

# Package ‘semseeker’

August 17, 2023

**Type** Package

**Title** Stochastic Epigenetic Mutations SEM Seeker

**Version** 0.9.7

**Author** Luigi Corsaro, Davide Sacco

**Maintainer** Luigi Corsaro <lcorsaro69@gmail.com>

**Description** Stochastic epimutation and enriched region upstream and downstream tool for EWAS.

**License** AGPL-3

**Encoding** UTF-8

**URL** <https://github.com/drake69/semseeker>

**BugReports** <https://github.com/drake69/semseeker/issues>

**Imports** coxed, dplyr, doFuture, doRNG, FactoMineR, factoextra, foreach, FSA, fst, future, future.apply, ggplot2, gtools, Hmisc, lqmm, openxlsx, plyr, progressr, quantreg, readxl, reshape2, R.utils, rlang, stats, stringr, utils, withr, zoo

**RoxygenNote** 7.2.3

**Suggests** pathfindR, GEOquery, stringi, testthat

**Depends** R (>= 4.3.0)

**LazyData** true

**LazyDataCompression** gzip

**SysDataCompression** bzip2

**Config/testthat/parallel** false

**Config/testthat/edition** 3

**NeedsCompilation** no

## R topics documented:

analyze_population . . . . .	2
analyze_single_sample . . . . .	3
annotate_bed . . . . .	4
apply_stat_model . . . . .	4
association_analysis . . . . .	5
build_data_set_from_geo . . . . .	6

compute_qr_beta_boot_p . . . . .	6
compute_quantreg_beta_boot_np . . . . .	7
create_heatmap . . . . .	7
data_preparation . . . . .	8
delta_single_sample . . . . .	8
deltar_single_sample . . . . .	9
dir_check_and_create . . . . .	10
dump_sample_as_bed_file . . . . .	10
glm_model . . . . .	11
init_env . . . . .	11
manhattan_plot_per_area . . . . .	12
mutations_get . . . . .	12
pivot_to_long_format . . . . .	13
pp_tot . . . . .	14
PROBES . . . . .	14
PROBES_CHR_CHR . . . . .	14
quantreg_model . . . . .	15
quantreg_summary . . . . .	15
range_beta_values . . . . .	16
read_multiple_bed . . . . .	16
semseeker . . . . .	17
sort_by_chr_and_start . . . . .	18
test_match_order . . . . .	18
test_model . . . . .	19

## Index 20

---

analyze_population	<i>Calculate stochastic epi mutations from a methylation dataset as outcome report of pivot</i>
--------------------	---

---

## Description

Calculate stochastic epi mutations from a methylation dataset as outcome report of pivot

## Usage

```
analyze_population(
  methylation_data,
  sliding_window_size,
  sample_sheet,
  beta_thresholds,
  bonferroni_threshold = 0.05,
  probe_features
)
```

## Arguments

**methylation\_data**  
whole matrix of data to analyze.

**sliding\_window\_size**  
size of the sliding widows to compute epilesions default 11 probe\_features.

sample\_sheet      name of samplesheet's column to use as control population selector followed by selection value,  
 beta\_thresholds      thresholds defined to calculate epimutations lesions definition  
 bonferroni\_threshold      threshold to define which pValue accept for  
 probe\_features      probe\_features detail from 27 to EPIC illumina dataset

### Value

files into the result folder with pivot table and bedgraph.

---

analyze\_single\_sample      *analyze\_single\_sample*

---

### Description

analyze\_single\_sample

### Usage

```
analyze_single_sample(  
  values,  
  sliding_window_size,  
  thresholds,  
  figure,  
  sample_detail,  
  bonferroni_threshold = 0.05,  
  probe_features  
)
```

### Arguments

values              values of methylation  
 sliding\_window\_size      size of window sliding to calculate hypergeometric  
 thresholds              threshold to use for comparison  
 figure              which figure's of sasample will be analyzed HYPO or HYPER  
 sample\_detail      details of the sample to analyze  
 bonferroni\_threshold      bonferroni threshold to validate pValue  
 probe\_features      probe\_features details to be used

### Value

list of lesion count and probe\_features count

---

annotate_bed	<i>create an annotated file for each marker, figure, area and subarea, each file has all the sample_groups used to calculate epimutation</i>
--------------	--

---

**Description**

create an annotated file for each marker, figure, area and subarea, each file has all the sample\_groups used to calculate epimutation

**Usage**

```
annotate_bed()
```

**Value**

nothing

---

apply_stat_model	<i>Title</i>
------------------	--------------

---

**Description**

Title

**Usage**

```
apply_stat_model(
  tempDataFrame,
  g_start,
  family_test,
  covariates = NULL,
  key,
  transformation,
  dototal,
  session_folder,
  independent_variable,
  depth_analysis = 3,
  ...
)
```

**Arguments**

tempDataFrame	data frame to apply association
g_start	index of starting data
family_test	family of test to run
covariates	vector of covariates
key	key to identify file to elaborate
transformation	transformation to apply to covariates, burden and independent variable

dototal            do a total per area  
 session\_folder   where to save log file  
 independent\_variable  
                     independent variable name  
 depth\_analysis   depth's analysis  
 ...                extra parameters

---

association\_analysis   *Association analysis of SEMseeker's results*

---

## Description

Association analysis of SEMseeker's results

## Usage

```
association_analysis(  
  inference_details,  
  result_folder,  
  maxResources = 90,  
  parallel_strategy = "multisession",  
  ...  
)
```

## Arguments

**inference\_details**  
                     independent variable: deve essere nella sample sheet passata a semseeker quando lo abbiamo eseguito la prima volta tipo di regressioni: gaussian, poisson, binomial, quantreg\_tau\_runs(both as number) eg quantreg\_0.25\_2000 tipi di test: wilcoxon, stats::t.test, tipi di correlazioni: pearson, kendall, spearman MUTATIONS\_\* ~ tcdd\_mother + exam\_age transformation to be applied to dependent variable (mutations and lesions): scale, log, log2, log10, exp, none, quantile\_quantiles(as number) eg quantile\_3 depth analysis: 1: sample level 2: type level (gene, DMR, cpgisland) (includes 1) 3: genomic area: gene, body, gene tss1550, gene whole, gene tss200, (includes 1 and 2) filter\_p\_value report after adjusting saves only significant nominal p-value

**result\_folder**    where semseeker's results are stored, the root folder

**maxResources**    percentage of max system's resource to use

**parallel\_strategy**  
                     which strategy to use for parallel execution see future vignete: possible values, none, multisession, sequential, multicore, cluster

...                other options to filter elaborations

---

```
build_data_set_from_geo
      build_data_set_from_geo
```

---

### Description

build\_data\_set\_from\_geo

### Usage

```
build_data_set_from_geo(GEOgse, workingFolder, downloadFiles = 0)
```

### Arguments

GEOgse	geo accession dataset identification
workingFolder	where sample sheet and files will be saved
downloadFiles	0 means download all files from Gene Expression Ombibus (GEO), different than zero means how many download

### Value

samplesheet, and sample's file saved and samplesheet csv

---

```
compute_qr_beta_boot_p
      Title
```

---

### Description

Title

### Usage

```
compute_qr_beta_boot_p(sig.formula, tau, localDataFrame)
```

### Arguments

sig.formula	formula to use for regression application
tau	tau to apply the quantile regression
localDataFrame	dataframe to apply th regression model

---

compute_quantreg_beta_boot_np	
	<i>Title</i>

---

### Description

Title

### Usage

```
compute_quantreg_beta_boot_np(sig.formula, df, tau, lqm_control)
```

### Arguments

sig.formula	formula to apply
df	dataframe to use
tau	tau at which apply the wuantile regression
lqm_control	specification of the lqmm package

---

create_heatmap	<i>create_heatmap load the multiple bed resulting from analysis organized into files and folders per marker and produce a pivot</i>
----------------	---

---

### Description

create\_heatmap load the multiple bed resulting from analysis organized into files and folders per marker and produce a pivot

### Usage

```
create_heatmap()
```

### Value

nothing

---

data_preparation	<i>Title</i>
------------------	--------------

---

**Description**

Title

**Usage**

```
data_preparation(
  family_test,
  transformation,
  tempDataFrame,
  independent_variable,
  g_start,
  dototal,
  covariates,
  depth_analysis
)
```

**Arguments**

family_test	test or regression to apply
transformation	transformation to apply to data
tempDataFrame	data frame to use for test/regression
independent_variable	regressor
g_start	starting column of the dataframe
dototal	boolean to calculate the total burden test/regression
covariates	vector of covariates to be found in the sample sheet
depth_analysis	1 only sample, 2 chr, 3 alle genomic areas

---

delta_single_sample	<i>delta_single_sample</i>
---------------------	----------------------------

---

**Description**

delta\_single\_sample

**Usage**

```
delta_single_sample(
  values,
  high_thresholds,
  low_thresholds,
  sample_detail,
  beta_medians,
  probe_features
)
```



**Arguments**

values	values of methylation
high_thresholds	highest threshold to use for comparison
low_thresholds	lowest threshold to use for comparison
sample_detail	details of sample to analyze
beta_medians	median to use for calculation
probe_features	genomic position of probe_features

**Value**

summary detail about the analysis

---

deltar_single_sample	<i>delta_single_sample</i>
----------------------	----------------------------

---

**Description**

delta\_single\_sample

**Usage**

```
deltar_single_sample(
  values,
  high_thresholds,
  low_thresholds,
  sample_detail,
  beta_medians,
  probe_features
)
```

**Arguments**

values	values of methylation
high_thresholds	highest threshold to use for comparison
low_thresholds	lowest threshold to use for comparison
sample_detail	details of sample to analyze
beta_medians	median to use for calculation
probe_features	genomic position of probe_features

**Value**

summary detail about the analysis

---

dir_check_and_create	<i>dir_check_and_create</i>
----------------------	-----------------------------

---

**Description**

dir\_check\_and\_create

**Usage**

```
dir_check_and_create(baseFolder, subFolders)
```

**Arguments**

baseFolder	folder to look in
subFolders	sub folders to create, complete tree

**Value**

full path

---

dump_sample_as_bed_file	<i>given data and colnames dump as bed file</i>
-------------------------	---

---

**Description**

given data and colnames dump as bed file

**Usage**

```
dump_sample_as_bed_file(data_to_dump, fileName)
```

**Arguments**

data_to_dump	data frame to dump into bed file with CHR, START, END
fileName	name of the file to save data in

**Value**

nothing

---

glm_model	<i>Title</i>
-----------	--------------

---

**Description**

Title

**Usage**

```
glm_model(family_test, tempDataFrame, sig.formula)
```

**Arguments**

family_test	regression model to apply
tempDataFrame	data frame to use for the model
sig.formula	formula to apply the model

---

init_env	<i>init ssEnvonment</i>
----------	-------------------------

---

**Description**

init ssEnvonment

**Usage**

```
init_env(
  result_folder,
  maxResources = 90,
  parallel_strategy = "multicore",
  ...
)
```

**Arguments**

result_folder	where result of semseeker will bestored
maxResources	percentage of how many available cores will be used default 90 percent, rounded to the lowest integer
parallel_strategy	which strategy to use for parallel executio see future vignete: possibile values, none, multisession,sequential, multicore, cluster
...	other options to filter elaborations

**Value**

the working ssEnvonment

---

manhattan_plot_per_area	<i>Title</i>
-------------------------	--------------

---

**Description**

Title

**Usage**

```
manhattan_plot_per_area(  
  marker,  
  figure,  
  group,  
  subgroup,  
  family,  
  adjust_method,  
  phenotype,  
  only_significant_areas = FALSE  
)
```

**Arguments**

- marker            investigated marker eg. MUTATIONS, DELTAR, DELTAQ
- figure            HYPO, HYPER
- group            genomic area (eg. GENE, ISLAND, DMR)
- subgroup        sub genomic area (TSS1550), depending on the genomic area
- family           fullname of the family used for the association analysis
- adjust\_method   colnames of the pvalue adjusted to use
- phenotype        variable to select from the sample\_sheet to use for coloring point
- only\_significant\_areas  
                  TRUE if filter for pvalue < 0.05

---

mutations_get	<i>mutations_get</i>
---------------	----------------------

---

**Description**

mutations\_get

**Usage**

```
mutations_get(values, figure, thresholds, probe_features, sampleName)
```

**Arguments**

values	values of methylation
figure	figure to get Mutaions of HYPO or HYPER methylation
thresholds	threshold to use for comparison
probe_features	probe_features features probe, chr, start,end
sampleName	name of the sample

**Value**

mutations

---

`pivot_to_long_format`    *Get the pivot in long format instead of wide format*

---

**Description**

Get the pivot in long format instead of wide format

**Usage**

```

pivot_to_long_format(
  marker,
  figure,
  group,
  subgroup,
  phenotype_column,
  sample_sheet,
  areas_selection = NULL
)

```

**Arguments**

marker	marker to filer HYPER, HYPO, BOTH
figure	DELTAS, DELTAQ,DELTAR, MUTATIONS
group	GENE, DMR ...
subgroup	TSS1500 ...
phenotype_column	column from the sample sheet to pair to each sample
sample_sheet	sample sheet of samples
areas_selection	genomic area to select, if NULL all areas will be selected

**Value**

the pivot in a long format of 3 columnns, the phontype column with name phenotype, the value of the marker and the area investigated

---

pp_tot	<i>PROBES_CHR_CHR</i>
--------	-----------------------

---

**Description**

Full data set probe\_features as defined by Illumina

**Usage**

pp\_tot

**Format**

A data frame with five variables: year, sex, name, n and prop (n divided by total number of applicants in that year, which means proportions are of people of that sex with that name born in that year).

---

PROBES	<i>PROBES_CHR_CHR</i>
--------	-----------------------

---

**Description**

Full data set probe\_features as defined by Illumina

**Usage**

PROBES

**Format**

A data frame with five variables: year, sex, name, n and prop (n divided by total number of applicants in that year, which means proportions are of people of that sex with that name born in that year).

---

PROBES_CHR_CHR	<i>PROBES_CHR_CHR</i>
----------------	-----------------------

---

**Description**

Full data set probe\_features as defined by Illumina

**Usage**

PROBES\_CHR\_CHR

**Format**

A data frame with five variables: year, sex, name, n and prop (n divided by total number of applicants in that year, which means proportions are of people of that sex with that name born in that year).

---

quantreg_model	<i>Title</i>
----------------	--------------

---

**Description**

Title

**Usage**

```
quantreg_model(  
  family_test,  
  sig.formula,  
  tempDataFrame,  
  independent_variable,  
  boot_success,  
  tests_count  
)
```

**Arguments**

family_test	family lqmm, quantreg
sig.formula	formula of the model
tempDataFrame	data
independent_variable	name of regressor
boot_success	number of success tests to calculate corrected confidence interval
tests_count	count of total executed tests

---

quantreg_summary	<i>Quantile regression result value, confidence interval and pvalue</i>
------------------	---

---

**Description**

Quantile regression result value, confidence interval and pvalue

**Usage**

```
quantreg_summary(  
  boot_vector,  
  estimate,  
  conf.level,  
  boot_success = 0,  
  tests_count = 1  
)
```

**Arguments**

boot_vector	vector of boot statistic beta regression
estimate	beta regression
conf.level	confidence intervals alpha level
boot_success	number of success respecting the null hypothesis
tests_count	how many tests were done

**Value**

ci and pvalue with BCA method

---

range_beta_values	<i>calculate the range of beta values to define the outlier</i>
-------------------	---

---

**Description**

calculate the range of beta values to define the outlier

**Usage**

```
range_beta_values(populationMatrix, iqrTimes = 3)
```

**Arguments**

populationMatrix	matrix of methylation for the population under calculation
iqrTimes	inter quartile ratio used to normalize

**Value**

methylation matrix as normalized distribution

---

read_multiple_bed	<i>read multiple bed with annotated data as per input parameter</i>
-------------------	---

---

**Description**

read multiple bed with annotated data as per input parameter

**Usage**

```
read_multiple_bed(sample_group, marker, figure)
```

**Arguments**

sample_group	name of the population used to build the data path
marker	marker definition used to label folder and files eg MUTATIONS, LESIONS
figure	figures like hypo/hyper to built the data path



**Value**

list of pivot by column identified with column Label and by Sample

---

semseeker	<i>Calculate stochastic epi mutations from a methylation dataset as outcome report of pivot</i>
-----------	---

---

**Description**

Calculate stochastic epi mutations from a methylation dataset as outcome report of pivot

**Usage**

```
semseeker(
  sample_sheet,
  methylation_data,
  result_folder,
  bonferroni_threshold = 0.05,
  maxResources = 90,
  iqrTimes = 3,
  parallel_strategy = "multisession",
  ...
)
```

**Arguments**

sample_sheet	dataframe with at least a column Sample_ID to identify samples
methylation_data	matrix of methylation data
result_folder	where the result will be saved
bonferroni_threshold	= 0.05 #threshold to define which pValue adjusted to define an epilesion
maxResources	percentage of how many available cores will be used default 90 percent, rounded to the lowest integer
iqrTimes	how many times below the first quartile and over the third quartile the interquartile is "added" to define the outlier
parallel_strategy	which strategy to use for parallel executio see future vignete: possibile values, none, multisession,sequential, multicore, cluster
...	other options to filter elaborations

**Value**

files into the result folder with pivot table and bedgraph.

---

sort_by_chr_and_start	<i>sort the dataframe using CHR and START sorting column first for CHR and after for START</i>
-----------------------	--

---

**Description**

sort the dataframe using CHR and START sorting column first for CHR and after for START

**Usage**

sort\_by\_chr\_and\_start(dataframe)

**Arguments**

dataframe          dataframe to be sorted

**Value**

sorted dataframe

---

test_match_order	<i>Title</i>
------------------	--------------

---

**Description**

Title

**Usage**

test\_match\_order(x, y)

**Arguments**

x                      vector to compare  
y                      vector to compare

**Value**

true if the order matches otherwise is false

---

test_model	<i>Title</i>
------------	--------------

---

**Description**

Title

**Usage**

```
test_model(  
  family_test,  
  tempDataFrame,  
  sig.formula,  
  burdenValue,  
  independent_variable  
)
```

**Arguments**

- family\_test      which family test to apply
- tempDataFrame   data frame to use with the test
- sig.formula      formula to apply
- burdenValue      burden colon name
- independent\_variable      independent variable for regressor

# Index

## \* datasets

- pp\_tot, [14](#)
- PROBES, [14](#)
- PROBES\_CHR\_CHR, [14](#)

- analyze\_population, [2](#)
- analyze\_single\_sample, [3](#)
- annotate\_bed, [4](#)
- apply\_stat\_model, [4](#)
- association\_analysis, [5](#)

- build\_data\_set\_from\_geo, [6](#)

- compute\_qr\_beta\_boot\_p, [6](#)
- compute\_quantreg\_beta\_boot\_np, [7](#)
- create\_heatmap, [7](#)

- data\_preparation, [8](#)
- delta\_single\_sample, [8](#)
- deltar\_single\_sample, [9](#)
- dir\_check\_and\_create, [10](#)
- dump\_sample\_as\_bed\_file, [10](#)

- glm\_model, [11](#)

- init\_env, [11](#)

- manhattan\_plot\_per\_area, [12](#)
- mutations\_get, [12](#)

- pivot\_to\_long\_format, [13](#)
- pp\_tot, [14](#)
- PROBES, [14](#)
- PROBES\_CHR\_CHR, [14](#)

- quantreg\_model, [15](#)
- quantreg\_summary, [15](#)

- range\_beta\_values, [16](#)
- read\_multiple\_bed, [16](#)

- semseeker, [17](#)
- sort\_by\_chr\_and\_start, [18](#)

- test\_match\_order, [18](#)
- test\_model, [19](#)