C. Chase Bolt, Ph.D.

orcid.org/0000-0002-3544-3552 christopher.bolt@epfl.ch http://chasebolt.com EPFL SV ISREC UPDUB SV 2843 (Bâtiment SV) Station 19 Lausanne CH 1015 +41 (0)21 693 06 06 (work) +41 (0)79 139 45 17 (mobile)

Post-Doctoral Work

École Polytechnique Fédérale de Lausanne - Laboratory of Denis Duboule

April 2015 - Present

Department: Swiss Institute for Experimental Cancer Research

Research -

Regulation of the HoxD cluster during mouse digit and reproductive development

Education

Doctorate of Philosophy - Laboratory of Lisa Stubbs

Aug 2008 - May 2015

Department: University of Illinois - Cell and Developmental Biology Research –

Regulatory dynamics of the Tbx18 locus in mouse urogenital development

available at http://hdl.handle.net/2142/78608

Teaching -

Teaching Assistant: MCB 410 – Developmental Biology (Discussions)

Spring 2014

Teaching Assistant: MCB 250 – Molecular Genetics (Discussions and Special Tutoring)
Ranked "Excellent" on TA reviews by students

Fall 2012

Teaching Assistant: CDB 495 – Genetics and Genomics (unofficial)

Spring 2010

Bachelor of Science, Cum Laude – Laboratory of Nicholas Stover

Jan 2005 - May 2008

Major: Bradley University - Cellular and Molecular Biology

Research -

Characterization of a novel non-receptor tyrosine kinase in the cnidarian, Hydra magnipapillata

Teaching -

Teaching Assistant: General Biology I and II Teaching Assistant: General Chemistry I 2007 - 2008 Fall 2007

Publications

Bolt CC, Negi S, Guimarães-Camboa N, Zhang H, Troy JM, Lu X, et al. *Tbx18* Regulates the Differentiation of Periductal Smooth Muscle Stroma and the Maintenance of Epithelial Integrity in the Prostate. PLoS ONE. 2016;11: e0154413–20. PMID 27120339.

Bolt CC, Elso CM, Lu X, Pan F, Kispert A, Stubbs L. A distant downstream enhancer directs essential expression of *Tbx18* in urogenital tissues. Developmental Biology. Elsevier; 2014 Aug 15;392(2):483–93. PMID 24854998.

Conferences & Symposia	
Society for Developmental Biology 75th Annual Meeting – Boston, MA Regulation of the HoxD cluster in the urogenital system	Aug 2016
Institute for Genomic Biology Fellows Symposium – Urbana, IL Dissecting the <i>Tbx18</i> locus regulatory structure and its role in mouse prostate development	May 2013
Cellular & Molecular Biology Training Grant Symposium – Urbana, IL Developmental dynamics of the <i>Tbx18</i> locus and its downstream transcriptional targets	Nov. 2012
GSA Mouse Molecular Genetics Conference – Pacific Grove, CA Developmental dynamics of the <i>Tbx18</i> locus and its downstream transcriptional targets	Oct. 2012
Scholarships & Awards	
NIH Ruth L. Kirchstein NRSA Predoctoral Training Fellowship	2010 - 2012
Bradley Biology Scholarship awarded for academic excellence	2005 - 2008
Bradley University Dean's List awarded for academic excellence	2005 - 2008
Mentoring	
John Elue Advised his senior thesis research.	2014 - 2015
Currently at Rush Medical College	
Andrew Park Advised his undergraduate research project and senior thesis Currently at University of Illinois College of Medicine	2009 - 2012
Drafassianal Davalanment 9 Carries	
Professional Development & Service MCB Graduate Student Advisory Panel Provide advice to first year graduate students in the program	Dec. 2013
Career Professionalization Seminar Committee Nominate, invite, and host guest speakers	Aug. 2012 – Mar. 2013
Cellular & Developmental Biology Department Seminar Student Committee Invite and host guest speakers working in a field of personal or professional interest, with relevance to the department. Blanche Capel, Ph.D., Duke University (Oct. 2012) Janet Rossant, Ph.D., University of Toronto (April 2014)	2011-2013
MCB Graduate Student Advisory Panel Provide advice to first year graduate students in the program	Oct. 2011
UIUC CMBTG Symposium Chair Managed weekly meetings, organized and delegated tasks, introduced speakers during speaking sessions	May – Oct. 2011

Animal Work and Training

Module 1 Certified by Réseau des Animaleries Lémaniques (RESAL)

June 2015

20 hours theoretical and 20 hours practical training on animal research models. Program accredited by the Federation of European Laboratory Animal Science Associations (FELASA) and recommended by the Swiss Federation of Cantonal Veterinary Surgeons (ASCV).

Mouse colony maintenance

Aug. 2008 - April 2015

During my time in the Stubbs lab I maintained my own mouse colony of approximately 300 mice. My duties included strain preservation, genotyping, earmarking, and weaning.

Professional Memberships

Society for Developmental Biology 2015 - Present

Genetics Society of America 2012 - 2014

Graduate Coursework

RNA-Seg Data Analysis Workshop June 2013

Pipeline construction for the analysis of RNA-Seq data analysis.

MCB 529 – Genomics and Gene Networks Spring 2010

Tools for the analysis of genomics data.

MCB 571 — Bioinformatics Fall 2009

Overview of the tools available for manipulation of bioinformatics data.

MCB 529LS – Genome Annotation Spring 2009

Using Apollo to build gene models from sequencing data.

MCB 529AB – Advanced Cell Biology Spring 2009

In depth study of cell biology and experimental techniques

MCB 502 – Advanced Molecular Genetics Fall 2008

Study of The Central Dogma and associated experimental techniques.

MCB 501 – Advanced Biochemistry Fall 2008

Advanced study of biochemical pathways and associated chemistry.

Skills & Techniques

Mouse colony management, RT-qPCR, Cloning, 5' and 3' RACE, Transcription Factor and Histone ChIP-Seq, RNA-Seq, Immunohistochemistry, *In situ* hybridization (whole mount, paraffin embedded, and frozen), Tissue culture, Histopathology, Western blot, siRNA knockdown, FACS, X-Gal staining, Whole genomic amplification, Histopathology, 4C-Seq, CRISPR Transgenesis.

Summary of Research

The expression of developmental genes is carefully controlled because they affect dramatic cellular changes such as lineage commitment and cellular differentiation. The developmental genes that facilitate these important biological transitions are frequently associated with an array of *cis*-acting regulatory elements which provide a platform for transcription factors proteins to control the expression of the target gene. The cell controls the accessibility of these elements using an assortment of epigenetic modifications. Together with chromatin architectural proteins, such as CTCF, these elements and transcription factors form a local compartment of chromatin loops that maximise enhancer-promoter contacts resulting in expression changes. The properties of regulatory elements located within and around developmental genes are frequently difficult to disambiguate because they can have redundant or additive functions.

Particular types of DNA structural mutations, such as chromosome translocations and inversions allow for a simple two-part dissection of regulatory elements from their target gene. In the Duboule Lab, we utilise a collection of structural mutations in the *HoxD* locus to study how the local chromatin architecture influences the transcription of these genes *in vivo*. The *HoxD* cluster contains nine *Hoxd* genes that are expressed in multiple domains of the embryo during a coordinated wave of transcription from one end of the cluster to the other. This remarkable property, termed collinearity, is controlled by regulatory elements disbursed around the cluster of genes. By disrupting the *cis*-regulation of these genes, dramatic morphological changes emerge in the embryo. This effect has been observed in species as ancient as the fruitfly, and is a unique property of the *Hox* genes, demonstrating their powerful influence on the formation of body patterns.

In pursuit of this knowledge, the primary goal of my research is to understand how long-range genetic regulation modulates expression of genes during embryonic development. Specifically, my current work in the Duboule Lab is focused on two projects. The first is an analysis of *Hox* gene function during the formation of the urogenital system. The formation of these tissues begins around embryonic day E10.5, which is concurrent with the establishment of *Hox* gene expression domains along the mesoderm, from which these organs arise. Interestingly, the anterior boundaries of *Hoxd* and *Hoxa* genes delineate the boundaries of nearly every portion of the urinary and reproductive systems. Loss-of-function mutations in the *HoxA* or *HoxD* clusters cause homeotic transformations in these tissues, several of which are similar to human congenital

malformations. I am working to understand how the *HoxD* genes are regulated during the formation and maintenance of these structures.

My second project is focused on the gene *Hoxd13* during formation of the handplate (autopod). The expression of this gene is very tightly controlled during limb formation because premature activation of *Hoxd13* abruptly terminates the collinearity program resulting in abbreviated arms and legs. Our previous work has shown that a chromatin structure sequesters *Hoxd13* from the activating mechanism that works on the adjacent genes in the cluster. The same work has shown that *Hoxd13* is in constitutive contact with many positions in the adjacent gene desert but when the gene is activated, a few additional elements gain chromatin contact with *Hoxd13*. To parse the regulatory structure controlling *Hoxd13*, we are utilising the CRISPR system to precisely modify elements that control *Hoxd13* in vivo. A main goal for this work is to determine how enhancers act on specific target genes in a complex chromatin landscape and how enhancer elements control the properties of the adjacent chromatin.