

# One-Way Indicator Variable Regression Handout

## 1 Initialization

```
> library(NCStats)
```

## 2 Salmon Sperm Example

### 2.1 Data Preparation

You must change the directory to where the following file is located. I also removed three outliers as discussed in the simple linear regression handout.

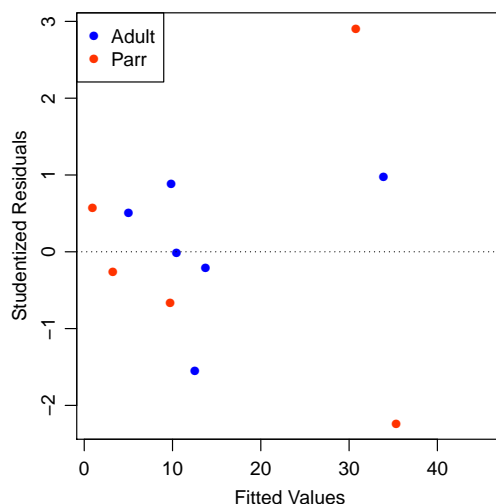
```
> ss <- read.table("SalmonSperm.txt",head=TRUE)
> ss1 <- ss[-c(1,10,11),]
> str(ss1)

'data.frame':      11 obs. of  3 variables:
 $ step.len : num  2.94 3 3.02 3.17 3.18 3.2 3.27 3.31 3.72 3.84 ...
 $ fert.succ: num   3 2.2 7 7 13.5 10.4 6.7 12.8 37.8 30 ...
 $ mat      : Factor w/ 2 levels "Adult","Parr": 2 2 1 2 1 1 1 1 2 2 ...

> xlbl <- "Sperm Tail End Piece Length (um)"
> ylbl <- "Fertilization Success"
```

### 2.2 Fitting the Linear Model

```
> lm1 <- lm(fert.succ~step.len*mat,data=ss1)
> residPlot(lm1,legend="topleft")
```



```
> adTest(lm1$residuals)
```

Anderson-Darling normality test

data: lm1\$residuals

A = 0.1784, p-value = 0.8932

## 2.3 Model Exploration and Summarization

```
> summary(lm1)
```

Call:

```
lm(formula = fert.succ ~ step.len * mat, data = ss1)
```

Residuals:

Min	1Q	Median	3Q	Max
-5.8463	-1.8829	-0.0417	2.0147	7.0466

Coefficients:

	Estimate	Std. Error	t value	Pr(> t )
(Intercept)	-85.769	20.266	-4.232	0.00388
step.len	30.066	6.066	4.956	0.00164
matParr	-25.661	27.273	-0.941	0.37809
step.len:matParr	8.155	8.148	1.001	0.35022

Residual standard error: 4.547 on 7 degrees of freedom

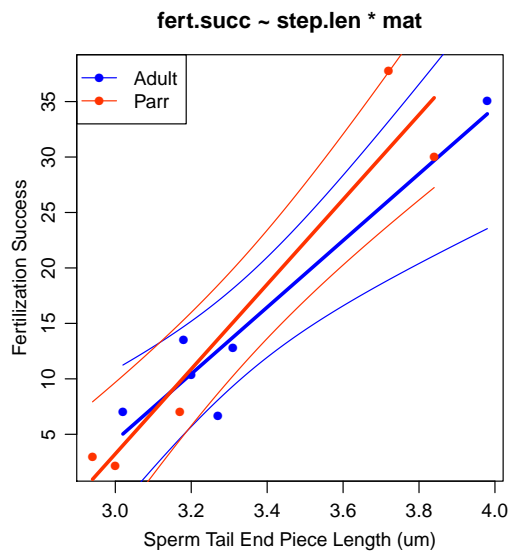
Multiple R-squared: 0.9139, Adjusted R-squared: 0.877

F-statistic: 24.78 on 3 and 7 DF, p-value: 0.0004207

```
> confint(lm1)
```

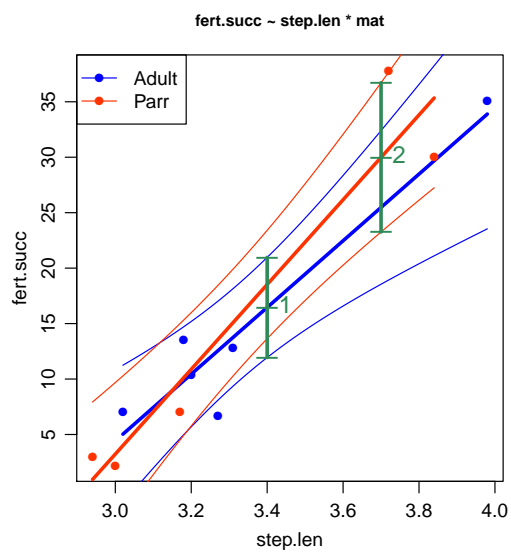
	2.5 %	97.5 %
(Intercept)	-133.68997	-37.84811
step.len	15.72132	44.41036
matParr	-90.15097	38.82930
step.len:matParr	-11.11233	27.42328

```
> fitPlot(lm1, interval="c", xlab=xlbl, ylab=ylbl, legend="topleft")
```



```
> nd <- data.frame(step.len=c(3.4,3.7),mat=c("Adult","Parr"))
> predictionPlot(lm1,nd,interval="c",legend="topleft")
```

```
obs step.len mat fit lwr upr
1 1 3.4 Adult 16.45483 11.94067 20.96899
2 2 3.7 Parr 29.98900 23.25902 36.71899
```



## 2.4 ANOVA Demonstration

```
> anova(lm1)
```

Analysis of Variance Table

Response: fert.succ

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
step.len	1	1510.23	1510.23	73.0316	5.966e-05
mat	1	6.11	6.11	0.2953	0.6037
step.len:mat	1	20.72	20.72	1.0017	0.3502
Residuals	7	144.75	20.68		

```
> lm2 <- lm(fert.succ~step.len+mat,data=ss1)
```

```
> anova(lm2)
```

Analysis of Variance Table

Response: fert.succ

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
step.len	1	1510.23	1510.23	73.0157	2.709e-05
mat	1	6.11	6.11	0.2952	0.6017
Residuals	8	165.47	20.68		

```
> lm3 <- lm(fert.succ~step.len,data=ss1)
```

```
> anova(lm3)
```

Analysis of Variance Table

Response: fert.succ

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
step.len	1	1510.23	1510.23	79.22	9.35e-06
Residuals	9	171.58	19.06		

```
> anova(lm3,lm2,lm1)
```

Analysis of Variance Table

Model 1: fert.succ ~ step.len

Model 2: fert.succ ~ step.len + mat

Model 3: fert.succ ~ step.len \* mat

	Res.Df	RSS	Df	Sum of Sq	F	Pr(>F)
1	9	171.57				
2	8	165.47	1	6.106	0.2953	0.6037
3	7	144.75	1	20.715	1.0017	0.3502

## 3 Fish Energy Density Example

### 3.1 Data Preparation

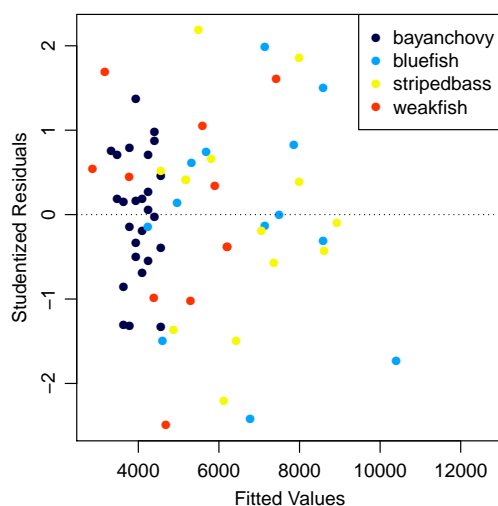
You must change the directory to where the following file is located.

```
> FED <- read.table("FishEnergyDensity.txt",head=TRUE)
> str(FED)

'data.frame':      64 obs. of  3 variables:
 $ species: Factor w/ 4 levels "bayanchovy","bluefish",...: 2 2 2 2 2 2 2 2 2 2 ...
 $ dw      : int   39 34 34 32 31 30 30 29 26 25 ...
 $ ed      : int 10000 9000 8500 8100 7500 7100 7700 6100 5900 5500 ...
```

### 3.2 Assumption Checking and Diagnostics

```
> lm1 <- lm(ed~dw*species,data=FED)
> residPlot(lm1)
```

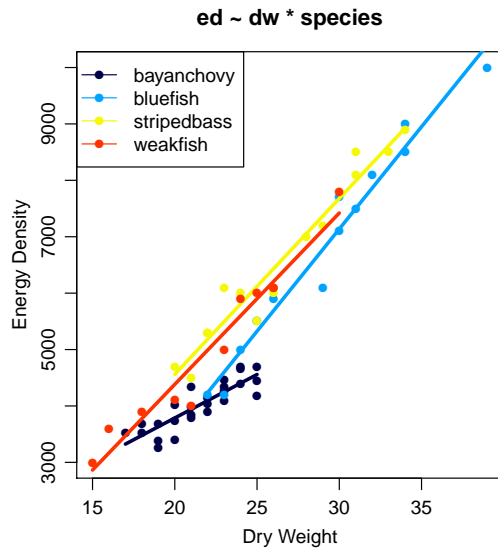


```
> adTest(lm1$residuals)
```

Anderson-Darling normality test

```
data:  lm1$residuals
A = 0.353, p-value = 0.4549
```

```
> fitPlot(lm1,xlab="Dry Weight",ylab="Energy Density",legend="topleft")
```



### 3.3 Model Exploration and Summarization

```
> anova(lm1)
```

Analysis of Variance Table

Response: ed

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
dw	1	170693154	170693154	1858.966	< 2.2e-16
species	3	10592036	3530679	38.452	1.258e-13
dw:species	3	4105617	1368539	14.904	3.002e-07
Residuals	56	5142008	91822		

```
> compSlopes(lm1)
```

Multiple comparison control procedures used: fdr

Multiple Slope Comparisons

	comparison	diff	lwr	upr	raw.p	adj.p
1	bluefish-bayanchovy	208.3095	145.628	270.99117	0.00000	0.00000
2	stripedbass-bayanchovy	157.6206	94.298	220.94336	0.00001	0.00003
3	weakfish-bayanchovy	149.6038	83.209	215.99852	0.00003	0.00006
4	stripedbass-bluefish	-50.6890	-101.086	-0.29227	0.04873	0.05848
5	weakfish-bluefish	-58.7057	-112.912	-4.49955	0.03430	0.05145
6	weakfish-stripedbass	-8.0168	-62.963	46.92958	0.77116	0.77116

Slope Information

	level	slopes	lwr	upr	raw.p	adj.p
1	bayanchovy	154.19	102.23	206.15	0	0
4	weakfish	303.79	262.46	345.13	0	0
3	stripedbass	311.81	275.61	348.01	0	0
2	bluefish	362.50	327.44	397.56	0	0

```
> FED1 <- Subset(FED, species!="bayanchovy")
```

```
> lm2 <- lm(ed~dw*species, data=FED1)
```

```
> anova(lm2)
```

## Analysis of Variance Table

Response: ed

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
dw	1	104962559	104962559	838.6208	< 2.2e-16
species	2	2583571	1291785	10.3210	0.0003476
dw:species	2	556352	278176	2.2225	0.1247887
Residuals	32	4005150	125161		

```
> compIntercepts(lm2)
```

Tukey HSD on adjusted means assuming parallel lines.

	comparison	diff	lwr	upr	p.adj
1	stripedbass-bluefish	631.40	291.11	971.69	0.00018433
2	weakfish-bluefish	506.47	144.53	868.42	0.00441609
3	weakfish-stripedbass	-124.92	-480.89	231.05	0.66939670

Mean adjusted values at a covariate value of 26.11

	bluefish	stripedbass	weakfish
	5828.7	6460.1	6335.1

```
> fitPlot(lm2,xlab="Dry Weight",ylab="Energy Density",legend="topleft")
```

