RJafroc documentation

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Contents

Chapter 1

Preface

- This book, an extended documentation of the **RJafroc** package, is undergoing extensive edits.
- It should not be used by the casual user until I give the go ahead.
- It bypasses the file size limits of **CRAN**, currently 5 MB, which severely limits the extent of the documentation that can be included with the CRAN version of the package.
- I welcome corrections and comments by the not-so-casual-user.
- Please use the GitHub website to raise issues and comments:

    – https://github.com/dpc10ster/RJafrocBook

Chapter 2

Introduction

- This is the book desribing the **RJafroc** packages.
- The name of the book is RJafrocBook
- Modality and treatment are used interchangeably.
- Reader is a generic radiologist, or a computer aided detection algorithm, or any algorithmic "reader"
- TBA

Chapter 3

ROC data format

3.1 Introduction

- In the receiver operating characteristic (**ROC**) paradigm (Metz, 1978) the observer's task is to **rate** (i.e., assign an ordered label representing the degree of suspicion) each case for confidence in presence of disease. The rating is frequently called a *confidence level*.
- The rating can be an integer or quasi- continuous (e.g., $0 - 100$), or a floating point value, as long as higher numbers represent greater confidence in presence of one or more lesions in the case [1].
- For human observer studies a 6-point rating scale is recommended, collected via two questions (Chakraborty, 2017):

    - Is the case diseased?

        * Binary response: *Yes* or *No*.

    - What is your confidence in the preceding decisions?

        * Three level response: *Low*, *Medium* or *High*.

- With algorithmic readers, e.g., computer aided detection algorithms a floating point rating, if possible, should be retained.
- In the most common study design, termed multiple-reader multiple-case (**MRMC**) the rating collection procedure is repeated for all cases, treatments and readers.

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[1]The directionaliy of the rating is not a limitation. If lower values correspond to increased confidence level, it is only necessary to transform the observed rating by subtracting it from a constant value. The constant value can be chosen arbitrarily, typically as the maximum of all observed ratings, thereby ensuring that the transformed value is always non-negative.

3.2   An actual MRMC ROC dataset

An actual MRMC ROC dataset (Van Dyke et al., 1993) is included as `dataset02`. It has the following structure:

```
str(dataset02)
#> List of 8
#>  $ NL        : num [1:2, 1:5, 1:114, 1] 1 3 2 3 2 2 1 2 3 2 ...
#>  $ LL        : num [1:2, 1:5, 1:45, 1] 5 5 5 5 5 5 5 5 5 5 ...
#>  $ lesionNum : int [1:45] 1 1 1 1 1 1 1 1 1 1 ...
#>  $ lesionID  : num [1:45, 1] 1 1 1 1 1 1 1 1 1 1 ...
#>  $ lesionWeight: num [1:45, 1] 1 1 1 1 1 1 1 1 1 1 ...
#>  $ dataType  : chr "ROC"
#>  $ modalityID : Named chr [1:2] "0" "1"
#>   ..- attr(*, "names")= chr [1:2] "0" "1"
#>  $ readerID   : Named chr [1:5] "0" "1" "2" "3" ...
#>   ..- attr(*, "names")= chr [1:5] "0" "1" "2" "3" ...
```

3.2.1   Overview of the data structure

- The `dataset` structure is a `list` variable with 8 members [2].
  - Ratings of non-diseased cases are stored in the `NL` list member.
  - Ratings of diseased cases are stored in the `LL` list member.
  - The `lesionNum` list member is an array of length 45, filled with ones. It lists the number of lesions per case, which for ROC data, is always unity. The length of this array equals the number of diseased cases K2, see below.
  - The `lesionID` list member is a `[45 x 1]` array, also filled with ones. [3]
  - The `LesionWeight` list member is also a `[45 x 1]` array filled with ones.
  - The `dataType` list member equals the string `"ROC"`, identifying it as an ROC dataset.
  - The `modalityID` list member is a string array identifying the names of the treatments (see below).
  - The `readerID` list member is a string array, identifying the names of the readers (see below).

3.2.2   Details of the `modalityID` and `readerID` list members

- The names of the treatments are in the `modalityID` list member:

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[2]This is true for ROC, FROC and ROI datasets. LROC datasets have 9 `list` members.

[3]The second "unnecessary" dimension is necessary for compatibility with FROC datasets.

```
attributes(dataset02$modalityID)
#> $names
#> [1] "0" "1"
```

For example, the name of the first treatment is `"0"`. The names can be longer strings, but use of very long string names may mess up the output formats of the analysis report. As per the **KISS** principle [4], keep the names short.

- The names of the readers are in the `readerID` array:

```
attributes(dataset02$readerID)
#> $names
#> [1] "0" "1" "2" "3" "4"
```

For example, the name of the second reader is `"1"`. A similar caveat regarding long reader names applies.

3.2.3 Details of the `NL` and `LL` list members

- For either `NL` or `LL` list members, the fourth dimension has unit length. This dimension, which is strictly speaking unnecessary for ROC data, is retained for ease of generalizability to the FROC and ROC paradigms, where more than one rating per case is possible.
- `dataset02` is a 2-treatment 5-reader dataset (the lengths of the first and second dimensions, respectively, of the `NL` and `LL` list members).

3.2.3.1 Numbers of non-diseased and diseased cases

```
K <- length(dataset02$NL[1,1,,1])
K2 <- length(dataset02$LL[1,1,,1])
K1 <- K - K2
```

- `K1` is the number of non-diseased cases, while `K2` is the number of diseased cases.

- The third dimension of the `NL` array is the total number of **all** cases, i.e., $K = 114$, and the third dimension of the `LL` array, i.e., $K2 = 45$, is the total number of diseased cases.

- Subtracting the number of diseased cases from the number of all cases yields the number of non-diseased cases.

- Therefore, in this dataset, there are **45** diseased cases and **69** non-diseased cases.

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[4]For those not familiar, KISS stands for Keep It Simple, Stupid.

3.2.3.2   Why dimension the `NL` array for the total number of cases?

- Again, this is for ease of generalizability to the FROC and ROI paradigms.

3.2.3.3   Ratings on a non-diseased case

- For ROC data a non-diseased case can have only one, and exactly one, `NL` rating.
- For treatment 1, reader 1 and case 1 (the first non-diseased case), the NL rating is `"1"`:

```
dataset02$NL[1,1,1,1]
#> [1] 1
mean(dataset02$NL[,,1:K1,1])
#> [1] 1.784058
```

- This study utilized a 5-point rating scale, 1 thru 5, so non-diseased cases are expected to have low ratings; in this case the lowest rating was observed.
- The mean rating over all non-diseased cases, treatments and readers, is 1.784058.

3.2.3.4   Ratings on a diseased case

- For ROC data a diseased case can have only one, and exactly one, `LL` rating.
- For treatment 1, reader 1, case 1 (the first diseased case) the LL rating is:

```
dataset02$LL[1,1,1,1]
#> [1] 5
mean(dataset02$LL)
#> [1] 4.297778
```

- As noted previously, this study utilized a 5-point rating scale, 1 thru 5, so diseased cases are expected to have high ratings; in this case the highest rating was observed.
- The mean rating over all diseased cases, treatments and readers, is 4.2977778.
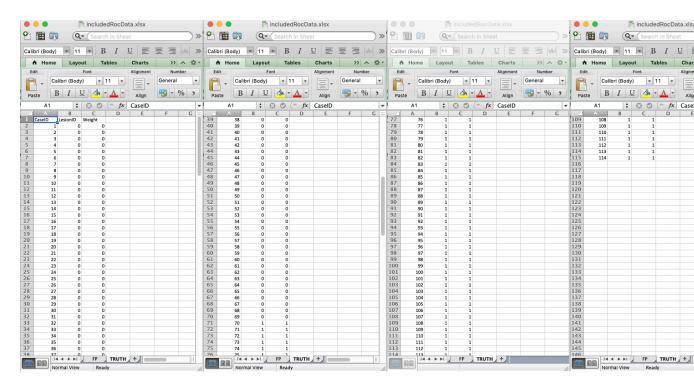
3.3   The ROC Excel data file

- An Excel file in JAFROC format containing ROC data corresponding to `dataset02`, is included with the distribution. The first command (below)

finds the location of the file and the second command reads it and saves it to a dataset object `ds`.

```
fileName <- system.file(
    "extdata", "includedRocData.xlsx", package = "RJafroc", mustWork = TRUE)
ds <- DfReadDataFile(fileName)
```
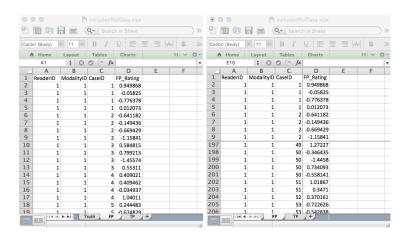
- It contains three worksheets, `Truth`, `TP` and `FP`.
- The `Truth` worksheet defines the ground-truth of each case. It indicates which cases are diseased and which are non-diseased.
- The `CaseID` column lists the numeric labels identifying each case. Again, string names are possible, but keep them short.
- A `1` in the `LesionID` column denotes a diseased case.
- A `0` in the `LesionID` column denotes a non-diseased case.
- The `Weight` column is irrelevant for ROC data [5].
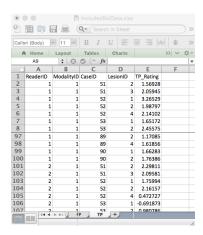- The contents of the `Truth` worksheet corresponding to `dataset02` are displayed next:

+ There are 69 non-diseased cases (labeled 1-69) under column `CaseID`.
+ There are 45 diseased cases (labeled 70-114).

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[5]It is only needed for FROC data.

+ The `LesionID` field for each non-diseased case (e.g., `CaseID` = 1) is zero.
+ The `LesionID` field for each diseased case (e.g., `CaseID` = 70) is unity.
+ The `Weights` field is irrelevant for ROC datasets. For convenience it is filled with

- The FP (or NL) worksheet - this lists the ratings of ROI-level-non-diseased regions.

    - For ReaderID $=$ 1, ModalityID $=$ 1 and CaseID $=$ 1 there are 4 rows, corresponding to the 4 ROI-level-non-diseased regions in this case. The corresponding ratings are 1. The pattern repeats for other treatments and readers, but the rating are, of course, different.

    - Each CaseID is represented in the FP worksheet (a rare exception could occur if a case-level abnon-diseased case has 4 abnon-diseased regions).

- The `TP` (or `LL`) worksheet - this lists the ratings of ROI-level-abnon-diseased regions.

  - Because non-diseased cases generate TPs, one does not find any entry with `CaseID` = 1-50 in the `TP` worksheet.

  - The lowest `CaseID` in the `TP` worksheet is 51, which corresponds to the first abnon-diseased case.

  - There are two entries for this case, corresponding to the two ROI-level-abnon-diseased regions present in this case. Recall that corresponding to this `CaseID` in the `Truth` worksheet there were two entries with `LesionID` = 2 and 3. These must match the `LesionID`'s listed for this case in the `TP` worksheet. Complementing these two entries, in the `FP` worksheet for `CaseID` = 51, there are 2 entries corresponding to the two ROI-level-non-diseased regions in this case.

  - One should be satisfied that for each abnon-diseased case the sum of the number of entries in the `TP` and `FP` worksheets is always 4.

Chapter 4

FROC data format

4.1 Introduction

- In the free-response ROC (**FROC**) paradigm (Bunch et al., 1978) the observer's task is to indicate (i.e., **mark** the location of) and **rate** (i.e., assign an ordered label - or confidence level - representing the degree of suspicion) regions in the image that are perceived as suspicious for presence of disease. Accordingly, FROC data consists of **mark-rating pairs**, where each mark indicates a region [1] that was considered suspicious for presence of a localized lesion and the rating is the corresponding confidence level. The number of mark-rating pairs on any particular case is a-priori unpredictable. It is a non-negative random integer (i.e., 0, 1, 2, ...) that depends on the case, the reader and the modality. The relatively unstructured nature of FROC data makes FROC paradigm data more difficult to analyze than ROC paradigm data [2].

- By adopting a proximity criterion, each mark is classified by the investigator as a lesion localization (LL) - if it is close to a real lesion - or a non-lesion localization (NL) otherwise.

- The rating can be an integer or quasi- continuous (e.g., $0 - 100$), or a floating point value, as long as higher numbers represent greater confidence in presence of one or more lesions in the ROI [3].

---

[1] In order to avoid confusion with the ROI-paradigm, I do not like to use the term ROI to describe the marks made by the observer.

[2] Recall that the ROC paradigm always yields a single rating per case.

[3] The directionaliy of the rating is not a limitation. If lower values correspond to increased confidence level, it is only necessary to transform the observed rating by subtracting it from a constant value. The constant value can be chosen arbitrarily, typically as the maximum of all observed ratings, thereby ensuring that the transformed value is always non-negative.

- Region-level-normal ratings are stored in the `NL` field and region-level-abnormal ratings are stored in the `LL` field.

4.2   An actual FROC dataset

An actual FROC dataset (Zanca et al., 2009) is included as `dataset04`, which has the following `dataset` structure:

```
str(dataset04)
#> List of 8
#>  $ NL          : num [1:5, 1:4, 1:200, 1:7] -Inf -Inf 1 -Inf -Inf ...
#>  $ LL          : num [1:5, 1:4, 1:100, 1:3] 4 5 4 5 4 3 5 4 4 3 ...
#>  $ lesionNum   : int [1:100] 1 1 1 1 1 1 1 1 1 1 ...
#>  $ lesionID    : num [1:100, 1:3] 1 1 1 1 1 1 1 1 1 1 ...
#>  $ lesionWeight: num [1:100, 1:3] 1 1 1 1 1 1 1 1 1 1 ...
#>  $ dataType    : chr "FROC"
#>  $ modalityID  : Named chr [1:5] "1" "2" "3" "4" ...
#>   ..- attr(*, "names")= chr [1:5] "1" "2" "3" "4" ...
#>  $ readerID    : Named chr [1:4] "1" "3" "4" "5"
#>   ..- attr(*, "names")= chr [1:4] "1" "3" "4" "5"
```

Examination of the output reveals that:

- The `dataset` structure is a list with 8 members.
- This is a 5-treatment 4-reader dataset (the lengths of the first and second dimensions, respectively, of the `NL` and `LL` arrays). The names of the treatments are in the `modalityID` array:

```
attributes(dataset04$modalityID)
#> $names
#> [1] "1" "2" "3" "4" "5"
```

For example, the name of the second treatment is `"2"`.

- The names of the readers are in the `readerID` array:

```
attributes(dataset04$readerID)
#> $names
#> [1] "1" "3" "4" "5"
```

For example, the name of the second reader is `"3"`. Apparently reader `"2"` "dropped out" of the study.

4.2.1 Numbers of non-diseased and diseased cases

```
length(dataset04$NL[1,1,,1])
#> [1] 200
length(dataset04$LL[1,1,,1])
#> [1] 100
```

- The third dimension of the `NL` array is the total number of **all** cases, i.e., 200, and the third dimension of the `LL` array, i.e., 100, is the total number of diseased cases.

- Subtracting the number of diseased cases from the number of all cases yields the number of non-diseased cases.

- Therefore, in this dataset, there are 100 diseased cases and 100 non-diseased cases.

4.2.2 Why dimension the `NL` array for the total number of cases?

- Because, in addition to `LLs`, `NLs` are possible on diseased cases.
- Only `LLs` are possible on diseased cases.
- Only `NLs` are possible on non-diseased cases.
- The missing values are filled in with `-Inf`.

4.2.3 Ratings on a non-diseased case

- For treatment 1, reader 1 and case 1 (the first non-diseased case), the NL ratings are:

```
dataset04$NL[1,1,1,]
#> [1] -Inf -Inf -Inf -Inf -Inf -Inf -Inf
```

4.2.4 The meaning of a negative infinity rating

- Obviously, a real rating cannot be negative infinity [4]. This value is reserved for **missing ratings**, and more generally, **missing marks** [5]. For

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[4]If an observer is so highly confident in the *absence* of a localized lesion, he will simply *not mark* the location in question; if he did, then, logically, he should mark *all* areas in the image that are definitely not lesions; in the FROC paradigm only regions with a reasonable degree of suspicion are marked. The radiologist only wishes to draw attention to regions that are reasonably suspicious; the definition of "reasonable" is determined by clinical considerations.

[5]Since there is a one-to-one correspondence between marks and ratings.

example, since all values in the above code chunk are negative infinities, this means this treatment-reader-case combination did not yield any mark-rating pairs. This possibility, alluded to above, is only possible with FROC data. All other paradigms (ROC, LROC and ROI) yield at least one rating per case.

- The length of the fourth dimension of the `NL` array is determined by that treatment-reader-case combination yielding the maximum number of `NL`s. Consider the following chunk:

```r
for (i in 1:5)
  for (j in 1:4)
    for (k in 1:200)
      if (all(dataset04$NL[i,j,k,] != -Inf))
        cat(i, j, k, all(dataset04$NL[i,j,k,] != -Inf),"\n")
#> 5 4 192 TRUE
```

- This shows that the fourth dimension of the `NL` array has to be of length 7 because *one, and only reader*, specifically reader "4", made 7 `NL` marks on a diseased case in treatment "5"!

4.2.5   Ratings on a non-diseased case

Unlike non-diseased cases, diseased cases can have both `NL` and `LL` ratings.

- For treatment 1, reader 1, case 51 (the 1st diseased case) the NL ratings are:

```r
dataset04$NL[1,1,51,]
#> [1] -Inf -Inf -Inf -Inf -Inf -Inf -Inf
dataset04$lesionNum[1]
#> [1] 1
dataset04$LL[1,1,1,]
#> [1]    4 -Inf -Inf
mean(is.finite(dataset04$LL))
#> [1] 0.3043333
```

. There are only two finite values because this case has two ROI-level-abnormal regions, and 2 plus 2 makes for the assumed 4-regions per case. The corresponding `$lesionNum` field is 1.

```r
mean(is.finite(dataset04$NL[,,1:50,]))
#> [1] 0.05942857
dataset04$NL[1,1,51,]
```

```
#> [1] -Inf -Inf -Inf -Inf -Inf -Inf -Inf
dataset04$lesionNum[1]
#> [1] 1
dataset04$LL[1,1,1,]
#> [1]    4 -Inf -Inf
mean(is.finite(dataset04$LL))
#> [1] 0.3043333
```

```
mean(is.finite(dataset04$NL[,,1:50,]))
#> [1] 0.05942857
dataset04$NL[1,1,51,]
#> [1] -Inf -Inf -Inf -Inf -Inf -Inf -Inf
dataset04$lesionNum[1]
#> [1] 1
dataset04$LL[1,1,1,]
#> [1]    4 -Inf -Inf
mean(is.finite(dataset04$LL))
#> [1] 0.3043333
```

- The ratings of the 2 ROI-level-abnormal ROIs on this case are 4. The mean rating over all ROI-level-abnormal ROIs is 3.6785323.

```
mean(is.finite(dataset04$NL[,,1:50,]))
#> [1] 0.05942857
dataset04$NL[1,1,51,]
#> [1] -Inf -Inf -Inf -Inf -Inf -Inf -Inf
dataset04$lesionNum[1]
#> [1] 1
dataset04$LL[1,1,1,]
#> [1]    4 -Inf -Inf
mean(is.finite(dataset04$LL))
#> [1] 0.3043333
```
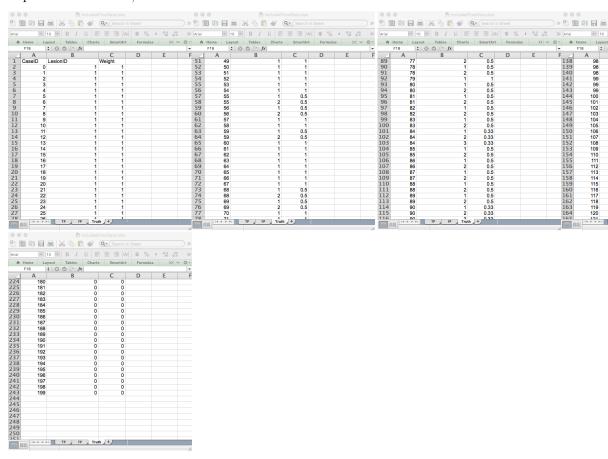
4.3 The FROC Excel data file

An Excel file in JAFROC format containing simulated ROI data corresponding to `dataset04`, is included with the distribution. The first command (below) finds the location of the file and the second command reads it and saves it to a dataset object `ds`.

```
fileName <- system.file(
    "extdata", "includedFrocData.xlsx", package = "RJafroc", mustWork = TRUE)
```

```
ds <- DfReadDataFile(fileName)
ds$dataType
#> [1] "FROC"
```
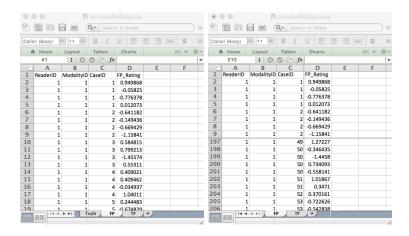
The `DfReadDataFile` function automatically recognizes that this is an *ROI*
dataset. Its structure is similar to the JAFROC format Excel file, with some
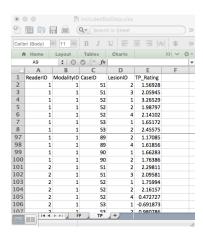important differences, noted below. It contains three worksheets:

- The `Truth` worksheet - this indicates which cases are diseased and which
  are non-diseased and the number of ROI-level-abnormal region on each
  case.

  - There are 50 normal cases (labeled 1-50) under column `CaseID` and
    40 abnormal cases (labeled 51-90).

  - The `LesionID` field for each normal case (e.g., `CaseID` = 1) is zero
    and there is one row per case. For abnormal cases, this field has

a variable number of entries, ranging from 1 to 4. As an example, there are two rows for `CaseID = 51` in the Excel file: one with `LesionID = 2` and one with `LesionID = 3`.

– The `Weights` field is always zero (this field is not used in ROI analysis).

- The `FP` (or `NL`) worksheet - this lists the ratings of ROI-level-normal regions.

  – For `ReaderID = 1`, `ModalityID = 1` and `CaseID = 1` there are 4 rows, corresponding to the 4 ROI-level-normal regions in this case. The corresponding ratings are . The pattern repeats for other treatments and readers, but the rating are, of course, different.

  – Each `CaseID` is represented in the `FP` worksheet (a rare exception could occur if a case-level abnormal case has 4 abnormal regions).

- The `TP` (or `LL`) worksheet - this lists the ratings of ROI-level-abnormal regions.

  - Because normal cases generate TPs, one does not find any entry with `CaseID` = 1-50 in the `TP` worksheet.

  - The lowest `CaseID` in the `TP` worksheet is 51, which corresponds to the first abnormal case.

  - There are two entries for this case, corresponding to the two ROI-level-abnormal regions present in this case. Recall that corresponding to this `CaseID` in the `Truth` worksheet there were two entries with `LesionID` = 2 and 3. These must match the `LesionID`'s listed for this case in the `TP` worksheet. Complementing these two entries, in the `FP` worksheet for `CaseID` = 51, there are 2 entries corresponding to the two ROI-level-normal regions in this case.

  - One should be satisfied that for each abnormal case the sum of the number of entries in the `TP` and `FP` worksheets is always 4.

Chapter 5

ROI data format

5.1 ROI paradigm

- One can think of the ROI paradigm as similar to the FROC paradigm, but with localization accuracy restricted to belonging to a region (one cannot distinguish multiple lesions within a region). The ROIs are defined prior to the study and made known to all observers participating in the study. Unlike the FROC paradigm, a **rating is required for every ROI**.

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