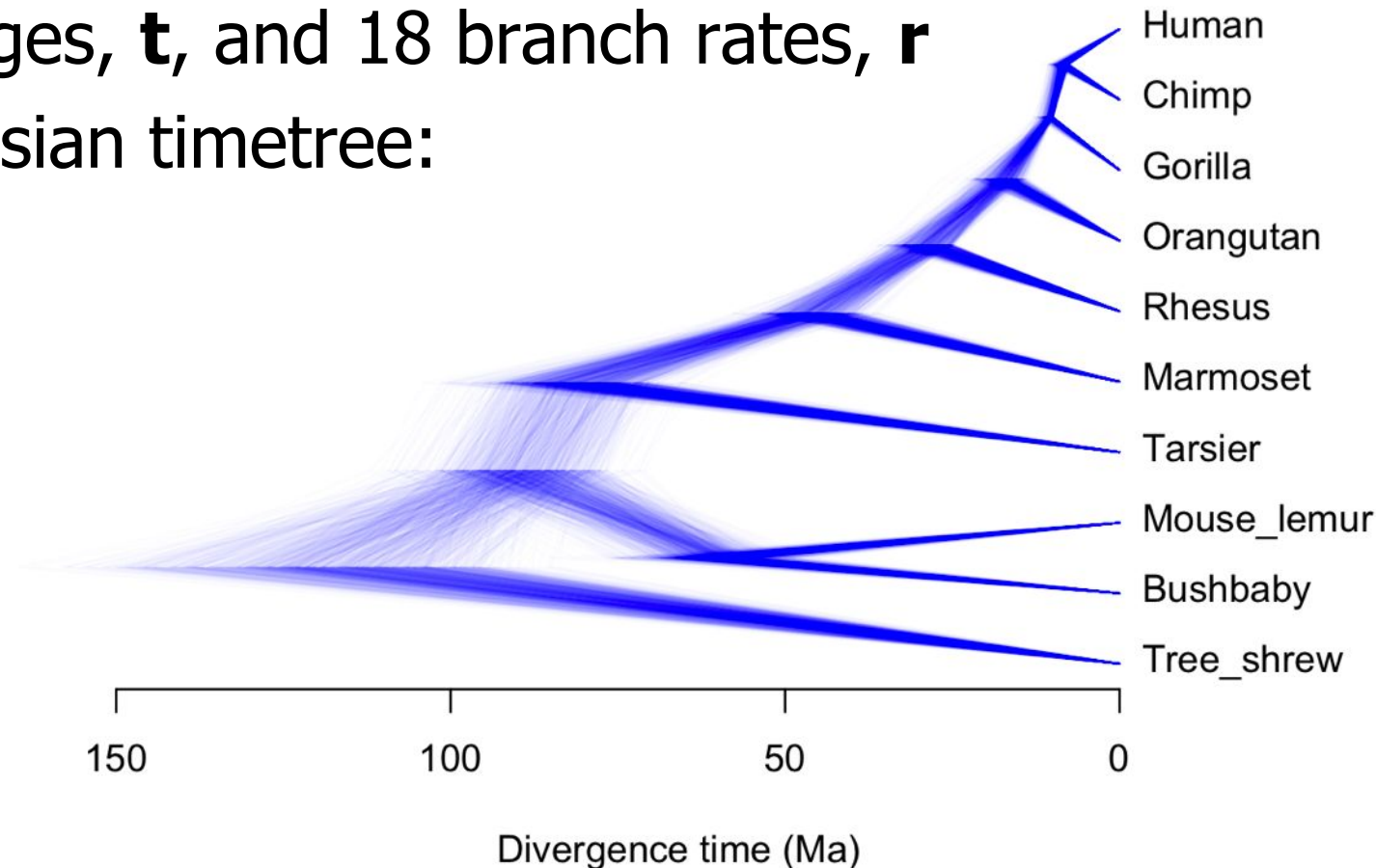


# Bayesian molecular-clock dating using genome-scale datasets

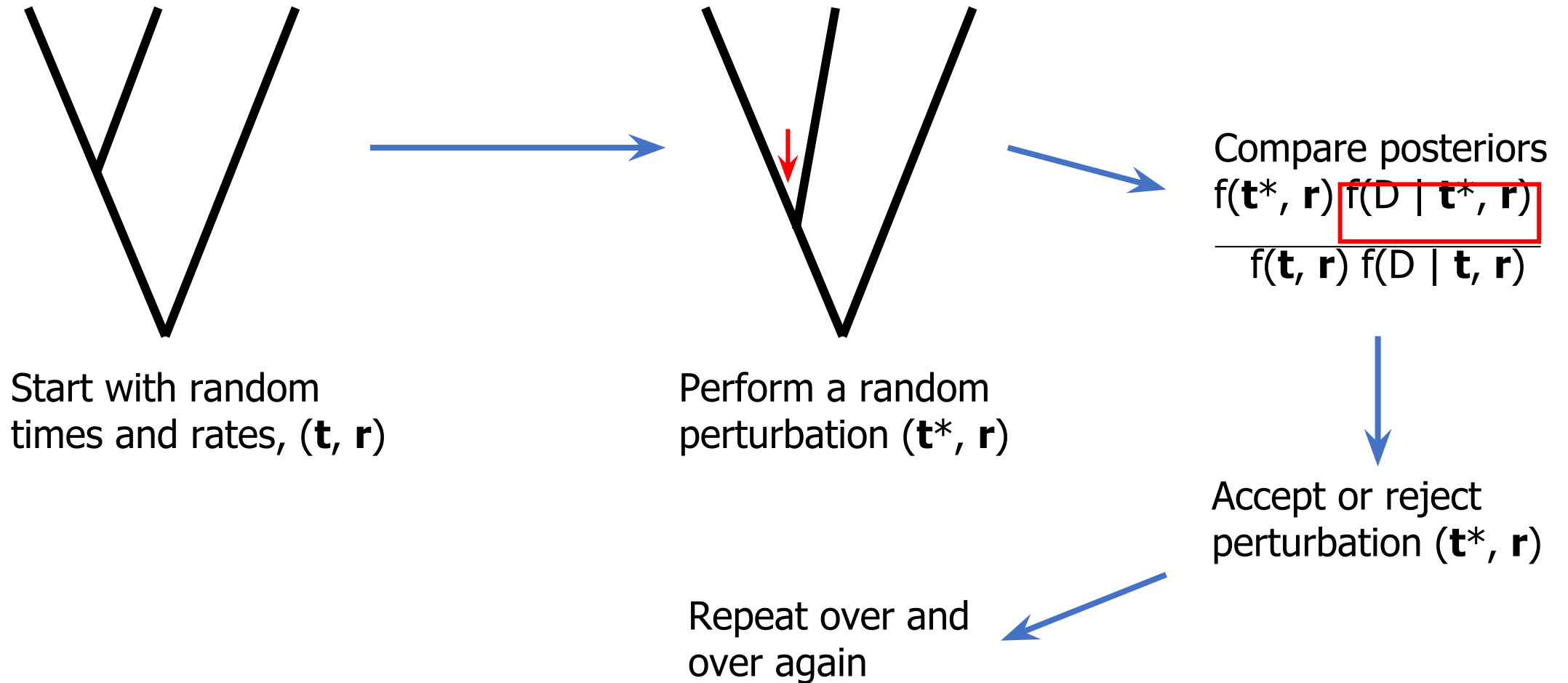
Mario dos Reis – Queen Mary University of London

# Bayesian molecular-clock dating using genome-scale datasets

- Today we will use **MCMCtree** to infer the timetree of 10 primates using a genome-scale alignment (3.4 million base pairs)
- 9 node ages, **t**, and 18 branch rates, **r**
- Our Bayesian timetree:



# MCMCtree uses the MCMC algorithm



# Likelihood calculation is expensive

- We must calculate the likelihood for every site in the alignment:
- $f(D \mid \mathbf{t}, \mathbf{r}) = \prod_i f(D_i \mid \mathbf{t}, \mathbf{r})$ , where  $D_i$  is the  $i$ -th alignment site
- Thus, for genome-scale alignments this multiplication can become very large
- It gets worse if there are many species in the alignment because the algorithm must visit all nodes in the tree to calculate  $f(D \mid \mathbf{t}, \mathbf{r})$
- MCMC sampling of using genome-alignments would take months or years

# Bayesian molecular-clock dating using genome-scale datasets

- MCMCtree implements a method to approximate the likelihood during MCMC sampling of timetrees
- To use the approximation we need to:
  - Work on a fixed tree topology
  - Estimate the branch lengths, **b**, (in substitution per site) on the tree by ML
  - Estimate the gradient, **g**, and Hessian, **H**, of the likelihood
- Then, by using **b**, **g**, and **H**, MCMCtree can provide a surprisingly accurate approximation of the likelihood during MCMC sampling
- $\Delta \log f(D \mid \mathbf{b}) \approx \Delta \mathbf{b}^T \mathbf{g} + \Delta \mathbf{b}^T \mathbf{H} \Delta \mathbf{b} / 2$
- Cost of approximate calculations depends on the number of species (which determines size of **b**, **g**, **H**) but not on alignment length

# Advantages of the approximation

- The approximation gives speedups of over 1,000x compared to MCMC sampling with exact likelihood calculations
- For example, in today's example, our 3.4 million-base alignment can be sampled in about **2.5 min** with the approximation
- However, it would take **~15,000 min** (10 days) with exact likelihood
- This is a **6,000x** computation speedup
- MCMCtree is now the de facto standard for timetree inference in large datasets

# MCMCtree best for very large data

Science

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Article | Published: 2019







**Integrated genomics  
illuminates the evolution of  
eukaryote organelles**

Holly C. Betts, Mark N. Burdett,  
Donoghue & Davide Pisani


*Nature Ecology & Evolution*

RESEARCH ARTICLE

**Phylogenomics reveals the evolutionary  
timing and pattern of butterflies and moths**

 Akito Y. Kawahara,  David Plotkin, Marianne Espeland, Karen Meusemann,  
 Emmanuel F. A. Toussaint,  Alexander Donath, France Gimnich, Paul B. Frandsen,  
Andreas Zwick,  Mario dos Reis, Jesse R. Barber, Ralph S. Peters, Shanlin Liu, Xin Zhou,  
 Christoph Mayer, Lars Podsiadlowski, Caroline Storer, Jayne E. Yack, Bernhard Misof, and  
Jesse W. Breinholt

PNAS November 5, 2019 116 (45) 22657–22663; first published October 21, 2019  
<https://doi.org/10.1073/pnas.1907847116>



**PNAS**

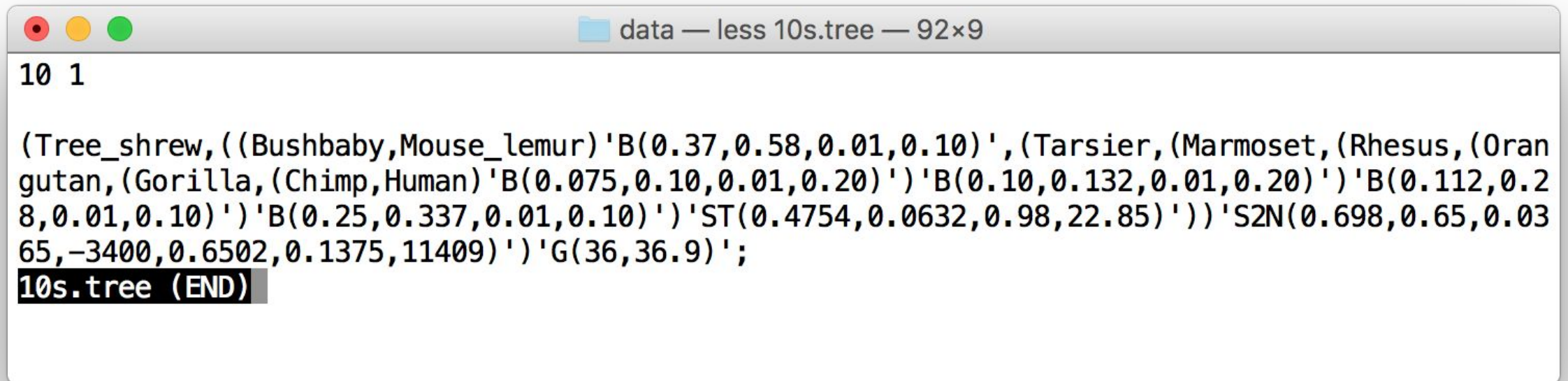
# Tutorial: Primates divergence times

- dos Reis and Yang (2019) **Bayesian Molecular Clock Dating Using Genome-Scale Datasets**. In: Evolutionary Genomics.  
[https://doi.org/10.1007/978-1-4939-9074-0\\_10](https://doi.org/10.1007/978-1-4939-9074-0_10)
- All data files are available from my GitHub:
- <https://github.com/mariodosreis/divtime>
- **MCMCtree** can be downloaded from:
- <http://abacus.gene.ucl.ac.uk/software/paml4.9j.tgz>
- It is recommended to have the following software: **R** and **RStudio**, **Tracer**, and **FigTree**.
- These slides: <https://dosreislabs.github.io/assets/slides/applnL.pdf>



# Data files: data/10s.tree

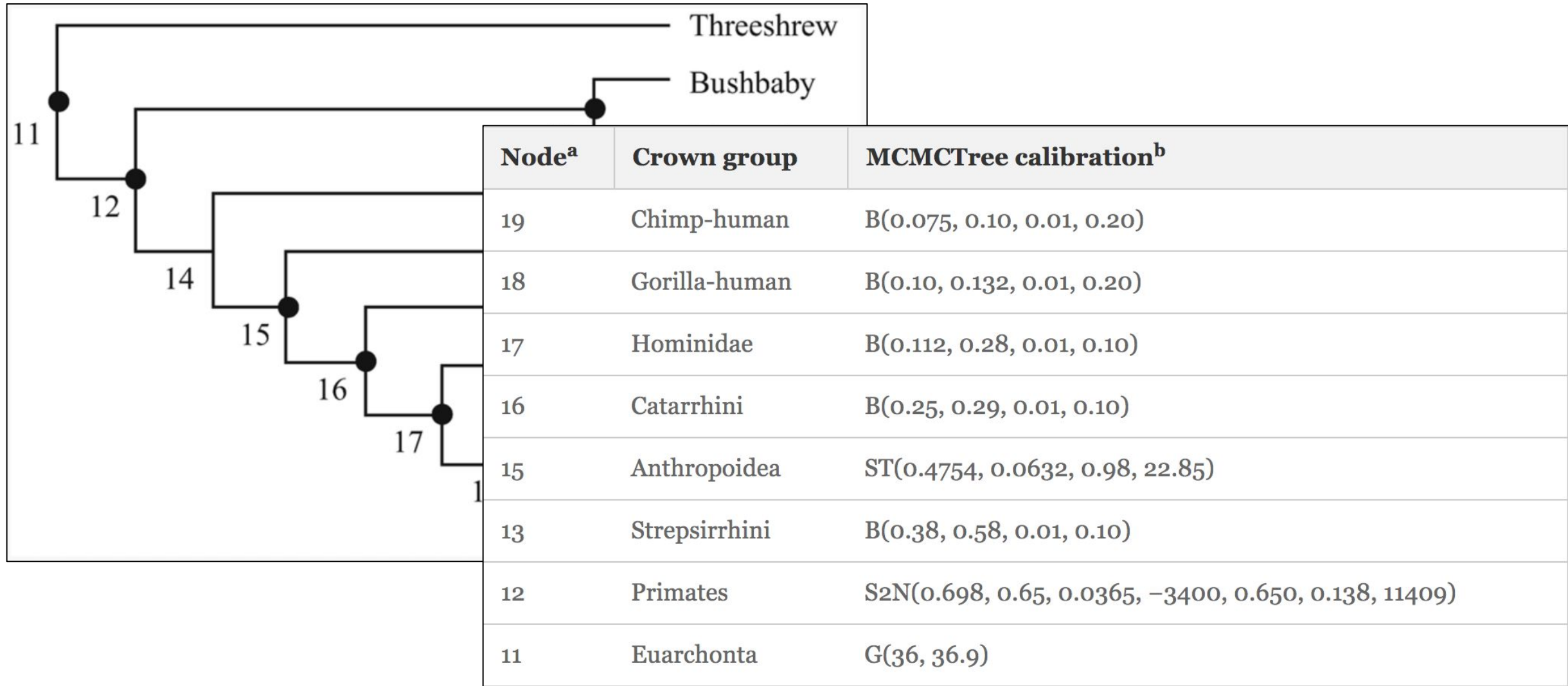
- Tree is in Newick format and has fossil calibrations
- You can open data/10s.tree in a text editor (Notepad, TextEdit, etc.)



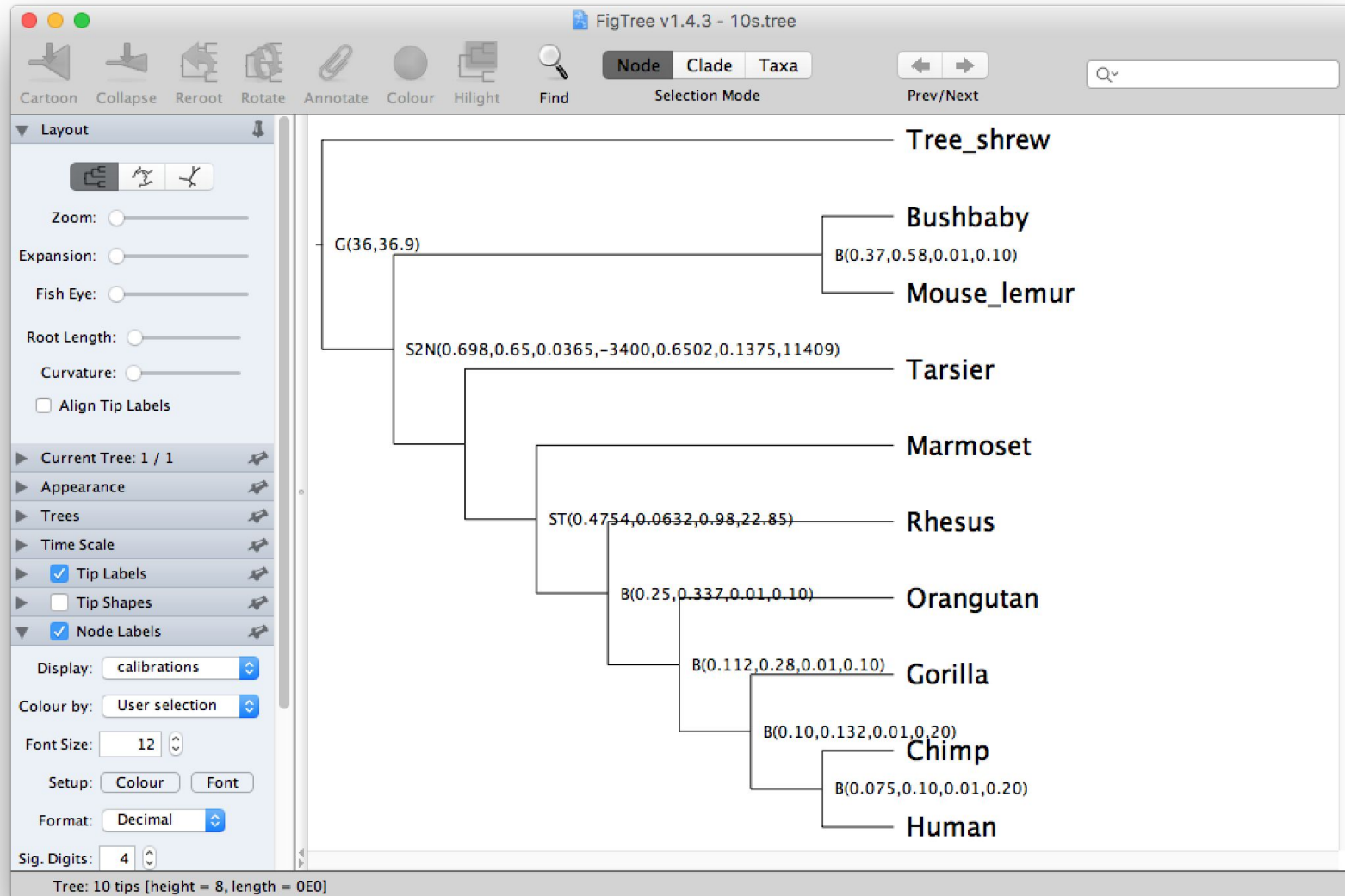
```
10 1

(Tree_shrew, ((Bushbaby, Mouse_lemur) 'B(0.37, 0.58, 0.01, 0.10)', (Tarsier, (Marmoset, (Rhesus, (Oran
gutan, (Gorilla, (Chimp, Human) 'B(0.075, 0.10, 0.01, 0.20)') 'B(0.10, 0.132, 0.01, 0.20)') 'B(0.112, 0.2
8, 0.01, 0.10)') 'B(0.25, 0.337, 0.01, 0.10)') 'ST(0.4754, 0.0632, 0.98, 22.85)')) 'S2N(0.698, 0.65, 0.03
65, -3400, 0.6502, 0.1375, 11409)') 'G(36, 36.9)';
10s.tree (END)
```

# Data files: data/10s.tree



# You may open data/10s.tree with FigTree



# Data files: `data/10s.phys`

- Alignment is in Phylip sequential format with site-patterns
- Alignment is divided into two partitions:
  - First and second codon positions (8,011 site-patterns)
  - Third codon positions (14,507 site-patterns)
- You can also open the alignment using a text editor

# Data files: data/10s.phys

data — less 10s.phys — 94×24

108011P

Tree\_shrew

Bushbaby

Mouse\_lemur

Tarsier

Marmoset

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Orangutan

Gorilla

Chimp

Human

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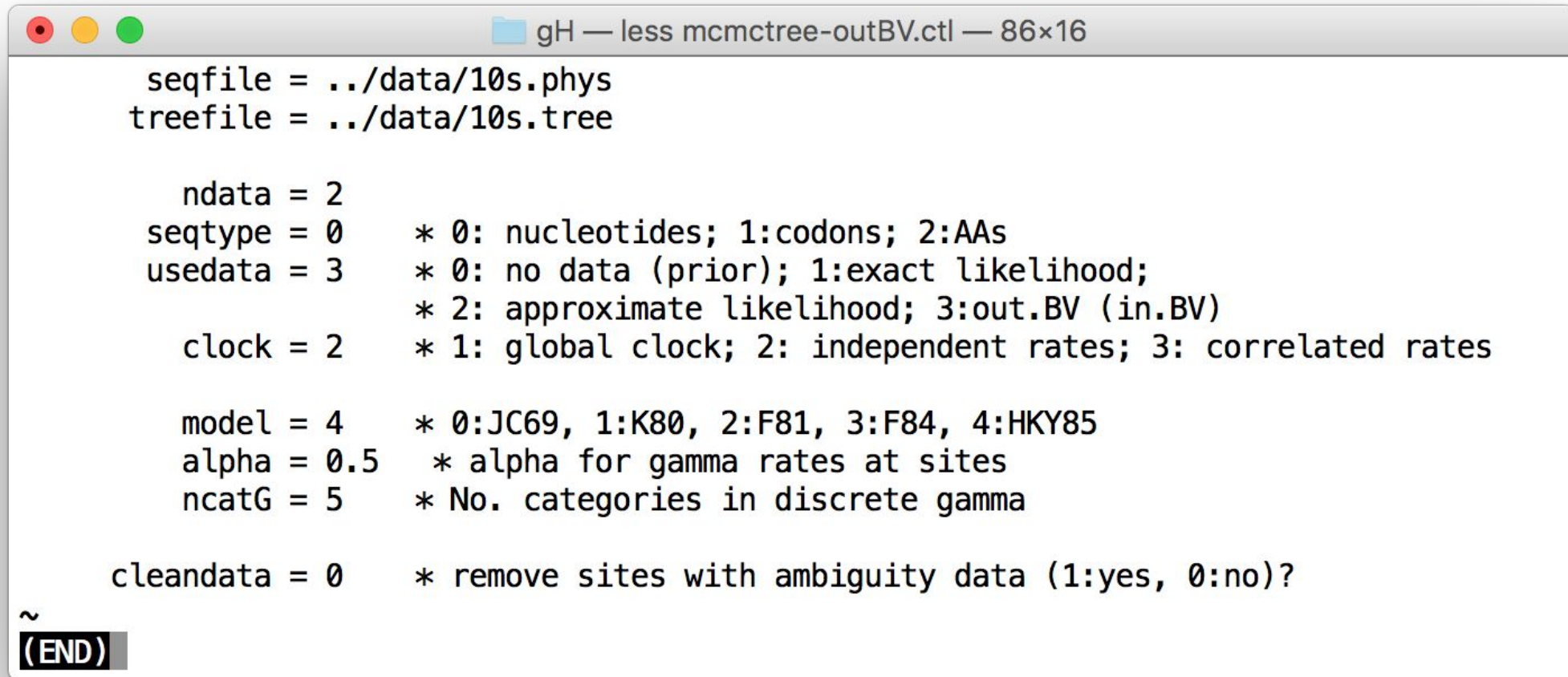
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# Step 1: Estimation of b, g and H

control file: gH/mcmctree-outBV.ctl



```
seqfile = ../data/10s.phys
treefile = ../data/10s.tree

ndata = 2
seqtype = 0    * 0: nucleotides; 1:codons; 2:AAs
usedata = 3    * 0: no data (prior); 1:exact likelihood;
               * 2: approximate likelihood; 3:out.BV (in.BV)
clock = 2      * 1: global clock; 2: independent rates; 3: correlated rates

model = 4      * 0:JC69, 1:K80, 2:F81, 3:F84, 4:HKY85
alpha = 0.5    * alpha for gamma rates at sites
ncatG = 5      * No. categories in discrete gamma

cleandata = 0  * remove sites with ambiguity data (1:yes, 0:no)?

~
(END)
```

# Step 1: Estimation of **b**, **g** and **H**

- In a terminal window, go to the `gH/` directory and type

```
$ mcmctree mcmctree-outBV.ctl
```

- MCMCtree will prepare an unrooted tree, a `baseml.ctl` file, and then call BASEML to calculate **b**, **g**, and **H**
- These parameters will be saved to a file called `gH/out.BV`
- Copy `gH/out.BV` as `mcmc/in.BV`

# Step 1: file gH/out.BV

```
gH — less -S out.BV — 87×23
:
10
((Bushbaby: 0.029524, Mouse_lemur: 0.019653): 0.006547, (Tarsier: 0.030898, (Marmoset:
0.006547 0.029524 0.019653 0.002123 0.030898 0.011754 0.015183 0.003426 0.008
-2.088785 -1.981107 7.873266 -24.785755 -4.465100 -5.585153 -2.139822 -18.493240 -14.

Hessian
-2.033e+08 -2.59e+06 -9.717e+06 -4.363e+07 1.799e+06 -5.457e+06 2.055e+06 -1.285e+0
-2.59e+06 -5.71e+07 2.235e+06 1.475e+06 3.315e+06 1.651e+06 3.436e+06 2.134e+0
-9.717e+06 2.235e+06 -8.733e+07 -2.954e+06 2.79e+06 7.275e+05 3.371e+06 1.512e+0
-4.363e+07 1.475e+06 -2.954e+06 -4.622e+08 -5.059e+06 -2.658e+07 3.701e+06 -5.156e+0
1.799e+06 3.315e+06 2.79e+06 -5.059e+06 -5.473e+07 7.951e+05 3.437e+06 2.28e+0
-5.457e+06 1.651e+06 7.275e+05 -2.658e+07 7.951e+05 -1.403e+08 3.724e+06 -1.163e+0
2.055e+06 3.436e+06 3.371e+06 3.701e+06 3.437e+06 3.724e+06 -1.25e+08 -1.69e+0
-1.285e+04 2.134e+06 1.512e+06 -5.156e+06 2.28e+06 -1.163e+07 -1.69e+07 -4.756e+0
3.483e+06 4.548e+06 4.413e+06 -1.405e+05 4.463e+06 2.246e+06 1.979e+06 1.699e+0
8.345e+05 2.861e+06 2.023e+06 1.605e+06 2.021e+06 -5.676e+05 -8.423e+05 -1.722e+0
:
```



# Step 2: Posterior MCMC sampling

control file: `mcmc/mcmctree.ctl`

```
mcmc — less -S mcmctree.ctl — 88x31

seed = -1
seqfile = ../data/10s.phys
treefile = ../data/10s.tree
mcmcfile = mcmc.txt
outfile = out.txt

ndata = 2
seqtype = 0    * 0: nucleotides; 1:codons; 2:AAs
usedata = 2    * 0: no data (prior); 1:exact likelihood;
               * 2:approximate likelihood; 3:out.BV (in.BV)
clock = 2      * 1: global clock; 2: independent rates; 3: correlated rates
RootAge = '<1.0' * safe constraint on root age, used if no fossil for root.

model = 4      * 0:JC69, 1:K80, 2:F81, 3:F84, 4:HKY85
alpha = 0.5    * alpha for gamma rates at sites
ncatG = 5      * No. categories in discrete gamma

cleandata = 0   * remove sites with ambiguity data (1:yes, 0:no)?

BDparas = 1 1 0 * birth, death, sampling
kappa_gamma = 6 2 * gamma prior for kappa
alpha_gamma = 1 1 * gamma prior for alpha

rgene_gamma = 2 40 1 * gammaDir prior for rate for genes
sigma2_gamma = 1 10 1 * gammaDir prior for sigma^2 (for clock=2 or 3)

print = 1      * 0: no mcmc sample; 1: everything except branch rates 2: everything
burnin = 20000
sampfreq = 100
nsample = 20000
```

## Step 2: Posterior MCMC sampling

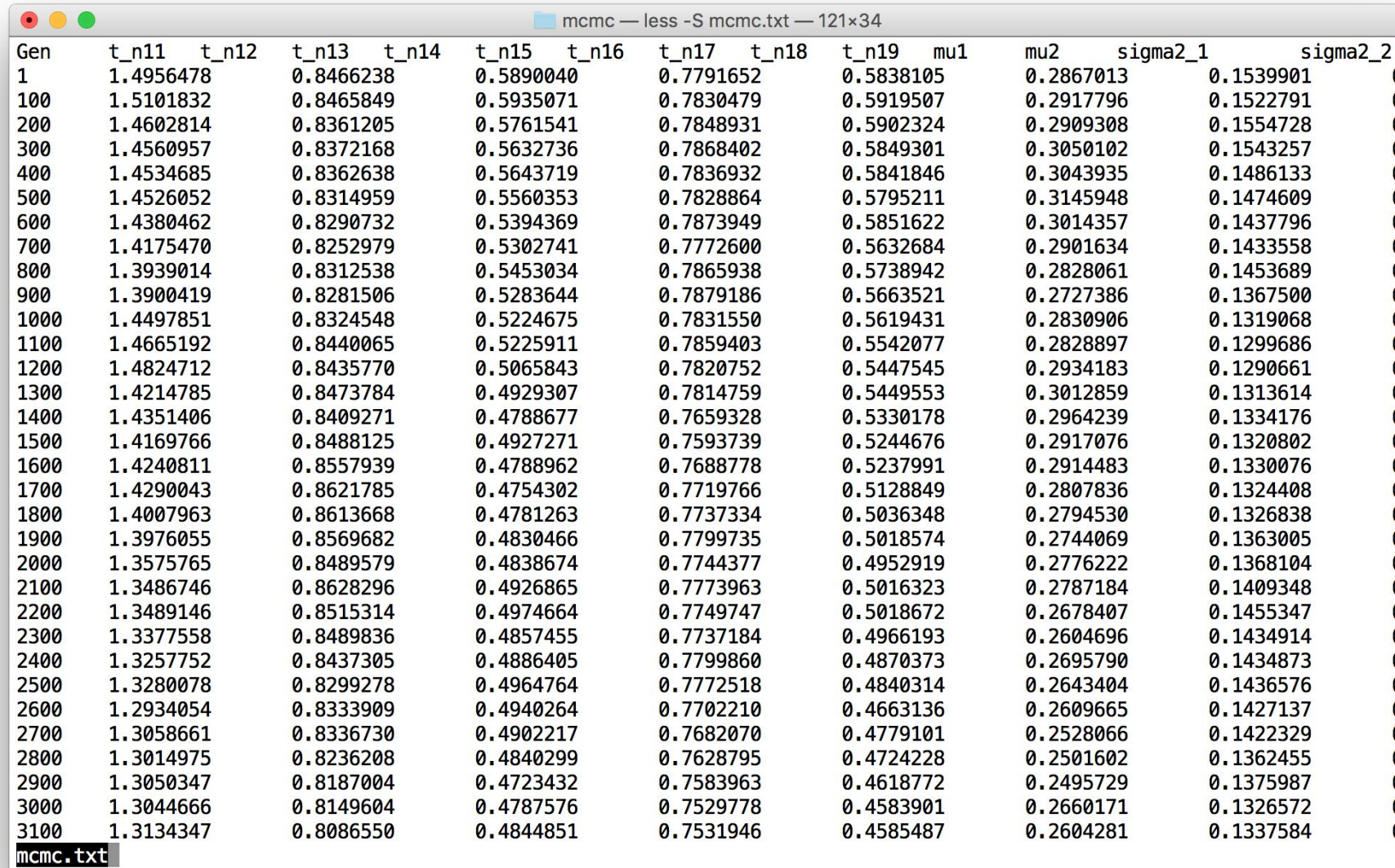
- In a terminal window, go to the `mcmc/` directory and type

```
$ mcmctree mcmctree.ctl
```

- MCMCtree will start the MCMC chain to obtain a posterior sample of primate timetrees
- Once the run is finished, MCMCtree will have created `mcmc/mcmc.txt`, `mcmc/out.txt`, and `mcmc/FigTree.tre` files
- Rename the files as `mcmc/mcmc1.txt`, `mcmc/out1.txt`, and `mcmc/FigTree1.tre` and run MCMCtree again
- This is necessary to assess convergence

# Step 2: Posterior MCMC sampling

## File mcmc/mcmc.txt

A terminal window titled 'mcmc — less -S mcmc.txt — 121x34' displays the contents of a file named 'mcmc.txt'. The file contains a table of MCMC sampling results. The table has 14 columns: 'Gen' (Generation), 't\_n11', 't\_n12', 't\_n13', 't\_n14', 't\_n15', 't\_n16', 't\_n17', 't\_n18', 't\_n19', 'mu1', 'mu2', 'sigma2\_1', and 'sigma2\_2'. The 'Gen' column lists generations from 1 to 3100 in increments of 100. The other columns contain numerical values representing parameters at each generation. The values for 'mu1' and 'mu2' fluctuate around 0.78 and 0.29 respectively. The values for 'sigma2\_1' and 'sigma2\_2' fluctuate around 0.15 and 0.13 respectively. The terminal window has a standard macOS-style title bar with red, yellow, and green window control buttons.

Gen	t_n11	t_n12	t_n13	t_n14	t_n15	t_n16	t_n17	t_n18	t_n19	mu1	mu2	sigma2_1	sigma2_2
1	1.4956478		0.8466238		0.5890040		0.7791652		0.5838105		0.2867013	0.1539901	0
100	1.5101832		0.8465849		0.5935071		0.7830479		0.5919507		0.2917796	0.1522791	0
200	1.4602814		0.8361205		0.5761541		0.7848931		0.5902324		0.2909308	0.1554728	0
300	1.4560957		0.8372168		0.5632736		0.7868402		0.5849301		0.3050102	0.1543257	0
400	1.4534685		0.8362638		0.5643719		0.7836932		0.5841846		0.3043935	0.1486133	0
500	1.4526052		0.8314959		0.5560353		0.7828864		0.5795211		0.3145948	0.1474609	0
600	1.4380462		0.8290732		0.5394369		0.7873949		0.5851622		0.3014357	0.1437796	0
700	1.4175470		0.8252979		0.5302741		0.7772600		0.5632684		0.2901634	0.1433558	0
800	1.3939014		0.8312538		0.5453034		0.7865938		0.5738942		0.2828061	0.1453689	0
900	1.3900419		0.8281506		0.5283644		0.7879186		0.5663521		0.2727386	0.1367500	0
1000	1.4497851		0.8324548		0.5224675		0.7831550		0.5619431		0.2830906	0.1319068	0
1100	1.4665192		0.8440065		0.5225911		0.7859403		0.5542077		0.2828897	0.1299686	0
1200	1.4824712		0.8435770		0.5065843		0.7820752		0.5447545		0.2934183	0.1290661	0
1300	1.4214785		0.8473784		0.4929307		0.7814759		0.5449553		0.3012859	0.1313614	0
1400	1.4351406		0.8409271		0.4788677		0.7659328		0.5330178		0.2964239	0.1334176	0
1500	1.4169766		0.8488125		0.4927271		0.7593739		0.5244676		0.2917076	0.1320802	0
1600	1.4240811		0.8557939		0.4788962		0.7688778		0.5237991		0.2914483	0.1330076	0
1700	1.4290043		0.8621785		0.4754302		0.7719766		0.5128849		0.2807836	0.1324408	0
1800	1.4007963		0.8613668		0.4781263		0.7737334		0.5036348		0.2794530	0.1326838	0
1900	1.3976055		0.8569682		0.4830466		0.7799735		0.5018574		0.2744069	0.1363005	0
2000	1.3575765		0.8489579		0.4838674		0.7744377		0.4952919		0.2776222	0.1368104	0
2100	1.3486746		0.8628296		0.4926865		0.7773963		0.5016323		0.2787184	0.1409348	0
2200	1.3489146		0.8515314		0.4974664		0.7749747		0.5018672		0.2678407	0.1455347	0
2300	1.3377558		0.8489836		0.4857455		0.7737184		0.4966193		0.2604696	0.1434914	0
2400	1.3257752		0.8437305		0.4886405		0.7799860		0.4870373		0.2695790	0.1434873	0
2500	1.3280078		0.8299278		0.4964764		0.7772518		0.4840314		0.2643404	0.1436576	0
2600	1.2934054		0.8333909		0.4940264		0.7702210		0.4663136		0.2609665	0.1427137	0
2700	1.3058661		0.8336730		0.4902217		0.7682070		0.4779101		0.2528066	0.1422329	0
2800	1.3014975		0.8236208		0.4840299		0.7628795		0.4724228		0.2501602	0.1362455	0
2900	1.3050347		0.8187004		0.4723432		0.7583963		0.4618772		0.2495729	0.1375987	0
3000	1.3044666		0.8149604		0.4787576		0.7529778		0.4583901		0.2660171	0.1326572	0
3100	1.3134347		0.8086550		0.4844851		0.7531946		0.4585487		0.2604281	0.1337584	0



# Step 2: Posterior MCMC sampling

## File mcmc/out.txt

```
mcmc — less -S out.txt — 121x34

# constant sites: 624389 (55.42%)
ln Lmax (unconstrained) = -4636133.236961

Time used: 2:18

mean of parameters using all iterations
  1.18274  0.90672  0.61149  0.82265  0.45967  0.29192  0.17809  0.10453  0.08527  0.02697  0.11132  0.13604

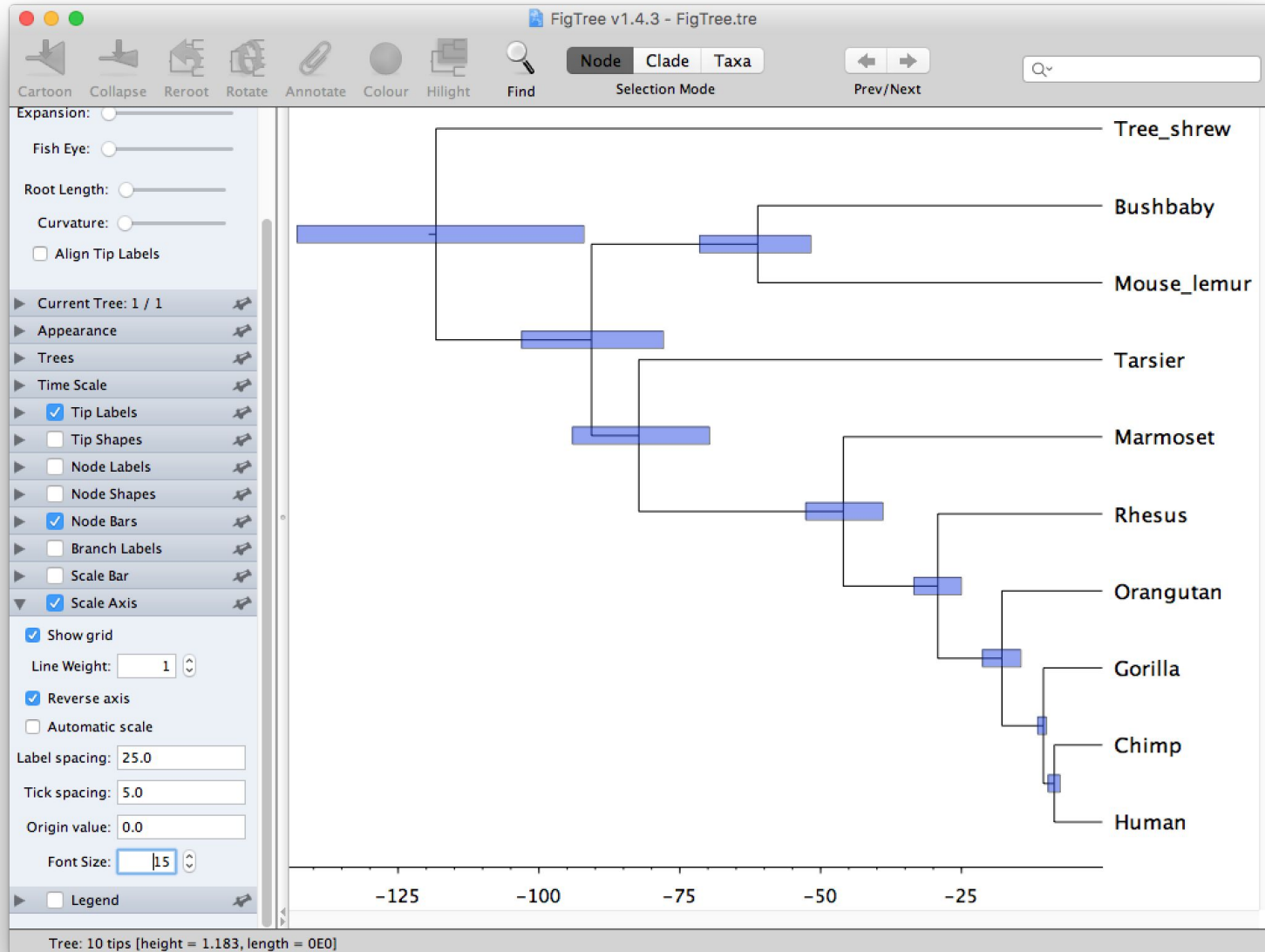
Species tree for FigTree. Branch lengths = posterior mean times; 95% CIs = labels
(1_Tree_shrew, ((2_Bushbaby, 3_Mouse_lemur) 13 , (4_Tarsier, (5_Marmoset, (6_Rhesus, (7_Orangutan, (8_Gorilla, (9_Chimp,
(Tree_shrew: 1.182735, ((Bushbaby: 0.611490, Mouse_lemur: 0.611490): 0.295229, (Tarsier: 0.822650, (Marmoset: 0.459674, (
(Tree_shrew: 1.182735, ((Bushbaby: 0.611490, Mouse_lemur: 0.611490) [&95%HPD={0.517085, 0.714825}]: 0.295229, (Tarsier: 0

Posterior mean (95% Equal-tail CI) (95% HPD CI) HPD-CI-width

t_n11      1.1828 ( 0.9272, 1.4391) ( 0.9199, 1.4291) 0.5092 (Jnode 18)
t_n12      0.9067 ( 0.7808, 1.0335) ( 0.7789, 1.0306) 0.2517 (Jnode 17)
t_n13      0.6115 ( 0.5212, 0.7209) ( 0.5171, 0.7148) 0.1977 (Jnode 16)
t_n14      0.8226 ( 0.7005, 0.9449) ( 0.6970, 0.9404) 0.2434 (Jnode 15)
t_n15      0.4597 ( 0.3912, 0.5286) ( 0.3891, 0.5262) 0.1371 (Jnode 14)
t_n16      0.2919 ( 0.2526, 0.3392) ( 0.2500, 0.3344) 0.0844 (Jnode 13)
t_n17      0.1781 ( 0.1473, 0.2157) ( 0.1444, 0.2127) 0.0682 (Jnode 12)
t_n18      0.1045 ( 0.0995, 0.1175) ( 0.0989, 0.1148) 0.0159 (Jnode 11)
t_n19      0.0853 ( 0.0757, 0.0987) ( 0.0746, 0.0962) 0.0215 (Jnode 10)
mu1         0.0270 ( 0.0221, 0.0336) ( 0.0216, 0.0329) 0.0113
mu2         0.1113 ( 0.0896, 0.1392) ( 0.0875, 0.1361) 0.0487
sigma2_1    0.1368 ( 0.0610, 0.2801) ( 0.0492, 0.2513) 0.2022
sigma2_2    0.1649 ( 0.0764, 0.3236) ( 0.0643, 0.2949) 0.2306
lnL        -17.0202 (-26.0530, -9.9180) (-25.2970, -9.4360) 15.8610
(END)
```

# Step 2: Visualization with FigTree

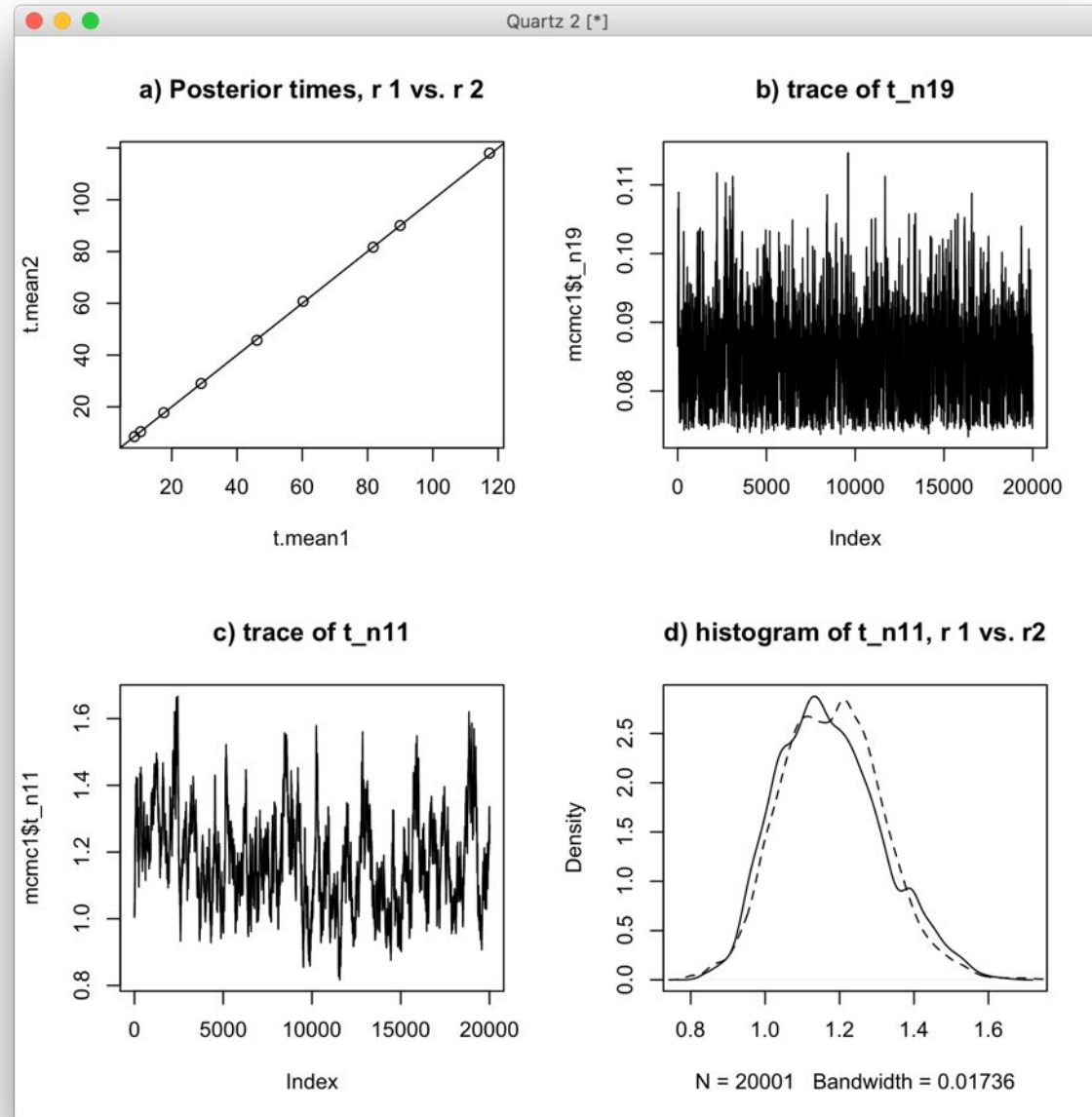
## File mcmc/FigTree.tre



# Step 3: Convergence diagnostics

- MCMC is a stochastic algorithm
- Thus, MCMC analyses must be run several times to verify results are consistent
- For example, you can carry out convergence diagnostics using Tracer or R

# Step 3: Convergence plots in R



# Exercises

- **Exercise 1:** Following the instructions in the book chapter, carry out MCMC sampling from the prior (**Hint:** `usedata = 0`)
- **Exercise 2:** The analysis we just carried out used the independent log-normal rates model. Repeat the analysis using auto-correlated rates (**Hint:** `clock = 3`)
- **Exercise 3:** Using a text editor, open the treefile, `data/10s.tree`, and modify the fossil calibrations. Use  $B(0.057, 0.10, 0.01, 0.2)$  for the human-chimp divergence, and use  $B(0.615, 1.315, 0.01, 0.05)$  for the root. Run the analysis again
- **Exercise 4:** Repeat the analysis using exact likelihood (**Hint:** `usedata = 1`, you need to reduce the number of iterations)



# Further Exercises

- **Exercise 5:** Date the 330 species primate dataset (data/330s.phy and data/330s.tree). MCMC sampling of this datasets takes several hours.
- **Exercise 6:** MCMCtree now implements marginal likelihood calculation for Bayesian model selection of tree topology, substitution model and relaxed-clock model. You can follow the tutorial to choose the clock model here:  
<https://dosreislabs.github.io/2017/10/24/marginal-likelihood-mc3r.html>