 4 discuss the implementation of these algorithms.

This project introduces a system for simulating three molecular algorithms for solving **SATISFIABILITY**. The system provides standard operations for molecular computing that

we introduce in Chapter 2. It also records runtime metrics, including counts of molecular operators, solution memory footprints, and execution times. These metrics let us analyze algorithmic performance of each molecular algorithm.

Molecular Simulation, the simulation system introduced in this project, automates execution of DIMACS CNF instances. Simulation of each of the algorithms measures metrics for a set of randomly generated 3-SAT expressions. The 3-SAT instances span discrete clause-variable ratios from 0.2 to 14.0 in increments of 0.2, creating a sweep of SATISFIABILITY instances. This experimental setup generates SATISFIABILITY problem instances with both SATISFIABLE and UNSATISFIABLE configurations.

## 1.3 Report Overview

In the following chapters, we describe molecular algorithms for solving SATISFIABILITY. We begin Chapter 2 with an introduction to gene sequencing technologies and molecular biology. We define molecular operations for operating on DNA or RNA. Next, we introduce SATISFIABILITY as a language and as a Boolean circuit.

Chapters 3 and 4 introduce each of the three molecular algorithms for solving SATISFIABILITY. In Chapter 3, we discuss Lipton’s [15, 12] and Ogihara and Ray’s [18, 19, 25] algorithms for SATISFIABILITY. The chapter concludes with a discussion of existing simulation frameworks and physical implementations of these molecular algorithms. Chapter 4 introduces the Distribution algorithm.

Chapters 5 and 6 discuss the project implementation. In Chapter 5, we introduce our software, Molecular Simulation, for simulating molecular algorithms. Chapter 6 describes the experimental workflow for importing SATISFIABILITY instances for each of the three molecular algorithms we study.

Chapter 7 provides a discussion of algorithm performance based test results. Chapter

8 concludes with a summary of contributions of this project and future directions for molecular computation.

## Chapter 2

# Background

This chapter provides a background on molecular computation techniques. We begin with an introduction to nanotechnology and then provide an example of how information is encoded with molecular matter. Following this example, we introduce Adleman’s molecular operators for solving an instance of HAMILTONIAN PATH. The operators provide an instruction set for molecular computation, and provide the primitives for constructing molecular algorithms.

In the second half of this chapter, we provide an introduction to SATISFIABILITY. We define SATISFIABILITY as a circuit. We then view SATISFIABILITY as a language. We also discuss practical matters related to efficiently evaluating SATISFIABILITY, such as how to encode input and output, and how to classify instances of SATISFIABILITY in the tests that we perform.

### 2.1 On nanotechnology and construction of molecules

Richard Feynman founded the field of nanotechnology in his 1959 talk ‘There’s Plenty of Room at the Bottom’ [6]. Examples of applied nanotechnology include the manufactur-

ing of graphene [23] and DNA nanopores [16]. Graphene consists of a planer arrangement of carbon atoms that provides desirable physical and electrical properties [23]. DNA nanopores use graphene to create a physical channel for reading genetic sequences [10]. Gene sequencing technologies provide an example of applied nanotechnology [10, 14, 20].

Smaller and cost-effective DNA sequencers provide the ability to read the contents of a gene. Benchtop sequencers [14, 20] allow doctors to treat patients at the genome level from their office. Life Technologies and Oxford Nanopore offer gene sequencers based on solid-state semiconductor technology [14, 20].

## 2.2 On microbiology and computation

Microbiology studies the interactions among organic molecules. In this project, we explore the use of applied genetics as a means for generalized computation. Molecular computation encodes data as sequences of DNA or RNA.

Arbitrary encodings that represent mappings from variables to physical oligonucleotides may have undesirable structure and functionality. Conventional techniques for DNA computing employ variable mappings from a library of oligonucleotides.

An *oligonucleotide* is a short string of genetic information. There are several configurations for DNA and RNA; these include +RNA, −RNA, +DNA, −DNA, ±RNA, ±DNA, and +mRNA [2]. The polarity of DNA denotes the direction of the genetic information. ‘+DNA’ is denoted 5′—3′ and ‘−DNA’ is denoted 3′—5′. We focus on +DNA and −DNA as the substrate for computational states. The computational states, in our setting, encode candidate witnesses for SATISFIABILITY.

Suppose that we would like to encode the sequence of integers  $S$  as an equivalent oligonucleotide representation  $O_1$ . Representing an integer sequence requires a systematic mapping of an oligonucleotide entry with an integer counterpart. A fixed width represen-



Table 2.1: A mapping of the integers  $[0, 5]$  with arbitrary oligonucleotide definitions.

Integer	Oligonucleotide	Reverse-complement
0	5'TCTCCC3'	3'AGAGGG5'
1	5'AAACCC3'	3'TTGGG5'
2	5'GGTAAA3'	3'CCATTT5'
3	5'CCCTCC3'	3'GGGAGG5'
4	5'CTTTTC3'	3'GAAAAG5'
5	5'CCTTCC3'	3'GGAAGG5'

tation map independent sequences on a readable boundary. Now we explore an example for encoding an integer sequence with a sequence of oligonucleotides with the definitions in Table 2.1.

We have, e.g.,

$$S = [1, 3, 4, 3, 2, 0]$$

and

$$O_1 = 5'AAACCC \mid CCCTCC \mid CTTTTC \mid CCCTCC \mid GGTAAA \mid TCTCCC3'.$$

Recovering the sequence  $S$  from  $O_1$  can be done several ways. Because the definition of the sequence exists, we may use the reverse complement to match sequences. Another method splits the sequence  $O_1$  on the encoding width. In this case, the encoding width is six base pairs. Gene sequencing tools permit one to read and decode data according to Table 2.1.

Molecular computation encodes genetic information for both storing and operating on a problem state. These operations include matching and replication. Although this setting describes and artificial processes, DNA in natural settings also share the same mechanics that we exploit here. Interactions between genetic molecules are the fundamental mechanism for generic computation with oligonucleotides.

In the following chapters, we describe molecular algorithms for SATISFIABILITY. In the

next section, we introduce techniques from Adleman’s molecular toolbox [1].

## 2.3 Adleman’s molecular toolbox for solving HAMILTONIAN PATH

In 1994, Leonard Adleman performed the first molecular computation using recombinant DNA in a bench laboratory setting [1]. This experiment solved a six vertex instance of HAMILTONIAN PATH, an NP-complete problem. In this section, we describe the techniques used in this experiment. We provide definitions for the following operations from Adleman’s molecular toolbox: append, extract, mix, split, and purify.

### Definition 2.3.1. HAMILTONIAN PATH

*Given an undirected graph  $G$ , does there exist a path that visits every vertex exactly once?*

Adleman uses oligonucleotides for defining each vertex for encoding a graph. His scheme for encoding a graph’s vertices shares a similar definition for our example of encoding a sequence of integers, given in Table 2.1. Representing edges requires a reverse-complement oligonucleotide, this string connects the suffix of the vertex  $v_i$  with the prefix of  $v_j$ . Let us consider an example. Let

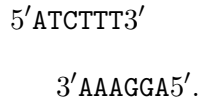
$$v_1 = 5'ATCTTT3'$$

$$v_2 = 5'CCTATA3'.$$

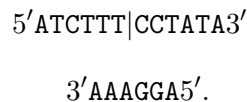
From the definition of  $v_1$  and  $v_2$ , we can construct an edge  $e_{1,2}$  as

$$e_{1,2} = 3'AAAGGA5'.$$

Appending  $v_2$  to  $v_1$  is accomplished by first attaching the edge  $e_{1,2}$  to the vertex  $v_1$



Next we attach  $v_2$  to the resulting complex, yielding



Finally the edge may be removed and we have the sequence

$$v_1 \cdot v_2 = 5'ATCTTT|CCTATA3'.$$

The sequence  $v_1 \cdot v_2$  represents the path  $v_1$  to  $v_2$ , and can be obtained with the *append* operation. A test tube  $T$  stores possible solutions. The tube  $T$  starts as an empty tube. To solve HAMILTONIAN PATH, we introduce equimolar portions of each oligonucleotide vertex for a starting configuration, using the *mix* operation.

**Definition 2.3.2.** *Mix*

$T \leftarrow \text{mix}(T_1, \dots, T_n)$  — combine  $n$  test tubes of information. The output consists of a single set  $T = T_1 \cup \dots \cup T_n$ .

A small initial set may be amplified with *polymerase chain reaction* (PCR). PCR thermocycles the contents of the tube to replicate the contents. Introducing each vertex representation to the contents randomly generates all potential paths. A set of DNA configurations are generated to represent the set of all witness candidates for Hamiltonian Paths in a graph instance. This set of DNA configurations will be filtered to only include

configurations that witness Hamiltonian Paths in  $G$ .

*Append* attaches a string to each string contained in a test tube. *Split* portions a tube into multiple portions. In Chapter 3, we will use split-mix synthesis as a means for generating a combinatorial space.

**Definition 2.3.3.** *Append*

$T' \leftarrow \text{append}(T, s)$  — the concatenation of the oligonucleotide  $s$  with each element in  $T$ .

**Definition 2.3.4.** *Split*

$[T', T''] \leftarrow \text{split}(T)$  — distributes  $T$  into two tubes. Each of the resulting tubes,  $T'$  and  $T''$ , contain the same representative elements of  $T$ .

The initial and terminal conditions for the graph get fulfilled by extracting, from the tube  $T$ , only paths that begin with  $V_{in}$  and end with  $V_{out}$ . Extracting only strings from  $T$  that match these conditions constrain the number of potential strings to only those that satisfy the conditions of the graph instance.

**Definition 2.3.5.** *Extract*

$T' \leftarrow \text{extract}(T, s)$  — separates all oligonucleotides from  $T$  containing the sequence  $s$ . The output consists of a set  $T'$  of those oligonucleotides containing  $s$ .

The tube  $T$  consists of possible encodings that have the correct starting and ending vertices. We select only strings of length  $n$ , where  $n$  is the number of vertices in  $G$ , to ensure that all vertices get traversed. This can be performed with *gel electrophoresis*, a technique for sorting molecules by mass. Next, we ensure that each vertex occurs exactly once. If a vertex occurs multiple times in a path, then the string representation gets discarded.

Finally, we check  $T$  with *detect* to determine if any valid paths remain. If valid paths exist, then each string may be read for the path assignment.

**Definition 2.3.6.** *Detect*

$\text{detect}(T)$  — determine if any encodings are present in  $T$ . The output consists of true or false, for  $T \neq \emptyset$  or  $T = \emptyset$  respectively.

**2.3.1 Additional molecular operators**

In the following chapters, we will use the molecular operators for constructing molecular SATISFIABILITY solvers. The Distribution algorithm, introduced in Chapter 4, requires the *splice* operation.

**Definition 2.3.7.** *Splice*

$[a_1, a_2] \leftarrow \text{splice}(a, b)$  — cuts an oligonucleotide  $a = a_1 \cdot \bar{b} \cdot a_2$  with a subsequence  $b$  into two pieces by a restriction enzyme. These two pieces are  $a_1$  and  $a_2$ .

In the implementation of a simulation system, we avoid redundant string representations with the *purify* operation. This is a synthetic version of PCR. Purify balances the space representation of molecules with a uniform distribution.

**Definition 2.3.8.** *Purify*

$T' \leftarrow \text{purify}(T)$  — provides a uniform distribution from the contents of  $T$  as  $T'$ .

**2.4 Definition of SATISFIABILITY****Definition 2.4.1.** SATISFIABILITY

$$\text{SATISFIABILITY} = \{\langle \phi \rangle \mid \phi \text{ is a satisfiable Boolean formula}\}[22].$$

Cook and Levin independently introduced the canonical instance of a NP-complete language SATISFIABILITY [3, 13]. A NP-complete language is one that is in NP and NP-hard. A NP-hard language is at least as hard as any problem in NP.

Evaluation of a SATISFIABILITY instance requires validating a witness candidate with the instance definition. We introduce SATISFIABILITY evaluation with a circuit. Let us consider a three-layered circuit for SATISFIABILITY. This circuit consists of  $n$  inverters,  $m$  **OR** gates, and one **AND** gate with  $m$ -fan-in. This circuit behaves according to the internal wiring of the input expression  $\phi$ . Figure 2.1 contains a schematic for SATISFIABILITY.

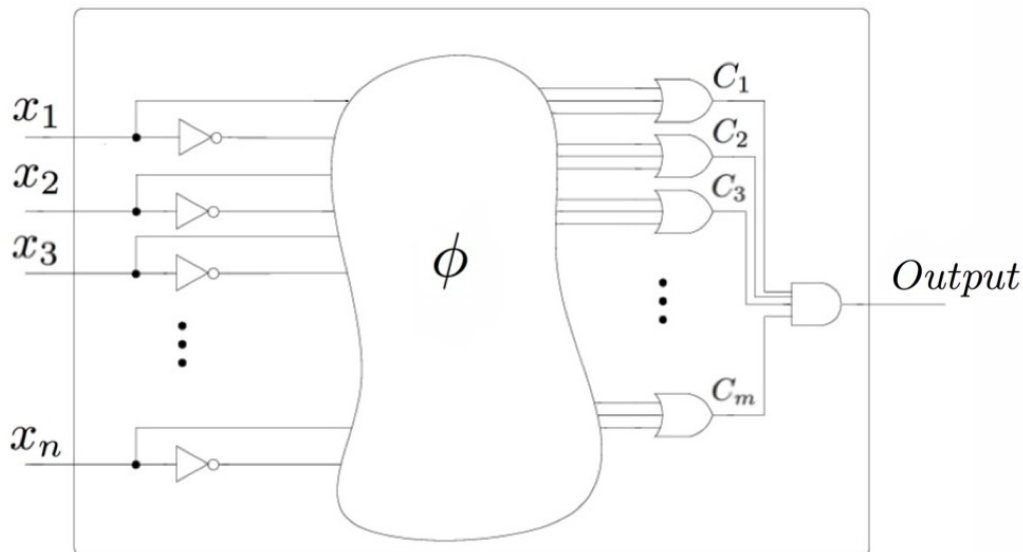


Figure 2.1: A circuit describing SATISFIABILITY.

The realization of SATISFIABILITY as a circuit shows two aspects of this problem. SATISFIABILITY can be implemented as a circuit proportional to the input. The worst case verification for all  $2^n$  possible witness candidates may be verified with the circuit in Figure 2.1. This circuit consists of the hardware equivalent version of the SATISFIABILITY validator CHECKSAT, as shown in Chapter 1. Subproblems of SATISFIABILITY include CNF,  $k$ -CNF, and  $k$ -SAT problem definitions.

**Definition 2.4.2.** *CNF*

*Conjunctive Normal Form consists of the intersection of sets of disjunctive literals.*

**Definition 2.4.3.** *k*-CNF

*Consists of a CNF expression with each disjunctive clause containing  $k$  literals.*

**Definition 2.4.4.** *k*-SAT

*Problem variant of SATISFIABILITY where each clause consists of  $k$  Boolean literals.  $k$ -CNF formula provide an equivalent representation.*

The next section considers standards adopted for SATISFIABILITY. This allows practitioners to apply SATISFIABILITY in various settings.

## 2.5 Evaluating SAT Solvers

In this section, we describe two standards for encoding the SATISFIABILITY problem that we adopt for the implementation. This includes the input and output standards from the SATISFIABILITY Competition [4, 21].

Next, we introduce problem instance classification scheme for SATISFIABILITY. Classification of SATISFIABILITY problem instances include randomly generated, combinatorial, and industrial [21]. Our experimental setup in Chapter 6 considers generation of random  $k$ -SAT input.

### 2.5.1 Input and output

The SAT Competition ranks implementations for SAT solvers for evaluating SATISFIABILITY [21]. SAT solvers demonstrate state-of-the-art techniques for solving three main tracks of SATISFIABILITY instances. The tracks exhibit applications for SATISFIABILITY, including: industrial applications, hard combinatorial, and random problem instances. The input and output standards for SATISFIABILITY allow common benchmarks for SAT solvers. We conform to the standards of this competition <http://www.satcompetition.org/>.

## Input

DIMACS CNF provides a standard input for SATISFIABILITY [4]. The format permits sharing of existing SATISFIABILITY benchmarks by encoding SATISFIABILITY in conjunctive normal form (CNF). We provide an example of this encoding in Section 5.6.

## Output

SAT Competition output consists of the status for a DIMACS CNF input instance [21]. This includes the known state, either SATISFIABLE, UNSATISFIABLE, or UNKNOWN. When a witnessing satisfying assignment occurs, the assignment is provided as a list of integers with the SATISFIABLE state. We provide an example along with a custom interface in Section 5.7.

### 2.5.2 Metrics for classifying SATISFIABILITY

SAT phase transition and SAT backbones are two classifying metrics for SATISFIABILITY. These metrics may be used to classify SATISFIABILITY expressions. We will use these metrics in the next section for defining a collection of random  $k$ -SAT instances.

**Definition 2.5.1.** *SAT phase transition*

*The ratio of  $m$  clauses to  $n$  variables  $\alpha = m/n$  provides a characterization for where phase transitions may occur in the space of all  $k$ -CNF formula [5, 11].*

The SAT phase transition is a region where both satisfiable and unsatisfiable instances are likely. This region separates trivially satisfiable and over-constrained unsatisfiable SATISFIABLE problem instances.

**Definition 2.5.2.** *SAT backbones*

*SAT backbones are the variable assignments present in all of the satisfying assignments to*



SATISFIABILITY *problem instances* [26].

SAT backbones contain a set of variables that occur in all satisfiable witnesses for an input expression. If there are no such variables in the set of all witnesses for a problem instance, then the set is empty.

### 2.5.3 SATISFIABILITY instances

There are several methods for generating SATISFIABILITY instances. We consider three classes of SATISFIABILITY based on random assignment, combinatorial, and industrial applications.

#### **Random $k$ -SAT**

A random  $k$ -SAT instance consists of  $m$  clauses with  $k$  literals per-clause from  $n$  variables [24]. An instance is drawn uniformly over the set of variables without replacement.

#### **Hard combinatorial SAT**

Combinatorial instances provide difficult benchmark cases. These instances represent other NP-complete problems. Including games and graph theoretic problems represented as SATISFIABILITY.

#### **Industrial SAT applications**

Industrial processes apply SATISFIABILITY to solve real world problems, including circuit layout, planning, logistics, circuit fault testing, and many other industrial NP-complete problems. Applications for industrial SAT will often apply heuristics and approximation techniques to relax the problem. This allows approximate solutions to be computed in an efficient amount of time.

## Chapter 3

# Existing molecular algorithms for SATISFIABILITY

In this chapter, we introduce two molecular algorithms for SATISFIABILITY. These algorithms are distinct in the resolution of a SATISFIABILITY instance. Lipton’s algorithm requires a combinatorial space of all witness candidates to be constructed before execution. Ogihara and Ray’s algorithm constructs a set of witness candidates during execution. Following the description, we explore physical implementation and simulation frameworks for these algorithms.

### 3.1 Lipton’s algorithm for SATISFIABILITY

Introduced in 1995 by Richard Lipton [15], this algorithm filters satisfiable witnesses from a combinatorial space of all witness candidates. Lipton’s algorithm is analogous to a conventional brute-force search for all witnesses for a SATISFIABILITY instance.

The algorithm begins by creating a combinatorial space of DNA configurations. These configurations are filtered by extracting potential solutions for each clause. The instance

$\phi$  has satisfiable witnesses if  $T$  is non-empty.

### 3.1.1 Description of Lipton’s algorithm

Lipton’s algorithm consists of two main procedures. The first phase constructs a combinatorial space of  $2^n$  independent vectors. Second, the combinatorial space gets filtered based on the input CNF instance.

The function COMBINATORIAL GENERATE( $n$ ) implements the split-mix synthesis technique [8, 9]. It returns a gel consisting of  $2^n$  independent oligos that correspond to a unique vector space. The space begins construction with an initial medium. An iterative loop elongates a growing solution with the split-mix synthesis. Each split corresponds with appending the tubes with a truth and false assignment. The two tubes are mixed and amplified to contain equimolar portions.

The amplification process gets modeled with a purification step. This eliminates all redundant strings for the simulated implementation. After the iteration completes, the complete combinatorial space gets returned. This space consists of  $2^n$  vectors of length  $n$ .

From the combinatorial space, we will begin to filter satisfying solutions to the input CNF formula. For each clause, we extract each of the variables present in the solution space. A disjunctive set  $T_C$  contains the satisfied string instances for each clause. LIPTON’S ALGORITHM iterates over each of the clauses. From the selected clause, the variables get extracted from the combinatorial space. Once complete, the remaining space,  $T$ , contains satisfiable instances for  $\phi$ .

### 3.1.2 Pseudocode for Lipton's algorithm

Algorithms 3.1.1 and 3.1.2 provide pseudocode for Lipton's algorithm. Appendix B lists a detailed execution trace for Lipton's algorithm.

**Algorithm 3.1.1:** COMBINATORIAL GENERATE( $n$ )

```
 $T_{comb} \leftarrow \emptyset$   
 $T_{comb} \leftarrow \text{mix}(T_{comb}, \text{start})$   
for  $v \leftarrow 1$  to  $n$   
  do  $\left\{ \begin{array}{l} [T_1, T_2] \leftarrow \text{split}(T_{comb}) \\ T_1 \leftarrow \text{append}(T_1, +v) \\ T_2 \leftarrow \text{append}(T_2, -v) \\ T_{comb} \leftarrow \text{mix}(T_1, T_2) \end{array} \right.$   
return  $(T_{comb})$ 
```

**Algorithm 3.1.2:** LIPTON'S ALGORITHM( $\phi$ ) $n$  number of variables in  $\phi$  $T \leftarrow \text{COMBINATORIAL\_GENERATE}(n)$ **for each** clause  $C$  in  $\phi$ 

$$\text{do } \left\{ \begin{array}{l} T_c \leftarrow \emptyset \\ \text{for each variable } v \text{ in } C \\ \quad \left\{ \begin{array}{ll} \text{if } v \text{ is a positive literal} & \\ \text{do } \left\{ \begin{array}{l} \text{then } \left\{ \begin{array}{l} T_P \leftarrow \text{extract}(T, +v) \\ T_c \leftarrow \text{mix}(T_P, T_c) \end{array} \right. \\ \text{else } \left\{ \begin{array}{l} T_N \leftarrow \text{extract}(T, -v) \\ T_c \leftarrow \text{mix}(T_N, T_c) \end{array} \right. \end{array} \right. \\ T \leftarrow T_c \end{array} \right.$$
**return** (detect( $T$ ))**3.2 Ogihara and Ray's algorithm for SATISFIABILITY**

Ogihara and Ray's algorithm consist of a breadth-first evaluation of clauses from a CNF formula [18, 19]. The algorithm constructs a set of potential solutions based on parsing a 3-CNF formula. In this section, we describe the preconditions and execution of Ogihara and Ray's algorithm.

**3.2.1 Description of Ogihara and Ray's algorithm**

Prior to execution of the algorithm it requires two attributes of CNF input:

1. All clauses consist of exactly three literals
2. All clauses must be sorted by variable

Attribute (1) gets fulfilled by considering only 3-SAT expressions. If models of  $k$ -SAT with  $k > 3$ , then a polynomial time reduction to 3-SAT must occur prior to execution.

Attribute (2) gets fulfilled by sorting the clauses prior to execution; providing the weak ordering

$$v_1 < \cdots < v_n,$$

where the polarity of each variable may consist of a positive or negative assignment.

The initial tube consists of potential states for the first two variables.

Expanding each partial assignment iterates over each clause in the input CNF. Construction of satisfiable expressions consider the possibilities of the clause ordering

$$x_u < x_v < x_w.$$

OGIHARA AND RAY'S ALGORITHM evaluates each subsequent variable and determines possible assignments. The possible assignments for the variables  $v_1$  and  $v_2$  get extracted if  $v_3$  matches. Effectively pruning only potential solutions. These potential solutions  $T_P$  and  $T_N$  get appended with the positive or negative string assignments. The algorithm continues until each variable gets evaluated. The remaining space  $T$  contains all solutions for the CNF instance  $\phi$  after the algorithm terminates.



### 3.2.2 Pseudocode for Ogihara and Ray's algorithm

Algorithm 3.2.1 provides pseudocode for Ogihara and Ray's algorithm. Appendix B lists a detailed execution trace for Ogihara and Ray's algorithm.

**Algorithm 3.2.1:** OGIHARA AND RAY'S ALGORITHM( $\phi$ )

$n$  number of variables in  $\phi$

Each variable of the reordered clause can be accessed by  $v_1$ ,  $v_2$ , and  $v_3$

Reorder variables by most frequent to least frequent literal appearance

Reorder each clause in increasing literal order

$T \leftarrow \{[+x_1 \cdot +x_2], [+x_1 \cdot -x_2], [-x_1 \cdot +x_2], [-x_1 \cdot -x_2]\}$

**for each** variable  $x_i$  in  $3 \leq i \leq n$

**do** {

$[T_P, T_N] \leftarrow \text{split}(T)$

**for each** clause  $C$  in  $\phi$

$[v_1, v_2, v_3] \leftarrow C$

**if**  $x_i = v_3$

**then** {

$T_{P1} \leftarrow \text{extract}(T_N, v_1)$

$T_{N1} \leftarrow \text{extract}(T_N, -v_1)$

$T_{P2} \leftarrow \text{extract}(T_{N1}, v_2)$

$T_N \leftarrow \text{mix}(T_{P1}, T_{P2})$

**if**  $\neg x_i = v_3$

**then** {

$T_{P1} \leftarrow \text{extract}(T_P, v_1)$

$T_{N1} \leftarrow \text{extract}(T_P, -v_1)$

$T_{P2} \leftarrow \text{extract}(T_{N1}, v_2)$

$T_P \leftarrow \text{mix}(T_{P1}, T_{P2})$

$T_P \leftarrow \text{append}(T_P, +x_i)$

$T_N \leftarrow \text{append}(T_N, -x_i)$

$T \leftarrow \text{mix}(T_P, T_N)$

25

**return** (detect( $T$ ))



### 3.3 Implementations of molecular SATISFIABILITY solvers

In this section, we describe physical and simulated implementations for molecular SATISFIABILITY algorithms. This includes simulation of Lipton’s and Ogihara and Ray’s algorithms. We see a physical implementation of Ogihara and Ray’s algorithm with manual laboratory procedures.

#### 3.3.1 Physical implementations

Yoshida and Suyama implemented Ogihara and Ray’s algorithm with manual molecular biology techniques [25]. This experiment solved a 3-CNF instance with four variables and 10 clauses.

#### 3.3.2 Simulations

Martn-Mateos et al. introduced a simulation for Lipton’s algorithm [17]. Molecular operations get implemented with ACL2, a Common Lisp variant. The framework for this system implemented test cases for Lipton’s algorithm.

Ogihara provides test results for implementation of his original molecular algorithm [18]. This simulation provides a comparison with Lipton’s algorithm for practical length restrictions.

## Chapter 4

# A new molecular algorithm for SATISFIABILITY

This chapter introduces a new molecular algorithm for SATISFIABILITY. The distribution algorithm parses an input CNF expression into growing and self regulated set of possible combinations.

### 4.1 Distribution algorithm for SATISFIABILITY

The distribution algorithm parses an input CNF expression into growing and self regulated set of possible combinations. A possible combination begins with all members of the first clause. Variables get inserted into an expanding set of valid assignments. A clause gets eliminated when an assignment contains a conflict.

#### 4.1.1 Description of the Distribution algorithm

Initially the algorithm starts with the variable assignments of a clause. Evaluation of subsequent clauses extends the solution space with the INSERT VARIABLE subroutine. During

each insertion, the variable gets inserted into a potential solution vector. Table 4.1 lists the four possibilities for variable assignment.

Table 4.1: Configurations for the INSERT VARIABLE subroutine

Case	Return state	State
1	$v \cdot s$	if $v$ is less than all elements in $s$
2	$s \cdot v$	if $v$ is greater than all elements in $s$
3	$s_1 \cdot v \cdot s_2$	if $v$ is between two elements in $s$
4	$\emptyset$	if $v$ conflicts with $-v$ in $s$
5	$s$	if $v$ exists in $s$

During this phase, each variable from a disjunctive clause gets considered, incrementally constructing a partial solution space. Cases (1), (2), and (3) place a variable  $v$  into an existing sequence  $s$ . Each of these cases represents when the variable  $v$  get inserted in a non-decreasing sequence.

A variable conflict occurs when both positive and negative assignments of a variable occur in a sequence  $s$ . In this case (4), the sequence  $s$  gets removed from the set potential solutions. If the sequence  $s$  contains the variable  $v$ , case (5), then the existing sequence  $s$  gets returned unmodified.

Redundant vectors get removed after insertion of the next disjunctive clause. Any remaining witnesses in the solution space contain non-conflicting variable assignments. This does not immediately require that each witness to be a complete satisfiable assignment. Satisfiable witnesses remain in a non-empty satisfying solution space.

Vectors that are of equal magnitude of the number of variables in the problem instance are satisfiable witnesses. However, there may exist solutions that span only the required

satisfiable assignments; that is activate each of the independent clauses with at least one non-conflicting assignment. This assignment may be the minimum witness for the expression, in the case that the backbone consists of the variables of the maximum witness.

#### 4.1.2 Pseudocode for Distribution algorithm

Algorithms 4.1.1 and 4.1.2 provide pseudocode for the Distribution algorithm. Appendix B lists a detailed execution trace for the Distribution algorithm.

**Algorithm 4.1.1:** INSERT VARIABLE( $T, v$ )

```

 $T_R \leftarrow \emptyset$ 
for each string  $s$  in  $T$ 
    do {
        case (1) :  $v < s$ 
            then  $s' \leftarrow \text{append}(v, s)$ 
        case (2) :  $v > s$ 
            then  $s' \leftarrow \text{append}(s, v)$ 
        case (3) :  $s_1 < v < s_2$ 
            then {
                 $[s_1, s_2] \leftarrow \text{splice}(s, v)$ 
                 $s_1 \leftarrow \text{append}(s_1, v)$ 
                 $s' \leftarrow \text{append}(s_1, s_2)$ 
            }
        case (4) :  $\neg v \in s$ 
            then  $s' \leftarrow \emptyset$ 
        case (5) :  $v \in s$ 
            then  $s' \leftarrow s$ 
         $T_R \leftarrow \text{mix}(T_R, s')$ 
    }
return ( $T_R$ )

```

**Algorithm 4.1.2:** DISTRIBUTION SAT( $\phi$ )

$m$  number of clauses

$k$  number of variables in each clause

Initialize with the variables from the first clause

$T \leftarrow \{C_1\}$

**for**  $i \leftarrow 2$  to  $m$

**do**  $\left\{ \begin{array}{l} T_C \leftarrow \emptyset \\ \textbf{for each} \text{ variable } v \text{ in } C_i \\ \textbf{do} \left\{ \begin{array}{l} T_I \leftarrow \text{INSERT VARIABLE}(T, v) \\ T_C \leftarrow \text{mix}(T_C, T_I) \end{array} \right. \\ T \leftarrow T_C \end{array} \right.$

**return** ( $\text{detect}(T)$ )

## Chapter 5

# Molecular Simulation: A system for molecular computation

This chapter introduces Molecular Simulation: A system for molecular computation. We provide an overview of the software and download location for Molecular Simulation and its documentation. We provide tools for use with Molecular Simulation. This includes Perl execution scripts and visualization for output data. We provide examples for Molecular Simulation's input and output. Invocation of Molecular Simulation from the command line provides user configurable options. The next chapter describes the usage of Molecular Simulation with automated execution.

### 5.1 Overview

Molecular Simulation provides a molecular lab for operating on DNA. The present simulation implements three molecular algorithms for SATISFIABILITY. The included `Perl` scripts process DIMACS CNF input directories with invocations to Molecular Simulation.

Molecular Simulation may be executed directly or invoked with the assistance of an

execution script. The system requirements to execute or design a molecular experiment are listed in this section.

This program is a simulated molecular lab for experimenting with DNA operations. Implementation of three molecular algorithms for solving SATISFIABILITY include Lipton's algorithm, Ogihara and Ray's algorithm, and the Distribution algorithm. Chapters 3 and 4 provide a background and pseudocode for these algorithms.

## 5.2 Download

Molecular Simulation can be downloaded from:

<https://github.com/dncarley/MolecularSimulation>.

## 5.3 Requirements

Requirements for Molecular Simulation are specified in this section. This includes the hardware and software requirements for running Molecular Simulation on your system.

### 5.3.1 Hardware requirements

Molecular Simulation requires a 64-bit processor with 2 GB of RAM.

### 5.3.2 Software requirements

`gcc` (GNU Compiler Collection) must be installed on your system.

`Perl` must be installed on your system to automate build and execution of Molecular Simulation.

## 5.4 Documentation

The project website contains detailed documentation for Molecular Simulation. The documentation provides an overview of Molecular Simulation that may be used independently of Chapters 5 and 6 for getting started. The online documentation provides detailed datatype, function, and class definitions.

## 5.5 Tools

This project uses several tools for automating tasks and execution. In this section, we introduce tools to automate execution and visualize output from Molecular Simulation.

### 5.5.1 Perl utilities

The source directory includes several `Perl` scripts to assist in building and initiation of tests for Molecular Simulation. Table 5.1 documents the basic usage for build and testbench execution scripts. Each script provides detailed execution options.

### 5.5.2 Data Visualization

A SAT datapoint visualization for Molecular Simulation’s output can be downloaded from:

`https://github.com/dncarley/VisualizeSatDatapoints`

Ben Fry’s example in Chapter 4 of *Visualizing Data* [7] provides a framework for importing output from Molecular Simulation. The visualization project directory contains a README for usage.



Table 5.1: Perl execution commands and descriptions.

Perl script	Usage	Description
<code>build.pl</code>	<code>\$ perl build.pl</code>	Compiles Molecular Simulation and generates an executable in the directory <code>./execute/simulation</code> .
<code>buildGenerate.pl</code>	<code>\$ perl buildGenerate.pl</code>	Generates a sweep of CNF formulas over a range of $k$ -SAT ratios. Program uses a modified random $k$ -SAT generator from Microsoft Research.
<code>executeMolecularSat.pl</code>	<code>\$ perl executeMolecularSat.pl</code>	Executes Molecular Simulation for a directory of SATISFIABILITY expressions with desired algorithms. If no options are specified, then each of the three algorithms are executed and output is generated in the same test directory.
<code>runSimulation.pl</code>	<code>\$ perl runSimulation.pl</code>	Executes <code>build.pl</code> followed by <code>executeMolecularSat.pl</code> . Any command line arguments get passed to <code>executeMolecularSat.pl</code>

## 5.6 Input

Input to Molecular Simulation consists of a DIMACS CNF file. The definition of the \*.cnf filetype can be accessed from: <ftp://dimacs.rutgers.edu/pub/challenge/satisfiability/doc/>.

```
c comments begin with a 'c'
c
c cnf input is designated with 'p cnf'
c   followed by number of variables <n>, and clauses <m>
c
p cnf <n> <m>
c
c A clause is represented by a sequence of <k> integers,
c   separated by whitespace and ending with a '0'.
c Each variable is represented by the integer sequence,
c   negative polarity is represented by '-'.
c
-3 9 14 0
6 -9 -12 0
-2 11 17 0
3 -13 -17 0
```

## 5.7 Output

Output from Molecular Simulation, by default, conforms to the 2011 SAT Competition rules. The rules can be accessed from: <http://www.satcompetition.org/2011/rules>.

pdf.

```
c comments begin with a 'c'

c

s SATISFIABLE

c

c A line beginning with a 's' marks the status.

c This can be either 'UNSATISFIABLE', 'SATISFIABLE', or 'UNKNOWN'.

c

v -3 -9 11 13 0

c

c A satisfiable witness begins with a 'v' and ends with a '0'.

c      A sequence of integers, between 'v' and '0', encodes a satisfiable assignment.
```

Table 5.2 describes an extended custom output. This output reports parameters for metric performance evaluation.

Table 5.2: Molecular Simulation output logging.

Parameter	Description
c algorithmType:	Display the algorithm type: <b>Lipton</b> , <b>Ogihara-Ray</b> , <b>Distribution</b>
c algorithmTime:	Display the algorithm execution time in seconds.
c solutionMemory:	Display the solution space memory footprint in Bytes.
c mixCount:	Display the number of <b>mixes</b> required during algorithm execution.
c extractCount:	Display the number of <b>extracts</b> required during algorithm execution.
c appendCount:	Display the number of <b>appends</b> required during algorithm execution.
c splitCount:	Display the number of <b>splits</b> required during algorithm execution.
c spliceCount:	Display the number of <b>splices</b> required during algorithm execution.
c purifyCount:	Display the number of <b>purifications</b> required during algorithm execution.
c numVar:	Display the number of <b>variables</b> in the input CNF expression.
c numClause:	Display the number of <b>clauses</b> in the input CNF expression.

## 5.8 Execution

Invocation of Molecular Simulation can be performed from the command line.

```
$ ./execute/simulation i [input] [options]
```

The [input] consists of a DIMACS CNF file. Command line [options] may be a combination of the options in Table 5.3.

Table 5.3: Command line options for Molecular Simulation

Argument	Parameters	Description
-a		Algorithm select
	d	Distribution algorithm
	l	Lipton's algorithm
	o	Ogihara and Ray's algorithm
-d		Debug
i	[input]	Input DIMACS CNF file
-w	[output]	Write output to file Output filename

Let us consider an example. Suppose that we would like to execute Ogihara and Ray's algorithm for a DIMACS CNF file. We would like to execute the instance `test1.cnf` located in the directory `/molecularSimulation/testbench`. We output the results `test1-o.out` in the same directory as the input CNF. We invoke Molecular Simulation with the following

command.

```
$ ./execute/simulation i ../testbench/test1.cnf -a o -w ../testbench/test1-o.out
```

In the next chapter, we will describe the automation for a random  $k$ -SAT sweep with each of the algorithms. The provided Perl scripts are the recommended method for building and execution of Molecular Simulation.

## Chapter 6

# Experimental Setup

This chapter describes the use of Molecular Simulation for evaluation of a set of DIMACS CNF SATISFIABILITY instances. We discuss configuration for generation of random  $k$ -SAT instances. Further, any existing DIMACS CNF benchmark may be imported for test. We provide example configuration options for automating the execution of Molecular Simulation. The example continues with an analysis of runtime metrics for each test instance. The next chapter provides the results from the  $k$ -SAT sweep experiment.

### 6.1 Setup

In this section, we describe prerequisites for executing a test bench with Molecular Simulation. Molecular Simulation requires a 64-bit architecture with a UNIX like system with `gcc` and `Perl`. The target system must meet the minimum requirements.

Building Molecular Simulation can be performed by invoking the `Perl` script `build.pl` from the command line.

```
$ perl build.pl
```

This script generates an executable `simulation` in the directory `molecularSimulation\execute`. The next sections describe invocation of Molecular Simulation with desired options. We begin with the creation and importation of DIMACS CNF datasets.

## 6.2 Create dataset

We will create a sweep of random  $k$ -SAT instances to observe SAT phase transition. David Wilson's `ksat.c` generates random  $k$ -SAT instances in DIMACS CNF format. The program takes four arguments to create a unique DIMACS CNF instance. Invocation of the program can be performed with the following command.

```
./execute/ksat k n m s > output.cnf
```

This generates *output.cnf* in DIMACS CNF format with  $k$  variables per clause  $n$  variables,  $m$  clauses, and random seed  $s$ .

We use automated Perl scripts to create a sweep of DIMACS CNF instances. Setup for a sweep configuration includes specifying a set of ratios. Invocation of the script generates a set of random  $k$ -SAT instances. The redirected output gets stored in the target directory with the previous file naming convention. We use the following command to invoke the construction of a sweep of  $k$ -SAT instances.

```
$ perl buildGenerate.pl
```

## 6.3 Import dataset

Datasets of DIMACS CNF input may be provided for batch processing. This includes random  $k$ -SAT instances generated from the previous section, or importing existing DIMACS CNF instances.

DIMACS CNF benchmarks are available for download from: <ftp://dimacs.rutgers.edu/pub/challenge/satisfiability/>.

## 6.4 Configure test

The previous chapter described a single execution of Molecular Simulation. Now we provide the automated invocation for processing datasets with each of the algorithms.

The provided Perl script `executeMolecularSat.pl` allows execution for a directory of DIMACS CNF input. Executing the script from the command line without arguments processes the experimental setup and saves output to the same directory.

```
$ perl executeMolecularSat.pl [options]
```

The options for `executeMolecularSat.pl` can be a combination of the options in Table 6.1.

Table 6.1: Command line options for `executeMolecularSat.pl`

Argument	Parameters	Description
-d -l -o		Distribution algorithm Lipton's algorithm Ogihara and Ray's algorithm
-debug		Debug
-p	[CNF file path]	Specify CNF file path. Default path: <code>data/testCNF</code>
-f		Write output to file



## 6.5 Execution and collection of data

The output can be analyzed after the automated tests have completed. The output consists of the standard SAT Competition output appended with custom runtime metric logging. We discuss viewing output directly during execution and reading saved output files. Collections of output files may be read by the data visualization program and exported into a condensed table.

### 6.5.1 Execution output

Molecular Simulation, by default, writes output to standard output on the console. With the `-f` option, output may be saved to a file. With the `-f` option specified, output gets saved with `[filename]-<a>.out`. The `[filename]` consists of the DIMACS CNF name and `<a>` specifies the algorithm type: `d`, `l` or `o`.

Output directed to standard output conforms to the SAT Competition rules. This output may be used during testing, or redirected to an external stream. The debug option `-debug` provides detailed information about the execution. The debug option writes verbose content based on the program execution.

Reading output metrics from the saved output, as defined in Table 5.2, allows for analysis of collected data. The data visualization reads a directory of output and condenses it as a `*.tsv` file. Subsequent datapoint browsing and the online view use the `*.tsv` file for condensed reading and transmission. In the next chapter, we provide the results of the experimental setup and discuss the design decisions for a general purpose molecular computer.

## Chapter 7

# Results

This chapter provides results of the  $k$ -SAT execution test from the previous chapter. We consider the results of the test and provide analysis of the algorithm metrics.

### 7.1 Algorithm metric comparison

This section provides results from the simulation. We provide the analysis for the molecular operations. These include counts of append, extract, mix, purify, splice, and split. Presentation of actual computation time and required memory for the solution representation allow for comparison of algorithms.

**Append** is an operation that concatenates molecules.

The Distribution algorithm is exponential in the number of appends. The operation count for append depends on the parsing order of the CNF expression.

Lipton's and Ogihara-Ray's algorithms use a fixed amount of appends. This depends on the number of variables and clauses present in the CNF expression.

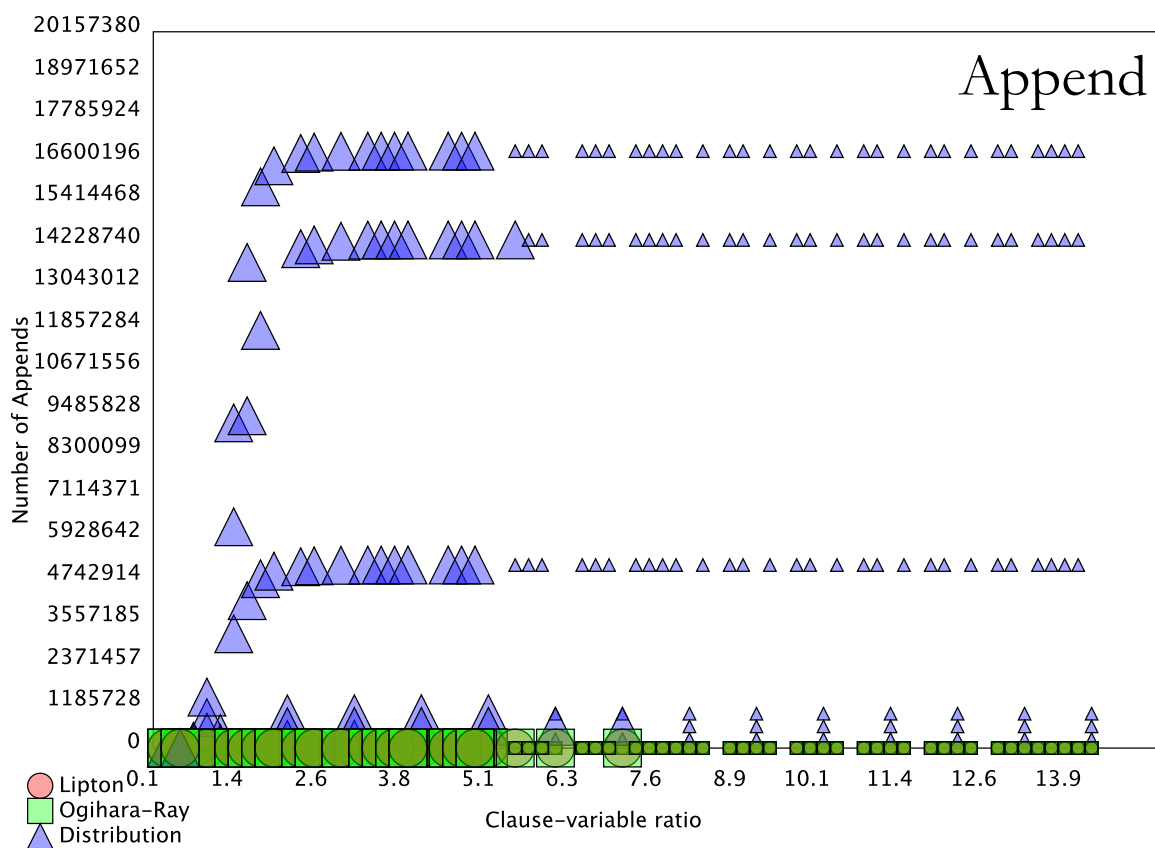


Figure 7.1: Clause to variable ratio  $\alpha$  vs. Number of appends

**Extract** is an operation that filters strings.

Ogihara-Ray's algorithm requires the greatest amount of extracts. Lipton's algorithm is linear on  $\alpha$  and varies a constant amount from Ogihara-Ray's algorithm.

The Distribution algorithm does not require extract.

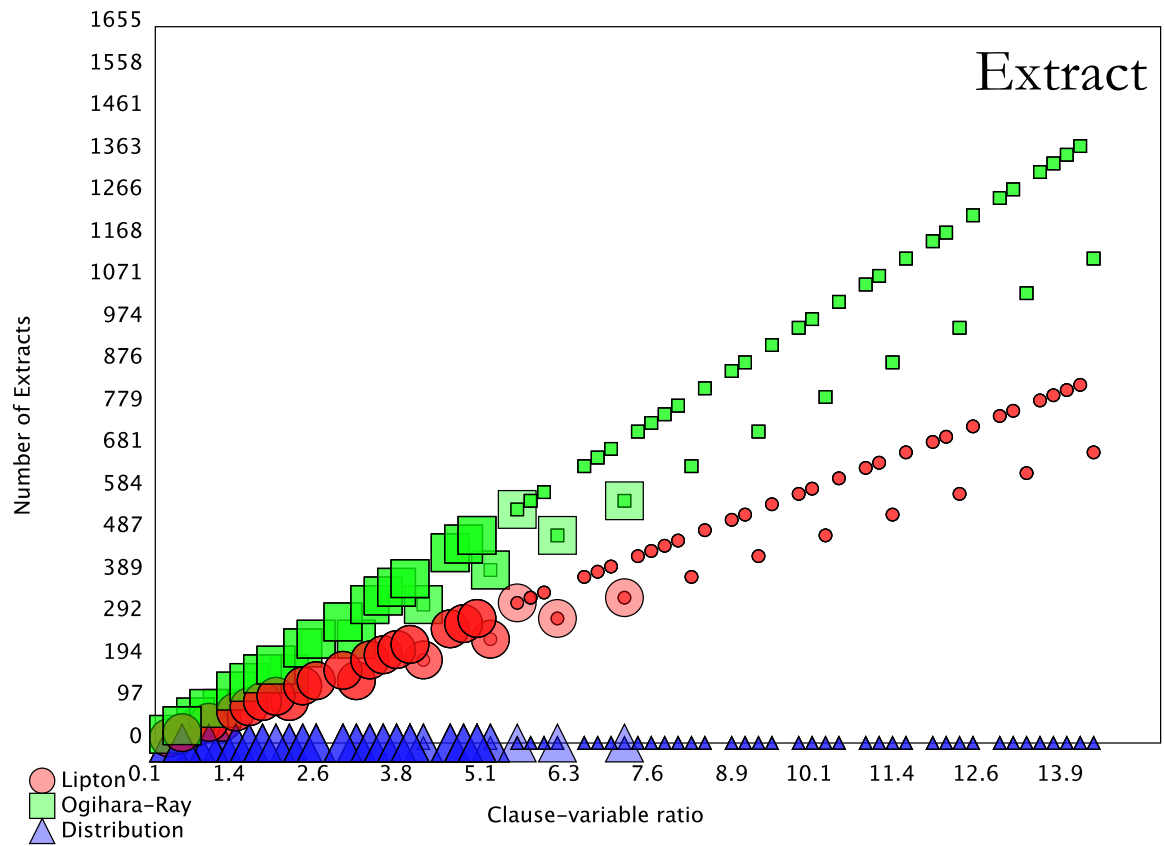


Figure 7.2: Clause to variable ratio  $\alpha$  vs. Number of extracts

**Mix** is an operation that combines two tubes.

Lipton's algorithm requires a linear amount of mixes on  $\alpha$ . The Distribution algorithm also requires a linear number of mixes, varying by a constant factor from Lipton's algorithm.

Ogihara-Ray's algorithm requires a constant amount of mixes on  $\alpha$ .

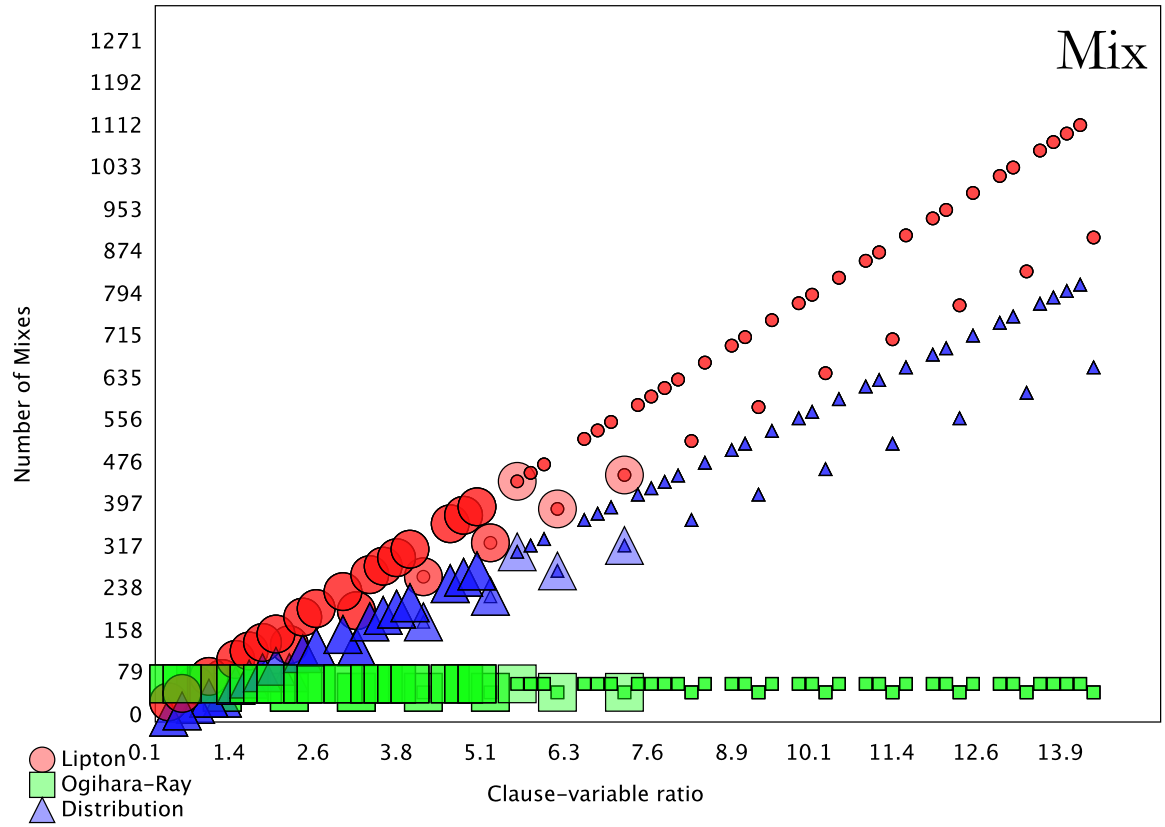


Figure 7.3: Clause to variable ratio  $\alpha$  vs. Number of mixes

**Purify** is an operation that ensures equal portions of each independent string.

All three algorithms operate with a linear number of purifications on  $\alpha$ . Ogihara-Ray's algorithm requires the greatest amount of purifications. The purifications vary by a constant amount when compared with Lipton's and the Distribution algorithms.

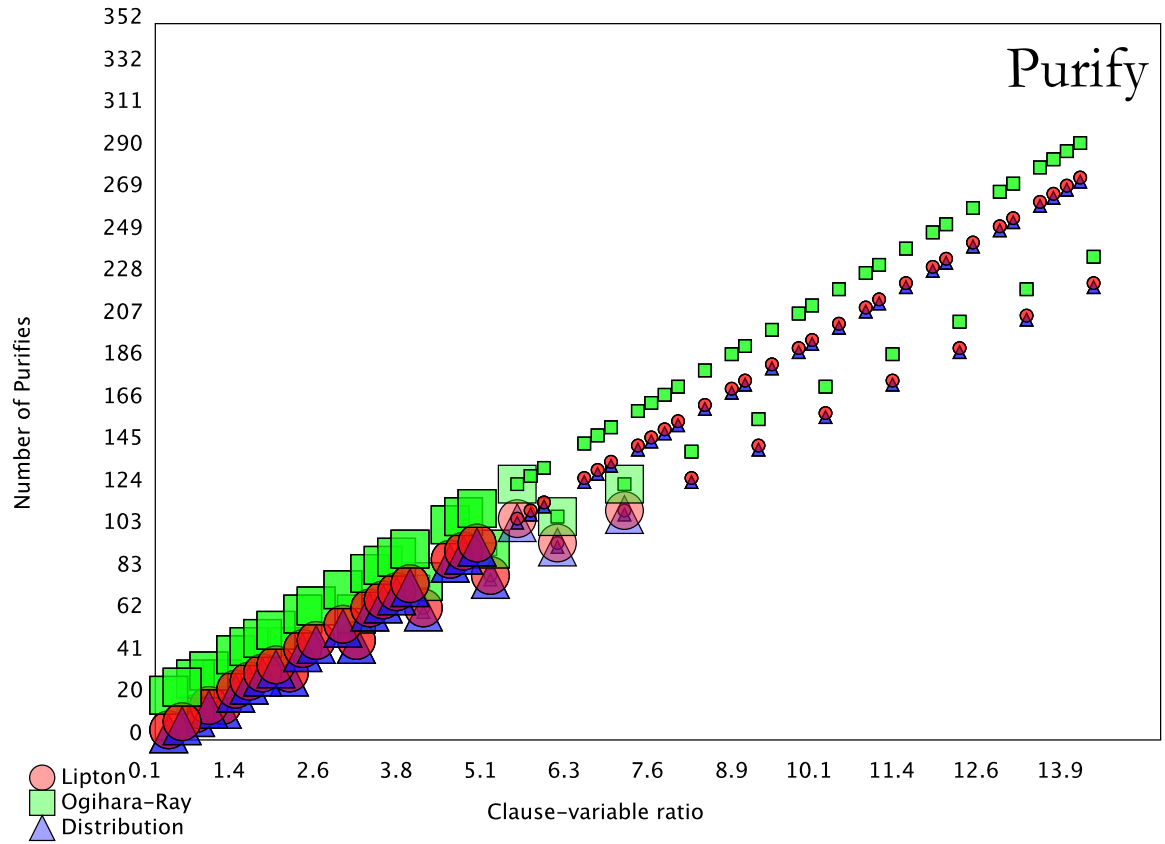


Figure 7.4: Clause to variable ratio  $\alpha$  vs. Number of purifies

**Splice** is an operation that inserts a string at a targeted location.

The Distribution algorithm is exponential in the number of splices. The number of splices depends on the parsing order of the CNF expression. Each split requires reassembly, accomplished with two appends. Figure 7.1 shows the number of appends.

Lipton's and Ogihara-Ray's algorithms do not require splice the splice operator.

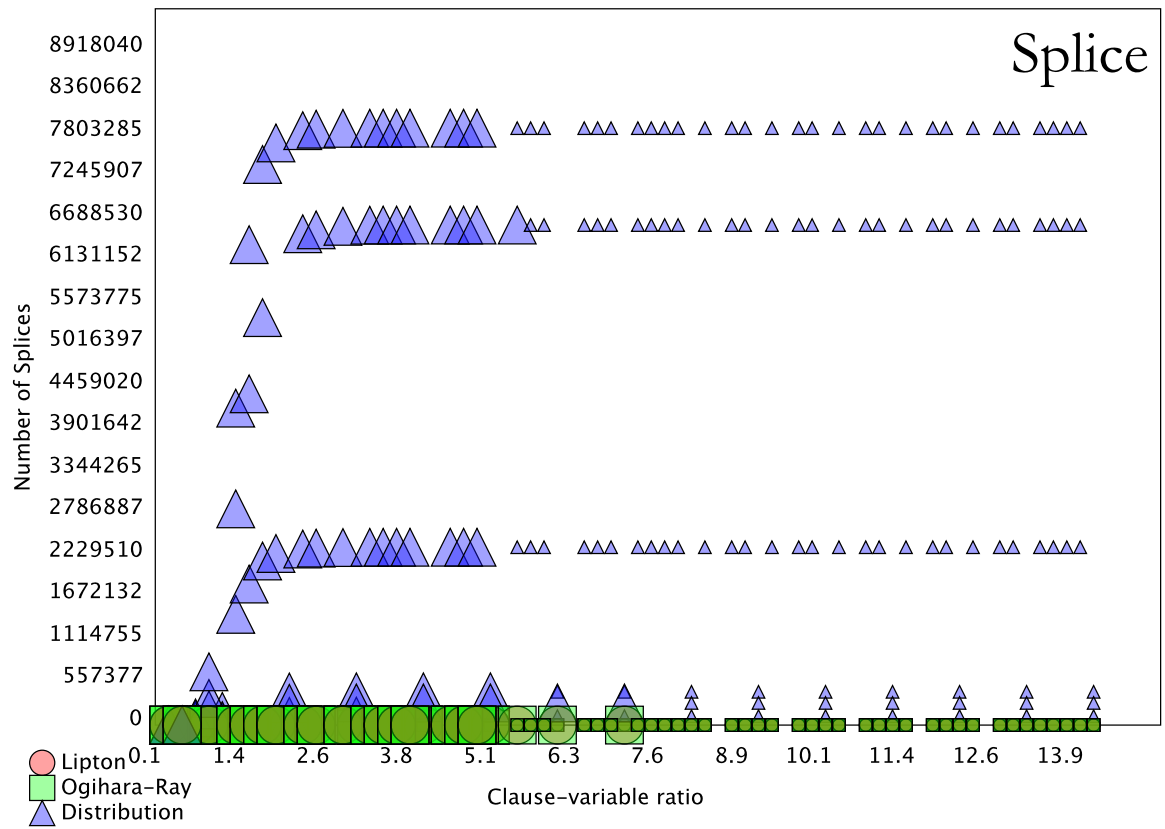


Figure 7.5: Clause to variable ratio  $\alpha$  vs. Number of splices

**Split** is an operation that portions a tube into two exact copies.

Distribution requires a linear number of splits.

Lipton's and Ogihara-Ray's algorithms are constant in splits based the number of variables.

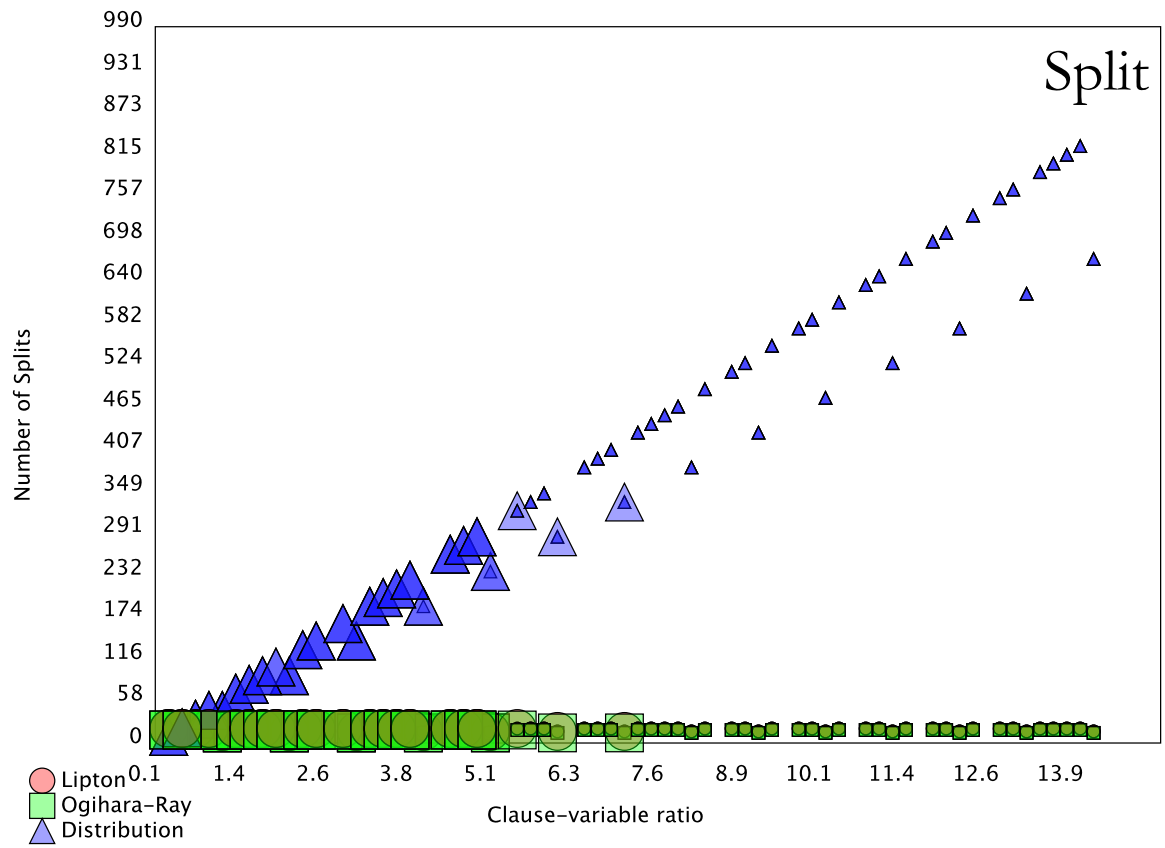


Figure 7.6: Clause to variable ratio  $\alpha$  vs. Number of splits



**Time** is a measurement of algorithm execution in seconds.

Ogihara-Ray's algorithm requires the least amount of time. In cases where the SATISFIABILITY instance is under-constrained, where more possible solutions occur, the algorithm takes the greatest amount of time. Less pruning occurs in over-constrained instances, reducing the execution time of test instances.

Lipton's algorithm executes in exponential time with  $\alpha \approx [4.2, 8.2]$  taking the longest. This is within the phase-transition region for 3-SAT.

The Distribution algorithm executes in exponential time, and performs better than Lipton's algorithm for low conflict ratios. However over the entire sweep performs worse than both Lipton's and Ogihara-Ray's algorithms. It shares the same  $\alpha \approx [4.2, 8.2]$  during the 3-SAT phase-transition.

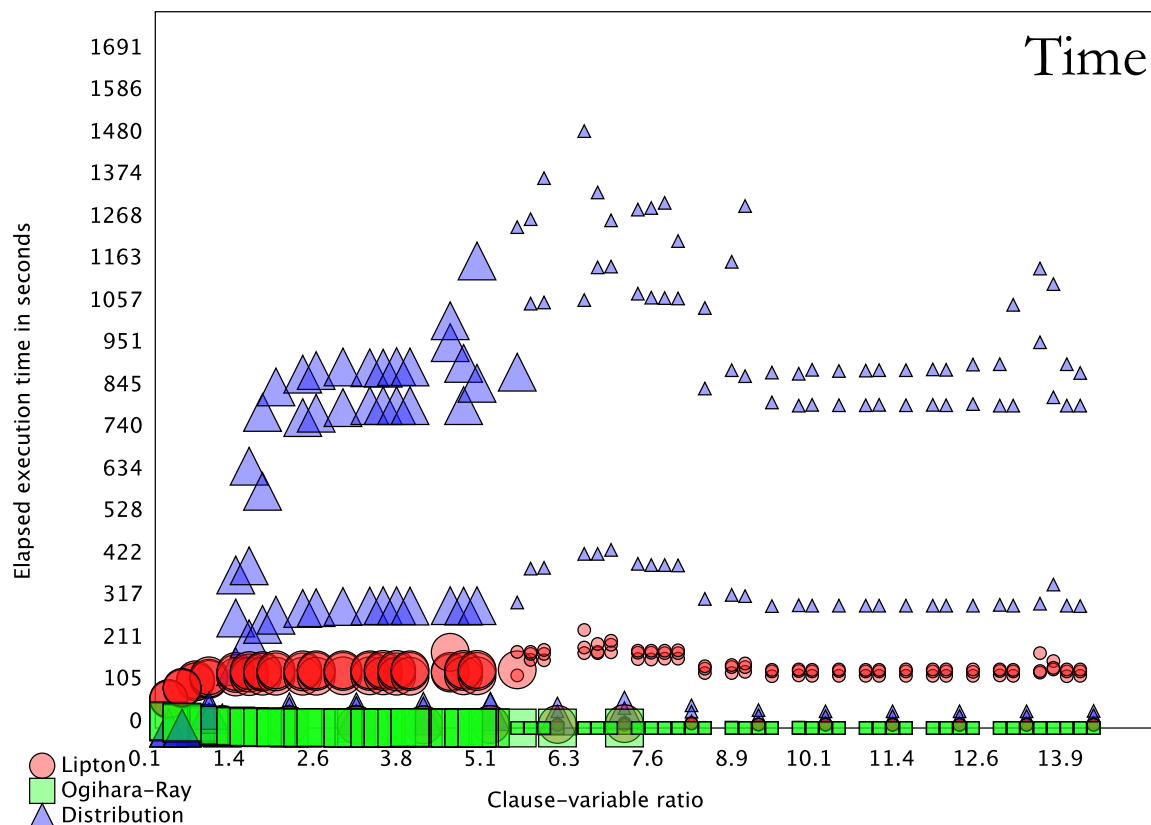


Figure 7.7: Clause to variable ratio  $\alpha$  vs. execution time in seconds

**Memory** is a measurement of the satisfiable instance footprint returned by each algorithm measured in Bytes.

Lipton's and Ogihara-Ray's algorithms share the same solution footprint.

The Distribution algorithm contains a larger solution footprint after the trivially satisfiable instances with  $\alpha \approx [0.2, 0.8]$ . The space provides a set of non-conflicting assignments from  $\alpha \approx [0.8, 2.9]$ . Non-conflicting assignments consist of witnesses for only necessary variables.

Each SATISFIABILITY instance has a constrained solution space during the phase-transition region. All three algorithms share the same footprint. There are no satisfiable instances in this test with  $\alpha > 7.2$ . The axis in Figure 7.8 scales accordingly.

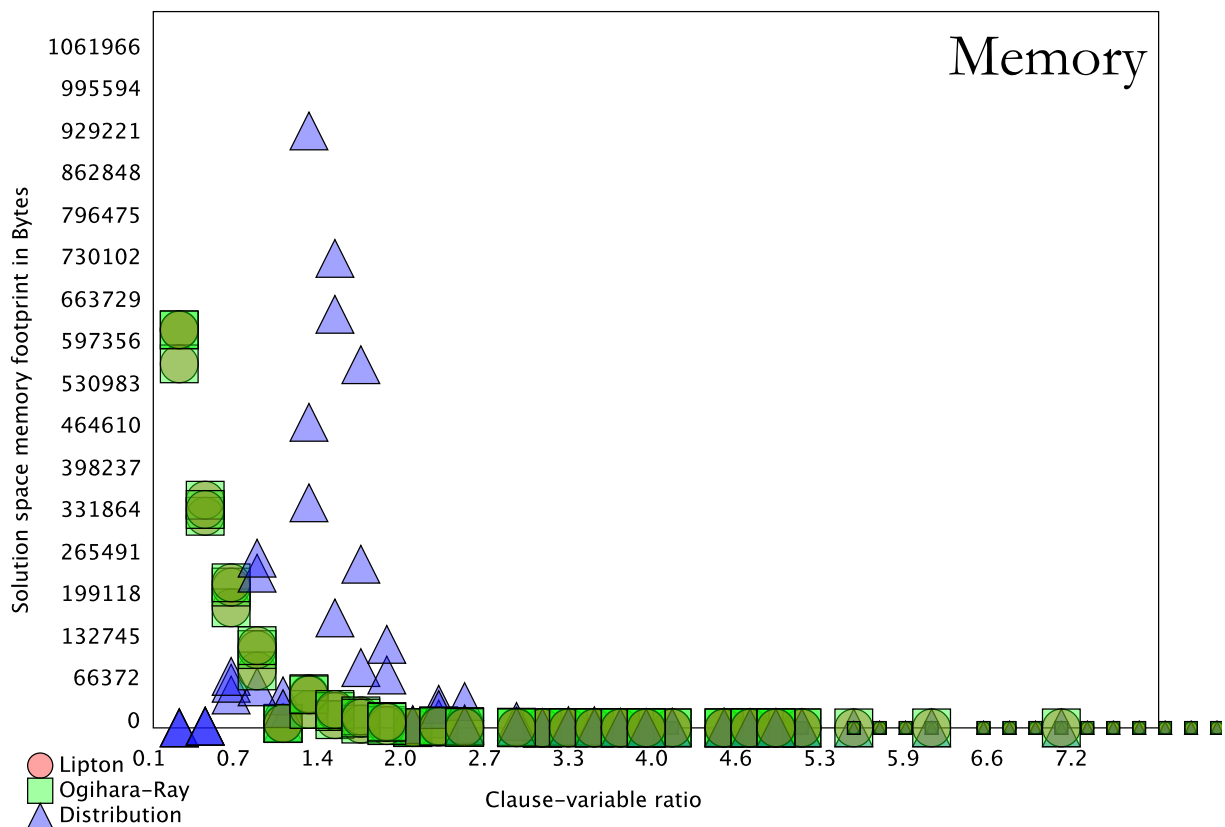


Figure 7.8: Clause to variable ratio  $\alpha$  vs. satisfiable solution footprint in Bytes

## Chapter 8

# Conclusions

This project considered SATISFIABILITY as a problem for general computation. We considered three molecular algorithms for SATISFIABILITY and simulated their execution with a conventional computing implementation. In this chapter, we state the contributions of this project and directions molecular computation will take.

### 8.1 Contributions

We developed several contributions for molecular computing during this project. This includes introducing the molecular Distribution algorithm for SATISFIABILITY in Chapter 4. We introduced Molecular Simulation in Chapter 5 and collected data from simulations of three molecular SATISFIABILITY algorithms described in Chapter 6.

### 8.2 Future work

Nanopore sequencers have been designed for reading molecules and diagnosing patients in a medical setting. Extending the sequencing capability to a configurable molecular

laboratory permits generalized computation.

SATISFIABILITY provides a canonical input format for combinatorial problems. Molecular algorithms for SATISFIABILITY may be constructed in gene sequencers designed as a configurable molecular laboratory.

# Bibliography

- [1] ADLEMAN, L. M. Molecular computation of solutions to combinatorial problems. *Science* 266 (November 1994), 1021–1024.
- [2] BALTIMORE, D. Expression of animal virus genomes. *Bacteriol Rev* 35, 3 (1971), 235–41.
- [3] COOK, S. A. The complexity of theorem-proving procedures. In *Proceedings of the third annual ACM symposium on Theory of computing* (New York, NY, USA, 1971), STOC '71, ACM, pp. 151–158.
- [4] DIMACS. SATISFIABILITY suggested format. Accessed from <ftp://dimacs.rutgers.edu/pub/challenge/satisfiability/>. DIMACS 1993. Accessed from <ftp://dimacs.rutgers.edu/pub/challenge/satisfiability/>. DIMACS 1993., May 1993.
- [5] DOHERTY, P., AND KVARNSTRÖM, J. *The Handbook of Knowledge Representation*. Elsevier, 2008.
- [6] FEYNMAN, R. There's Plenty of Room at The Bottom. Accessed from: <http://resolver.caltech.edu/CaltechES:23.5.0>. *Caltech Engineering and Science* 23, 5 (1960).

- [7] FRY, B. *Visualizing Data*. O'Reilly Media Inc., 2008.
- [8] FURKA, A. Study on possibilities of systematic searching for pharmaceutically useful peptides. *Notarized on May 29, 1982. Accessed from <http://szerves.chem.elte.hu/furka/>* (May 1982).
- [9] FURKA, A. *Combinatorial Chemistry Combinatorial Chemistry Principles and Techniques*. -, 2007.
- [10] GARAJ, S., HUBBARD, W., REINA, A., KONG, J., BRANTON, D., AND GOLOVCHENKO, J. A. Graphene as a subnanometre trans-electrode membrane. *Nature* 467, 7312 (Sept. 2010), 190–193.
- [11] GENT, I. P., AND WALSH, T. The SAT phase transition. In *ECAI* (1994), John Wiley & Sons, pp. 105–109.
- [12] IGNATOVA, Z., MARTINEZ-PEREZ, I., AND ZIMMERMAN, K.-H. *DNA Computing Models*. Springer, 2008.
- [13] LEVIN, L. Universal search problems (in Russian). *Problemy Peredachi Informatsii* 9, 3 (1973), 115–116.
- [14] LIFE TECHNOLOGIES. Ion Torrent. Accessed from <http://www.iontorrent.com/>.
- [15] LIPTON, R. Using DNA to solve NP-complete problems. *Science* 268 (1995), 542–545.
- [16] LOUGHRAN, M. IBM Research Aims to Build Nanoscale DNA Sequencer to Help Drive Down Cost of Personalized Genetic Analysis. Accessed from: <http://www-03.ibm.com/press/us/en/pressrelease/28558.wss>, October 2009.

- [17] MARTÍN-MATEOS, F., ALONSO, J. A., PEREZ-JIMENEZ, M., AND SANCHO-CAPARRINI, F. Molecular computation models in ACL2: a simulation of Lipton's experiment solving SAT, 2002.
- [18] OGIHARA, M. Breadth first search 3-SAT algorithms for DNA computers. Tech. rep., University of Rochester, Rochester, NY, USA, 1996.
- [19] OGIHARA, M., AND RAY, A. DNA-based parallel computation by "counting". Tech. rep., University of Rochester, 1997.
- [20] OXFORD NANOPORE TECHNOLOGIES. Oxford Nanopore Technologies. Accessed from <http://www.nanoporetech.com/>.
- [21] SATCOMP ORGANIZING COMMITTEE. The international SAT Competitions web page. Accessed from <http://satcompetition.org/>.
- [22] SIPSER, M. *Introduction to the Theory of Computation, Second Edition*. Course Technology, 2006.
- [23] STANKOVICH, S., DIKIN, D. A., DOMMETT, G. H. B., KOHLHAAS, K. M., ZIMNEY, E. J., STACH, E. A., PINER, R. D., NGUYEN, S. T., AND RUOFF, R. S. Graphene-based composite materials. *Nature* 442, 7100 (2006), 282–6.
- [24] WILSON, D. Random  $k$ -SAT generator. Accessed from: <http://research.microsoft.com/en-us/um/people/dbwilson/ksat/default.htm> (2011).
- [25] YOSHIDA, H., AND SUYAMA, A. Solution to 3-SAT BY BREADTH FIRST SEARCH. IN *DNA Based Computers V* (2000), E. WINFREE AND D. GIFFORD, EDS., VOL. 54 OF *DIMACS: Series in Discrete Mathematics and Theoretical Computer Science*, PP. 9–22.

- [26] ZHANG, W. PHASE TRANSITIONS AND BACKBONES OF 3-SAT AND MAXIMUM 3-SAT.  
IN *Principles and Practice of Constraint Programming — CP 2001*, T. WALSH, ED.,  
VOL. 2239 OF *Lecture Notes in Computer Science*. SPRINGER BERLIN / HEIDELBERG,  
2001, PP. 153–167.



# Appendix A

## Source

### A.1 Contributed

Download Molecular Simulation:

- <https://github.com/dncarley/MolecularSimulation>
- Documentation
  - Online Documentation:
    - \* <http://www.cs.rit.edu/~dnc6813/project/generatedDocs/index.html>
  - Offline Documentation:
    - \* <http://www.cs.rit.edu/~dnc6813/project/refman.pdf>

Download SAT Datapoints Visualization:

- <https://github.com/dncarley/VisualizeSatDatapoints>

## A.2 External

Download David Wilson's  $k$ -SAT Generator:

- <http://research.microsoft.com/en-us/um/people/dbwilson/ksat/default.htm>

Download Doxygen:

- <http://www.stack.nl/~dimitri/doxygen/>

Download Ben Fry's examples for *Visualizing Data*:

- <http://benfry.com/writing/archives/3>

## Appendix B

# Molecular algorithm trace

### B.1 Example SATISFIABILITY instance

$$\phi = (x_1 \vee x_2 \vee \neg x_3) \wedge (x_2 \vee x_3 \vee \neg x_4) \wedge (\neg x_1 \vee \neg x_3 \vee \neg x_4)$$

### B.2 Lipton's Algorithm

$$T = \text{COMBINATORIAL GENERATE}(4)$$

$$T =$$

TTTT FTTT TFTT FFTT TTFT FTFT TFFT FFFT

TTTF FTTF TTF FTF TTFF FTFF TFFF FFFF

Next select Clause 1:

$$C_1 = (x_1 \vee x_2 \vee \neg x_3)$$

TTTT FTTT TFTT FTTT TTFT FTFT TFFT FFFT  
 TTTF FTTF TFTF FTF FTF TTF TFF FFF

Extract  $x_1$ :

TTTT TFTT TTFT TFFT  
 TTTF TTF TTF TFF

Extract  $x_2$ :

TTTT FTTT TTFT FTFT  
 TTTF FTF TTF FTF

Extract  $\neg x_3$ :

TTFT FTFT TFFT FFFT  
 TTF FTF TFF FFF

Mix contents:

TTTT FTTT TFTT TTFT FTFT TFFT FFFT  
 TTTF FTF TTF FTF TTF FFF

Next select Clause 2:

$$C_2 = (x_2 \vee x_3 \vee \neg x_4)$$

TTTT FTTT TFTT TTFT FTFT TFFT FFFT  
 TTTF FTF TTF FTF TTF FFF

Extract  $x_2$ :

TTTT FTTT	TTFT FTFT
TTTF FTTF	TTFF FTFF

Extract  $x_3$ :

TTTT FTTT TFFT
TTTF FTTF TFTF

Extract  $\neg x_4$ :

TTTF FTTF TFTF	TTFF FTFF TFFF FFFF
----------------	---------------------

Mix contents:

TTTT FTTT TFFT	TTFT FTFT
TTTF FTTF TFTF	TTFF FTFF TFFF FFFF

Finally, select Clause 3:

$$C_3 = (\neg x_1 \vee \neg x_3 \vee x_4)$$

TTTT FTTT TFFT	TTFT FTFT
TTTF FTTF TFTF	TTFF FTFF TFFF FFFF

Extract  $\neg x_1$ :

FTTT	FTFT	
FTTF	FTFF	FFFF

Extract  $\neg x_3$ :

TTFT FTFT
TTFF FTFF TFFF FFFF

Extract  $x_4$ :

TTTT FTTT TFTT      TTFT FTFT

Mix contents:

TTTT FTTT TFTT      TTFT FTFT  
 FTTF      TTFF FTFF TFFF FFFF

### B.3 Ogihara and Ray's Algorithm

Initialize the tube  $T$  with initial vector assignments for variables  $x_1$  and  $x_2$

$$T = \{\text{TT}, \text{TF}, \text{FT}, \text{FF}\}$$

Iterate variable  $x_3$ :

$$C_1 = (x_1 \vee x_2 \vee \neg x_3)$$

$\neg x_3$  matches  $v_3$

$$T_{P1} = \{\text{TT}, \text{TF}\}$$

$$T_{N1} = \{\text{FT}, \text{FF}\}$$

$$T_{P2} = \{\text{FT}\}$$

$$T_P = \{\text{TT}, \text{TF}, \text{FT}\}$$

$$C_2 = (x_2 \vee x_3 \vee \neg x_4)$$

$x_3$  or  $\neg x_3$  does not match  $v_3$

$$C_3 = (\neg x_1 \vee \neg x_3 \vee x_4)$$

$x_3$  or  $\neg x_3$  does not match  $v_3$

Append

$$T_P = \{\text{TTT}, \text{TFT}, \text{FTT}\}$$

$$T_N = \{\text{TTF}, \text{TFF}, \text{FTF}, \text{FFF}\}$$

Mix

$$T = \{\text{TTT}, \text{TFT}, \text{FTT}, \text{TTF}, \text{TFF}, \text{FTF}, \text{FFF}\}$$

Iterate variable  $x_4$ :

$$C_1 = (x_1 \vee x_2 \vee \neg x_3)$$

$x_4$  or  $\neg x_4$  does not match  $v_3$

$$C_2 = (x_2 \vee x_3 \vee \neg x_4)$$

$\neg x_4$  matches  $v_3$

$$T_{P1} = \{\text{TTT}, \text{FTT}, \text{TTF}, \text{FTF}\}$$

$$T_{N1} = \{\text{TFT}, \text{TFF}, \text{FFF}\}$$

$$T_{P2} = \{\text{TFT}\}$$

$$T_P = \{\text{TTT}, \text{FTT}, \text{TTF}, \text{FTF}, \text{TFT}\}$$

$$C_3 = (\neg x_1 \vee \neg x_3 \vee x_4)$$

$x_4$  matches  $v_3$

$$T_{P1} = \{\text{FTT}, \text{FTF}, \text{FFF}\}$$

$$T_{N1} = \{\text{TTT}, \text{TFT}, \text{TTF}, \text{TFF}\}$$

$$T_{P2} = \{\text{TTF}, \text{TFF}\}$$

$$T_N = \{\text{FTT}, \text{FTF}, \text{FFF}, \text{TTF}, \text{TFF}\}$$

Append

$$T_P = \{\text{TTT}, \text{TFT}, \text{FTT}\}$$

$$T_N = \{\text{TTF}, \text{TFF}, \text{FTF}, \text{FFF}\}$$

Mix



$$T = \{\text{TTT}, \text{TFT}, \text{FTT}, \text{TTF}, \text{TFF}, \text{FTF}, \text{FFF}\}$$

## B.4 Distribution Algorithm

Initialize the tube  $T$  with the variables from the first clause

$$T = \{[1], [2], [-3]\}$$

Select Clause 2

$$T_1 = \text{INSERTVARIABLE}(T, 2)$$

$$T_1 = \{[1, 2], [2], [2, -3]\}$$

$$T_2 = \text{INSERTVARIABLE}(T, 3)$$

$$T_1 = \{[1, 3], [2, 3]\}$$

$$T_3 = \text{INSERTVARIABLE}(T, -4)$$

$$T_3 = \{[1, -4], [2, -4], [-3, -4]\}$$

$$T = \text{mix}(T_1, T_2, T_3)$$

$$T = \{[1, 2], [2], [2, -3], [1, 3], [2, 3], [1, -4], [2, -4], [-3, -4]\}$$

Select Clause 3

$$T_1 = \text{INSERTVARIABLE}(T, -1)$$

$$T_1 = \{[-1, 2], [-1, 2, -3], [-1, 2, 3], [-1, 2, -4], [-1, -3, -4]\}$$

$$T_2 = \text{INSERTVARIABLE}(T, -3)$$

$$T_2 = \{[1, 2, -3], [2, -3], [2, -3], [1, -3, -4], [2, -3, -4], [-3, -4]\}$$

$$T_2 = \text{INSERTVARIABLE}(T, 4)$$

$$T_3 = \{[1, 2, 4], [2, 4], [2, -3, 4], [1, 3, 4], [2, 3, 4]\}$$

$$T = \text{mix}(T_1, T_2, T_3)$$

$$\begin{aligned} T = \{ & [-1, 2], [-1, 2, -3], [-1, 2, 3], [-1, 2, -4], [-1, -3, -4], \\ & [1, 2, -3], [2, -3], [1, -3, -4], [2, -3, -4], [-3, -4], \\ & [1, 2, 4], [2, 4], [2, -3, 4], [1, 3, 4], [2, 3, 4] \} \end{aligned}$$