CUT&RUN MDM - Figures

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Introduction

To assess whether MafB also regulates macrophage identity in humans, we performed CUT&RUN for MafB on human monocyte-derived mac (MDMs). CUT&RUN was performed with a CUTANA ChIC/CUT&RUN Kit (EpiCypher, 141048) according to manufacturer's instructions, with modifications. BMDMs from Mafbfl/fl or Lyz2CreMafbfl/fl mice were washed with ice-cold PBS and fixed with 0.1 % formaldehyde in PBS for 2 min at room temperature. Fixation was quenched by adding glycine (Merck, 104691000) to 0.125 M. For each CUT&RUN sample, 1×10^6 fixed cells and 0.5 µg antibodies were added:

IgG (EpiCypher, 13-0042) MafB2 (Cell Signaling Technology, 41019)

CUT&RUN libraries were prepared with a CUTANATM CUT&RUN Library Prep Kit (EpiCypher, 141001) according to manufacturer's instructions. These libraries were sequenced on an NovaSeq 6000 (Illumina) sequencer on an S4 flow cell at 10 million reads per sample.

Load Packages

```
suppressMessages({
    library(ggplot2)
    library(colorRamp2)
    library(ComplexHeatmap)
    library(readxl)
    library(futile.logger)
    library(grid)
    library(rtracklayer)
    library(dplyr)
    library(VennDiagram)
})
```

Warning: package 'colorRamp2' was built under R version 4.4.3

Figure S7

```
MafB2_peaks_rep1 <- read.table("MafB2_R1.seacr.peaks.stringent.bed",
    header = FALSE, sep = "\t")

MafB2_peaks_rep2 <- read.table("MafB2_R2.seacr.peaks.stringent.bed",
    header = FALSE, sep = "\t")

MafB2_peaks_consensus <- read.table("MafB2.seacr.peaks.consensus.bed",
    header = FALSE, sep = "\t")

# peaks rep1
nrow(MafB2_peaks_rep1)

## [1] 26327

# peaks rep2
nrow(MafB2_peaks_rep2)

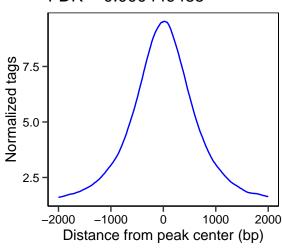
## [1] 30718

# shared peaks
nrow(MafB2_peaks_consensus)</pre>
```

Rep. 1

[1] 24755

Rep. 1: 26327 peaks FDR = 0.000449485



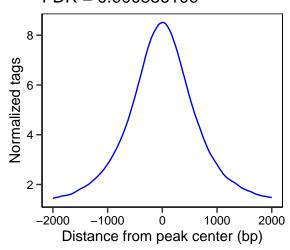
n peaks : 30718 FDR = 0.00038610038610043

```
ggsave("Figure_S7G_Rep1.pdf", width = 3, height = 3)
```

Rep 2

```
ggplot(data = MafB2_R2_hist, aes(x = Distance.from.Center, y = MafB2_R2..Coverage)) +
   geom_line(show.legend = FALSE, colour = "blue") + ggtitle("Rep. 2: 30718 peaks \nFDR = 0.000386100"
   xlab("Distance from peak center (bp)") + ylab("Normalized tags") +
   theme_classic() + theme(axis.text.x = element_text(color = "black"),
   axis.text.y = element_text(color = "black"), axis.ticks.length = unit(0.15,
        "cm"), panel.border = element_rect(fill = NA, color = "black",
        linetype = "solid"))
```

Rep. 2: 30718 peaks FDR = 0.000386100



```
ggsave("Figure_S7G_Rep2.pdf", width = 3, height = 3)
```

Figure S7H

```
samtools view -c -F 4 MafB2_R1.target.markdup.sorted.bam bedtools intersect -a MafB2_R1.target.markdup.sorted.bam -b MafB2_R1.seacr.peaks.stringent.bed -bed | w samtools view -c -F 4 MafB2_R2.target.markdup.sorted.bam bedtools intersect -a MafB2_R2.target.markdup.sorted.bam -b MafB2_R2.seacr.peaks.stringent.bed -bed | w
```

Figure S7I

```
MafB2_counts <- read.table("MafB2_counts.txt", header = TRUE,
    sep = "\t")

colnames(MafB2_counts)[colnames(MafB2_counts) == "PeakID..cmd.annotatePeaks.pl.MafB2_peaks_merged.bed.m

colnames(MafB2_counts)[colnames(MafB2_counts) == "IgG_R1..Tag.Count.in.given.bp..4149983.0.Total..norma

colnames(MafB2_counts)[colnames(MafB2_counts) == "MafB2_R1..Tag.Count.in.given.bp..7202841.0.Total..norma</pre>
```

```
colnames(MafB2_counts) [colnames(MafB2_counts) == "MafB2_R2..Tag.Count.in.given.bp..8260258.0.Total..nor
```

24755 merged peaks 12.5 10.0 _og2(norm tags + 1) 7.5 5.0 2.5 0.0 7.5 0.0 2.5 5.0 10.0 12.5 Log2(norm tags + 1)Rep. 1

```
ggsave("Figure_S7I.pdf", width = 3, height = 3.225)
```

Figure 7A

Histogram

```
annotatePeaks.pl MafB2_peaks_merged.bed mm10 -size 4000 -hist 10 -d IgG_R1/ MafB2_R1/ MafB2_R2/ > MafB2_hist <- read.table("MafB2_hist.txt", header = TRUE, sep = "\t")

colnames(MafB2_hist)[colnames(MafB2_hist) == "Distance.from.Center..cmd.annotatePeaks.pl.MafB2_peaks_merged.bed mm10 -size 4000 -hist 10 -d IgG_R1/ MafB2_R1/ MafB2_R2...

MafB2_hist <- read.table("MafB2_hist.txt", header = TRUE, sep = "\t")

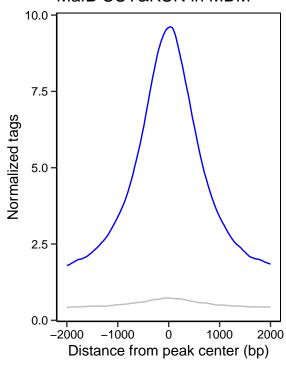
colnames(MafB2_hist)[colnames(MafB2_hist) == "Distance.from.Center..cmd.annotatePeaks.pl.MafB2_peaks_merged.bed mm10 -size 4000 -hist 10 -d IgG_R1/ MafB2_R1...

MafB2_hist <- read.table("MafB2_hist.txt", header = TRUE, sep = "\t")

colnames(MafB2_hist)[colnames(MafB2_hist) == "Distance.from.Center..cmd.annotatePeaks.pl.MafB2_peaks_merged.bed mm10 -size 4000 -hist 10 -d IgG_R1/ MafB2_R1...

MafB2_hist $\frac{1}{2} \text{ MafB2_hist} \text{ Coverage", "MafB2_R2...Coverage", "MafB2_R2...Coverage")]}</pre>
```

MafB CUT&RUN in MDM



```
ggsave("Figure_7A_MDM_Hist.pdf", width = 3, height = 4)
```

Heatmap

```
annotatePeaks.pl MafB2_peaks_merged.bed mm10 -size 4000 -hist 25 -ghist -d IgG_R1/ MafB2_R1/ MafB2_R2/
```

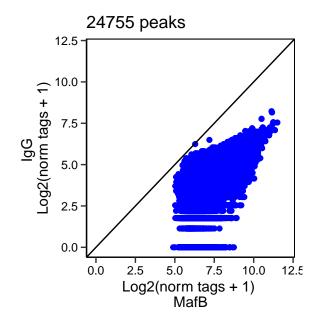
IgG

```
col_fun = colorRamp2(c(0, 20), c("white", "red"))
pdf("Figure_7A_heatmap_IgG.pdf", width = 2, height = 4)
Heatmap(IgG_R1_heatmap, col_fun, cluster_columns = F, cluster_rows = F,
   heatmap_legend_param = list(title = "Tags"), use_raster = TRUE,
   raster_quality = 10, , column_title = "IgG")
## Warning: The input is a data frame-like object, convert it to a matrix.
## 'magick' package is suggested to install to give better rasterization.
##
## Set 'ht_opt$message = FALSE' to turn off this message.
dev.off()
## pdf
## 2
MafB
col_fun = colorRamp2(c(0, 20), c("white", "red"))
pdf("Figure_7A_heatmap_MafB.pdf", width = 2, height = 4)
Heatmap(MafB2_heatmap, col_fun, cluster_columns = F, cluster_rows = F,
   heatmap_legend_param = list(title = "Tags"), use_raster = TRUE,
   raster_quality = 10, column_title = "MafB2")
## Warning: The input is a data frame-like object, convert it to a matrix.
## 'magick' package is suggested to install to give better rasterization.
## Set 'ht_opt$message = FALSE' to turn off this message.
dev.off()
## pdf
##
```

Figure 7B

```
MafB2_counts$MafB2 <- rowMeans(MafB2_counts[, c("MafB2_R1", "MafB2_R2")])

MafB2_counts$log2_IgG <- log2(MafB2_counts$IgG_R1 + 1)
MafB2_counts$log2_MafB2 <- log2(MafB2_counts$MafB2 + 1)
MafB2_counts$below_diag <- MafB2_counts$log2_IgG < MafB2_counts$log2_MafB2</pre>
```



```
ggsave("Figure_7B.pdf", width = 3, height = 3.225)
```

```
"TRUE", ]$PeakID)
MafB2_peaks <- MafB2_peaks_merged[MafB2_peaks_merged[, 4] %in%
    MafB2_PeakID, ]

write.table(MafB2_peaks, "MafB2_peaks.bed", sep = "\t", quote = FALSE,
    row.names = FALSE, col.names = FALSE)</pre>
```

Figure 7C

```
annotatePeaks.pl MafB2_peaks.bed genome.fa -gtf genes.gtf > MafB2_peaks_annot.txt
MafB2_peaks_annot <- read.table("MafB2_peaks_annot.txt", header = TRUE,
   sep = "\t")
MafB2_peaks_annot$Annotation.not.detailed <- sub(" .*", "", MafB2_peaks_annot$Annotation)
table(MafB2_peaks_annot$Annotation.not.detailed)
##
##
                 Intergenic
                                  intron promoter-TSS
                                                                TTS
           exon
           1076
                       5173
                                  11663
                                                               1237
##
                                                 5606
```

Figure 7D

Convert mouse MafB2 BMDM peaks to human

```
MafB2_BMDM_peaks_human <- read.table("MafB2_BMDM_peaks_human.bed",
    header = FALSE, sep = "\t")

MafB2_BMDM_peaks_human$V1 <- sub("^chr", "", MafB2_BMDM_peaks_human$V1)

write.table(MafB2_BMDM_peaks_human, "MafB2_BMDM_peaks_human.bed",
    sep = "\t", quote = FALSE, row.names = FALSE, col.names = FALSE)</pre>
```

check overlap

bedtools window -a MafB2_peaks.bed -b MafB2_BMDM_peaks_human.bed -w 100 > MafB2_peaks_conserved.bed

```
MafB2_peaks_conserved <- read.table("MafB2_peaks_conserved.bed",
    header = FALSE, sep = "\t")
nrow(MafB2_peaks)</pre>
```

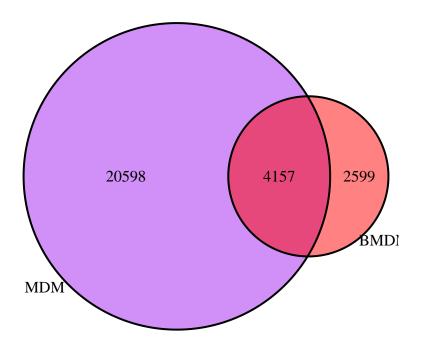
[1] 24755

```
nrow(MafB2_BMDM_peaks_human)
```

[1] 6756

```
nrow(MafB2_peaks_conserved)
```

[1] 4157

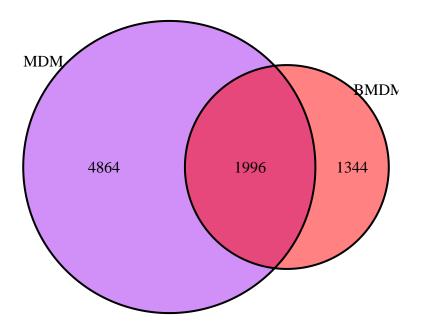


(polygon[GRID.polygon.846], polygon[GRID.polygon.847], polygon[GRID.polygon.848], polygon[GRID.polyg

```
# grid.newpage()
```

Figure 7E

```
# Helper function to display Venn diagram
display_venn <- function(x, ...) {</pre>
    grid.newpage()
    venn_object <- venn.diagram(x, filename = NULL, ...)</pre>
    grid.draw(venn_object)
}
MafB2_genes <- MafB2_peaks_annot[MafB2_peaks_annot[, "Distance.to.TSS"] >=
    -2000 & MafB2_peaks_annot[, "Distance.to.TSS"] <= 2000, ]
MafB2_genes <- unique(MafB2_genes$Gene.Name)</pre>
MafB2_genes <- na.omit(MafB2_genes)</pre>
MafB2_BMDM_peaks_annot <- read.table("MafB2_BMDM_peaks_annot.txt",</pre>
    header = TRUE, sep = "\t")
MafB2_BMDM_genes <- MafB2_BMDM_peaks_annot[MafB2_BMDM_peaks_annot[,</pre>
    "Distance.to.TSS"] >= -2000 & MafB2_BMDM_peaks_annot[, "Distance.to.TSS"] <=
    2000, ]
MafB2_BMDM_genes <- unique(MafB2_BMDM_genes$Gene.Name)
MafB2_BMDM_genes <- na.omit(MafB2_BMDM_genes)</pre>
write.table(MafB2_BMDM_genes, "MafB2_BMDM_genes.txt", sep = "\t",
    quote = FALSE, row.names = FALSE, col.names = FALSE)
MafB2_BMDM_genes_human <- read.csv("MafB2_BMDM_genes_human.csv")</pre>
MafB2_BMDM_genes_human <- na.omit(unique(MafB2_BMDM_genes_human$ortholog_name))
x <- list(MDM = MafB2_genes, BMDM = MafB2_BMDM_genes_human)
# pdf(file = 'Figure_7E_Venn.pdf', width = 4, height = 4)
display_venn(x, fill = c("purple", "red"))
```



dev.off()