# Lab13: Transcriptomics and the analysis of RNA-Seq data

Duy An Le (PID: A16400411)

The data from today's lab comes from a published RNA-seq experiment where airway smooth muscle cells were treated with dexamethasone, a synthetic glucocorticoid steroid with anti-inflammatory effects.

# **Import Data**

We need the raw counts and metadata for DESeq2.

```
counts <- read.csv("airway_scaledcounts.csv", row.names=1)
metadata <- read.csv("airway_metadata.csv")
head(counts)</pre>
```

S	SRR1039508	SRR1039509	SRR1039512	SRR1039513	SRR1039516
ENSG00000000003	723	486	904	445	1170
ENEGOCOCOCOCO	125	400	304	440	1170
ENSG0000000005	0	0	0	0	0
ENSG00000000419	467	523	616	371	582
ENSG00000000457	347	258	364	237	318
ENSG00000000460	96	81	73	66	118
ENSG00000000938	0	0	1	0	2
S	SRR1039517	SRR1039520	SRR1039521		
ENSG00000000003	1097	806	604		
ENSG00000000005	0	0	0		
ENSG00000000419	781	417	509		
ENSG00000000457	447	330	324		
ENSG00000000460	94	102	74		
ENSG00000000938	0	0	0		
			· -		

#### head(metadata)

```
id dex celltype geo_id
1 SRR1039508 control N61311 GSM1275862
2 SRR1039509 treated N61311 GSM1275863
3 SRR1039512 control N052611 GSM1275866
4 SRR1039513 treated N052611 GSM1275867
5 SRR1039516 control N080611 GSM1275870
6 SRR1039517 treated N080611 GSM1275871
```

The counts are organized with one gene per row and experiment per column.

```
nrow(counts)
[1] 38694
sum(metadata$dex=="control")
```

# [1] 4

#### Check on match of metadata and coldata names

```
all(colnames(counts) == metadata$id)
[1] TRUE
```

### **Analysis**

I want to start by comparing "control" and "treated" columns. To do this, I will find the average for each gene (row) in all "control" columns. Then I will find the average in the "treated' columns. Then I will compare them.

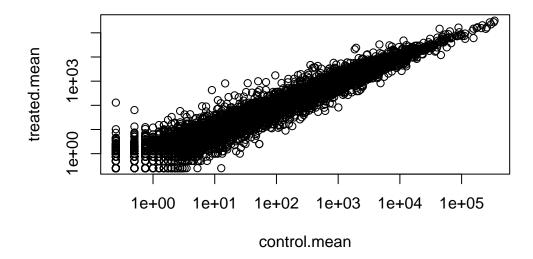
```
library(dplyr)
```

Attaching package: 'dplyr'

```
The following objects are masked from 'package:stats':
    filter, lag
The following objects are masked from 'package:base':
    intersect, setdiff, setequal, union
  control <- metadata %>% filter(dex=="control")
  control.counts <- counts %>% select(control$id)
  control.mean <- rowSums(control.counts)/4</pre>
  treated <- metadata %>% filter(dex=="treated")
  treated.counts <- counts %>% select(treated$id)
  treated.mean <- rowSums(treated.counts)/4</pre>
  meancounts <- data.frame(control.mean, treated.mean)</pre>
  head(meancounts)
                control.mean treated.mean
                                    658.00
ENSG00000000003
                      900.75
ENSG00000000005
                        0.00
                                     0.00
                                    546.00
ENSG00000000419
                      520.50
ENSG00000000457
                      339.75
                                    316.50
ENSG00000000460
                       97.25
                                     78.75
                                      0.00
ENSG00000000938
                        0.75
"Normalize" the data
  colSums(meancounts)
control.mean treated.mean
    23005324
                 22196524
Create the initial scatter plot
  plot(meancounts, log="xy")
Warning in xy.coords(x, y, xlabel, ylabel, log): 15032 x values <= 0 omitted
```

from logarithmic plot

Warning in xy.coords(x, y, xlabel, ylabel, log): 15281 y values <= 0 omitted from logarithmic plot



## Calculate $\log 2 \mathrm{fc}$ of mean counts

meancounts\$log2fc <- log2(meancounts[,"treated.mean"]/meancounts[,"control.mean"])
head(meancounts)</pre>

	${\tt control.mean}$	${\tt treated.mean}$	log2fc
ENSG0000000003	900.75	658.00	-0.45303916
ENSG0000000005	0.00	0.00	NaN
ENSG00000000419	520.50	546.00	0.06900279
ENSG00000000457	339.75	316.50	-0.10226805
ENSG00000000460	97.25	78.75	-0.30441833
ENSG00000000938	0.75	0.00	-Inf

Filter out genes with 0 expression

```
zero.vals <- which(meancounts[,1:2]==0, arr.ind=TRUE)
to.rm <- unique(zero.vals[,1])</pre>
```

```
mycounts <- meancounts[-to.rm,]
head(mycounts)</pre>
```

	${\tt control.mean}$	${\tt treated.mean}$	log2fc
ENSG0000000003	900.75	658.00	-0.45303916
ENSG00000000419	520.50	546.00	0.06900279
ENSG00000000457	339.75	316.50	-0.10226805
ENSG00000000460	97.25	78.75	-0.30441833
ENSG00000000971	5219.00	6687.50	0.35769358
ENSG0000001036	2327.00	1785.75	-0.38194109

Define where the log2fc of gene expression is > 2 and < -2

```
up.ind <- mycounts$log2fc > +2
down.ind <- mycounts$log2fc < (-2)
sum(up.ind)

[1] 250
sum(down.ind)</pre>
```

[1] 367

# **DESeq** analysis

We need to do this analysis properly with our inner stats person kept happy.

converting counts to integer mode

Warning in DESeqDataSet(se, design = design, ignoreRank): some variables in design formula are characters, converting to factors

```
class: DESeqDataSet
dim: 38694 8
metadata(1): version
assays(1): counts
rownames(38694): ENSG00000000003 ENSG0000000005 ... ENSG00000283120
  ENSG00000283123
rowData names(0):
colnames(8): SRR1039508 SRR1039509 ... SRR1039520 SRR1039521
colData names(4): id dex celltype geo_id
Run DEseq analysis
  dds <- DESeq(dds)
estimating size factors
estimating dispersions
gene-wise dispersion estimates
mean-dispersion relationship
final dispersion estimates
fitting model and testing
Get the results
  res <- results(dds)
  head(res)
log2 fold change (MLE): dex treated vs control
Wald test p-value: dex treated vs control
DataFrame with 6 rows and 6 columns
                  baseMean log2FoldChange
                                               lfcSE
                                                                  pvalue
                                                          stat
                 <numeric>
                                <numeric> <numeric> <numeric> <numeric>
```

```
ENSG00000000003 747.194195
                            -0.3507030 0.168246 -2.084470 0.0371175
ENSG00000000005
                0.000000
                                   NA
                                            NA
                                                     NA
                                                              NA
ENSG00000000419 520.134160
                             ENSG00000000457 322.664844
                             0.0245269 0.145145 0.168982 0.8658106
ENSG00000000460 87.682625
                            -0.1471420 0.257007 -0.572521 0.5669691
ENSG00000000938
                            -1.7322890 3.493601 -0.495846 0.6200029
                0.319167
                   padj
              <numeric>
ENSG0000000000 0.163035
ENSG00000000005
ENSG0000000419 0.176032
ENSG00000000457 0.961694
ENSG00000000460
               0.815849
ENSG00000000938
                    NA
```

I want to make a figure showing overview of all my results. A plot of log2 fold change vs the p-value (using adjusted p-value)

```
# Setup our custom point color vector
mycols <- rep("gray", nrow(res))
mycols[ abs(res$log2FoldChange) > 2 ] <- "red"

inds <- (res$padj < 0.01) & (abs(res$log2FoldChange) > 2 )
mycols[ inds ] <- "blue"

# Volcano plot with custom colors
plot( res$log2FoldChange, -log(res$padj),
    col=mycols, ylab="-Log(P-value)", xlab="Log2(FoldChange)" )

# Cut-off lines
abline(v=c(-2,2), col="gray", lty=2)
abline(h=-log(0.1), col="gray", lty=2)</pre>
```

