Supplementary material

Chromatin looping and the probability of transcription

Qiliang Li, Gráinne Barkess and Hong Qian

Division of Medical Genetics, Department of Medicine, Department of Applied Mathematics, University of Washington, Seattle, WA 98195, USA

Corresponding author: Li, Q. (li111640@u.washington.edu).

This was written by H. Qian as an appendix to the article by Li et al., published in the April 2006 issue of *Trends in Genetics*.

According to the worm-like chain model of polymers, the end-to-end distance (n) of a polymer chain follows the probability distribution [1,2]:

$$P_{wc}(r; L, \ell_p) 4\pi r^2 dr = \frac{3\sqrt{3}r^2 dr}{2\sqrt{\pi} (L\ell_p)^{3/2}} e^{-3r^2/4L\ell_p} (1 - w(r, L, \ell_p))$$

where

$$w(r,L,\ell_p) = \frac{5\ell_p}{4L} - \frac{2r^2}{L^2} + \frac{33r^4}{80L^3\ell_p} + \frac{79\ell_p^2}{160L^2} + \cdots$$

in which L is the contour length of the polymer, and ℓ_p is the persistence length of the polymer. According to the Jacobson-Stockmayer theory of loop formation [3], the probability of forming a loop of contour length L is proportional to

$$P_{loop}(L; \ell_p) \propto \left(\frac{\ell_p}{L}\right)^{3/2} - \frac{5}{4} \left(\frac{\ell_p}{L}\right)^{5/2} - \frac{79}{160} \left(\frac{\ell_p}{L}\right)^{7/2}$$
 (Eqn 1)

Figure 1 shows P_{loop} as functions of L/l_p and Eqn 1, which is essentially the second equation of Ringrose et~al. [4], and is valid for both long and short polymer chains. The optimal length of loop formation increases with the persistence length. One can find the optimal ring length r* from Eqn 1 by differentiation $dP_{loop}/dL=0$: $r*=2.537\ell_p$.

This model deals with a polymer with free ends. In the present application, however, the two ends of a section of chromatin in the middle of the genome are not free. Hence, we have the formation of 'a loop within a loop' (Figure 2a) with a probability of $P_{loop}(L, \ell_p) P_{loop}(L_T - L, \ell_p)$, where LT is the total contour length of the section of chromatin. That is:

$$\left\{ \left(\frac{\ell_p}{L} \right)^{3/2} - \frac{5}{4} \left(\frac{\ell_p}{L} \right)^{5/2} - \frac{79}{160} \left(\frac{\ell_p}{L} \right)^{7/2} \right\} \left\{ \left(\frac{\ell_p}{L_T - L} \right)^{3/2} - \frac{5}{4} \left(\frac{\ell_p}{L_T - L} \right)^{5/2} - \frac{79}{160} \left(\frac{\ell_p}{L_T - L} \right)^{7/2} \right\}$$

(Eqn 2). Figure 2b shows several curves for different persistence lengths.

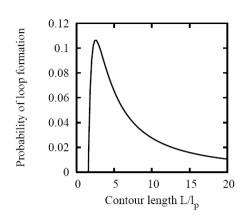


Figure 1. Probability distribution for the loop formation (Eqn 1) with contour length L, measured in terms of the persistence length lp. When the flexibility is constant, the probability of loop formation is determined by the distance between two points.

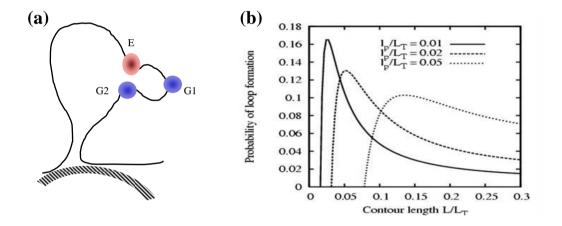


Figure 2. Formation of a loop within a loop. (a) A schematic diagram of loop formation within a section of chromatin anchoring at a nuclear substructure. An enhancer (E) is required to activate the cognate genes G1 and G2 (round). (b) Probability distributions for the loop formation (Eqn 2) within a section of chromatin with several different values for the persistence length. The *x*-axis is expressed in terms of the ratio of lp/LT. When LT is constant, the curves show the probability of loop formation between different distances at a given flexibility.

Acknowledgements
We thank H-X.Zhou for useful discussions.

References

- 1 Zhou, H.-X. (2004) Loops, linkages, rings, catenanes, cages, and crowders: entropy-based strategies for stabilizing proteins. Acc. Chem. Res. 37, 123-130.
- 2 Gobush, W. et al. (1972) Statistical mechanics of wormlike chains: 1 Asymptotic behavior. J. Chem. Phys. 57, 2839-2843
- 3 Yamakawa, H. (1971) Modern Theory of Polymer Solutions, Harper & Row
- 4 Ringrose, L. et al. (1999). Quantitative comparison of DNA looping in vitro and in vivo: chromatin increases effective DNA flexibility at short distances. Embo J. 18, 6630-6641