



Mecor: An R package for measurement error correction in linear regression models with a continuous outcome



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ABSTRACT

Measurement error in a covariate or the outcome of regression models is common, but is often ignored, even though measurement error can lead to substantial bias in the estimated covariate-outcome association. While several texts on measurement error correction methods are available, these methods remain seldomly applied. To improve the use of measurement error correction methodology, we developed **mecor**, an R package that implements measurement error correction methods for regression models with a continuous outcome. Measurement error correction requires information about the measurement error model and its parameters. This information can be obtained from four types of studies, used to estimate the parameters of the measurement error model: an internal validation study, a replicates study, a calibration study and an external validation study. In the package **mecor**, regression calibration methods and a maximum likelihood method are implemented to correct for measurement error in a continuous covariate in regression analyses. Additionally, methods of moments methods are implemented to correct for measurement error in the continuous outcome in regression analyses. Variance estimation of the corrected estimators is provided in closed form and using the bootstrap.

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1. Introduction

Measurement error is common across research fields, affecting the measurement of outcomes as well as important covariates. When left uncorrected, this can lead to severely biased and inefficient estimates of associations between covariates and outcome variables. Several texts have been published describing the impact of measurement error, and measurement error correction methodology [1–4]. However, recent reviews by Brakenhoff et al. [5] and Shaw et al. [6] show that, in biomedical research, measurement error correction methods remain seldomly applied. Keogh et al. [7] suggest that one of the main barriers to the use of correction methods may be the lack of accessible software. Moreover, as exemplified in [8], measurement is not only common in biomedical research, but in bioinformatics, chemistry, astronomy and econometrics as well. Therefore, to facilitate and encourage the use of measurement error correction methodology, we developed **mecor**,

an R package that provides measurement error correction methods for linear models with a continuous outcome.

Several approaches to measurement error correction have been developed in the past decade. Examples include, simulation-extrapolation (SIMEX) by Cook et al. [9], multiple imputation for measurement error by Cole et al. [10], Bayesian correction (e.g., [4,11]), maximum likelihood-based methods (e.g., [12,13]), method of moments (MM) (e.g., [1]), and regression calibration (RC) introduced by Gleser [14] and Carroll et al. [15]. Of all these measurement error correction methods, RC is among the most commonly applied in biomedical research [6], possibly because of its relative simplicity and the possibility to implement it in conjunction with a variety of analysis types, e.g., linear regression [14,15], survival analysis [16], logistic regression [17] and other generalized linear models [2,18].

In R [19], covariate measurement error correction by means of SIMEX is implemented in the package **simex** by Lederer et al. [20]. The R package **simexaft** by He et al. [21] provides SIMEX covariate measurement error correction for accelerated failure time models. A special issue of the *Stata* [22] Journal was published in 2003 and dedicated to measurement error models [23]. Three

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Table 1

Data structure of internal validation studies. The true covariate or outcome is observed in a subset of the individuals from the main study. The superscript * indicates that there is random or systematic measurement error in the variable.

(a) Covariate-validation study			
Y	X*	Z	X
y ₁	x ₁ *	z ₁	x ₁
⋮	⋮	⋮	⋮
⋮	⋮	⋮	⋮
⋮	⋮	⋮	x _{n_{sub}}
⋮	⋮	⋮	⋮
⋮	⋮	⋮	⋮
y _n	x _n *	z _n	⋮

(b) Outcome-validation study			
Y*	X	Z	Y
y ₁ *	x ₁	z ₁	y ₁
⋮	⋮	⋮	⋮
⋮	⋮	⋮	⋮
⋮	⋮	⋮	y _{n_{sub}}
⋮	⋮	⋮	⋮
⋮	⋮	⋮	⋮
y _n *	x _n	z _n	⋮

different methods were introduced for correction of measurement error in covariates in a generalized linear model. The `rca1` and `eivreg` procedure were introduced for RC by Hardin et al. [24], the `simex` and `simexplot` procedure were introduced for SIMEX by Hardin et al. [25] and, the `cme` procedure was introduced by Rabe-Hesketh et al. [26] for measurement error correction using a maximum likelihood approach. In SAS, multiple macros have been developed for measurement error correction. These macros include `%blinplus`, implementing the method by Rosner et al. [17]), `%relibpls8`, implementing the method by Rosner et al. [27], and `%rrc`, implementing the method by Liao et al. [28]), and the NCI method macros, implementing the methods by Kipnis et al. [29]. An overview of available software including useful web links can be found in Table 4 and 5 of the paper by Keogh et al. [7]. Although several measurement error correction methods are available in Stata and SAS, to date RC-like methods for measurement error correction in a covariate have not been implemented in an R package. Moreover, no method for measurement error correction in a continuous outcome has been implemented in R.

In this paper we present and describe **mecor**, an R package for measurement error correction in linear regression models with a continuous outcome. Several methods (i.e., RC, MM and maximum likelihood) are implemented to correct covariate-outcome associations for measurement error in a covariate, or in the outcome. The package **mecor** is flexible regarding the information that can be used to enable the measurement error correction, which can be of either of four types of measurement validation studies: an internal validation study, a replicates study, a calibration study and an external validation study. For each of these types of validation studies, standard RC, validation RC, efficient RC by Spiegelman et al. [30] and a maximum likelihood approach by Bartlett et al. [12] are implemented for measurement error correction in a covariate. For outcome measurement error correction, standard MM [1] and efficient MM [31] are available, for all different types of validation studies except replicates studies. The package **mecor** allows for random or systematic measurement error in a covariate, systematic measurement error in the outcome and, additionally, differ-

ential outcome measurement error in a univariable analysis. This broad spectrum of validation study types, measurement error models and correction methods in our easy-to-use software package should improve the application of measurement error corrections in research practice.

This paper is organized as follows. Section 2 introduces several measurement error models and the data structures of the four validation study types that can be used to estimate the parameters of the measurement error model. Section 3 outlines the measurement error correction methods. Section 4 introduces the functions in the package **mecor**. Section 5 demonstrates how the package **mecor** can be used in different settings using simulated example data.

2. Measurement error: notation, types and data structures

In this section, we introduce notation, derive expressions for the impact of measurement error on covariate-outcome associations and introduce the data structure of four different types of studies, that provide input for measurement error correction methods. Throughout, it is assumed that there is a continuous outcome Y , a continuous covariate X and a vector of k other covariates $\mathbf{Z} = (Z_1, Z_2, Z_3, \dots, Z_k)$. We consider measurement error in one variable at a time, i.e., in the covariate, X , or in the outcome, Y and assume that the other variables in the model are measured without error. Since our focus is on studies in which we aim to estimate the covariate-outcome association, the covariate X could be the main exposure of interest or a variable that confounds the relation between the main exposure and the outcome (one of the Z variables). The parameters of interest are $\beta = (\beta_X, \beta_0, \beta_Z)$ (with β_Z a $1 \times k$ matrix) from the linear model,

$$Y = \beta_X X + \beta_0 + \beta_Z \mathbf{Z}' + e, \quad \text{Var}(e) = \sigma^2, \quad (1)$$

where we assume that $E(e) = 0$ and $\text{Cov}(e, X) = \text{Cov}(e, \mathbf{Z}) = 0$. This model will be referred to as the **outcome model**.

2.1. Types of measurement error and their impact

To quantify the impact of measurement error, we first define the assumed measurement error models. Subsequently, we outline the impact of measurement error in a covariate and the outcome on the estimates of the outcome model parameters, separately.

2.1.1. Covariate measurement error

Let X^* denote the error-prone substitute measure of the error-free reference measure X , following the measurement error model,

$$X^* = \theta_0 + \theta_1 X + U, \quad \text{Var}(U) = \tau^2, \quad (2)$$

and assume that $E(U) = 0$ and $\text{Cov}(U, X) = 0$. We assume non-differential covariate measurement error (i.e., $X^* \perp\!\!\!\perp Y | X, \mathbf{Z}$ or, equivalently, that the errors U are independent of the errors e in Eq. (1)). The measurement error is called 'classical' or 'random' if $\theta_0 = 0$ and $\theta_1 = 1$. The terms *classical measurement error* and *random measurement error* are used interchangeably in the literature. In this paper, we use the term random measurement error to refer to this type of measurement error. The measurement error is called 'systematic' for all other values of θ_0 and θ_1 (where $\theta_1 \neq 0$).

Suppose that there is one covariate $\mathbf{Z} = Z_1$ in the outcome model in (1), and that data on Y , X^* and Z_1 are available to fit the linear model,

$$E(Y | X^*, Z_1) = \beta_X^* X^* + \beta_0^* + \beta_Z^* Z_1. \quad (3)$$

In this model, the least squares estimators $\hat{\beta}^* = (\hat{\beta}_X^*, \hat{\beta}_0^*, \hat{\beta}_Z^*)$, are biased for β , and consistent and unbiased estimators for $\beta \mathbf{A}$ [2], where \mathbf{A} is the 3×3 **calibration model matrix**:

$$\mathbf{A} = \begin{pmatrix} \lambda_{X^*} & \lambda_0 & \lambda_{Z_1} \\ 0 & 1 & 0 \\ 0 & 0 & 1 \end{pmatrix}.$$

A well-known special case of the calibration model matrix is the attenuation factor. In particular, when there is random measurement error in the substitute error-prone measure X^* , we have $\beta_X^* = \lambda_{X^*} \beta$, where λ_{X^*} is called the attenuation factor [32] or regression dilution factor [33,34]. When there is more than one \mathbf{Z} covariate in the outcome model defined by Eq. (1), the **calibration model matrix** generalizes to the following $(2+k) \times (2+k)$ matrix:

$$\mathbf{A} = \begin{pmatrix} \lambda_{X^*} & \lambda_0 & \lambda_{\mathbf{Z}} \\ \mathbf{0} & \mathbf{I} & \end{pmatrix}, \quad (4)$$

where $\lambda_{\mathbf{Z}}$ is a $1 \times k$ matrix, $\mathbf{0}$ is a $(1+k) \times 1$ null matrix and \mathbf{I} is a $(1+k) \times (1+k)$ identity matrix.

2.1.2. Outcome measurement error

Let Y^* denote the error-prone substitute measure of the error-free reference measure Y , following the measurement error model,

$$Y^* = \theta_0 + \theta_1 Y + U, \quad \text{Var}(U) = \tau^2, \quad (5)$$

and assume that $E(U) = 0$ and $\text{Cov}(U, Y) = 0$. We assume non-differential outcome measurement error (i.e., $Y^* \perp\!\!\!\perp X | Y, \mathbf{Z}$ or, equivalently, that the errors U are independent of the errors e in Eq. (1)), unless specified otherwise. Random and systematic outcome measurement error are defined analogously to random and systematic covariate measurement error, respectively [35,36].

Suppose, again, that there is one covariate $\mathbf{Z} = Z_1$ in the outcome model in (1) and that data on Y^* , X and Z_1 are available to fit the linear model,

$$E[Y^* | X, Z_1] = \beta_X^* X + \beta_0^* + \beta_{Z_1}^* Z_1. \quad (6)$$

If the measurement error in Y^* is random, the least squares estimators $\hat{\beta}^* = (\hat{\beta}_X^*, \hat{\beta}_0^*, \hat{\beta}_{Z_1}^*)$ are unbiased for β . In contrast, if the error in Y^* is systematic, the least squares estimators $\hat{\beta}^* = (\hat{\beta}_X^*, \hat{\beta}_0^*, \hat{\beta}_{Z_1}^*)$ are biased for β [1,31,36]. In order to identify consistent estimators for β by matrix multiplication, we add the integer 1 to the vector $\hat{\beta}^*$. Then, $(\hat{\beta}^*, 1)$ are consistent and unbiased estimators for $(\beta, 1)\Theta$ where Θ is the 4×4 outcome **measurement error model matrix**:

$$\Theta = \begin{pmatrix} \theta_1 & 0 & 0 & 0 \\ 0 & \theta_1 & 0 & 0 \\ 0 & 0 & \theta_1 & 0 \\ 0 & \theta_0 & 0 & 1 \end{pmatrix}.$$

When there is more than one \mathbf{Z} covariate in the outcome model defined in Eq. (1), the calibration model matrix generalizes to the following $(2+k+1) \times (2+k+1)$ outcome **measurement error model matrix**:

$$\Theta = \begin{pmatrix} \theta_1 & \dots & \dots & 0 \\ \vdots & \ddots & & \vdots \\ \vdots & & \theta_1 & \vdots \\ 0 & \theta_0 & \dots & 1 \end{pmatrix}, \quad (7)$$

where $\hat{\Theta}$ contains all zero's except on the diagonal and the $(2+k+1, 2)$ th element.

2.1.3. Differential outcome measurement error in univariable analyses

We assume non-differential measurement error in the outcome in all but the following special case. Suppose exposure X is binary (e.g., in a two-arm controlled randomised trial) and that there are no other covariates \mathbf{Z} in the outcome model defined by Eq. (1). Further, suppose that the measurement error in Y is differential such that the measurement error in the unexposed individuals (i.e.,

$X = 0$) is different from the measurement error in the exposed individuals (i.e., $X = 1$). Equivalently, let Y^* be the error-prone substitute measure of the error-free reference measure Y , with mean $E(Y^* | Y, X) = \theta_{X0} + \theta_{X1} Y$ and variance τ^2 , for $X = 0, 1$. Suppose now that data on Y^* and X are available to fit the linear model,

$$E[Y^* | X] = \beta_X^* X + \beta_0^*.$$

In this model, the least squares estimators $\hat{\beta}^* = (\hat{\beta}_X^*, \hat{\beta}_0^*)$ are biased for β [31,36]. In order to identify consistent estimators for β by matrix multiplication, we again add the integer 1 to the vector $\hat{\beta}^*$. Then, $(\hat{\beta}^*, 1)$ are consistent and unbiased estimators for $(\beta, 1)\Theta$ where, Θ is the following 3×3 differential outcome **measurement error model matrix**:

$$\Theta = \begin{pmatrix} \theta_{11} & 0 & 0 \\ \theta_{11} - \theta_{10} & \theta_{10} & 0 \\ \theta_{01} - \theta_{00} & \theta_{00} & 1 \end{pmatrix}. \quad (8)$$

2.2. Validation study data structures for measurement error correction

Four types of validation studies can be used to estimate the calibration model matrix or outcome measurement error model matrix defined in Section 2.1: an internal validation study, a replicates study, a calibration study or an external validation study [7,37]. The first three validation studies make use of information internal to the study cohort, whereas the fourth makes use of information external to the study cohort.

2.2.1. Internal validation study

In an internal validation study, the error-free reference covariate values X or outcome values Y are observed in a subset of individuals (Table 1). Table 1a shows the structure of an internal validation study for covariate measurement error. In the main study, the outcome Y , the error-prone substitute covariate X^* and the covariates \mathbf{Z} are measured in all n individuals. Additionally, in n_{sub} individuals ($n_{\text{sub}} < n$) the true covariate X is measured, assumed a random subset of the main study. As an example, suppose the true exposure of interest is visceral adipose tissue measurements (i.e., X) but that this is too expensive to obtain on all study participants and the error-prone substitute measure of waist circumference is instead collected for everyone (i.e., X^*) [42]. The same structure holds for an internal validation study for outcome measurement error, as shown in Table 1b.

Replicates study

A replicates study can be used if the measurement error in a covariate is random, denoted by X^{*r} . We will only use this type of study for covariate measurement error since random measurement error in an outcome does not result in biased association estimates (Section 2.1). In a replicates study, the error-prone substitute covariate X^{*r} is repeatedly measured (i.e., m times, where $m \geq 2$) in all or in a random subset of individuals (Table 2). The repeated measures are denoted by $X_1^{*r}, \dots, X_m^{*r}$. We assume that, in each individual, the same number of repeated measures was observed. Further, we assume that the measurement error in the replicates is jointly independent. Table 2a and 2b show the structure of a replicates study with full and partial replicates, respectively. In the main study, the outcome Y , the error-prone substitute covariate X_1^{*r} and the covariates \mathbf{Z} are measured in all n individuals. Additionally, $n_{\text{sub}} \leq n$ individuals have m replicates of the error-prone substitute measure X_j^{*r} for $j = 2 \dots m$. An example is the repeated measurement of several coronary risk factors in the Framingham Heart study, such as serum cholesterol, blood glucose, and systolic blood pressure [27].

Table 2

Data structure of a covariate-replicates study for full or partial replicates. The error-prone covariate is measured m times in all or a subset of individuals. The superscript $*_r$ indicates random measurement error.

(a) Full replicates study					
Y	X_1^{*r}	Z	X_2^{*r}	...	X_m^{*r}
y_1	x_{11}^{*r}	z_1	x_{12}^{*r}	...	x_{1m}^{*r}
\vdots	\vdots	\vdots	\vdots	\vdots	\vdots
\vdots	\vdots	\vdots	\vdots	\vdots	\vdots
\vdots	\vdots	\vdots	\vdots	\vdots	\vdots
\vdots	\vdots	\vdots	\vdots	\vdots	\vdots
y_n	x_{n1}^{*r}	z_n	x_{n2}^{*r}	...	x_{nm}^{*r}
(b) Partial replicates study					
Y	X_1^{*r}	Z	X_2^{*r}	...	X_m^{*r}
y_1	x_{11}^{*r}	z_1	x_{12}^{*r}	...	x_{1m}^{*r}
\vdots	\vdots	\vdots	\vdots	...	\vdots
\vdots	\vdots	\vdots	\vdots	...	\vdots
\vdots	\vdots	\vdots	$x_{n_{sub}2}^{*r}$...	$x_{n_{sub}m}^{*r}$
\vdots	\vdots	\vdots	-	...	-
\vdots	\vdots	\vdots	\vdots	\vdots	\vdots
y_n	x_{n1}^{*r}	z_n	-	...	-

Table 3

Data structure of calibration studies. Two types of error-prone measurement methods are used to measure the covariate or outcome. The superscripts $*_r$ and $*_s$ indicate random and systematic measurement error, respectively.

(a) Covariate-calibration study					
Y	X^{*s}	Z	X_1^{*r}	...	X_m^{*r}
y_1	x_1^{*s}	z_1	x_{11}^{*r}	...	x_{1m}^{*r}
\vdots	\vdots	\vdots	\vdots	...	\vdots
\vdots	\vdots	\vdots	\vdots	...	\vdots
\vdots	\vdots	\vdots	$x_{n_{sub}1}^{*r}$...	$x_{n_{sub}m}^{*r}$
\vdots	\vdots	\vdots	-	...	-
\vdots	\vdots	\vdots	\vdots	\vdots	\vdots
y_n	x_n^{*s}	z_n	-	...	-
(b) Outcome-calibration study					
Y^{*s}	X	Z	Y_1^{*r}	...	Y_m^{*r}
y_1^{*s}	x_1	z_1	y_{11}^{*r}	...	y_{1m}^{*r}
\vdots	\vdots	\vdots	\vdots	...	\vdots
\vdots	\vdots	\vdots	\vdots	...	\vdots
\vdots	\vdots	\vdots	$y_{n_{sub}1}^{*r}$...	$y_{n_{sub}m}^{*r}$
\vdots	\vdots	\vdots	-	...	-
\vdots	\vdots	\vdots	\vdots	\vdots	\vdots
y_n^{*s}	x_n	z_n	-	...	-

Calibration study

A calibration study is a special type of sub-study where two types of error-prone substitute measurement methods are used to measure the covariate or outcome: a substitute measurement prone to systematic measurement error and a substitute measurement prone to random measurement error (Table 3). Table 3a shows the structure of a calibration study for covariate measurement error. All n individuals in the main study have obtained measures of the outcome Y , the error-prone substitute covariate X^{*s}

Table 4

Data structure of external validation studies. An error-prone covariate or outcome is measured in the main study and the true covariate or outcome is measured in a small external set. The superscript $*$ indicates that there is random or systematic measurement error in the variables.

(a1) External covariate-validation study (main study)		
Y	X^*	Z
y_1	x_1^*	z_1
\vdots	\vdots	\vdots
\vdots	\vdots	\vdots
y_n	x_n^*	z_n
(a2) External covariate-validation study (external part)		
X	X^*	Z
x_1	x_1^*	z_1
\vdots	\vdots	\vdots
$x_{n_{ex}}$	$x_{n_{ex}}^*$	$z_{n_{ex}}$
(b1) External outcome-validation study (main study)		
Y^*	X	Z
y_1^*	x_1	z_1
\vdots	\vdots	\vdots
\vdots	\vdots	\vdots
y_n^*	x_n	z_n
(b2) External outcome-validation study (external part)		
Y	Y^*	
y_1	y_1^*	
\vdots	\vdots	
$y_{n_{ex}}$	$y_{n_{ex}}^*$	

and the covariates Z . The error-prone substitute covariate X^{*s} is systematically different from X , or, $E(X^{*s}|X) \neq X$ (systematic measurement error). Additionally, a random subset of n_{sub} individuals ($n_{sub} < n$) have m replicates of the error-prone substitute measure X_j^{*r} , where $E(X_j^{*r}|X) = X$ for $j = 1 \dots m$ (random measurement error). The same structure holds for a calibration study for outcome measurement error, as shown in Table 3b. An example of an calibration study for outcome measurement error is a study of sodium intake measured by a 24-hour recall (assumed systematic measurement error) and urinary biomarkers (assumed random measurement error) [31].

External validation study

In an external validation study the error-free reference covariate values X or outcome values Y are observed in a small set of individuals not included in the main study (Table 4). Table 4a shows the structure of an external validation study for covariate measurement error (Table 4a1 shows the main study and Table 4a2 the external part). In all n individuals in the main study measures are obtained of outcome Y , the error-prone substitute covariate X^* and the covariates Z . Additionally, there is an external data set comprising of individuals on whom measures are obtained of the error-free reference covariate X , the error-prone substitute covariate X^* and the other covariates Z . Table 4b shows the structure of an external validation study for outcome measurement error (Table 4b1 shows the main study and Table 4b2 shows the external part). In this setting, there is an external data set comprising of individuals of whom measures are obtained of the error-free reference outcome Y and the error-prone substitute outcome Y^* . The

external data set does not need to comprise measures of the covariates. An example of an external validation study for outcome measurement error is a trial designed to study the efficacy of iron supplementation in pregnant women where haemoglobin is measured in capillary blood samples (error-prone substitute measure) instead of in venous blood samples (error-free reference measure) [36].

3. Measurement error correction

In Section 2.1, the calibration model matrix Λ and the measurement error model matrix Θ were introduced. These matrices quantify the bias in the naive analysis, i.e., the analysis that does not take the measurement error in X^* or Y^* into account. In the following sections, measurement error correction methods are introduced that utilize the matrices Λ and Θ .

The standard method for covariate measurement error correction that uses the calibration model matrix Λ is *standard regression calibration (RC)* [14,15]. *Standard RC* can be applied in all four types of studies from the previous section. In addition, *validation RC*, an adapted version of *standard RC* for internal validation studies, is the standard covariate measurement error correction method for internal validation studies [2]. Further, the standard method for outcome measurement error correction that uses the measurement error model matrix Θ is *standard method of moments (MM)* [1]. *Standard MM* can be applied in internal and external validation studies, and calibration studies.

Standard RC and *standard MM* do not make the most efficient use of the information available in internal validation studies and calibration studies [2]. More efficient methods for measurement error correction methods are therefore implemented in **meCOR**. A more efficient RC estimator, called *efficient RC*, was introduced by Spiegelman et al. [30]. A more efficient MM estimator was introduced by Keogh et al. [31], which is called the Buonaccorsi approach using the method of moments. For simplicity, we will refer to this method as *efficient MM*.

Likewise, in replicates studies, *standard RC* does not make the most efficient use of the information available [33]. The *standard RC* method is sub-optimal in terms of efficiency, since the method depends on the ordering of the replicate measurements [33]. This can be intuitively understood as follows. The *standard RC* regresses the mean of all but the first replicate on the first replicate, but this could as easily be exchanged with the second replicate. Therefore, different approaches are possible (e.g., maximum likelihood) [33][12], showed how a standard random-intercepts model can be used to obtain *maximum likelihood (ML)* estimates that are more efficient than *standard RC*, at the cost of some additional parametric assumptions, discussed in Section 3.3.

Section 3.1 introduces *standard RC* and *validation RC* for covariate measurement error correction, and *standard MM* for outcome measurement error correction. *Efficient RC* and *efficient MM* are introduced in Section 3.2 and the maximum likelihood approach for replicates studies is introduced in Section 3.3. When no information is available to estimate the parameters of the measurement error model, a *sensitivity analysis* or *quantitative bias analysis* can be used to analyse the sensitivity of study results to measurement error [38,39]. An approach for conducting *sensitivity analyses* is discussed in Section 3.4.

3.1. Standard measurement error correction

3.1.1. Covariate measurement error

In *standard RC*, the biased least squares estimator $\hat{\beta}^*$ is multiplied by the inverse of an estimate of the calibration model matrix Λ to give a consistent and unbiased estimator of β , denoted $\hat{\beta}_{RC}$:

$$\hat{\beta}_{RC} = \hat{\beta}^* \hat{\Lambda}^{-1} \quad (9)$$

Standard RC can be applied using all four types of validation studies (Section 2.2).

To construct the calibration model matrix Λ (see equation (4)), we estimate its components $\lambda = (\lambda_{X^*}, \lambda_0, \lambda_Z)$, from the linear calibration model:

$$E(X|X^*, Z) = \lambda_{X^*} X^* + \lambda_0 + \lambda_Z Z', \quad (10)$$

using least squares. Here, λ_Z is a $1 \times k$ matrix. Throughout, we assume that the calibration model matrix is correctly specified. To obtain estimates of the parameters of interest λ in an internal validation study (Table 1a) and external validation study (Table 4a), the error-free reference measure X is regressed on the error-prone substitute measure X^* and the other covariates Z . To obtain estimates of the parameters of interest λ in a replicates study (Table 2a), the mean of all replicates except the first replicate (i.e., $X_2^{*r}, \dots, X_m^{*r}$) is regressed on the first replicate X_1^* and the other covariates Z . To obtain estimates of the parameters of interest λ in a calibration study (Table 3a), the mean of the replicates $X_1^{*r}, \dots, X_m^{*r}$ with random measurement error is regressed on the measurement X^* s with systematic measurement error and the other covariates Z .

An adapted version of *standard RC* in internal validation studies is *validation RC* [2]. In *validation RC*, the outcome Y is regressed on the calibrated values X_{cal} and Z . The calibrated values X_{cal} are constructed as follows: if X is observed, $X_{cal} = X$, and if X is not observed, $X_{cal} = E(X|X^*, Z)$. The parameters from the regression of Y on X_{cal} and Z are estimates of our parameters of interest β in Eq. (5). Note that *standard RC* described above is identical to using $X_{cal} = E(X|X^*, Z)$ for all X [7].

3.1.2. Outcome measurement error

In *standard MM*, the biased least squares estimator $\hat{\beta}^*$ is multiplied by the inverse of an estimate of the outcome measurement error model matrix Θ to give a consistent and unbiased estimator of β , denoted $\hat{\beta}_{MM}$:

$$\hat{\beta}_{MM} = (\hat{\beta}^*, 1) \hat{\Theta}^{-1}. \quad (11)$$

Standard MM can be applied using internal and external validation studies, and calibration studies (Section 2.2).

To construct the outcome measurement error model matrix Θ (see Eq. (7)), we estimate its components $\theta = (\theta_0, \theta_1)$ from the linear measurement error model $E(Y^*|Y) = \theta_0 + \theta_1 Y$ using least squares. Throughout, we assume that the measurement error model matrix is correctly specified. To obtain estimates of the parameters of interest θ in an internal validation study (Table 1b) and an external validation study (Table 4b), the error-prone substitute measurement Y^* is regressed on the error-free reference measurement Y . To obtain estimates of the parameters of interest θ in a calibration study (Table 3b), the measurement Y^* s with systematic measurement error is regressed on the mean of the replicates $Y_1^{*r}, \dots, Y_m^{*r}$ with random measurement error, thereby correcting for the measurement error bias in the estimated $\hat{\theta}$ using standard RC (implying that $m > 1$).

3.1.3. Differential outcome measurement error in univariable analyses

For the special case of differential measurement error, the outcome measurement error model matrix Θ (see Eq. (8)), can be constructed as follows. We estimate its components $\theta = (\theta_{00}, \theta_{01}, \theta_{10}, \theta_{11})$ from the measurement error model $E(Y^*|Y, X) = \theta_{00} + (\theta_{01} - \theta_{00})X + \theta_{10}Y + (\theta_{11} - \theta_{10})XY$. This model can be fitted directly in an internal validation study (Table 1b), provided that the random internal subset includes exposed (i.e., $X = 1$) and non-exposed individuals (i.e., $X = 0$). The model can be fitted in an external validation study (Table 4b), provided that X is measured, and that exposed and non-exposed individuals are included in the external set.

3.1.4. Variance estimation

The variance of the *standard RC* estimator can be estimated using the multivariate delta method [17] or the zero-variance method [40]. Confidence intervals can then be obtained by constructing Wald-type confidence intervals using one of the former two methods. Additionally, confidence intervals can be obtained by the stratified bootstrap, by sampling the observations in the internal subset separately from the observations outside the internal subset. The variance of the *standard MM* estimator can also be estimated with the multivariate delta method, the zero-variance method or the stratified bootstrap. Additionally, for *standard RC*, confidence intervals for $\hat{\beta}_{X_{RC}}$ (the first element of the $\hat{\beta}_{RC}$) can be obtained by the Fieller method [33]. For *standard MM*, confidence intervals for $\hat{\beta}_{X_{MM}}$ and $\hat{\beta}_{Z_{MM}}$ (the first two elements of the $\hat{\beta}_{MM}$) can be obtained by the Fieller method [36]. Details of these procedures can be found in the appendix Section A.1.

3.2. More efficient measurement error correction

3.2.1. Covariate measurement error

Efficient RC can be used in internal validation studies or calibration studies [30]. It pools the *standard RC* estimate with an internal estimate for β obtained in the internal validation study or calibration study.

In internal validation studies, the error-free reference covariate X is obtained in an internal subset of the main study (Table 1a). By regressing the outcome Y on X and the other covariates Z using least squares in the internal subset, one obtains an unbiased estimate for our parameters of interest β . Denote this estimator by $\hat{\beta}_I$. This internal estimator $\hat{\beta}_I$ can then be combined with the *standard RC* estimator $\hat{\beta}_{RC}$ defined in Eq. (9), by taking the inverse variance weighted mean of the two estimates:

$$\hat{\beta}_{ERC} = \left[\hat{\Sigma}_{\beta_{RC}}^{-1} + \hat{\Sigma}_{\beta_I}^{-1} \right]^{-1} \left[\hat{\Sigma}_{\beta_{RC}}^{-1} \hat{\beta}_{RC} + \hat{\Sigma}_{\beta_I}^{-1} \hat{\beta}_I \right], \quad (12)$$

where $\hat{\Sigma}_{\beta_{RC}}^{-1}$ is the variance-covariance matrix obtained from the multivariate delta method and $\hat{\Sigma}_{\beta_I}$ is the standard variance-covariance matrix of a least squares estimator. The *efficient RC* estimator defined above is an unbiased, consistent and the most efficient estimator for β if sampling into the internal validation set is unbiased (e.g., if the validation study is a random subset of participants) [30].

In calibration studies, the covariate X is observed with random measurement error in an internal subset of the main study (Table 3a). If at least 2 replicates are available, an unbiased estimator for β can be obtained by using the *standard RC* estimator for a replicates study (see Section 3.1) in the internal subset. Again, denote this estimator by $\hat{\beta}_I$. Then, the estimate obtained from the internal subset can be pooled with the *standard RC* estimate following Eq. (12). Alternatively, an unbiased estimator for β using the replicates in the internal subset can be obtained by using the ML estimation discussed in Section 3.3. Again, this estimate can then be pooled with the *standard RC* estimate following Eq. (12).

3.2.2. Outcome measurement error

Efficient MM can be used in internal validation studies or calibration studies [31]. It pools the *standard MM* estimate with an internal estimate for β obtained in the internal validation study or calibration study.

In internal validation studies, the error-free reference outcome Y is obtained in an internal subset of the main study (Table 1b). By regressing Y on the covariates X and Z using least squares in the internal subset, one obtains an unbiased estimator for β . Denote this estimator by $\hat{\beta}_I$. In calibration studies, the outcome is observed with random measurement error in an internal subset of

the main study (Table 3b). The internal estimator $\hat{\beta}_I$ is obtained by regressing the outcome $Y^{*,r}$ with random measurement error on the covariates X and Z using least squares in the internal subset. Using the outcome with random measurement error will lead to the unbiased estimation of the association under study since random outcome measurement error does not bias the association. A single measurement with random measurement error (i.e., $m = 1$ in Table 1b) is sufficient to obtain an internal estimate. However, if the outcome with random measurement error is observed more than once, the mean of the measures $Y_1^{*,r}, \dots, Y_m^{*,r}$ can be used and regressed on the covariates X and Z . Subsequently, the estimate obtained from the internal subset in an internal validation study or calibration study can be pooled with the *standard MM* estimate following Eq. (12), by replacing the *standard RC* estimate with the *standard MM* estimate in the equation.

3.2.3. Differential outcome measurement error in univariable analyses

In internal validation studies, the internal estimator $\hat{\beta}_I$ can be obtained by regressing Y on the covariates X and Z using least squares. We assume that the internal subset is a random subset of the main study, and hence that exposed and unexposed are included in the internal subset. Subsequently, the estimate obtained from the internal subset in an internal validation study can be pooled with the *standard MM* estimate following Eq. (12), by replacing the *standard RC* estimate with the *standard MM* estimate in the equation.

Variance estimation

The variance of the *efficient RC* estimator can be obtained from the following:

$$\hat{\Sigma}_{\beta_{ERC}} = \left[\hat{\Sigma}_{\beta_{RC}}^{-1} + \hat{\Sigma}_{\beta_I}^{-1} \right]^{-1}.$$

The variance of the *efficient RC* estimator can also be obtained by stratified bootstrapping, by sampling the observations in the internal subset separately from the observations outside the internal subset. Confidence intervals can be obtained by constructing Wald-type confidence intervals using one of the former two variances or by stratified percentile bootstrap. The same applies for the *efficient MM* estimator.

3.3. Maximum likelihood estimation for replicates studies

The use of a standard random-intercepts model to obtain maximum likelihood (ML) estimates for β in replicates studies was introduced by Bartlett et al. [12]. To explain the ML method for replicates studies, we add the index $i = 1, \dots, n$ to our notation in the outcome model:

$$Y_i = \beta_X X_i + \beta_0 + \beta_Z Z_i' + e_i, \quad \text{Var}(e_i) = \sigma^2,$$

where we again assume that $E(e_i) = 0$ and $\text{Cov}(e_i, X_i) = \text{Cov}(e_i, Z_i) = 0$. Further, $Z_i = (Z_{i1}, \dots, Z_{ik})$ and β_Z is again a $1 \times k$ matrix. On top of these assumptions, we also assume that the e_i are normal and independently distributed. Additionally, assume that X_i is normally distributed given Z_i , with,

$$E(X_i | Z_i) = \rho_0 + \rho_Z Z_i' \quad \text{and} \quad \text{Var}(X_i | Z_i) = \sigma_{X_i | Z_i}^2,$$

where ρ_Z is a $1 \times k$ matrix. In a replicates study, X_i is not observed. Instead, m replicates of the error-prone measurement $X_i^{*,r} = (X_{i1}^{*,r}, \dots, X_{im}^{*,r})$ are observed, for $i = 1, \dots, n$. In a full-replicates study (Table 2a), we assume that the number of replicate measurements $m \geq 2$ is constant for every individual. In a partial-replicates study (Table 2b), we assume that the number of replicates $m \geq 2$ is constant in the replicate sub-study and $m = 1$ in the main study. These measurements are assumed to follow the following random measurement error model:

$$X_{ij}^{*,r} = X_i + U_{ij}, \quad \text{Var}(U_{ij}) = \tau^2, \quad j = 1, \dots, m,$$

where we again assume that $E(U_{ij}) = 0$, $\text{Cov}(U_{ij}, X_i) = 0$, and that the measurement error in non-differential, i.e., the errors U_{ij} are independent of the errors e_i in the outcome model described above. In addition, we also assume that the errors U_{ij} are normal and independently distributed.

We consider the likelihood function when only Y_i , X_i^{*r} and Z_i are observed. The log likelihood can be factorized as follows:

$$\ell(\theta|Y_i, X_i^{*r}, Z_i) = \log(f(Y_i|Z_i, \theta)) + \log(f(X_i^{*r}|Y_i, Z_i, \theta)), \quad (13)$$

where $\theta = (\beta_X, \beta_0, \beta_Z, \sigma^2, \rho_0, \rho_Z, \sigma_{X|Z}^2, \tau^2)$. From the assumptions that $X_i|Z_i$ is normally distributed, the e_i are normally distributed and that $X_i|Z_i$ and e_i are independent, Bartlett et al. show in [12] that Y_i given Z_i is normal with mean $\delta_0 + \delta_Z Z_i$ and variance $\sigma_{Y|Z}^2$, where δ_Z is a $1 \times k$ matrix. Furthermore, since $X_i|Z_i$ and $Y_i|Z_i$ are jointly normal, $X_i|Y_i, Z_i$ is also normal. Bartlett et al. show in [12] that we can therefore write:

$$X_i = \kappa_0 + \kappa_Y Y_i + \kappa_Z Z_i + b_i,$$

where $b_i \sim N(0, \sigma_{X|Y,Z}^2)$. Then, since $X_{ij}^* = X_i + U_{ij}$, it follows from the above equation that,

$$X_{ij}^* = \kappa_0 + \kappa_Y Y_i + \kappa_Z Z_i + b_i + U_{ij},$$

where $U_{ij} \sim N(0, \tau^2)$ is independent of b_i [12] and κ_Z is a $1 \times k$ matrix. Hence, X_{ij}^{*r} given Y_i and Z_i follows a random-intercepts model with fixed effects of Y_i and Z_i , random intercepts variance $\sigma_{X|Y,Z}^2$ and within subject variance τ^2 .

The parameter vector $\zeta = (\delta_0, \delta_Z, \sigma_{Y|Z}^2, \kappa_0, \kappa_Y, \kappa_Z, \sigma_{X|Y,Z}^2, \tau^2)$ is a one-to-one function of the original model parameter vector $\theta = (\beta_X, \beta_0, \beta_Z, \sigma^2, \rho_0, \rho_Z, \sigma_{X|Z}^2, \tau^2)$. Accordingly, Bartlett et al. show in [12] that the ML estimate for ζ can be obtained by maximizing the two likelihood components of Eq. (13) separately. The likelihood component corresponding to $f(Y_i|Z_i, \zeta)$ in Eq. (13) can be maximized by fitting the least squares regression of Y_i on Z_i . The likelihood component corresponding to $f(X_i^{*r}|Y_i, Z_i, \zeta)$ in Eq. (13) can be maximized by fitting a random-intercepts model for X_i^{*r} given Y_i and Z_i .

An ML estimate for β can now be obtained by the following formulas:

$$\begin{aligned} \beta_X &= \kappa_Y \times \frac{\sigma_{Y|Z}^2}{\sigma_{X|Y,Z}^2 + \kappa_Y^2 \sigma_{Y|Z}^2}, \\ \beta_0 &= \delta_0 - \beta_X \rho_0 = \delta_0 - \beta_X \{\kappa_0 + \kappa_Y \delta_0\}, \\ \beta_Z &= \delta_Z - \beta_X \rho_Z = \delta_Z - \beta_X \{\kappa_Z + \kappa_Y \delta_Z\}. \end{aligned}$$

The estimator $\hat{\beta}_{ML} = (\hat{\beta}_{X_{ML}}, \hat{\beta}_{0_{ML}}, \hat{\beta}_{Z_{ML}})$ can be obtained by replacing the parameters from parameter vector ζ by their estimates in the above equations.

Variance estimation

The variance of the maximum likelihood estimator can be estimated with the multivariate delta method [12]. Confidence intervals can then be obtained by constructing Wald-type confidence intervals. Confidence intervals can also be obtained by stratified bootstrap, by sampling the observations in the internal subset separately from the observations outside the internal subset. Details of these procedures can be found in the appendix Section A.2.

3.4. Sensitivity analyses

Information from a validation study may not always be available. In that case, a formal correction is not possible. Nevertheless, when measurement error in a covariate or the outcome is

expected, one may check how sensitive study results are to that measurement error. Literature or expert knowledge can be used to inform this sensitivity analysis, e.g., by hypothesizing possible ranges for the parameter values of the measurement model.

When random covariate measurement error is expected, speculation is needed of the values of τ^2 , i.e., the variance of the random measurement error. Additionally, when systematic covariate measurement error is suspected, speculation is needed about the parameter values of the calibration model described by Eq. (10). When systematic outcome measurement error is suspected, speculation is needed about the parameter values of the outcome measurement error model, described in Eq. (5).

4. The R package mecor

The R package **mecor** offers functionality to correct for measurement error in a continuous covariate or outcome in linear models with a continuous outcome. The main model fitting function in **mecor** is `mecor`:

```
mecor(formula, data, method, B)
```

The function fits the linear model defined in `formula`, corrected for the measurement error in one of the variables. The arguments are as follows:

- `formula` a formula object, with the response on the left of a '~' operator and the terms, separated by + operators, on the right. This argument takes the form `outcome ~ MeasError(substitute, reference, replicate, differential) + covariates` for covariate measurement error, and `MeasError(substitute, reference, replicate, differential) ~ covariates` for outcome measurement error. The `MeasError` object can be used for measurement error correction in internal validation, replicates and calibration studies. For external validation studies or sensitivity analyses of systematic measurement error, the object `MeasErrorExt(substitute, model)` is used instead of a `MeasError` object. For sensitivity analyses of random measurement error, the object `MeasErrorRandom(substitute, error)` is used.
- `data` a `data.frame` containing the variables in the model specified by `formula`.
- `method` specifies the method used for measurement error correction. The options are 'standard' for standard RC and standard MM, 'valregcal' for validation RC, 'efficient' for efficient RC and efficient MM, and 'mle' for maximum likelihood estimation.
- `B` number of bootstrap samples used for standard error estimation. The default is set to 0.

An object of class `mecor` can be summarised using the `summary` function:

```
summary(object, alpha, zero var, fieller)
```

The arguments are as follows:

- `object` an object of class `mecor`.
- `alpha` a numeric indicating the probability of obtaining a type II error. Defaults to 0.05.
- `zero var` a boolean indicating whether confidence intervals using the zero-variance method [40] must be printed. Only available for `mecor` objects fitted with `method` equal to 'standard'. Defaults to FALSE.
- `fieller` a boolean indicating whether confidence intervals using the fieller method [33,36] must be printed. Only available for `mecor` objects fitted with `method` equal to 'standard'. Defaults to FALSE.

The default summary object of an object of class `mecor` prints standard errors and confidence intervals obtained by the `delta` method. See the various 'Variance estimation' paragraphs in [Section 3](#) for a description of the methods for variance estimation.

The formula argument in `mecor` contains a `MeasError` object, a `MeasErrorExt` object or a `MeasErrorRandom` object. All three objects are described below.

4.1. The *MeasError* object

To correct for measurement error using an internal validation study, a replicates study or a calibration study, the formula argument in `mecor` contains a `MeasError` object on the right-hand side (covariate measurement error) or left-hand side (outcome measurement error). The `MeasError` object can be used for random and systematic measurement error correction, depending on the method used to correct for the measurement error in `mecor`:

`MeasError(substitute, reference, replicate, differential)` with the arguments being described as follows:

- `substitute` the error-prone measurement;
- `reference` the gold-standard reference measurement, to be used in case of an internal validation study, else `NULL`;
- `replicate` (a vector of) the replicate measurement of the error-prone substitute measurement, to be used in case of a replicates study or calibration study, else `NULL`;
- `differential` the binary exposure on which the outcome measurement error structure is dependent, to be used for differential outcome measurement error in univariable analyses, else `NULL`.

Depending on the type of validation study used, either argument `reference` (internal validation study) or `replicate` (replicates study or calibration study) should be used, but never both.

4.2. The *MeasErrorExt* object

To correct for measurement error using an external validation study, the formula object in `mecor` contains a `MeasErrorExt` object on the right-hand side (covariate measurement error) or left-hand side (outcome measurement error):

`MeasErrorExt(substitute, model)` with the arguments being described as follows:

- `substitute` the error-prone measurement;
- `model` a fitted `lm` object of the calibration model in [Eq. \(10\)](#) (covariate measurement error) or the measurement error model in [Eq. \(5\)](#) (outcome measurement error). Or alternatively, a list with named arguments `coef` containing a vector of the coefficients of the calibration model or measurement error model and named argument `vcov` containing a matrix of the corresponding variance-covariance matrix. The argument `vcov` is not required.

The argument `model` is also used for conducting a sensitivity analysis by making informed guesses about the parameters of the calibration model (covariate measurement error) or measurement error model (outcome measurement error).

4.3. The *MeasErrorRandom* object

When random measurement error in a covariate is suspected but cannot be quantified, the `MeasErrorRandom` object can be used to conduct a sensitivity analysis:

`MeasErrorRandom(substitute, variance)` with the arguments being described as follows:

- `substitute` the error-prone measurement;
- `variance` a numeric indicating the random measurement error variance in the substitute measurement, i.e., the parameter value of τ^2 in [Eq. \(2\)](#).

5. Examples

Six simulated datasets are included in the package **mecor**. These datasets mimic real datasets and represent the data structures described in [Section 2.2](#). There is an internal validation study with covariate measurement error (`vat`), an internal validation study with outcome measurement error (`haemoglobin`), a replicates study (`bloodpressure`) and a calibration study with outcome measurement error (`sodium`). The dataset `vat_ext` provides an external validation study for the `vat` dataset, and the dataset `haemoglobin_ext` provides an external validation study for the `haemoglobin` dataset. These datasets are described and analysed in the following sections.

5.1. Internal validation study

The dataset `vat` is a simulated dataset, representing the structure of the internal covariate-validation study shown in [Table 1a](#). The dataset is inspired by the Netherlands Epidemiology of Obesity (NEO) study [\[41\]](#) and was used as the motivating example in a study investigating measurement error correction by Nab et al. [\[42\]](#). The dataset represents a cross-sectional study of the association between visceral adipose tissue and insulin resistance. Visceral adipose tissue measures are expensive and therefore only available in 40% of the study population. Waist circumference measures however provide a simple proxy for visceral adipose tissue and are observed in the full study population. The dataset `vat` contains 650 observations of the natural logarithm of the outcome insulin resistance (`ir_ln`, fasting glucose (mmol/L) x fasting insulin (mU/L) / 22.5), the standardised error-prone substitute measurement of the exposure waist circumference (`wc`, cm), the covariates `sex` (`sex`, 0 = male, 1 = female), `age` (`age`, years), and standardised total body fat (`tbtf`, %), and the standardised error-free measurement of the exposure visceral adipose tissue (`vat`, cm²).

```
R> head(vat)
R> data("vat", package = "mecor")

      ir_ln      wc sex age      tbtf vat
1 -0.09341837 -1.3136816 1  48 -0.6571345 NA
2  0.16820894 -2.0336624 0  54 -1.5882163 NA
3  0.57299976 -0.2611214 0  46 -1.1033709 NA
4  0.63677178  0.8631987 0  55 -1.4785869 0.5083247
5  0.92908882 -1.2054861 1  61  0.9020136 NA
6 -0.72410039 -2.5032852 1  47 -0.9584166 NA
```


By ignoring the measurement error in wc, a linear model can be fitted to the data as follows:

```
R> lm(ir_ln ~ wc + sex + age + tbf, data = vat)

Call:
lm(formula = ir_ln ~ wc + sex + age + tbf, data = vat)
Coefficients:
(Intercept)      wc      sex      age      tbf
 0.50976 0.09697 -0.70953 0.01133 0.38783
```

The coefficients of this model will however be biased due to the measurement error in wc. The measurement error in wc can be corrected for using standard regression calibration (RC) as follows:

```
R> mecor(ir_ln ~ MeasError(wc, reference = vat) + sex + age + tbf,
+       data = vat,
+       method = "standard")
```

```
Call:
mecor(formula = ir_ln ~ MeasError(wc, reference = vat) + sex +
      age + tbf, data = vat, method = "standard")
```

```
Coefficients Corrected Model:
(Intercept)      vat      sex      age      tbf
0.473398350 0.207598087 -0.438453038 0.009477677 0.270864391
```

```
Coefficients Uncorrected Model:
(Intercept)      wc      sex      age      tbf
0.50976395 0.09697045 -0.70952736 0.01132712 0.38782671
```

Stratified percentile bootstrap confidence intervals of the coefficients of the corrected model can be obtained by using the argument B in the function mecor. To obtain standard errors and confidence intervals using the Fieller method or zero-variance method, the arguments zerovar and fieller of the summary object are set to TRUE:

```
R> set.seed(20210526)
R> mecor_fit <-
+   mecor(ir_ln ~ MeasError(wc, reference = vat) + sex + age + tbf,
+   data = vat,
+   method = "standard",
+   B = 999
+ )
R> summary(mecor_fit, zerovar = TRUE, fieller = TRUE)
```

```
Call:
mecor(formula = ir_ln ~ MeasError(wc, reference = vat) + sex +
      age + tbf, data = vat, method = "standard", B = 999)
```

```
Coefficients Corrected Model:
      Estimate      SE SE (btstr) SE (zerovar)
(Intercept) 0.473398 0.146766 0.134792 0.126665
vat         0.207598 0.034210 0.035302 0.029534
sex        -0.438453 0.079596 0.077277 0.069276
age         0.009478 0.002598 0.002409 0.002236
tbf         0.270864 0.036662 0.034541 0.031805
```

```
95 % Confidence Intervals:
      Estimate      LCI      UCI LCI (btstr) UCI (btstr)
(Intercept) 0.473398 0.185743 0.761054 0.214303 0.721416
vat         0.207598 0.140549 0.274648 0.147096 0.284406
sex        -0.438453 -0.594458 -0.282448 -0.569175 -0.258816
age         0.009478 0.004385 0.014570 0.004666 0.013956
tbf         0.270864 0.199007 0.342721 0.197417 0.329173
```

```
      LCI (zerovar) UCI (zerovar) LCI (fieller) UCI (fieller)
(Intercept) 0.225140 0.721657      NA      NA
vat         0.149712 0.265484 0.145068 0.281464
sex        -0.574231 -0.302675      NA      NA
age         0.005096 0.013860      NA      NA
tbf         0.208528 0.333201      NA      NA
```

Bootstrap Confidence Intervals are based on 999 bootstrap replicates using percentiles

The measurement error is corrected for by application of regression calibration

Coefficients Uncorrected Model:

	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	0.5097640	0.1264211	4.0323	6.185e-05
wc	0.0969705	0.0137957	7.0290	5.308e-12
sex	-0.7095274	0.0390086	-18.1890	< 2.2e-16
age	0.0113271	0.0022048	5.1374	3.695e-07
tbf	0.3878267	0.0201489	19.2481	< 2.2e-16

95% Confidence Intervals:

	Estimate	LCI	UCI
(Intercept)	0.509764	0.261517	0.758011
wc	0.096970	0.069881	0.124060
sex	-0.709527	-0.786127	-0.632928
age	0.011327	0.006998	0.015657
tbf	0.387827	0.348261	0.427392

Residual standard error: 0.3123469 on 645 degrees of freedom

In addition to standard RC, efficient RC (method = "efficient") or validation RC (method = "valregcal") can also be used to correct for the measurement error in the error-prone covariate wc.

The dataset haemoglobin is a simulated dataset, representing the structure of the internal outcome-validation study shown in Table 1b. The dataset is inspired by a trial investigating the efficacy of low-dose iron supplements [43] and was used as the motivating example for a study investigating measurement error correction in trial endpoints by Nab et al. [36]. The dataset represents a trial investigating the effect of low-dose iron supplements during pregnancy on haemoglobin levels at delivery. Haemoglobin levels were measured in venous blood in approximately 25% of the subjects (reference measurement), and were measured in capillary blood in all subjects (substitute measurement). The dataset haemoglobin contains 400 observations of the error-prone capillary haemoglobin levels (capillary, g/L), an indicator of whether the subject was randomised to receive the low-dose iron supplement (20 mg/d) (supplement, 0 = no, 1 = yes), and the error-free reference venous haemoglobin levels (venous, g/L).

```
R> data("haemoglobin", package = "mecor")
R> tail(haemoglobin)
```

	capillary	supplement	venous
395	124.0489	1	NA
396	127.1005	0	127.9526
397	132.1858	1	NA
398	123.4427	0	NA
399	125.2438	1	NA
400	124.0738	0	NA

The measurement error in capillary can be accounted for by using standard method of moments (MM) as shown in the following:

```
R> mecor(MeasError(capillary, reference = venous) ~ supplement,
+       data = haemoglobin,
+       method = "standard")
```

Call:

```
mecor(formula = MeasError(capillary, reference = venous) ~ supplement,
      data = haemoglobin, method = "standard")
```

Coefficients Corrected Model:

	supplement
(Intercept)	117.99341
supplement	6.97392

Coefficients Uncorrected Model:

	supplement
(Intercept)	124.452261
supplement	7.764702

In addition to standard MM, efficient MM (method = "efficient") can also be used to correct for the measurement error in the error-prone outcome Y_star.

When differential outcome measurement error in capillary haemoglobin measures is suspected, the argument differential of the MeasError object can be used to correct for differential measurement error as follows:

```
R> mecor(MeasError(capillary,
+                 reference = venous,
+                 differential = supplement) ~ supplement,
+       data = haemoglobin,
+       method = "standard")
```

```
Call:
mecor(formula = MeasError(capillary,
                        reference = venous,
                        differential = supplement) ~ supplement,
      data = haemoglobin,
      method = "standard")
```

```
Coefficients Corrected Model:
(Intercept) supplement
118.386903    6.080729
```

```
Coefficients Uncorrected Model:
(Intercept) supplement
124.452261    7.764702
```

Efficient MM (method = "efficient") can also be used to correct for the differential measurement error in the error-prone outcome capillary.

5.2. Replicates study

The dataset `bloodpressure` is a simulated dataset, representing the structure of the replicates study shown in Table 2a. The dataset represents a cross-sectional study of the association between blood pressure and creatinine in pregnant women [44]. Blood pressure measurements are prone to random measurement error. The dataset `bloodpressure` contains 450 observations of serum creatinine (creatinine, mmol/L), age (age, years), and systolic blood pressure (sbp, mm Hg). Systolic blood pressure is measured at 30, 60, 90 and 120 minutes.

```
R> data("bloodpressure", package = "mecor")
R> head(bloodpressure)
```

	creatinine	age	sbp30	sbp60	sbp90	sbp120
1	53.75670	27	120.7987	113.2812	118.0705	124.2282
2	63.08498	36	121.7254	106.8143	118.9882	115.1341
3	60.04718	31	108.8798	119.6577	106.5588	117.5473
4	62.42976	43	116.5566	117.4964	126.3625	121.7148
5	61.31801	25	123.3018	116.4629	112.0310	109.8754
6	50.60952	35	124.9119	129.0927	129.0224	114.0828

In a study estimating the association between serum creatinine and systolic blood pressure, corrected for age, the random measurement error in the error-prone systolic blood pressure measurement at 30 minutes can be accounted for as follows:

```
R> mecor(+ creatinine ~ MeasError(sbp30,
+   replicate = cbind(sbp60, sbp90, sbp120)) + age,
+   data = bloodpressure,
+   method = "standard"
+ )
```

```
Call:
mecor(formula = creatinine ~ MeasError(sbp30, replicate = cbind(sbp60,
  sbp90, sbp120)) + age, data = bloodpressure, method = "standard")
```

```
Coefficients Corrected Model:
(Intercept) cor_sbp30    age
32.3796021  0.1877343  0.1743760
```

```
Coefficients Uncorrected Model:
(Intercept)    sbp30    age
41.3050286  0.1165333  0.1650849
```

Maximum likelihood estimation (method = "mle") can also be used to correct for the measurement error in the error-prone exposure `sbp30`. Note that, in this example dataset, the coefficients of the corrected model using standard RC will differ when `MeasError(sbp60, replicate = cbind(sbp30, sbp90, sbp120))` is used instead of `MeasError(sbp30, replicate = cbind(sbp60, sbp90, sbp120))`. In contrast, the corrected estimated coefficients obtained using maximum likelihood estimation will not change when the order of replicates is changed.

5.3. Calibration study

The dataset `sodium` is a simulated dataset, representing the structure of the outcome calibration study, shown in [Table 3b](#). The dataset represents a randomised controlled trial designed to investigate whether a reduction in sodium intake results in satisfactory blood pressure control [\[45\]](#) and was used as the motivating example for a study investigating measurement error correction in dietary intake [\[31\]](#). Sodium intake of the subjects was measured by a 24h recall and in urine. Sodium intake measured by a 24h recall is assumed prone to systematic measurement error and sodium intake measured in urine is assumed prone to random measurement error. The dataset `sodium` contains 1000 observations of sodium intake measured by a 24h recall (`recall`, mg), an indicator of whether the subject was randomised to their usual diet or sodium-lowering diet (`diet`, 0 = usual, 1 = sodium-lowering), and two measures of urinary sodium (`urinary1`, `urinary2`, mg). The replicate urinary sodium are observed in approximately 50% of the subjects included in the trial.

```
R> data("sodium", package = "mecor")
R> tail(sodium)
```

	recall	diet	urinary1	urinary2
995	3.320633	1	NA	NA
996	3.496626	0	NA	NA
997	3.127590	1	3.818815	4.204880
998	4.363960	0	NA	NA
999	4.009316	1	4.719055	4.389111
1000	3.910490	0	NA	NA

The measurement error in the error-prone exposure `recall` can be accounted for as follows:

```
R> mecor(
+   MeasError(recall, replicate = cbind(urinary1, urinary2)) ~ diet,
+   data = sodium,
+   method = "standard"
+ )
```

Call:

```
mecor(formula = MeasError(recall, replicate = cbind(urinary1,
  urinary2)) ~ diet, data = sodium, method = "standard")
```

Coefficients Corrected Model:

(Intercept)	diet
4.6075011	-0.4843495

Coefficients Uncorrected Model:

(Intercept)	diet
3.8819732	-0.3051777

Efficient MM (`method = 'efficient'`) can also be used to correct for the measurement error in the error-prone outcome `recall`.

5.4. External validation study

The dataset `vat_ext` is a simulated dataset, representing the structure of the external part of the external covariate-validation study shown in [Table 4a](#). The dataset accompanies the dataset `vat` introduced in [Section 5.1](#). The dataset contains 100 observations of the error-free continuous exposure `vat`, the error-prone exposure `wc` and the covariates `sex`, `age` and `tbf`.

```
R> data("vat_ext", package = "mecor")
R> head(vat_ext)
```

	wc	vat	sex	age	tbf
1	-0.01357552	-1.69944962	1	50	-1.17103270
2	1.10201426	1.43889836	0	51	-0.99837467
3	1.23328072	1.24129099	0	54	-0.91030636
4	-0.07849380	0.05219091	0	55	-1.52766077
5	-0.47481715	-0.61165766	1	46	0.28706021
6	-1.33717429	-0.58193963	1	50	0.08718737

Suppose that in the dataset `vat`, the reference measure `vat` had not been observed. Using dataset `vat_ext`, we can correct for the measurement error in `wc` in dataset `vat`. The first step is to fit the calibration model in the external validation study as follows:

```
R> calmod_fit <- lm(vat ~ wc + sex + age + tbf,
+   data = vat_ext))
R> calmod_fit
```


Call:

```
lm(formula = vat ~ wc + sex + age + tbf, data = vat_ext)
Coefficients:
(Intercept)      wc      sex      age      tbf
 0.437466 0.571233 -0.984891 0.001111 0.488749
```

The second step is to use the calibration model `calmod_fit` in the `MeasErrorExt` object as follows:

```
R> data("vat", package = "mecor")
R> mecor(
+   ir_ln ~ MeasErrorExt(wc, calmod_fit) + sex + age + tbf,
+   data = vat,
+   method = "standard"
+ )
```

Call:

```
mecor(formula = ir_ln ~ MeasErrorExt(wc, calmod_fit) + sex +
      age + tbf, data = vat, method = "standard")
Coefficients Corrected Model:
(Intercept)      cor_wc      sex      age      tbf
 0.43550128 0.16975650 -0.54233566 0.01113844 0.30485834
Coefficients Uncorrected Model:
(Intercept)      wc      sex      age      tbf
 0.50976395 0.09697045 -0.70952736 0.01132712 0.38782671
```

Dataset `haemoglobin_ext` is a simulated dataset, representing the structure of the external part of the external outcome-validation study shown in [Table 4b](#). The dataset accompanies the dataset `haemoglobin` introduced in [Section 5.1](#). The dataset contains 100 observations of the error-free outcome venous and the error-prone outcome capillary.

```
R> data("haemoglobin_ext", package = "mecor")
R> head(haemoglobin)
```

```
  capillary  venous
1  104.7269 115.3023
2  133.9946 119.7616
3  104.0304 108.0562
4  119.0214 121.1780
5  114.3891 111.7864
6  111.7754 112.8943
```

Suppose that in the dataset `haemoglobin`, the reference venous haemoglobin levels had not been observed. Using dataset `haemoglobin_ext`, we correct for the measurement error in capillary in the dataset `haemoglobin`, by fitting the measurement error model, as follows:

```
R> memod_fit <- lm(capillary ~ venous, data = haemoglobin_ext)
R> data("iovs", package = "mecor")
R> mecor(MeasErrorExt(capillary, memod_fit) ~ supplement,
+       data = haemoglobin,
+       method = "standard")
```

Call:

```
mecor(formula = MeasErrorExt(capillary, memod_fit) ~ supplement,
      data = haemoglobin, method = "standard")
Coefficients Corrected Model:
(Intercept) supplement
 119.136649    7.227302
Coefficients Uncorrected Model:
(Intercept) supplement
 124.452261    7.764702
```

5.4.1. Sensitivity analyses

Suppose that there is no error-free measure and no external validation study available for dataset `vat`. To investigate the sensitivity of study results to measurement error in variable `vat`, informed guesses of the coefficients of the calibration model are needed. Suppose

one assumes that $E(\text{VAT}|\text{WC}, \text{sex}, \text{age}, \text{tbf}) = 0.4 + 0.6 \times \text{WC} - \text{sex} + 0 \times \text{age} + 0.5 \times \text{TBF}$. A sensitivity analysis could then be conducted as follows:

```
R> data("vat", package = "mecor")
R> mecor_fit_sens <-
+   mecor(ir_ln ~ MeasErrorExt(wc, list(coef = c(0.4, 0.6, -1, 0, 0.5))) +
+     sex + age + tbf,
+     data = vat,
+     method = "standard")
R> mecor_fit_sens
```

Call:

```
mecor(formula = ir_ln ~ MeasErrorExt(wc, list(coef = c(0.4, 0.6,
-1, 0, 0.5))) + sex + age + tbf, data = vat, method = "standard")
```

Coefficients Corrected Model:

(Intercept)	cor_wc	sex	age	tbf
0.44511698	0.16161742	-0.54790994	0.01132712	0.30701800

Coefficients Uncorrected Model:

(Intercept)	wc	sex	age	tbf
0.50976395	0.09697045	-0.70952736	0.01132712	0.38782671

The calibration model matrix used to correct for the measurement error in wc, is saved as matrix in the corfit object attached to mecor_fit_sens:

```
R> mecor_fit_sens$corfit$matrix
```

	Lambda1	Lambda0	Lambda3	Lambda4	Lambda5
Lambda1	0.6	0.4	-1	0	0.5
Lambda0	0.0	1.0	0	0	0.0
Lambda3	0.0	0.0	1	0	0.0
Lambda4	0.0	0.0	0	1	0.0
Lambda5	0.0	0.0	0	0	1.0

In the dataset bloodpressure discussed in [Section 5.2](#), random measurement error is suspected in systolic blood pressure. Suppose now that in the dataset bloodpressure, the three replicate measures sbp60, sbp90, sbp120 had not been observed. Suppose further that a measurement error variance of 30 mm Hg is assumed in the first systolic blood pressure measure sbp30. For measurement error correction, the MeasErrorRandom object can be used, here in combination with zerovariance estimation of standard errors (assuming that there is no uncertainty in the speculated value of the variance of the random measurement error sbp30):

```
R> mecor_fit_random <-
+   mecor(
+     creatinine ~ MeasErrorRandom(sbp30, variance = 30) + age,
+     data = bloodpressure,
+     method = "standard"
+   )
R> summary(mecor_fit_random, zerovar = T)
```

Call:

```
mecor(formula = creatinine ~ MeasErrorRandom(sbp30, variance = 30) +
age, data = bloodpressure, method = "standard")
```

Coefficients Corrected Model:

	Estimate	SE (zerovar)
(Intercept)	33.568149	9.909771
cor_sbp30	0.182509	0.080298
age	0.159752	0.094837

95% Confidence Intervals:

	Estimate	LCI (zerovar)	UCI (zerovar)
(Intercept)	33.568149	14.145355	52.990943
cor_sbp30	0.182509	0.025127	0.339890
age	0.159752	-0.026125	0.345628

The measurement error is corrected for by application of regression calibration

Coefficients Uncorrected Model:

	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	41.305029	6.758932	6.1112	2.155e-09
sbp30	0.116533	0.051271	2.2729	0.02351
age	0.165085	0.094705	1.7431	0.08200

95% Confidence Intervals:

	Estimate	LCI	UCI
(Intercept)	41.305029	28.021799	54.588258
sbp30	0.116533	0.015771	0.217296
age	0.165085	-0.021038	0.351208

Residual standard error: 9.897091 on 447 degrees of freedom

The calibration model matrix used to correct for the measurement error in sbp30, is again saved as `matrix` in the `corfit` object attached to `mecor_fit_random`:

```
R > mecor_fit_random$corfit$matrix
```

	Lambda1	Lambda0	Lambda3
Lambda1	0.6385083	42.39186	0.02922153
Lambda0	0.0000000	1.00000	0.00000000
Lambda3	0.0000000	0.00000	1.00000000

The sensitivity analyses could be expanded to ranges of possible coefficients of the calibration model or assumed variance of the random measurement error.

6. Conclusion

We demonstrated how measurement error correction methods can be applied using our R package **mecor**. These correction methods can be used in linear models with a continuous outcome when there is measurement error in the outcome or in a continuous covariate. The package accommodates measurement error correction methodology for a wide range of data structures: internal and external validation studies, replicates studies, and calibration studies. Various measurement error correction methods are implemented in the package: RC, MM and correction based on maximum likelihood estimation. For standard error estimation, the delta method and bootstrap are implemented for all methods. The package also facilitates sensitivity analysis or quantitative bias analysis when no data are available to estimate the parameters of the measurement error model, but the assumption of no measurement error is not warranted. A vast body of literature exists comparing the relative performance of the measurement error correction methods implemented in **mecor** [42,46] and in comparison, with other methods e.g., simulation-extrapolation [47,48], multiple imputation methods [49,50] and Bayesian methods [11]. We focused on studies in which interest lies in estimating a covariate-outcome association. In other types of studies, e.g., prediction studies, considerations for measurement error correction are different and may not even require corrections [51,52]. In future updates of the package, the measurement error correction methods may be extended to time-to-event [16] and binary outcomes, and multiple variables with measurement error [17,27].

Computational details

The results in this paper were obtained using R 4.0.2. R itself and **mecor** are available from the Comprehensive R Archive Network (CRAN) at <https://CRAN.R-project.org/>. The latest versions of **mecor** are available on www.github.com/LindaNab/mecor.

Declaration of Competing Interest

The authors declare no conflict of interest.

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Appendix A. Variance estimation

A1. Standard regression calibration

Covariate measurement error. The variance–covariance matrix for the standard regression estimator $\hat{\beta}_{RC}$ can be approximated by using the multivariate delta method by [17], given by

$$\hat{\Sigma}_{\beta_{RC}}(j_1, j_2) = (\hat{A}' \hat{\Sigma}_{\beta^*} \hat{A})_{j_1, j_2} + \hat{\beta}^* \hat{\Sigma}_{A, j_1, j_2} \hat{\beta}^{*'}', \quad j_1, j_2 = 1, \dots, (k+2), \quad (14)$$

where \hat{A} is the inverse of the calibration model matrix \hat{A} . Further, $\hat{\Sigma}_{\beta^*}$ is the variance–covariance matrix obtained from the naive regression defined in Eq. (2) in the main text and $\hat{\Sigma}_{A, j_1, j_2}$ is the $(k+2) \times (k+2)$ matrix relating the j_1 th and j_2 th column of A (we refer to Appendix of [17] for a derivation). Additionally, the so-called zero-variance variance–covariance matrix for $\hat{\beta}$ can be estimated by $\hat{A}' \Sigma_{\beta^*} \hat{A}$ (i.e., by omitting the variance in the calibration model matrix).

A $100(1 - \alpha)$ percent confidence interval for the j th element of $\hat{\beta}_{RC}$ is then

$$\hat{\beta}_{RC, j} \pm \sqrt{\text{Var}(\hat{\beta}_{RC, j})}, \quad (15)$$

where $\text{Var}(\hat{\beta}_{RC, j})$ is the j th element on the diagonal of $\hat{\Sigma}_{\beta_{RC}}$. The variance–covariance matrix $\hat{\Sigma}_{\beta_{RC}}$ can be obtained by either using the delta variance–covariance matrix or zero-variance variance–covariance matrix. In general, the zero-variance variance–covariance matrix will underestimate the true variance–covariance matrix and thus lead to too narrow confidence intervals.

Other methods to construct confidence intervals include stratified bootstrap [2] and the Fieller method [1,33,36,40]. In case of covariate measurement error, the Fieller method can only be applied to construct a $100(1 - \alpha)$ percent confidence interval for the first element of $\hat{\beta}_{RC}$, i.e., $\hat{\phi}_{RC}$. From [36] we obtain:

$$\{f_1 \pm \sqrt{f_1^2 - f_0 f_2 / f_2}\}, \quad (16)$$

where $f_0 = z_{\alpha/2}^2 \text{Var}(\hat{\phi}^*) - \hat{\phi}^*$, $f_1 = z_{\alpha/2}^2 \text{Cov}(\hat{\phi}^*, \hat{\lambda}_1) - \hat{\phi}^* \hat{\lambda}_1$, $f_2 = z_{\alpha/2}^2 \text{Var}(\hat{\lambda}_1) - \hat{\lambda}_1^2$. Where it is assumed that $\text{Cov}(\hat{\phi}^*, \hat{\lambda}_1)$ is null. If the $(1 - \alpha) \times 100\%$ confidence interval of $\hat{\lambda}_1$ includes 0, the Fieller method does not lead to bounded confidence intervals. Bootstrap confidence intervals are obtained by sampling the people in the validation set separately from the people not included in the validation set [2] and taking the $(100 - \alpha)$ percentiles of the obtained distribution.

Outcome measurement error. The variance–covariance matrix for the standard regression estimator $(\hat{\beta}_{RC}, 1)$ can be approximated by applying the multivariate delta method similar to the variance obtained for the corrected estimator for covariate measurement error,

$$\hat{\Sigma}_{(\beta_{RC}, 1)}(j_1, j_2) = (B' \hat{\Sigma}_{(\beta^*, 1)} B)_{j_1, j_2} + (\hat{\beta}^*, 1) \hat{\Sigma}_{B, j_1, j_2} (\hat{\beta}^*, 1)', \quad j_1, j_2 = 1, \dots, (k+3),$$

where \hat{B} is the inverse of the measurement error model matrix $\hat{\Theta}$. $\hat{\Sigma}_{(\beta^*, 1)}$ is a $(k+3) \times (k+3)$ matrix where the upper $(k+2) \times (k+2)$ comprises the variance–covariance matrix obtained from the uncorrected regression defined by model (6) and the last row and column contain zeros. Further, $\hat{\Sigma}_{B, j_1, j_2}$ is the $(k+3) \times (k+3)$ matrix relating the j_1 th and j_2 th column of B (similar to [17]). The so-called zero-variance variance–covariance matrix for $\hat{\beta}$ can be estimated by $B' \hat{\Sigma}_{(\beta^*, 1)} B$.

A $100(1 - \alpha)$ percent confidence interval can be obtained from Eq. (15). Further, $100(1 - \alpha)$ percent confidence intervals for $\hat{\phi}$ and $\hat{\gamma}$ can be approximated by the Fieller method as defined in model 16, where $f_0 = \hat{\phi}^* - z_{\alpha/2}^2 \text{Var}(\hat{\phi}^*)$, $f_1 = \hat{\phi}^* / \hat{\theta}_1 - z_{\alpha/2}^2 \text{Cov}(\hat{\phi}^*, 1 / \hat{\theta}_1)$, $f_2 = 1 / \hat{\lambda}_1^2 - z_{\alpha/2}^2 \text{Var}(1 / \hat{\lambda}_1)$ and idem for $\hat{\gamma}$. Additionally, bootstrap can be used to construct confidence intervals for $\hat{\beta}_{RC}$. Bootstrap confidence intervals are obtained by sampling the individuals in the internal adjustment set separately from the individuals not included in the internal adjustment set and taking the $(100 - \alpha)$ percentiles of the obtained distribution.

Differential outcome measurement error in univariable analyses. The variance–covariance matrix for the standard regression estimator ($\hat{\beta}_{RC}, 1$) can be estimated similar to non-differential outcome measurement error defined above (by using the measurement error matrices for differential outcome measurement error). Confidence intervals can then be obtained from Eq. (15). Bootstrap confidence intervals are obtained by sampling the individuals in the internal adjustment set separately from the individuals not included in the internal adjustment set and taking the $(100 - \alpha)$ percentiles of the obtained distribution.

A2. Maximum likelihood for replicates studies

The variance–covariance matrix for the maximum likelihood estimator $\hat{\beta}_{MLE}$ can be approximated by the multivariate delta method [12]. Denote $\zeta^* = (\delta_0, \delta_Z, \sigma_{Y|Z}^2, \kappa_0, \kappa_Y, \kappa_Z, \sigma_{X|Y,Z}^2)$, leaving the τ^2 from ζ in the main text (see Section 3.3) out as this parameter is not needed for the estimation of $\beta = (\alpha, \phi, \gamma)$. A standard result from linear mixed models is that the estimators of fixed parameters are asymptotically uncorrelated with the estimators of the variance component parameters [12]. If one further assumes that the estimators from the linear model of Y given Z are uncorrelated with the parameters estimated in the linear mixed model, it follows for large samples that $\hat{\zeta}^*$ is multivariate normal with mean ζ and variance covariance matrix $Var(\hat{\zeta})$ equal to:

$$\begin{pmatrix} Var(\hat{\delta}_0) & Cov(\hat{\delta}_0, \hat{\delta}_Z) & 0 & 0 & 0 & 0 & 0 \\ Cov(\hat{\delta}_Z, \hat{\delta}_0) & Var(\hat{\delta}_Z) & 0 & 0 & 0 & 0 & 0 \\ 0 & 0 & Var(\hat{\sigma}_{Y|Z}^2) & 0 & 0 & 0 & 0 \\ 0 & 0 & 0 & Var(\hat{\kappa}_0) & Cov(\hat{\kappa}_0, \hat{\kappa}_Y) & Cov(\hat{\kappa}_0, \hat{\kappa}_Z) & 0 \\ 0 & 0 & 0 & Cov(\hat{\kappa}_Y, \hat{\kappa}_0) & Var(\hat{\kappa}_Y) & Cov(\hat{\kappa}_Y, \hat{\kappa}_Z) & 0 \\ 0 & 0 & 0 & Cov(\hat{\kappa}_Z, \hat{\kappa}_0) & Cov(\hat{\kappa}_Z, \hat{\kappa}_Y) & Var(\hat{\kappa}_Z) & 0 \\ 0 & 0 & 0 & 0 & 0 & 0 & Var(\hat{\sigma}_{X|Y,Z}^2) \end{pmatrix}$$

If $g: \mathbb{R}^{5+2k} \rightarrow \mathbb{R}^{2+k}$ is the function that transforms ζ^* to $\beta_{ML} = (\alpha_{ML}, \phi_{ML}, \gamma_{ML})$, as defined in the main text, then by the multivariate delta method it follows that in large samples:

$$\hat{\beta}_{ML} \sim N(\beta_{ML}, JgVar(\hat{\zeta})(Jg)'),$$

Where J is the Jacobian matrix of g :

$$Jg = \begin{pmatrix} \frac{\partial \phi}{\partial \delta_0} & \frac{\partial \phi}{\partial \delta_Z} & \frac{\partial \phi}{\partial \sigma_{Y|Z}^2} & \cdots & \frac{\partial \phi}{\partial \sigma_{X|Y,Z}^2} \\ \frac{\partial \alpha}{\partial \delta_0} & \frac{\partial \alpha}{\partial \delta_Z} & \frac{\partial \alpha}{\partial \sigma_{Y|Z}^2} & \cdots & \frac{\partial \alpha}{\partial \sigma_{X|Y,Z}^2} \\ \frac{\partial \gamma}{\partial \delta_0} & \frac{\partial \gamma}{\partial \delta_Z} & \frac{\partial \gamma}{\partial \sigma_{Y|Z}^2} & \cdots & \frac{\partial \gamma}{\partial \sigma_{X|Y,Z}^2} \end{pmatrix}.$$

Confidence intervals can then be obtained from Eq. (15). Bootstrap confidence intervals are obtained by sampling the individuals in the internal adjustment set separately from the individuals not included in the internal adjustment set and taking the $(100 - \alpha)$ percentiles of the obtained distribution.

References

- [1] J.P. Buonaccorsi, *Measurement error: Models, Methods, and Applications*, Chapman & Hall/CRC, Boca Raton, 2010.
- [2] R.J. Carroll, D. Ruppert, L.A. Stefanski, C.M. Crainiceanu, *Measurement Error in Nonlinear Models: A Modern Perspective*, 2nd, Chapman & Hall/CRC, Boca Raton, 2006.
- [3] W.A. Fuller, *Measurement error models*, John Wiley & Sons, New York, 1987.
- [4] P. Gustafson, *Measurement error and misclassification in statistics and epidemiology: Impacts and bayesian adjustments*, Chapman & Hall/CRC, Boca Raton, 2004.
- [5] T.B. Brakenhoff, M. Mitroiu, R.H. Keogh, K. Moons, R. Groenwold, M. van Smeden, Measurement error is often neglected in medical literature: a systematic review, *J Clin Epidemiol* 98 (2018) 89–97, doi:10.1016/j.jclinepi.2018.02.023.
- [6] P.A. Shaw, V. Deffner, R.H. Keogh, J.A. Tooze, K.W. Dodd, H. Küchenhoff, V. Kipnis, L.S. Freedman, Epidemiologic analyses with error-prone exposures: review of current practice and recommendations, *Ann Epidemiol* 28 (11) (2018) 821–828, doi:10.1016/j.annepidem.2018.09.001.
- [7] R.H. Keogh, P.A. Shaw, P. Gustafson, R.J. Carroll, V. Deffner, K.W. Dodd, H. Küchenhoff, J.A. Tooze, M.P. Wallace, V. Kipnis, L.S. Freedman, STRATOS guidance document on measurement error and misclassification of variables in observational epidemiology: part 1—Basic theory and simple methods of adjustment, *Stat Med* 39 (16) (2020) 2197–2231, doi:10.1002/sim.8532.
- [8] X. Wang, B. Wang, Deconvolution estimation in measurement error models: the r package decon, *J Stat Softw* 39 (10) (2011) 1–24, doi:10.18637/jss.v039.i10.
- [9] J.R. Cook, L.A. Stefanski, Simulation-extrapolation estimation in parametric measurement error models, *J Am Stat Assoc* 89 (428) (1994) 1314–1328, doi:10.1080/01621459.1994.10476871.
- [10] S.R. Cole, H. Chu, S. Greenland, Multiple-imputation for measurement-error correction, *Int J Epidemiol* 35 (4) (2006) 1074–1081, doi:10.1093/ije/dyl097.
- [11] J.W. Bartlett, R.H. Keogh, Bayesian correction for covariate measurement error: a frequentist evaluation and comparison with regression calibration, *Stat Methods Med Res* 27 (6) (2018) 1695–1708, doi:10.1177/0962280216667764.
- [12] J.W. Bartlett, B.L. De Stavola, C. Frost, Linear mixed models for replication data to efficiently allow for covariate measurement error, *Stat Med* 28 (25) (2009) 3158–3178, doi:10.1002/sim.3713.
- [13] S. Rabe-Hesketh, A. Pickles, A. Skrondal, Correcting for covariate measurement in logistic regression using nonparametric maximum likelihood estimation, *Stat Modelling* 3 (3) (2003) 215–232, doi:10.1191/1471082X03st056oa.
- [14] L.J. Gleser, Improvements of the Naive Approach to Estimation in Nonlinear Errors-in-variables Regression Models, in: P.J. Brown, W.A. Fuller (Eds.), *Statistical Analysis of Measurement Error Models and Applications*, American Statistical Society, Providence, 1990, doi:10.1090/conm/112/1087101. 199–114.
- [15] R.J. Carroll, L.A. Stefanski, Approximate quasi-likelihood estimation in models with surrogate predictors, *Journal of American Statistical Association* 85 (411) (1990) 652–663, doi:10.1080/01621459.1990.10474925.
- [16] R.L. Prentice, Covariate measurement errors and parameter estimation in a failure time regression, *Biometrika* 69 (2) (1982) 331–342, doi:10.2307/2335407.
- [17] B. Rosner, D. Spiegelman, W.C. Willett, Correction of logistic regression relative risk estimates and confidence intervals for measurement error: the case

- of multiple covariates measured with error, *Am. J. Epidemiol.* 132 (4) (1990) 734–745, doi:[10.1093/oxfordjournals.aje.a115715](https://doi.org/10.1093/oxfordjournals.aje.a115715).
- [18] B. Armstrong, Measurement error in the generalised linear model, *Communications in Statistics - Simulation and Computation* 14 (3) (1985) 529–544, doi:[10.1080/03610918508812457](https://doi.org/10.1080/03610918508812457).
- [19] p. C. Team, R: A Language and Environment for Statistical Computing, R Foundation for Statistical Computing, Vienna, Austria, 2020. <https://www.R-project.org/>.
- [20] W. Lederer, H. Küchenhoff, A short introduction to the SIMEX and MCSIMEX, *R news* 6 (4) (1997) 26–31.
- [21] W. He, J. Xiong, G.Y. Yi, SIMEX R Package for Accelerated Failure Time Models with Covariate Measurement Error, *Journal of statistical Software* 46 (1) (2012) 1–14, doi:[10.18637/jss.v046.c01](https://doi.org/10.18637/jss.v046.c01).
- [22] StataCorp, Stata Statistical software: Release 16, StataCorp LLC, College Station, 2019. <https://www.stata.com>.
- [23] H.J. Newton, N.J. Cox, A special issue of the stata journal, *Stata J* 3 (4) (2003) 327, doi:[10.1177/1536867X0400300401](https://doi.org/10.1177/1536867X0400300401).
- [24] J.W. Hardin, H. Schmiediche, R.J. Carroll, The regression-calibration method for fitting generalized linear models with additive measurement error, *Stata J* 3 (4) (2003) 361–372, doi:[10.1177/1536867X0400300406](https://doi.org/10.1177/1536867X0400300406).
- [25] J.W. Hardin, H. Schmiediche, R.J. Carroll, The simulation extrapolation method for fitting generalized linear models with additive measurement error, *Stata J* 3 (4) (2003) 373–385, doi:[10.1177/1536867X0400300407](https://doi.org/10.1177/1536867X0400300407).
- [26] S. Rabe-Hesketh, A. Skrondal, A. Pickles, Maximum likelihood estimation of generalized linear models with covariate measurement error, *The Stata Journal* 3 (4) (2003) 386–411, doi:[10.1177/1536867X0400300408](https://doi.org/10.1177/1536867X0400300408).
- [27] B. Rosner, D. Spiegelman, W.C. Willett, Correction of logistic regression relative risk estimates and confidence intervals for random within-person measurement error, *Am. J. Epidemiol.* 136 (11) (1992) 1400–1413, doi:[10.1093/oxfordjournals.aje.a116453](https://doi.org/10.1093/oxfordjournals.aje.a116453).
- [28] X. Liao, D.M. Zucker, Y. Li, D. Spiegelman, Survival analysis with error-prone time-varying covariates: a risk set calibration approach, *Biometrics* 67 (1) (2011) 50–58, doi:[10.1111/j.1541-0420.2010.01423.x](https://doi.org/10.1111/j.1541-0420.2010.01423.x).
- [29] V. Kipnis, D. Midthune, D.W. Buckman, K.W. Dodd, P.M. Guenther, S.M. Krebs-Smith, A.F. Subar, J.A. Toozé, R.J. Carroll, L.S. Freedman, Modeling data with excess zeros and measurement error: application to evaluating relationships between episodically consumed foods and health outcomes, *Biometrics* 65 (4) (2009) 1003–1010, doi:[10.1111/j.1541-0420.2009.01223.x](https://doi.org/10.1111/j.1541-0420.2009.01223.x).
- [30] D. Spiegelman, R.J. Carroll, V. Kipnis, Efficient regression calibration for logistic regression in main study/internal validation study designs with an imperfect reference instrument, *Stat Med* 20 (1) (2001) 139–160, doi:[10.1002/1097-0258\(20010115\)20:1<139::AID-SIM644>3.0.CO;2-K](https://doi.org/10.1002/1097-0258(20010115)20:1<139::AID-SIM644>3.0.CO;2-K).
- [31] R.H. Keogh, R.J. Carroll, J.A. Toozé, S.I. Kirkpatrick, L.S. Freedman, Statistical issues related to dietary intake as the response variable in intervention trials, *Stat Med* 35 (25) (2016) 4493–4508, doi:[10.1002/sim.7011](https://doi.org/10.1002/sim.7011).
- [32] C. Spearman, The proof and measurement of association between two things, *American Journal of Psychology* 15 (1) (1904) 72–101, doi:[10.2307/1412159](https://doi.org/10.2307/1412159).
- [33] C. Frost, S. Thompson, Correcting for regression dilution bias: comparison of methods for a single predictor variable, *Journal of the Royal Statistical Society A* 163 (2) (2000) 173–189.
- [34] J.A. Hutcheon, A. Chiolerio, J.A. Hanley, Random measurement error and regression dilution bias, *Br Med J* 340 (2010) c2289, doi:[10.1136/bmj.c2289](https://doi.org/10.1136/bmj.c2289).
- [35] R.H. Keogh, I.R. White, A toolkit for measurement error correction, with a focus on nutritional epidemiology, *Stat Med* 33 (12) (2014) 2137–2155, doi:[10.1002/sim.6095](https://doi.org/10.1002/sim.6095).
- [36] L. Nab, R. Groenwold, P. Welsing, M. van Smeden, Measurement error in continuous endpoints in randomised trials: problems and solutions, *Stat Med* 38 (27) (2018) 5182–5196, doi:[10.1002/sim.8359](https://doi.org/10.1002/sim.8359).
- [37] R.H. Keogh, J.W. Bartlett, Measurement error as a missing data problem, 2019. Unpublished, <http://arxiv.org/abs/1910.06443>.
- [38] T.L. Lash, M.P. Fox, A.K. Fink, *Applying quantitative bias analysis to epidemiologic data*, Springer, New York, 2009.
- [39] L. Nab, R. Groenwold, M. van Smeden, R.H. Keogh, Quantitative bias analysis for a misclassified confounder: a comparison between marginal structural models and conditional models for point treatments, *Epidemiology* 31 (6) (2020) 796–805, doi:[10.1097/EDE.0000000000001239](https://doi.org/10.1097/EDE.0000000000001239).
- [40] V.H. Franz, Ratios: A short guide to confidence limits and proper use, 2007. Unpublished, <https://arxiv.org/abs/0710.2024>.
- [41] R. de Mutsert, M.d. Heijer, T.J. Rabelink, J. Smit, J.A. Romijn, J.W. Jukema, A. de Roos, C.M. Cobbaert, M. Kloppenburg, S. Le Cessie, S. Middeldorp, F.R. Rosendaal, The Netherlands epidemiology of obesity (NEO) study: study design and data collection, *Eur. J. Epidemiol.* 28 (2013) 513–523, doi:[10.1007/s10654-013-9801-3](https://doi.org/10.1007/s10654-013-9801-3).
- [42] L. Nab, M. van Smeden, R. de Mutsert, F.R. Rosendaal, R. Groenwold, Sampling strategies for internal validation samples for exposure measurement error correction: a study of visceral adipose tissue measures replaced by waist circumference measures, *Am. J. Epidemiol.* (2021), doi:[10.1093/aje/kwab114](https://doi.org/10.1093/aje/kwab114).
- [43] M. Makrides, C.A. Crowther, R.A. Gibson, R.S. Gibson, C.M. Skeaff, Efficacy and tolerability of low-dose iron supplements during pregnancy: a randomized controlled trial 13, *Am. J. Clin. Nutr.* 78 (1) (2003) 145–153.
- [44] E.A. McCarthy, T.A. Carins, Y. Hannigan, N. Bardien, A. Shub, S.P. Walker, Data from: effectiveness and safety of 1 vs 4h blood pressure profile with clinical and laboratory assessment for the exclusion of gestational hypertension and pre-eclampsia: a retrospective study in a university affiliated maternity hospital, Dryad (2015), doi:[10.5061/dryad.0bq15](https://doi.org/10.5061/dryad.0bq15).
- [45] L.J. Appel, M. Espeland, P.K. Whelton, T. Dolecek, S. Kumanyika, W.B. Applegate, W.H. Ettinger, J.B. Kostis, A.C. Wilson, C. Lacy, S.T. Miller, Trial of nonpharmacologic intervention in the elderly (TONE), *Ann Epidemiol* 5 (2) (1995) 119–129, doi:[10.1016/1047-2797\(94\)00056-Y](https://doi.org/10.1016/1047-2797(94)00056-Y).
- [46] S.W. Thurston, P.L. Williams, R. Hauser, H. Hu, M. Hernandez-Avila, D. Spiegelman, A comparison of regression calibration approaches for designs with internal validation data, *J Stat Plan Inference* 131 (1) (2005) 175–190, doi:[10.1016/j.jspi.2003.12.015](https://doi.org/10.1016/j.jspi.2003.12.015).
- [47] F. Perrier, L. Giorgis-Allemand, R. Slama, C. Philippat, Within-subject pooling of biological samples to reduce exposure misclassification in biomarker-based studies, *Epidemiology* 27 (3) (2016) 378–388, doi:[10.1097/EDE.0000000000000460](https://doi.org/10.1097/EDE.0000000000000460).
- [48] E. Batistatou, R. McNamee, Performance of bias-correction methods for exposure measurement error using repeated measurements with and without missing data, *Stat Med* 31 (28) (2012) 3467–3480, doi:[10.1002/sim.5422](https://doi.org/10.1002/sim.5422).
- [49] L.S. Freedman, D. Midthune, R.J. Carroll, V. Kipnis, A comparison of regression calibration, moment reconstruction and imputation for adjusting for covariate measurement error in regression, *Stat Med* 27 (25) (2008) 5195–5216, doi:[10.1002/sim.3361](https://doi.org/10.1002/sim.3361).
- [50] K. Messer, L. Natarajan, Maximum likelihood, multiple imputation and regression calibration for measurement error adjustment, *Stat Med* 27 (30) (2008) 6332–6350, doi:[10.1002/sim.3458](https://doi.org/10.1002/sim.3458).
- [51] K. Luijken, R. Groenwold, B. Van Calster, E.W. Steyerberg, M. van Smeden, Impact of predictor measurement heterogeneity across settings on the performance of prediction models: a measurement error perspective, *Stat Med* 38 (18) (2019) 3444–3459, doi:[10.1002/sim.8183](https://doi.org/10.1002/sim.8183).
- [52] K. Luijken, L. Wynants, M. van Smeden, B. Van Calster, E.W. Steyerberg, R. Groenwold, Collaborators, Changing predictor measurement procedures affected the performance of prediction models in clinical examples, *J Clin Epidemiol* 119 (2020) 7–18, doi:[10.1016/j.jclinepi.2019.11.001](https://doi.org/10.1016/j.jclinepi.2019.11.001).