Dating with constraints

A tutorial on McmcDate

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Abstract

McmcDate can date a phylogenetic tree with constraints. That is, it can estimate the ages of the ancestral nodes of a phylogenetic tree with node age calibrations, node order constraints, and node braces. McmcDate is fast because it approximates the phylogenetic likelihood with a multivariate normal distribution.

In this tutorial, we are going to date a phylogenetic tree of eukaryotes. We are going to use node age calibrations from fossils, and node order constraints from two possible successions of plastid endosymbiosis events.

This tutorial is a work in progress.

Definitions and explanations

We use the term *(rooted)* tree to denote a directed acyclic graph with node labels and branch lengths, and in which not more than one branch connects any two nodes. Usually, nodes correspond to (ancestral) species, and branch lengths to sequence distance or time. We use the term *(rooted)* topology to denote a (rooted) tree without information about branch lengths. We put the word rooted into parentheses, because it is sometimes omited. The root of the tree or topology is the only node with all branches pointing away to other nodes, and is the oldest node. Leaves are extant nodes with a single branch pointing towards them.

For a given topology, dating the tree¹ corresponds to finding branch lengths for this topology which are measured in real time (for example, in Million years), and which describe the data in the best way. Best can mean different things, and we will carefully analyze what best means in our case.

Similarly, we use the term *unrooted tree* to denote an undirected acyclic graph with node labels and branch lengths, and in which not more than one branch connects any two nodes. Finally, we use the term *unrooted topology* to denote an unrooted tree without information about branch lengths. We need unrooted trees and topologies because we will use reversible substitution models (for a review, see Yang 2006) to infer unrooted trees with branch lengths measured in expected number of substitutions per unit time. Reversible substitution models are unable to discriminate between the two directions in time.

¹We should probably say dating the topology but this phrase is not used.

Dating a tree

The pipeline for dating a topology is:

- 1. Prepare a multi sequence alignment and an unrooted topology.
- 2. For this alignment and unrooted topology, infer a distribution of unrooted trees with branch lengths measured in expected number of substitutions per unit time.
- 3. Find a rooted topology to date. Prepare auxiliary data such as node age calibrations or node order constraints.
- 4. Date the topology with McmcDate. In particular, infer a distribution of rooted trees with branch lengths measured in real time.

The following sections describe the steps of the pipeline in detail.

Step 1: Provision of alignment and unrooted topology

We are going to date a topology of eukaryotes (Strassert et al. 2021). The full published data set contains three alignments:

- 1. The alignment with the highest number of taxa comprises 733 eukaryotes with 62723 amino acids.
- 2. The authors used the first alignment to infer a tree with IQ-TREE (Minh et al. 2020) so they can filter taxa in an informed way (for example, keep slow evolving taxa). The reduced data set comprises 136 operational taxonomic units with 73460 amino acids.
- 3. A small data set with 63 operational taxonomic units and 73460 amino acids for tree inference with Bayesian models.

Here, we focus on the second alignment with 136 taxa. We use shorter taxon names and I have converted the file from FASTA format to Phylip format which is required by Phylobayes (Lartillot et al. 2013). I provide the alignment strassert-320genes-136taxa.phy in the data subfolder of this tutorial.

Further, we need a topology to date. Strassert et al. (2021) discuss two possible topologies. In this tutorial, we will choose the more plausible one: The root separates Amorphea and Diaphoretickes (see Figure 3 in Strassert et al. (2021)).

TODO: Continue here with the tree/topology.

Step 2: Phylogenetic inference

• Use Phylobayes (Lartillot et al. 2013).

• Decide on evolutionary model depending on the size of the data set and the computational requirements. Recommended models from preferred but slow and complex to fast and simple: GTR+CAT+G4, LG+CAT+G4, LG+EDM64+G4, LG+C60+G4, LG+G4.

We specify an evolutionary model with exchangeabilities EX, and across-site compositional heterogeneity model ASCH as EX+ASCH. All discussed evolutionary models used for simulations as well as inferences implicitly use discrete gamma rate heterogeneity with four components.

- GTR model (Tavaré 1986).
- CAT model (Lartillot and Philippe 2004).
- Gamma rate variation model (Yang 1993).
- LG model (Le and Gascuel 2008).
- EDM model (Schrempf et al. 2020).
- C60 model (Quang et al. 2008)

Step 3: Preparation of rooted topology and auxiliary data

- Node order calibrations (Yang and Rannala 2005).
- Relative node order constraints (Szöllősi et al. 2022).
- McmcDate can also brace nodes (Appendix Node braces).

Step 4: Dating with McmcDate

• McmcDate is a Haskell program (Appendix Internals of McmcDate).

Node braces

Internals of McmcDate

McmcDate is a Haskell program.

Recommend cabal, but there is also stack (-s) option.

The wrapper script used in this tutorial tries to make a good compromise between usability and customizability. It exposes some, but not all functionality of McmcDate. Most notably,

- Based on mcmc.
- Based on elynx-tree.
- Explain code a bit (I guess mostly proposals).

Haskell modules

Modules containing definitions specific to the analysis are in the app subfolder of the McmcDate repository.

More important modules

Definitions Proposals and monitors, configuration.

State State space. If you try to understand what is going on, or if you want to change analysis settings, this should be your starting point.

Other modules

Hamiltonian Hamiltonian proposal.

Main Functions to prepare the data, run and continue the Metropolis-Hasting-Green algorithm.

Monitor Prior specific monitoring functions.

Options Handle command line options.

Probability Prior and likelihood functions.

Tools Miscellaneous tools.

References

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