

SHORT COMMUNICATION

Precision of age estimates in striped snakehead *Channa striata* (Bloch, 1793) from the Ganga River and its tributaries (rivers Gomti and Yamuna)

M. Afzal Khan | Salman Khan | Shahista Khan

Section of Fishery Science & Aquaculture, Department of Zoology, Aligarh Muslim University, Aligarh, India

Correspondence

M. Afzal Khan, Section of Fishery Science & Aquaculture, Department of Zoology, Aligarh Muslim University, Aligarh, India.

Email: khanmafzal@yahoo.com

Funding information

Research Assistantship provided by the Council of Science and Technology, Lucknow, Uttar Pradesh, India

1 | INTRODUCTION

Precise age estimation is crucial for fisheries management. Age composition and parameters such as growth, mortality, maturity, recruitment and estimates of longevity are significant indicators to assess stock status. Ageing studies use various calcified structures including otoliths, scales, opercular bones, vertebrae, fin rays and others (Campana, 2001; Khan, 2014). Inaccuracy in ageing fish can lead to errors in fisheries management decisions (Beamish & McFarlane, 1983). Ageing errors can originate from lack of accuracy, or the precise reproducibility of repeated measurements on a given structure (Kalish et al., 1995). Age underestimates may result in high growth calculations and mortality rate estimates leading to serious over-exploitation of populations and their eventual collapse (Campana, 2001).

The freshwater striped murrel, *Channa striata*, (Channidae) is distributed throughout India and other Asian countries. It is a valuable food fish (Haniiffa, Marimuthu, Nagarajan, Jesu Arokiaraj, & Kumar, 2004). Several studies have compared age estimates from different bony structures to obtain precise data, avoiding potential biases associated with each structure. Well-studied species include *Cyprinus carpio* (Phelps, Edwards, & Willis, 2007), *Labeo rohita*, *Catla catla*, and *Channa marulius* (Khan & Khan, 2009), *Channa punctata* (Khan, Khan, & Miyan, 2013b), *Hoplias malabaricus* (Lozano, Vegh, Doma' nico, & Ros, 2014), and *Labeo bata* (Khan, Khan, & Miyan, 2015). Otoliths have been previously used for age and growth studies in *C. striata* (Jutagate, Phomikong, Avakul, & Saowakoon, 2013). However, there are no publications on the precision of age estimates from various calcified structures in *Channa striata*. Therefore, the objective of this study was to compare and evaluate the age estimates from different calcified structures (otoliths, scales, vertebrae, opercular bones, cleithra and fin rays)

in order to identify the most suitable calcified structure exhibiting a precise age estimate in *C. striata*.

2 | MATERIALS AND METHODS

2.1 | Sample collection

Channa striata (N = 486) were collected monthly from November 2012 to August 2015 in the River Ganga (N = 175) and two of its tributaries, the River Gomti (N = 160), and the River Yamuna (N = 151) in India (Figure 1). Fish were collected using cast nets (mesh size = 25 mm) and drag nets (mesh size = 28 mm), ensuring that the ageing structures were not damaged in the selected samples. Total length (TL) of each fish was measured to the nearest cm and body weight recorded to the nearest gram as total weight (TW) including gut and gonads. Scales, otoliths, opercular bones, vertebrae, cleithra and fin rays were removed from the fish and prepared for ageing according to Khan & Khan, 2009; Khan, Nazir, & Khan, 2016 and Sabah & Khan, 2014;. All structures were examined under a stereozoom microscope with 10×, 20× and 40× magnification under transmitted light (Figure 2). Each calcified structure was examined by **two readers independently** (486 samples × 7 structures × 2 readers × 1 replicate). Neither reader had prior information on the fish length, weight or sex, except for the collection date. An opaque and a translucent zone together were regarded as a year's growth; age was considered to be the total number of translucent zones.

2.2 | Calculations and statistical analysis

Age estimates were compared by calculating the average percentage error (APE), coefficient of variation (CV), and percentage agreement

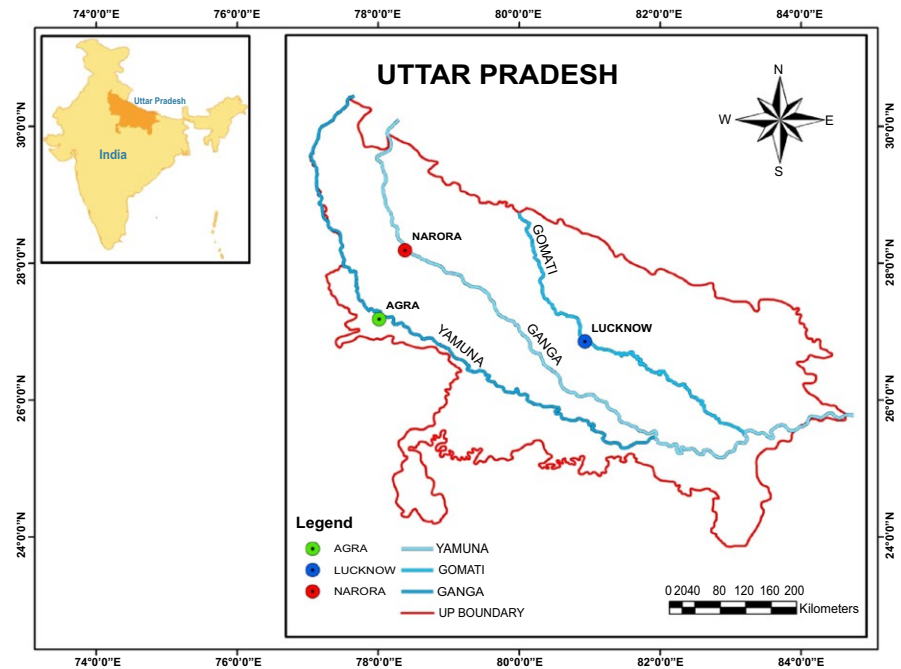


FIGURE 1 Map indicating the three sampling sites

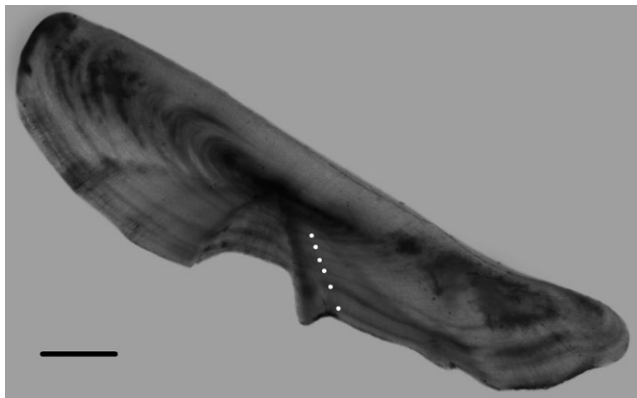


FIGURE 2 Sectioned otolith of *Channa striata* under transmitted light (dots indicate annuli). Scale bar = 1000 μ m

(PA) between the readers and between the pairs of ageing structures. To calculate APE the formula of Beamish and Fournier (1981) was used:

$$APE_j = 100\% \times \frac{1}{R} \sum_{i=1}^R \frac{|x_{ij} - x_j|}{x_j}$$

where x_{ij} is the i th age determination of the j th fish, x_j is the average age calculated for the j th fish, and R is the number of times each fish was aged.

The coefficient of variation (Campana, 2001) was calculated as the ratio of standard deviation over the mean, which can be written as

$$CV_j = 100\% \times \frac{\sqrt{\frac{\sum_{i=1}^R (x_{ij} - x_j)^2}{R-1}}}{x_j}$$

where CV_j is the age precision estimate for the j th fish.

Both APE and CV have been widely used as statistically sound measures of ageing precision in fishes (Campana, 2001). However, although PA is used as an index of ageing precision in fish by many researchers (Hoxmeier, Aday, & Wahl, 2001; Sabah & Khan, 2014; Stolarski & Hartman, 2008) it is not considered to be a suitable measure of precision by several authors (Beamish & Fournier, 1981; Campana, Annand, & McMillan, 1995; Chang, 1982). Percentage agreement may be expressed as the percentage of the number of observations showing similar age estimates to the total number of observations on age estimates. Percentage agreement was calculated using the templates for calculating ageing precision (Sutherland, 2006). Age bias graphs (Campana et al., 1995) were constructed to examine potential biases between readers and between pairs of ageing structures.

Mean age readings obtained from various bony structures were subjected to one-way analysis of variance (ANOVA) followed by Duncan's multiple range test (DMRT) (Gomez & Gomez, 1984), in order to explain whether the readings from different bony structures of the same species showed significant differences among themselves (Khan & Khan, 2009). Although the mean age estimate is not an indicator for the reliability of ageing structure, it may provide useful information regarding over- or under-estimation of age by a structure irrespective of fish size-class. This may prove useful in selecting the structure(s) that could be used for age estimation in the absence of the most suitable ageing structure.

3 | RESULTS

Average percentage error (1.28%) and CV (2.59%) values were lowest for the age estimates of sectioned otoliths followed by whole otoliths, scales, opercular bones, vertebrae, cleithra and fin rays. Percentage agreement between readers was highest for sectioned

Table 1 Comparison of percentage of agreement (PA), average percentage of error (APE) and coefficient of variation (CV) between the age readings of two readers and between pairs of ageing structures in *C. striata* collected from three Indian rivers $N = 486$

Hard parts	APE	CV	PA
Sectioned otoliths	1.28	2.59	89.7
Whole otoliths	2.14	3.89	85.0
Scales	3.66	5.15	79.2
Opercular bones	6.14	7.35	71.2
Vertebrae	8.75	9.92	60.9
Cleithra	11.2	12.62	53.5
Fin rays	14.8	15.4	43.2
Between structures			
Sectioned otoliths—whole otoliths	0.86	1.14	95.3
Sectioned otoliths—scales	1.98	2.03	91.4
Sectioned otoliths—opercular bones	3.99	3.63	86.4
Sectioned otoliths—vertebrae	5.77	5.26	80.5
Sectioned otoliths—cleithra	7.05	7.80	71.4
Sectioned otoliths—fin rays	10.10	8.76	70.6

otoliths (Table 1). Ageing structures exhibited variation in estimates of age composition for fish collected from three Indian rivers (Figure 3). When sectioned otoliths age estimates were compared with other ageing structures (i.e. whole otoliths, scales, opercular bones, vertebrae, cleithra and fin rays), the least variation was observed between sectioned otoliths versus whole otoliths age estimates, as indicated by lowest APE and CV values followed by sectioned otoliths versus scales, sectioned otoliths versus opercular bone, sectioned otoliths versus vertebrae, sectioned otoliths versus cleithra and sectioned otoliths versus fin rays (Table 1).

Age bias graphs between age estimates from sectioned otoliths and each of the other ageing structures are presented in Figure 4. Age estimates from sectioned otoliths were in good agreement with those

Table 2 Comparison of mean values of age estimates from different bony parts in *C. striata*

Ageing structure	Mean values of age estimates
Sectioned otoliths	4.1255 ^b
Whole otoliths	4.0864 ^b
Scales	4.0473 ^b
Opercular bones	3.9918 ^{ab}
Vertebrae	3.9568 ^{ab}
Cleithra	3.9444 ^{ab}
Fin rays	3.7922 ^a

Mean values with similar superscripts are insignificantly different to each other.

from whole otoliths but differed substantially when compared to the age estimates from other ageing structures. Mean values of age estimates from different structures, when compared using ANOVA followed by DMRT, showed that maximum age estimates obtained from sectioned otoliths were significantly ($p < 0.05$) higher from dorsal fin rays but comparable ($p > 0.05$) to the values obtained from whole otoliths, scales, opercular bones, vertebrae and cleithra (Table 2).

4 | DISCUSSION

Whole otoliths in *C. striata* exhibited clearer annuli that were easier to read than other structures. However, when otoliths were sectioned the annuli became more visible, requiring less time and effort by the readers. Sectioned otoliths have also been reported to provide precise age estimates in *Paralichthys dentatus* (Sipe & Chittenden, 2001) and *Sperata aor* (Khan et al., 2016). The suitability of otoliths for age estimation is supported by the fact that otoliths do not show reabsorption and their growth is acellular rather than by calcification (Secor, Trice, & Hornick, 1995). Otoliths are also reported to be metabolically inert and thus do not reflect physiological changes that might occur throughout the life of the fish (Phelps et al., 2007). Otoliths continue to grow and form annuli, even when the body growth slows and the asymptotic length is reached; annuli reabsorption also does not seem to occur during periods of food limitation or stress (DeVries & Frie, 1996). Otoliths were reported to be

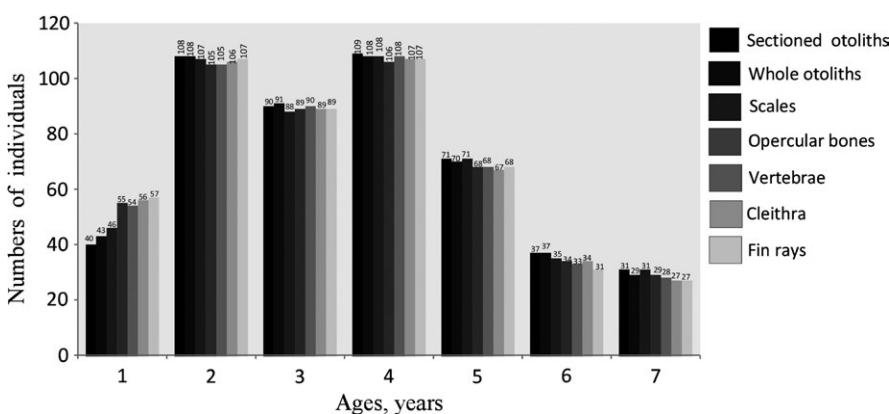


FIGURE 3 Age composition of *C. striata* derived from readings of different ageing structures. Total number of samples ($N = 486$) remains the same for each ageing structure

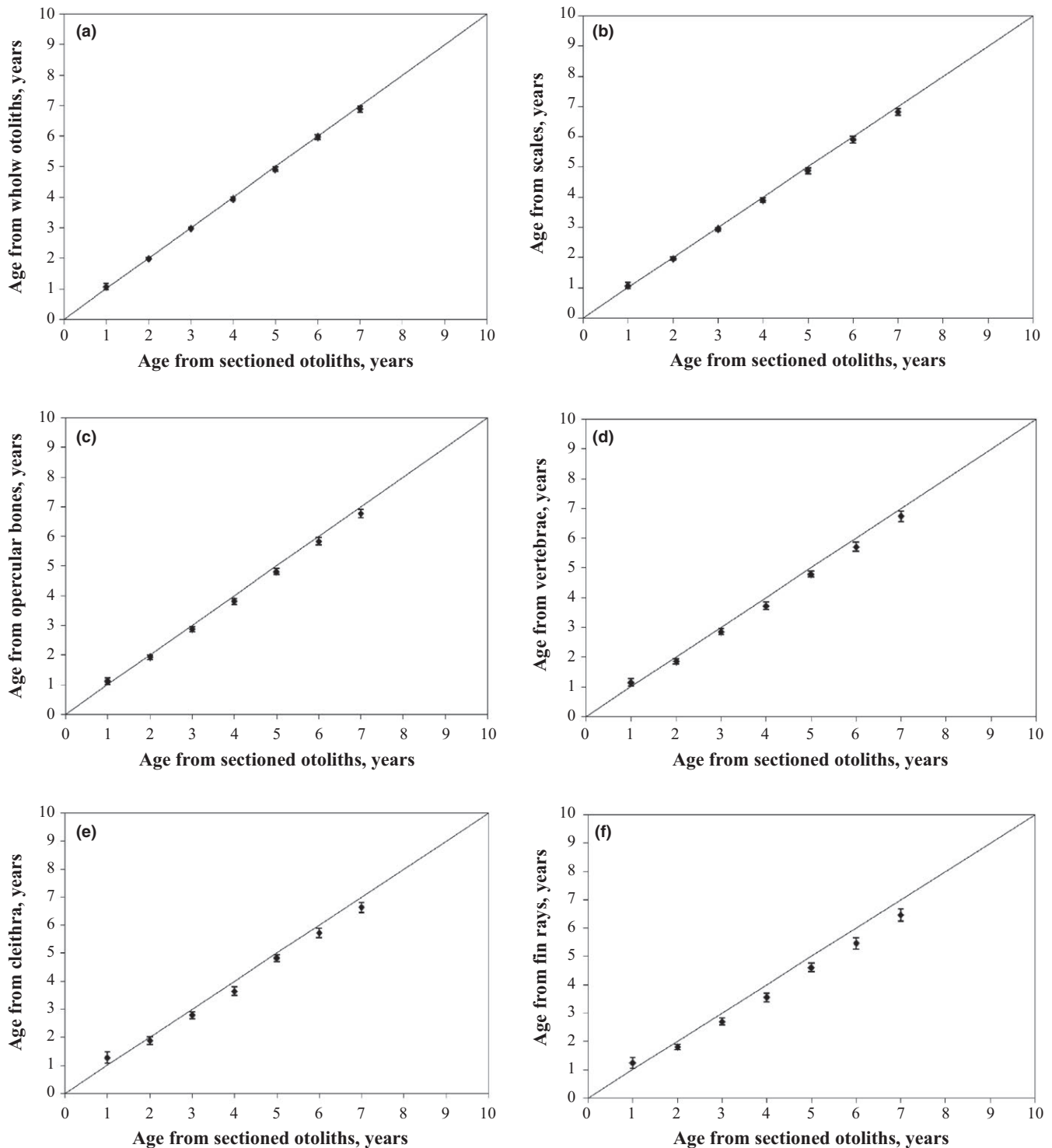


FIGURE 4 Age bias graphs for *C. striata* (N=486) age estimates from sectioned otoliths versus (a) whole otoliths, (b) scales, (c) opercular bones, (d) vertebrae, (e) cleithra and (f) fin rays. Each error bar represents the 95% confidence interval. The 1:1 equivalence (solid) line is also indicated. Points above the line indicate ages that were overestimated whereas a point below the line indicates ages that were underestimated

the most suitable ageing structure in a number of fish species such as *Pylodictis olivaris* (Nash & Irwin, 1999), *Ictalurus punctatus* (Buckmeier, Irwin, Betsill, & Prentice, 2002), *Clarias gariepinus* (Khan, Khan, & Miyan, 2011) and *Channa punctatus* (Khan et al., 2013b). Khan, Khan, and Miyan (2013a) reported otoliths as the most time-efficient and precise means of age estimates in *Clarias batrachus* compared

to vertebrae and pectoral spines. In the present study scales also showed a good value of agreement between age readers because annuli were clear and easy to read and have been the most widely used ageing practice for the majority of cyprinids (Kamilov, 1984), primarily due to advantages such as the ease of collection and preparation, and more importantly, because the method is non-destructive to fish

(DeVries & Frie, 1996). In the published literature on Indian freshwater fishes, scales have been exclusively used for age studies in *Catla catla* (Natarajan & Jhingran, 1963) and *C. mrigala* (Kamal, 1969). According to Kamal (1969), *Cirrhinus mrigala* scales showed clear annulations. In the first published report on precision of age estimates from different ageing structures of Indian freshwater fishes, it was observed that in *L. rohita*, *C. catla*, and *C. marulius*, among different structures (scales, otoliths, opercular bones, vertebrae, and dorsal fin rays), scales were the most suitable structure for ageing *L. rohita* and *C. marulius*, but that opercular bones were best for ageing *C. catla* (Khan & Khan, 2009). In corroboration with the observations of Beamish and McFarlane (1987), we also noted an underestimation of age by scales in older (5+ years) *C. striata*; however, annuli on the scales of younger fish were quite clearly legible. Opercular bones also provided age readings, as evident from the PA, APE, and CV values between structures. In the present study opercular bones did not show clear annuli in older fishes, possibly due to an overgrowth of dense bone near the hyomandibular socket. Similarly, in the long-nose sucker (*Catostomus catostomus*) this problem was observed by Perry and Casselman (2012), who concluded that when using the whole operculum technique, interpreters should be aware that as fish become older, increments and annuli located close to the origin become obscured. Also, in some fishes the opercular bones were found to be less reliable than other structures, e.g. in *Schizothorax o'connori* (Ma, Xie, Huo, Yang, & Li, 2011) *Labeo rohita* and *Channa marulius* (Khan & Khan, 2009). Khan et al. (2015) determined that the opercular bones were the second most suitable ageing structure for *Labeo bata* in the Ganga River. In the present study, rings on the vertebral centra were not very clear and showed numerous minute marks unrelated to cyclical events. Because we used whole vertebral centra for age estimation in *C. striata*, the degree of clarity and sharpness of the growth rings were low. Several researchers have reported that the rings on vertebral centra were not very clear and showed numerous minute marks unrelated to cyclic events in a number of fish species such as *Makaira nigricans* (Hill, Calliet, & Radtke, 1989), *Labeo rohita*, *Catla catla*, and *Channa marulius* (Khan & Khan, 2009), *Channa punctata* (Khan et al., 2013b). In the cleithra of *C. striata* it was very difficult to identify true rings because of the presence of numerous marks. Similarly, cleithra was the least preferred ageing structure in *Schizopyge curvifrons*, *Schizopyge niger* and *Schizothorax esocinus* compared to applying otoliths, scales, opercular bones, and vertebrae (Sabah & Khan, 2014). Fin rays underestimated age in *C. striata* and more so in the older individuals. Many investigators rejected fin rays because of difficulties in sample preparation, identification of the first annulus, and distinction between true and false annuli (Khan & Khan, 2009). Quinn and Ross (1982) questioned the reliability of age estimates from pectoral fin rays in fish older than 7 years because of the lack of annulus formation and the difficulty in interpreting fin ray sections. Thus, suitability of fin rays for ageing may vary among species and, potentially, among populations (Zymonas & McMahon, 2009). In conclusion, otoliths may provide precise age estimates in *C. striata*, especially when the otoliths are sectioned and polished.

ACKNOWLEDGEMENTS

The authors are thankful to the Chairman of the Department of Zoology, Aligarh Muslim University, Aligarh, India, for providing the necessary facilities for the study. The second author gratefully acknowledges the financial support in the form of a Research Assistantship provided by the Council of Science and Technology, Lucknow, Uttar Pradesh, India. The authors are also grateful to the organisers of the XV European Congress of Ichthyology, 2015 for accepting the manuscript for presentation.

REFERENCES

- Beamish, R. J., & Fournier, D. A. (1981). A method for comparing the precision of a set of age determinations. *Canadian Journal of Fisheries and Aquatic Science*, 38, 982–983.
- Beamish, R. J., & McFarlane, G. A. (1983). The forgotten requirements for age validation in fisheries biology. *Transactions of the American Fisheries Society*, 112, 735–743.
- Beamish, R. J., & McFarlane, G. A. (1987). Current trends in age determination methodology. In R. C. Summerfelt & G. E. Hall (Eds.), *Age and growth of fish* (pp. 15–42). Ames: Iowa State University Press.
- Buckmeier, D. L., Irwin, E. R., Betsill, R. K., & Prentice, J. A. (2002). Validity of otoliths and pectoral spines for estimating ages of channel catfish. *North American Journal of Fisheries Management*, 22, 934–942.
- Campana, S. E. (2001). Accuracy, precision and quality control in age determination, including a review of the use and abuse of age validation methods. *Journal of Fish Biology*, 59, 197–242.
- Campana, S. E., Annand, M. C., & McMillan, J. I. (1995). Graphical and statistical methods for determining the consistency of age determinations. *Transactions of the American Fisheries Society*, 124, 131–138.
- Chang, W. Y. B. (1982). A statistical method for evaluating the reproducibility of age determination. *Canadian Journal of Fisheries and Aquatic Science*, 39, 1208–1210.
- DeVries, D. R., & Frie, R. V. (1996). Determination of age and growth. In: B. R. Murphy & D. W. Willis (Eds.), *Fisheries techniques* (pp. 483–515). Bethesda, MD: American Fisheries Society.
- Gomez, K. A., & Gomez, A. A. (1984). *Statistical procedures for agricultural research*. Singapore: John Wiley & Sons.
- Haniffa, M. A., Marimuthu, K., Nagarajan, M., Jesu Arokiaraj, A., & Kumar, D. (2004). Breeding behavior and parental care of the induced bred spotted murrel *Channa punctatus* under captivity. *Current Science*, 86, 1375–1376.
- Hill, K. T., Calliet, G. M., & Radtke, R. L. (1989). A comparative-analysis of growth zones in 4 calcified structures of Pacific blue marlin, *Makaira nigricans*. *Fishery Bulletin*, 87, 829–843.
- Hoxmeier, R. J. H., Aday, D. D., & Wahl, D. H. (2001). Factors influencing precision of age estimation from scales and otoliths of bluegills in Illinois reservoirs. *North American Journal of Fisheries Management*, 21, 374–380.
- Jutagate, T., Phomikong, P., Avakul, P., & Saowakoon, S. (2013). Age and growth determinations of chevron snakehead *Channa striata* by otolith reading. Proc. 51st Kasetsart University Annual Conference. 011.
- Kalish, J. M., Beamish, R. J., Brothers, E. B., Casselman, J. M., Francis, R. I. C. C., Mosegaard, H., Panfili, J., Prince, E. D., Thresher, R. E., Wilson, C. A., & Wright, P. J. (1995). Glossary for otolith studies. In D. H. Secor, J. M. Dean & S. E. Campana (Eds.), *Recent developments in fish otolith research* (pp. 723–729). Columbia: University of South Carolina Press.
- Kamal, M. Y. (1969). Studies on the age and growth of *Cirrhina mrigala* (Hamilton) from the river Yamuna at Allahabad. *Proceedings of National Academy of Sciences India*, 35, 72–92.

- Kamilov, B. G. (1984). Morphology of growth structures in silver carp *Hypophthalmichthys molitrix*, in relation to estimation of age and growth rate. *Journal of Applied Ichthyology*, 6, 1003–1013.
- Khan, S. (2014). *Studies on some aspects of biology of selected freshwater teleost from river Ganga*. Ph.D. Thesis, Department of Zoology, Aligarh Muslim University, Aligarh, India.
- Khan, M. A., & Khan, S. (2009). Comparison of age estimates from scale, opercular bone, otolith, vertebrae and dorsal fin ray in *Labeo rohita* (Hamilton), *Catla catla* (Hamilton) and *Channa marulius* (Hamilton). *Fisheries Research*, 100, 255–259.
- Khan, S., Khan, M. A., & Miyan, K. (2011). Comparison of age estimates from otoliths, vertebrae, and pectoral spines in African sharp-tooth catfish, *Clarias gariepinus* (Burchell). *Estonian Journal of Ecology*, 60, 183–193.
- Khan, S., Khan, M. A., & Miyan, K. (2013a). Evaluation of ageing precision from different structures of three threatened freshwater fish species, *Clarias batrachus*, *Heteropneustes fossilis* and *Wallago attu*. *Folia Zoologica*, 62, 103–109.
- Khan, S., Khan, M. A., & Miyan, K. (2013b). Precision of age determination from otoliths, opercular bones, scales and vertebrae in the threatened freshwater snakehead, *Channa punctata* (Bloch, 1793). *Journal of Applied Ichthyology*, 29, 757–761.
- Khan, S., Khan, M. A., & Miyan, K. (2015). Precision of age estimates from different ageing structures of *Labeo bata* (Hamilton) collected from river Ganga, India. *Indian Journal of Fisheries*, 62, 110–114.
- Khan, M. A., Nazir, A., & Khan, S. (2016). Assessment of growth zones on whole and thin-sectioned otoliths in *Sperata aor* (Bagridae) inhabiting the River Ganga, India. *Journal of Ichthyology*, 56, 242–246.
- Lozano, I. E. S., Vegh, L., Doma' nico, A. A., & Ros, A. E. (2014). Comparison of scale and otolith age readings for trahira, *Hoplias malabaricus* (Bloch, 1794), from Paraná River, Argentina. *Journal of Applied Ichthyology*, 30, 130–134.
- Ma, B., Xie, C., Huo, B., Yang, X., & Li, P. (2011). Age validation, and comparison of otolith, vertebrae and opercular bone for estimating age of *Schizothorax o'connori* in the Yarlung Tsangpo river, Tibet. *Environmental Biology of Fishes*, 90, 159–169.
- Nash, M. K., & Irwin, E. R. (1999). Use of otoliths versus pectoral spines for ageing adult flathead catfish. In E. R. Irwin, W. A. Hubert, C. F. Rabeni, H. L. Schramm Jr & T. Coon (Eds.), *Proceedings of the international ictalurid symposium* (pp. 309–316). Bethesda, Maryland: American Fisheries Society, Symposium 24.
- Natarajan, A. V., & Jhingran, A. G. (1963). On the biology of *Catla catla* (Ham.) from the river Jamuna. *Proceedings of National Institute of Sciences India*, 29, 326–355.
- Perry, R. C., & Casselman, J. M. (2012). Comparisons of precision and bias with two age interpretation techniques for opercular bones of longnose sucker, a long-lived northern fish. *North American Journal of Fisheries Management*, 32, 790–795.
- Phelps, Q. E., Edwards, K. R., & Willis, D. W. (2007). Precision of five structures for estimating age of common carp. *North American Journal of Fisheries Management*, 27, 103–105.
- Quinn, S. P., & Ross, M. R. (1982). Annulus formation by white suckers and the reliability of pectoral fin rays for ageing them. *North American Journal of Fisheries Management*, 2, 204–208.
- Sabah, & Khan, M. A. (2014). Precise age estimation and growth of three *Schizothoracinae* fishes from Kashmir valley. *Zoology and Ecology*, 24, 16–25.
- Secor, D. H., Trice, T. M., & Hornick, H. T. (1995). Validation of otolith-based ageing and comparison of otolith and scale-based ageing in mark-recaptured Chesapeake Bay striped bass, *Morone saxatilis*. *Fishery Bulletin*, 93, 186–190.
- Sipe, A. M., Chittenden, M. E. Jr (2001). A comparison of calcified structures for ageing *Paralichthys dentatus*. *Fishery Bulletin*, 99, 628–640.
- Stolarski, J. T., & Hartman, K. J. (2008). An evaluation of the precision of fin ray, otolith, and scale age determinations for brook trout. *North American Journal of Fisheries Management*, 28, 1790–1795.
- Sutherland, S. J. (2006). *Templates for calculating ageing precision*. Retrieved from <http://www.nefsc.noaa.gov/fbi/ageprec/> (accessed 2015-09-16).
- Zymonas, N. D., & McMahon, T. E. (2009). Comparison of pelvic fin rays, scales and otoliths for estimating age and growth of bull trout, *Salvelinus confluentus*. *Fisheries Management and Ecology*, 16, 155–164.

How to cite this article: Afzal Khan M, Khan S, Khan S.

Precision of age estimates in striped snakehead *Channa striata* (Bloch, 1793) from the Ganga River and its tributaries (rivers Gomti and Yamuna). *J Appl Ichthyol*. 2017;33:230–235. <https://doi.org/10.1111/jai.13300>