

FluoroAnalysisTool README

Description of code directory:

- 'data' is intended to be example analysis. TBD.
- 'doc' is intended to be instructions/manual. TBD.
- 'lib' contains source-code downloaded from the internet to serve abstract purposes within the FluoroAnalysisTool.
- 'Projects' is intended to be space for the user to save their project state. TBD.
- 'src'
 - 00-Startup
 - Custom Data object classes for fluoros, types of bones,
 - GUI building routines, i.e. the "View" in Model-View-Controller paradigms.
 - main file ('main.m')
 - 01-Data_Management
 - Geometry and conversion routines for fluoro/image data.
 - Scripts for various tasks, which should eventually be included into a larger interface TBD.
 - 02-Procedures
 - Custom data object classes for the types of annotations that we do (e.g. DHS, PSHF).
 - 03-Neural_Net_Models (TBD)
 - Code for socketing MATABL to python and applying computer vision models.
 - 04-Non_GUI_Dependent_Scripts (TBD)
 - Niche scripts that perform tasks relevant to the interface, which should eventually be included into a larger interface TBD.
 - Image Processing algorithm for computing a precise wire width. Not sure if I ever implemented this TBD.
 - 05-Decision_Error_Processing (TBD)
 - Trevor's code for PSHF decision errors.
 - Helpers
 - Misc. code that should probably be moved to one of the previously-listed folders TBD.

Steps for using this tool:

0. Make sure that you have properly imported the dicom exam.
 - R:\Anderson_Colaborations\AHRQ - 11287500\Fluoroscopy
 - Open and run 'OpenMe.m'. Read the instructions for importing a case.
1. Open main.m
 - R:\Anderson_Colaborations\AHRQ - 11287500\FluoroAnalysisTool\src\00-Startup\main.m
2. Click 'Run' (or hit F5 when the main.m script is open and selected in the MATLAB interface.

- This will likely prompt MATLAB to ask you whether you want to change the directory.
 - Say/click 'Change Folder'.
- The FluoroAnalysisTool may take a second to start the first time.
- Pay attention for a prompt that asks whether you want to use AI tools.
 - Say/click 'no'.

3. Loading imported Data.

- Click 'Load Directory', select your DICOM folder.
- Click on one of the files in the folder.
- Then click 'Plot'.

4. Annotating data.

- In the 'Procedure' Panel, flick the drop-down and select your procedure.
 - Buttons for the annotation procedure should appear
 - I don't know if the DHS procedure still works, as of Nov 1, 2020.
- For PSHF:
 - Click 'Fracture Plane'
 - Left click two points to approximate the 3D plane in the 2D image.
 - Right click to confirm. Note that clicking more than 2 points may cause an error. If that happens, just try again. Worst case, restart the tool.
 - Click 'Wire N'
 - Beginning with the tip of the wire (pointy end), left click down the *centerline* of the wire.
 - Sometimes you only need 2 points.
 - When the wire is bent by the surgeon (so they can put the cast on), do not include this part of the wire.
 - Once you've traversed the length of the wire, (left) click one additional point *at the edge of the wire, perpendicular to your final centerpoint line*.
 - Right click to confirm.
 - Your "wire" should have at minimum 3 points, including the width point.
 - If you've done it correctly, the plot will show a rough outline of the width of the wire.
 - This won't be perfect, but that's ok. All we care about is the centerline for this procedure.
 - If you don't like where the algorithm roughly places the centerline, just click the same button again and redo it.
 - Repeat this until all 3 wires are defined.
 - Order *might* not be important (I think I have some code that deduces the left-to-right order of the wires along the fracture line), but just to be safe, make Wire 2 the middle wire.
 - When you have successfully defined all 3 wires and the fracture line, the 'Log' Panel will print off the relevant metrics.
 - Breadth of wires at fracture, mid-ratio
 - Make note of these in your excel sheet.

- For lateral images, since we are only concerned with the breadth between the lateral and medial wires, feel free to skip annotating 'Wire 2' on these images.
 - If you annotate all 3 wires, the data will be saved, but the corresponding Mid-Ratio is not something that we are currently interested in using for our analysis.

-----Ignore below this, as of Nov 1, 2020-----

Created Using:

<https://dillinger.io/>

Description

FluoroAnalysisTool is a MATLAB-built UI application for annotating DICOM image data.

- It currently supports *DHS* and *PSHF* surgeries.

New Features

- None haha. Hopefully some AI CV stuff for automatic feature detection.

LAST UPDATED: 09 SEPT. 2020

Installation

- Matlab 2018a
 - Originally written.
 - **May not work on earlier versions!**
 - Toolboxes:
 - ...
- Python 3.7.3
 - Installed via command line
 - Default PATH variable.
 - Libraries:
 - TensorFlow 1.14
 - Keras 2.24
 - h5py 2.80 (forced)
 - Numpy 1.16

Startup Routine

1. Navigate to the 'main.m' file [here](..\src\00-Startup\main.m)
2. Run script (F5) to use GUI.

Getting Started

1. Click the 'Project Directory' folder.
 - This is where your 'Results.json' text files will be saved.
 - If you do not specify a folder, the results will be saved somewhere else...I forget where. ***edit this***
2. Click the 'Login' Button on the toolbar toward the top-left
 - Enter your info

- This is optional, but helpful because it is printed in the Results.json files that you produce.

3. Load a directory of DICOMs
 - .dcm
4. Plot File
5. Select a procedure
6. Follow the instructions in the log
 - Bottom right.

Helpful Hints

- In the event of an error:
 - Screenshot both the UI and the MATLAB command window
 - Email Dominik with a description of what you clicked and what you were trying to do prior to the error.
 - Try restarting MATLAB and the application and try again. I ain't a perfect programmer, dawg.
- As of **September 2020** I (Dominik) disabled the AI stuff because of complications with socketing to the Python script that runs the neural network models.
 - If you know how to socket between MATLAB and Python and you know how to hide command prompts from the user, contact Dominik.
- If you are doing the procedure correctly, there will not be any red text.
 - Red text in the Log is not fatal to the program.
- The application should autosave your data in your Project Directory as '###_Results.json'