EDA of C. elegans dataset

Domenick Braccia 2/26/2019

Overview

This document will serve as an exploratory data analysis of data from this publication by Asija Diag et al.

Loading & munching data

Overview of the data present in each .xlsx file:

- S1: Hermaphrodite expression levels (gonads #1 & #2)
- S2: Hermaphrodite expression levels (normalized and averaged accross replicates see M&M)
- S3: Hermaphrodite dynamic transcript groups
- S4: Hermaphrodite dynamic transcripts (Profile I)
- S5: Analysis of comparison with NEXTDB
- S6: expression levels of X linked genes in hermaphrodites
- S8 : expression levels / transcriptional rates in hermaphrodites
- S9: Male transcript counts (gonads 1 and 2)
- S10 : Male transcript expression levels
- S11: transcript to gene name table

```
# read in necessary libraries
library(readxl)
library(tidyverse)
library(skimr)

### Importing and cleaning tables S1,S2,S8,S9,S10,S11 ###

# importing S1 - hermaphrodite expression table
hermExpNad <- read_excel("../data/S1_Table.xlsx", skip = 3)
colnames(hermExpNad) <- c("ORF", 1:10, 1:10, "WBGene")
glimpse(hermExpNad)</pre>
```

```
## Observations: 20,779
## Variables: 22
   <chr> "2L52.1", "2RSSE.1", "2RSSE.2", "3R5.1", "4R79.1", "4R7...
## $ ORF
## $ `1`
   ## $ `2`
   ## $ `3`
   <dbl> 0, 0, 0, 12, 0, 0, 0, 0, 0, 1, 0, 0, 0, 0, 0, 0, ...
## $ `4`
   ## $ `5`
   ## $ `6`
   ## $ `7`
   ## $ `8`
   ## $ `9`
   ## $ `10`
   ## $ `1`
   ## $ `2`
   ## $ `3`
```

```
## $ `4`
          ## $ `5`
          ## $ `6`
          ## $ `7`
          ## $ `8`
          ## $ `9`
          ## $ `10`
          <dbl> 0, 0, 0, 1, 0, 0, 0, 0, 0, 0, 2, 0, 0, 0, 0, 0, 0, 0, 2...
## $ WBGene <chr> "WBGene00007063", "WBGene00007064", "WBGene00044165", "...
# S2 Herm exp levels; normalized and averaged
hermExpNormed <- read_excel("../data/S2_Table.xlsx", skip = 3)
colnames(hermExpNormed)[1] <- "ORF"</pre>
colnames(hermExpNormed)[17] <- "Gene_Name"</pre>
glimpse(hermExpNormed)
## Observations: 20,687
## Variables: 18
             <chr> "2L52.1", "2RSSE.1", "2RSSE.2", "3R5.1", "4R79.1", ...
## $ ORF
## $ `1`
             <dbl> 0.0000000, 0.0000000, 0.0000000, 1.1285917, 0.00000...
## $ `2`
             <dbl> 0.000000, 0.000000, 0.000000, 4.934989, 0.000000, 0...
             <dbl> 0.0000000, 0.0000000, 0.0000000, 3.9633200, 0.00000...
## $ `3`
## $ `4`
             <dbl> 0.0000000, 0.0000000, 0.0000000, 3.4258657, 0.00000...
## $ `5`
             <dbl> 0.0000000, 0.0000000, 0.0000000, 2.2290508, 0.00000...
## $ `6`
             <dbl> 0.5280138, 0.0000000, 0.0000000, 2.7463885, 0.00000...
## $ `7`
             <dbl> 0.9536161, 0.0000000, 0.0000000, 0.4751858, 0.00000...
## $ `8`
             <dbl> 0.000000, 0.000000, 0.000000, 2.373090, 0.000000, 0...
## $ `9`
             <dbl> 0.0000000, 0.0000000, 0.0000000, 1.4237781, 0.00000...
## $ `10`
             <dbl> 0.000000, 0.000000, 0.000000, 1.197841, 0.000000, 0...
## $ Chromosome <chr> "CHROMOSOME II", "CHROMOSOME II", "CHROMOSOME II", ...
## $ Start
             <dbl> 1867, 15268136, 15274315, 13780127, 17486946, 17480...
## $ End
             <dbl> 4663, 15273238, 15275613, 13781032, 17490461, 17483...
             ## $ Strand
             <chr> "WBGene00007063", "WBGene00007064", "WBGene00044165...
## $ WBGene
## $ Gene_Name
             <chr> NA, NA, NA, "pot-3", "nas-6", NA, NA, "sri-20", "sp...
## $ Type
             <chr> "cds", "cds", "cds", "cds", "cds", "cds", "cds", "c...
# S8
hermRates <- read_excel("../data/S8_Table.xlsx", skip = 3)
colnames(hermRates) <- c("ORF", "Gene Name", "WormBase ID", "Chromosome", "Strand", "Ribo rep1", "Ribo :
glimpse(hermRates)
## Observations: 19,977
## Variables: 9
## $ ORF
              <chr> "B0019.1", "B0019.2", "B0025.1a", "B0025.2", "B002...
              <chr> "amx-2", NA, "vps-34", "csn-2", NA, NA, NA, NA, "a...
## $ Gene_Name
## $ WormBase_ID <chr> "WBGene00000138", "WBGene00007094", "WBGene0000693...
              ## $ Chromosome
## $ Strand
              <chr> "-", "+", "+", "+", "-", "+", "-", "+", "-", "+", ...
              <dbl> 6.5816939, 5.1639885, 13.8514375, 70.1363533, 19.9...
## $ Ribo_rep1
              <dbl> 4.4000493, 4.8461051, 13.9819546, 100.0447569, 34....
## $ Ribo_rep2
## $ mRNA rep1
              <dbl> 30.1078502, 86.9022041, 119.9181990, 321.2644476, ...
              <dbl> 18.9364571, 120.9614503, 137.9656158, 365.9760174,...
## $ mRNA_rep2
# S9
maleExpNad <- read_excel("../data/S9_Table.xlsx", skip = 3)</pre>
colnames(maleExpNad) <- c("ORF", 1:10, 1:10, "WBGene")</pre>
```

```
glimpse(maleExpNad)
## Observations: 20,779
## Variables: 22
         <chr> "2L52.1", "2RSSE.1", "2RSSE.2", "3R5.1", "4R79.1", "4R7...
## $ ORF
         ## $ `1`
## $ `2`
         ## $ `3`
         ## $ `4`
         <dbl> 0, 0, 0, 3, 0, 0, 0, 0, 2, 1, 0, 0, 0, 0, 0, 0, 0, 0, 0...
## $ `5`
         <dbl> 0, 0, 0, 1, 0, 0, 0, 0, 19, 5, 0, 0, 0, 0, 0, 0, 0, 0, ...
## $ `6`
         <dbl> 0, 0, 0, 0, 0, 0, 0, 10, 4, 0, 0, 0, 0, 0, 0, 0, ...
## $ `7`
         <dbl> 0, 0, 0, 0, 0, 0, 0, 13, 1, 0, 0, 0, 0, 0, 0, 0, ...
## $ `8`
         <dbl> 0, 0, 0, 0, 0, 0, 0, 0, 6, 0, 2, 0, 0, 0, 0, 0, 1, 2...
## $ `9`
         <dbl> 0, 0, 0, 0, 0, 0, 0, 0, 1, 0, 0, 0, 0, 0, 0, 0, 0...
## $ `10`
         ## $ `1`
         ## $ `2`
         ## $ `3`
         ## $ `4`
         <dbl> 0, 0, 0, 7, 0, 0, 0, 0, 0, 1, 0, 0, 0, 0, 0, 0, 0, 0, 0...
## $ `5`
         <dbl> 0, 0, 0, 6, 0, 0, 0, 0, 14, 4, 0, 0, 0, 0, 0, 0, 0, 0, ...
## $ `6`
         <dbl> 0, 0, 0, 0, 0, 0, 0, 13, 5, 0, 0, 0, 0, 0, 0, 0, ...
## $ `7`
         <dbl> 0, 0, 0, 0, 0, 0, 0, 0, 9, 2, 0, 0, 0, 0, 0, 0, 0, 0...
## $ `8`
         <dbl> 0, 0, 0, 0, 0, 0, 0, 15, 2, 0, 0, 0, 0, 0, 0, 0, ...
## $ `9`
         <dbl> 0, 0, 0, 0, 0, 0, 0, 0, 6, 0, 0, 0, 0, 0, 0, 0, 0, 0...
## $ `10`
         ## $ WBGene <chr> "WBGene00007063", "WBGene00007064", "WBGene00044165", "...
maleExpNormed <- read_excel(".../data/S10_Table.xlsx", skip = 3)</pre>
colnames(maleExpNormed)[1] <- "ORF"; colnames(maleExpNormed)[colnames(maleExpNormed) == "Gene Name"] <-
glimpse(maleExpNormed)
## Observations: 20,687
## Variables: 19
            <chr> "2L52.1", "2RSSE.1", "2RSSE.2", "3R5.1", "4R79.1", ...
## $ ORF
## $ `1`
            <dbl> 0.000000, 0.000000, 0.000000, 4.265513, 0.000000, 0...
## $ `2`
            <dbl> 0.0000000, 0.0000000, 0.0000000, 5.2675156, 0.00000...
            <dbl> 0.0000000, 0.0000000, 0.0000000, 4.8864558, 0.00000...
## $ `3`
## $ `4`
            <dbl> 0.0000000, 0.0000000, 0.0000000, 3.1359046, 0.00000...
## $ `5`
            <dbl> 0.000000, 0.000000, 0.000000, 1.848302, 0.000000, 0...
## $ `6`
            <dbl> 0.0000000, 0.0000000, 0.0000000, 0.0000000, 0.00000...
## $ `7`
            <dbl> 0.000000, 0.000000, 0.000000, 0.000000, 0.000000, 0...
## $ `8`
            <dbl> 0.0000000, 0.0000000, 0.0000000, 0.0000000, 0.00000...
            <dbl> 0.000000, 0.000000, 0.000000, 0.000000, 0.000000, 0...
## $ `9`
## $ `10`
            <dbl> 0.0000000, 0.0000000, 0.0000000, 2.6478937, 0.00000...
## $ diffExp
## $ Chromosome <chr> "CHROMOSOME_II", "CHROMOSOME_II", "CHROMOSOME_II", ...
```

<dbl> 1867, 15268136, 15274315, 13780127, 17486946, 17480...

<chr> "WBGene00007063", "WBGene00007064", "WBGene00044165...

<chr> NA, NA, NA, "pot-3", "nas-6", NA, NA, "sri-20", "sp...
<chr> "cds", "cds",

\$ Start ## \$ End

\$ Strand

\$ WBGene
\$ Gene_Name

\$ Type

To this point, I have imported and cleaned up most of the data I think we need. Some further steps may need to be taken to process data, but that can be done on an as needed basis.