# Simple Linear Regression Handout

#### 1 Initialization

> library(NCStats)

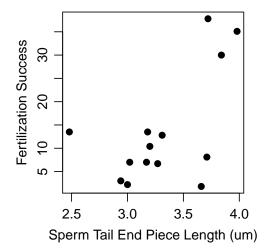
## 2 Salmon Sperm Example

#### 2.1 Data Preparation

You must change the directory to where the following file is located.

The next two commands save the name to be used for axes labels into objects to save typing in later commands.

```
> xlbl <- "Sperm Tail End Piece Length (um)"
> ylbl <- "Fertilization Success"
> plot(fert.succ~step.len,data=ss,xlab=xlbl,ylab=ylbl,pch=19)
```

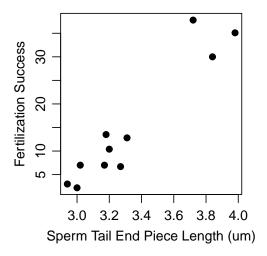


The following command was not evaluated in R because it can be used to interactively click on points on the plot above. When you hit the "STOP" button after clicking on the last point then it will give you a list of the row numbers for the selected points. This command must be issued while the plot above is still active in R. I used this command to identify the individuals that were apparent outliers.

```
> identify(fert.succ~step.len,data=ss)
```

I removed the three outliers from the data set with the commands below. I removed these outliers only to simplify this analysis so that you could remain focused on the concepts of simple linear regression. It is generally NOT good practice to simply remove outliers without considering them further.

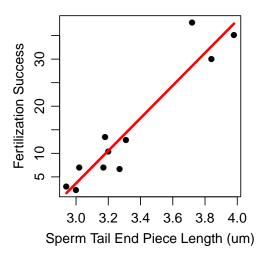
```
> ss1 <- ss[-c(1,10,11),]
> plot(fert.succ~step.len,data=ss1,xlab=xlbl,ylab=ylbl,pch=19)
```



#### 2.2 Lecture Support I – Model Fitting and Simple Predictions

```
> lm1 <- lm(fert.succ~step.len,data=ss1)</pre>
> summary(lm1)
lm(formula = fert.succ ~ step.len, data = ss1)
Residuals:
           1Q Median
                         30
                                Max
                             9.257
-6.269 -2.475 -1.424
                     2.068
Coefficients:
            Estimate Std. Error t value Pr(>|t|)
(Intercept) -100.205
                         13.015
                                 -7.699 3.00e-05
step.len
              34.610
                          3.889
                                  8.901 9.35e-06
Residual standard error: 4.366 on 9 degrees of freedom
Multiple R-squared: 0.898,
                                  Adjusted R-squared: 0.8866
F-statistic: 79.22 on 1 and 9 DF, p-value: 9.35e-06
> fitPlot(lm1,xlab=xlb1,ylab=ylb1)
```

## fert.succ ~ step.len



> predict(lm1,data.frame(step.len=3.5))

1 20.92912

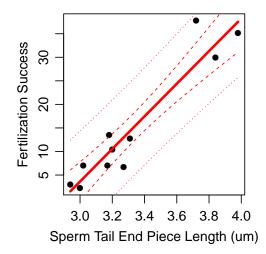
# 2.3 Lecture Support II – Sampling Variability

#### > confint(lm1)

```
2.5 % 97.5 % (Intercept) -129.64815 -70.76202 step.len 25.81336 43.40619
```

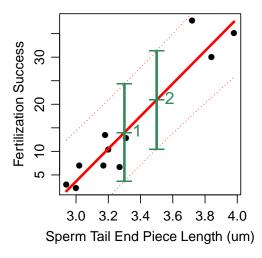
> fitPlot(lm1,xlab=xlb1,ylab=ylb1,interval="both")

## fert.succ ~ step.len



```
> predict(lm1,data.frame(step.len=3.5),interval="c")
       fit
              lwr
                        upr
1 20.92912 17.5967 24.26153
> predict(lm1,data.frame(step.len=3.5),interval="p")
      fit
                lwr
                         upr
1 20.92912 10.50502 31.35321
> predictionPlot(lm1,data.frame(step.len=c(3.3,3.5)),interval="p",xlab=xlbl,ylab=ylbl)
  obs step.len
                   fit
                              lwr
                                       upr
           3.3 14.00716 3.687506 24.32682
   1
1
           3.5 20.92912 10.505016 31.35321
```

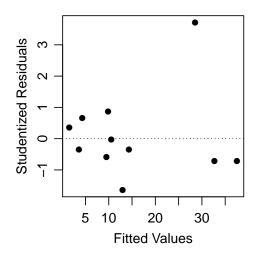
#### fert.succ ~ step.len



#### 2.4 Lecture Support III – Model Comparisons

#### 2.5 Lecture Support IV – Assumption Checking

```
> residPlot(lm1)
```

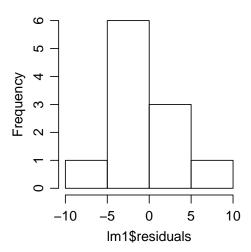


#### > adTest(lm1\$residuals)

Anderson-Darling normality test

data: lm1\$residuals
A = 0.4022, p-value = 0.2962

> hist(lm1\$residuals,main="")



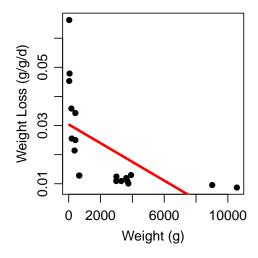
#### > outlierTest(lm1)

No Studentized residuals with Bonferonni p < 0.05 Largest |rstudent|: rstudent unadjusted p-value Bonferonni p 12 3.717896 0.0058892 0.064781

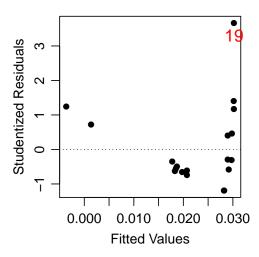
# 3 Petrels Example

The following assumes you have loaded the appropriate packages (as shown in Section 1) and changed the working directory to the folder containing the external data file.

## weight.loss ~ weight



### > residPlot(lm1)



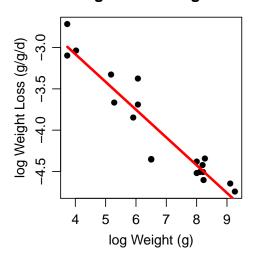
#### > with(petrels,max(weight)/min(weight))

#### [1] 251.8333

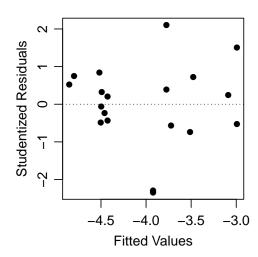
The following command is an interactive command and, thus, the result cannot be shown below. Because the max:min ratio for the weight variable was greater than 10 I set the *lambda.x* slider bar to 0 (for natural log) and then manipulated the *lambda.y* variable to find an appropriate transformation for the response variable. This process led me to use the natural log for *weight.loss* also.

- > transChooser(lm1)
- > petrels\$log.wt <- log(petrels\$weight)</pre>
- > petrels\$log.wtloss <- log(petrels\$weight.loss)</pre>
- > lm2 <- lm(log.wtloss~log.wt,data=petrels)</pre>
- > fitPlot(lm2,xlab="log Weight (g)",ylab="log Weight Loss (g/g/d)")

#### log.wtloss ~ log.wt



#### > residPlot(lm2)



```
> adTest(lm2$residuals)
       Anderson-Darling normality test
data: lm2$residuals
A = 0.3881, p-value = 0.3514
> anova(1m2)
Analysis of Variance Table
Response: log.wtloss
         Df Sum Sq Mean Sq F value Pr(>F)
         1 6.5113 6.5113 140.65 1.204e-09
log.wt
Residuals 17 0.7870 0.0463
> summary(1m2)
Call:
lm(formula = log.wtloss ~ log.wt, data = petrels)
Residuals:
   Min
            1Q Median
                            3Q
                                   Max
-0.4368 -0.1033 0.0447 0.1279 0.4008
Coefficients:
          Estimate Std. Error t value Pr(>|t|)
(Intercept) -1.73403 0.19792 -8.761 1.04e-07
          -0.33632 0.02836 -11.860 1.20e-09
Residual standard error: 0.2152 on 17 degrees of freedom
Multiple R-squared: 0.8922, Adjusted R-squared: 0.8858
F-statistic: 140.6 on 1 and 17 DF, p-value: 1.204e-09
> confint(lm2)
                2.5 %
                        97.5 %
(Intercept) -2.1516113 -1.3164546
          -0.3961507 -0.2764885
> p.log.wtloss <- predict(lm2,data.frame(log.wt=log(5000)),interval="c")
> p.log.wtloss
       fit.
               lwr
                           upr
1 -4.598532 -4.746569 -4.450495
> exp(p.log.wtloss)*exp(anova(lm2)[2,3]/2)
        fit
                   lwr
1 0.01030234 0.008884726 0.01194614
```