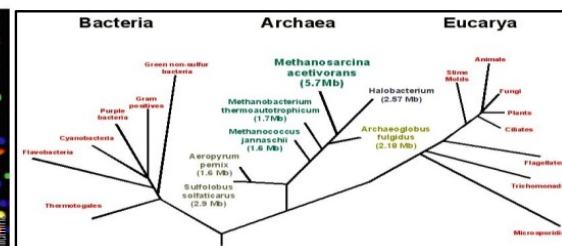
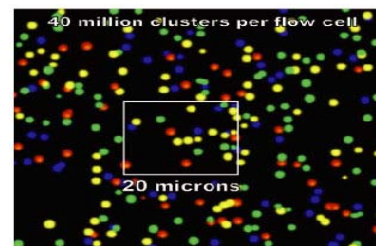




TAACCCTAACCCTAACCCTAACCCTAACCCTA
CCTAACCCTAACCCTAACCCTAACCCTAACC
CCCTAACCCTAACCCTAACCCTAACCCTAAC
AACCCTAACCCTAACCCTAACCCTAACCCTA
ACCCTAACCCTAACCCTAACCCTAACCCTAAC
CTACCCTAACCCTAACCCTAACCCTAACCCTA
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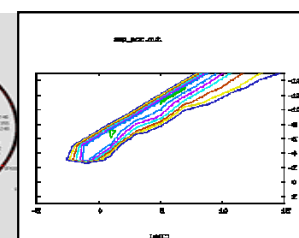
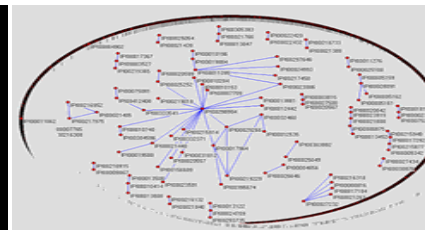
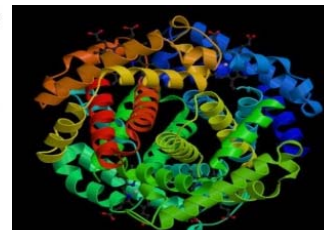
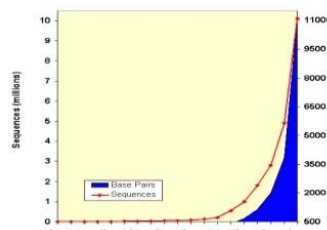
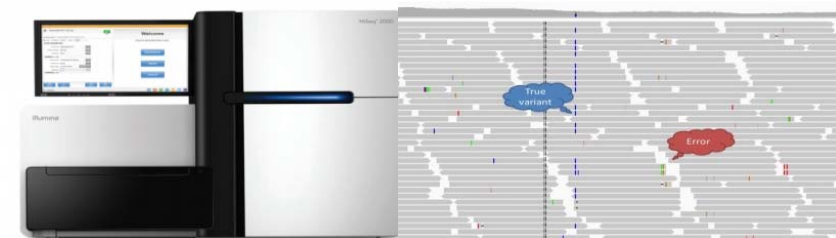


Next Generation Sequencing (NGS): Reads Mapping

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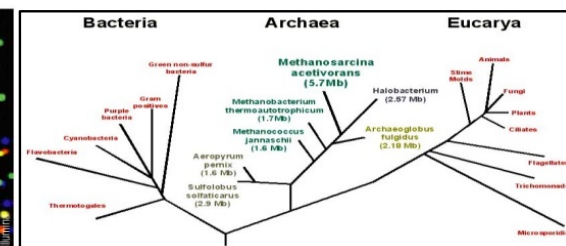
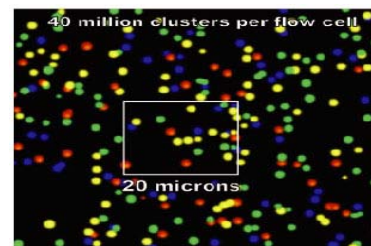
Ge Gao, Ph.D.

Center for Bioinformatics, Peking University





TAACCCTAACCCTAACCCTAACCCTAACCCTA
 CCTAACCCTAACCCTAACCCTAACCCTAACC
 CCTAACCCTAACCCTAACCCTAACCCTAACC
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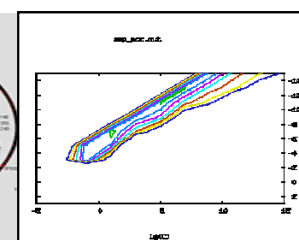
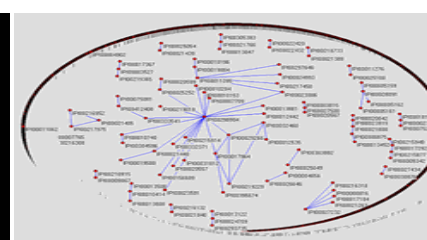
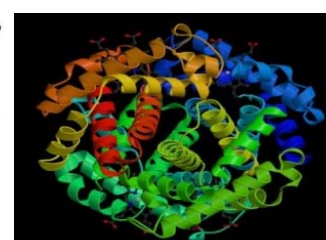
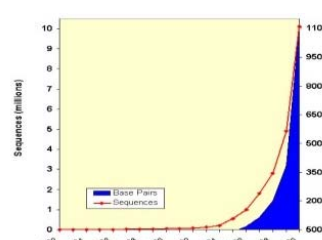
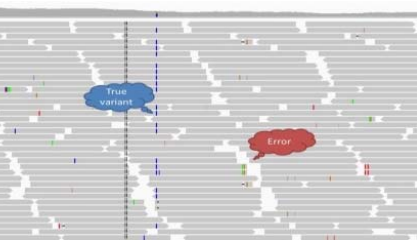


Unit 2: NGS: Reads Mapping

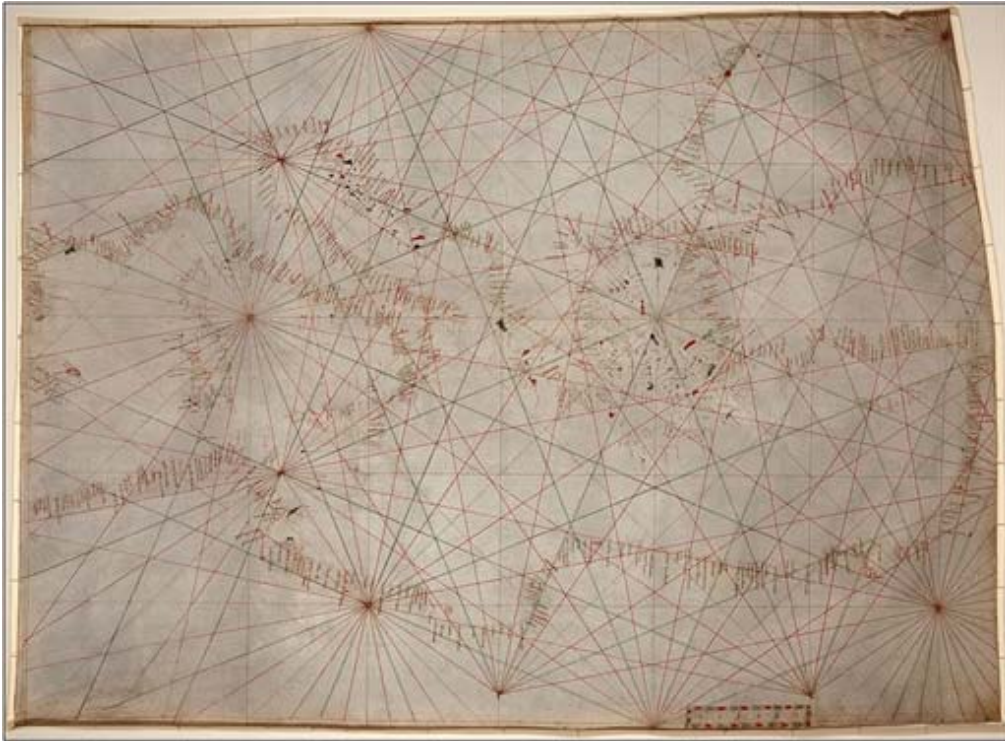
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Center for Bioinformatics, Peking University



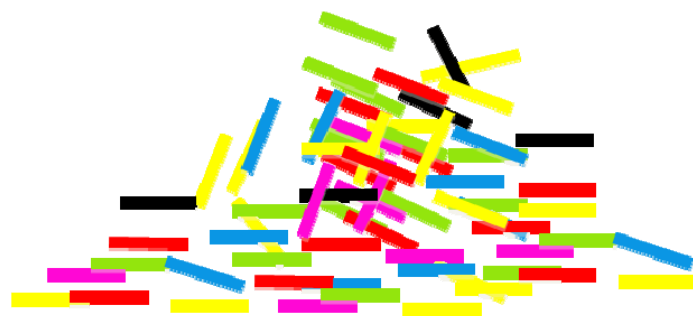
Reads Mapping



Map-Making / Cartography:
Establish relationship between locations
(<http://en.wikipedia.org/wiki/Cartography>)

Technological: Reads is
usual too short to be
used/assembled *de novo*

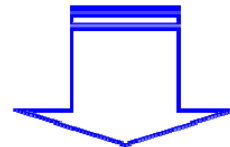
Scientific: Taking full
usage of existing
annotation/knowledge



mapping



Reference Genome



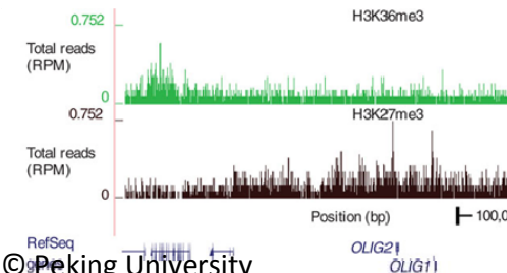
Mapped alignment



Calling
Genetic
Variants



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Measuring
Abundance:
RNA-Seq,
ChIP-Seq, etc.

Mapping: Input Data

- Reference Genome

- Nucleotide

- Length: Hundreds of Mb *per* chromosome

- ~3 Gb in total (for human genome)



Reference Genome

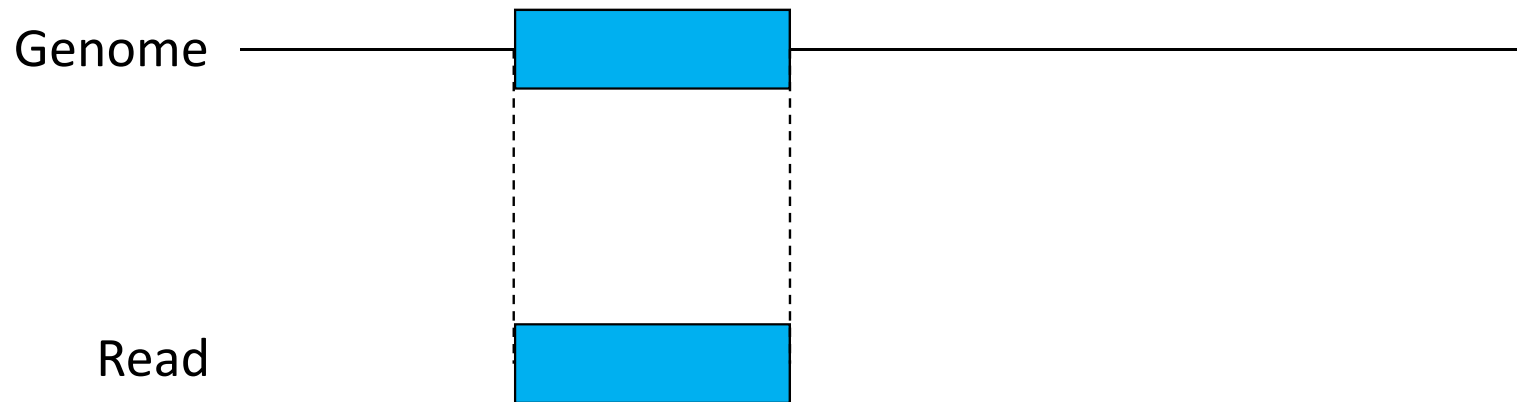
- Reads

- Nucleotide, with various qualities (relatively **high error rate**: $1e-2 \sim 1e-5$)

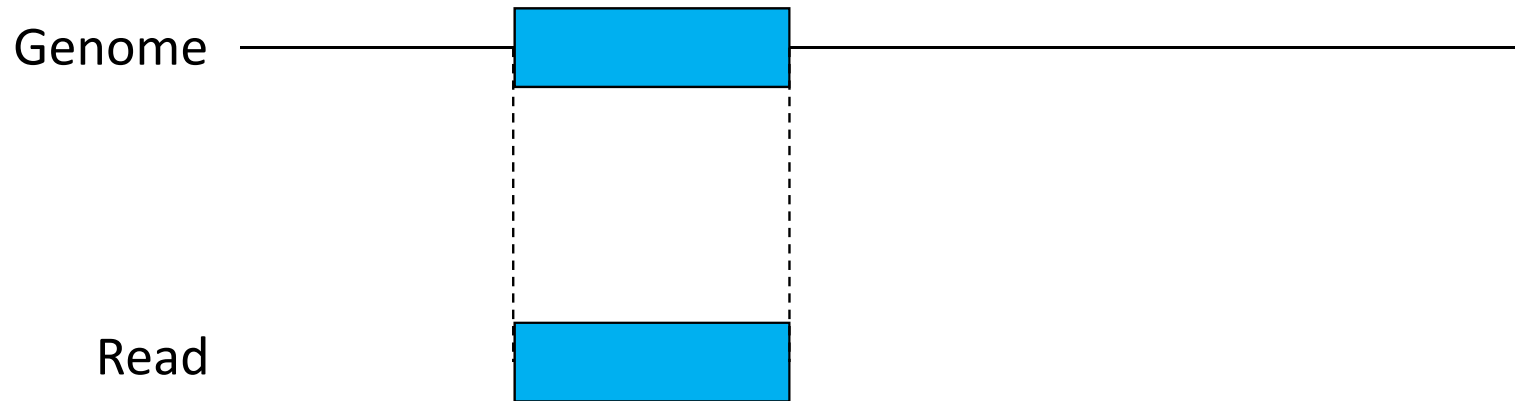
- Length: 36~80 bp *per* read

- Hundreds of Gbs *per* run

“Embedded” Alignment

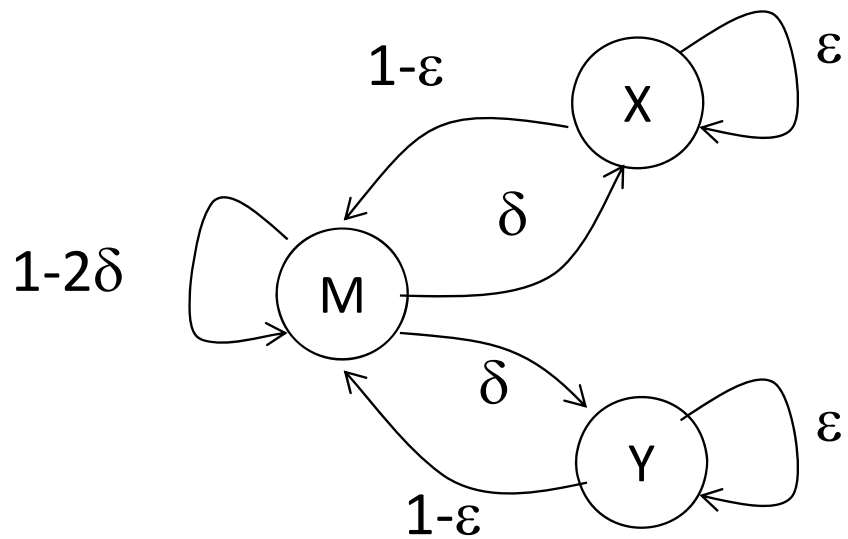


One sequence is “*embedded*” in the other sequence (NGS Reads, PCR primer, *etc.*)



What we need here is actually a hybrid “**global-local**” alignment

- ✓ “**Global**” for short sequence (i.e. NGS Read)
- ✓ But “**Local**” for long sequence (i.e. Reference Genome)
- ✓ In particular, the surrounding “overhang” gaps should be not penalized.



M	Match
X	Insert at sequence X (delete at sequence Y)
Y	Insert at sequence Y (delete at sequence X)

δ	Gap open
ε	Gap Extension

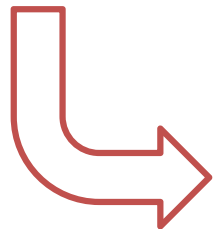
	M	X	Y
M	1-2 δ	δ	δ
X	1- ε	ε	0
Y	1- ε	0	0

Genomic chromosome: $m = \text{hundreds of Mb}$

Sequencing Read: $n = 36 \sim 80 \text{bp}$



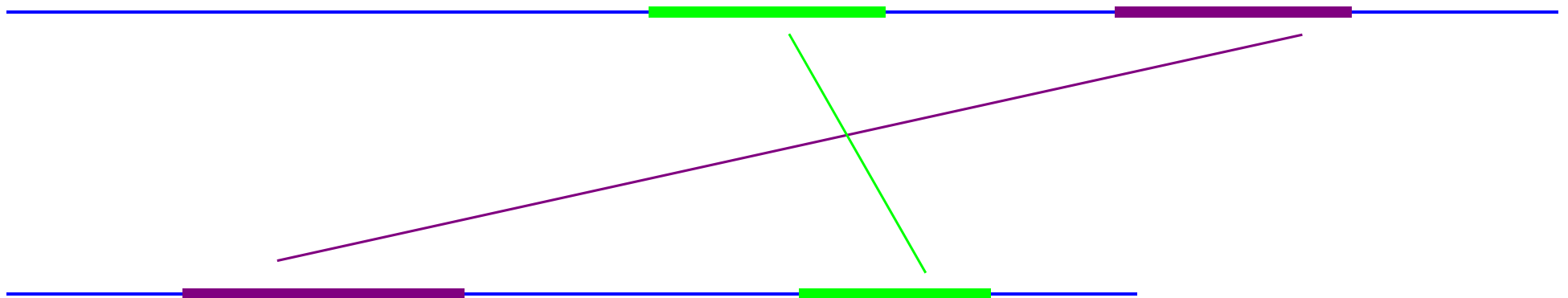
Most of paths will just fail eventually!



In real world, the speed will be a BIG problem!

BLAST Ideas: Seeding-and-extending

1. Find matches (**seed**) between the query and subject
2. Extend seed into High Scoring Segment Pairs (**HSPs**)
 - Run Smith-Waterman algorithm on the specified region only.
3. Assess the reliability of the alignment.



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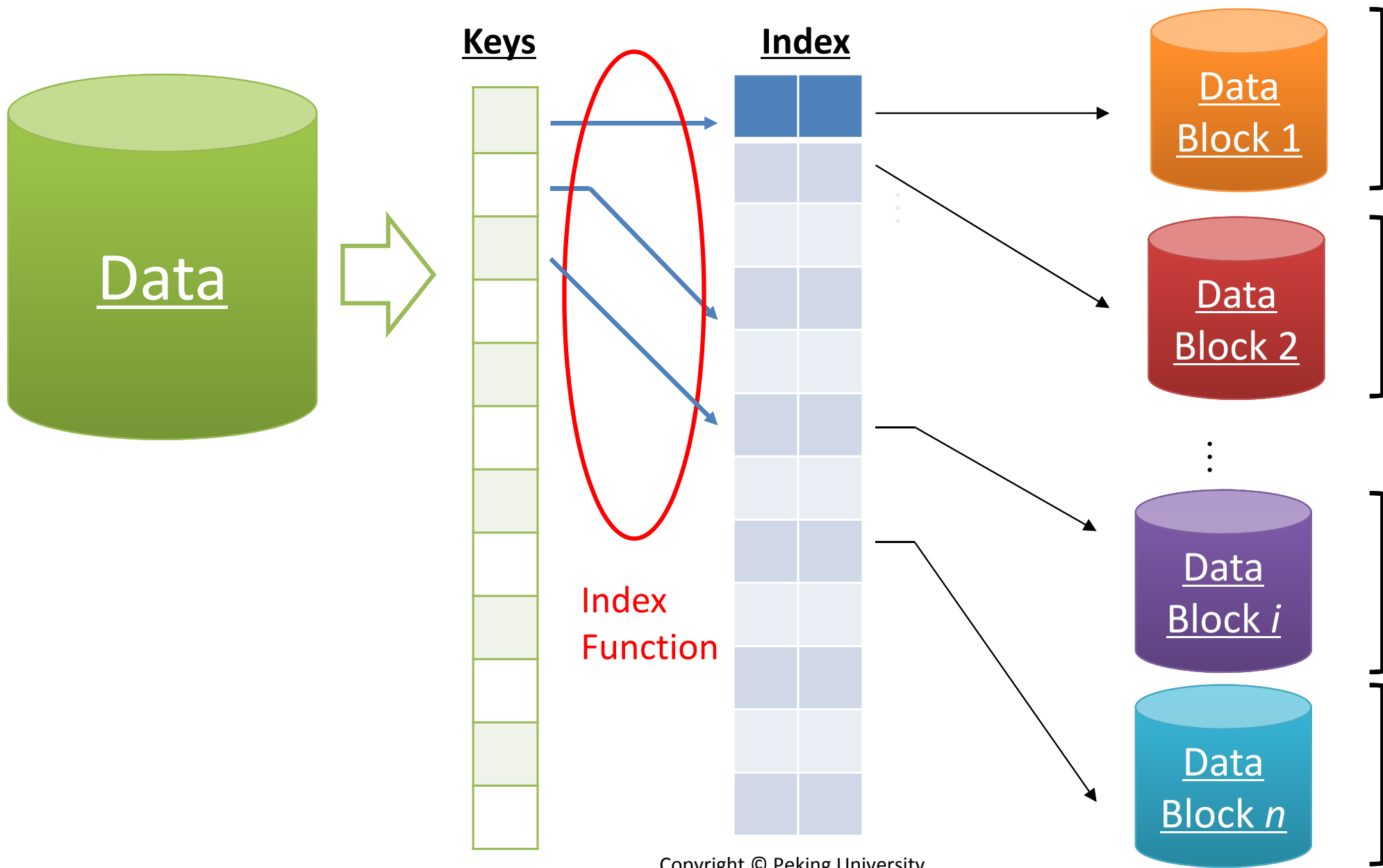
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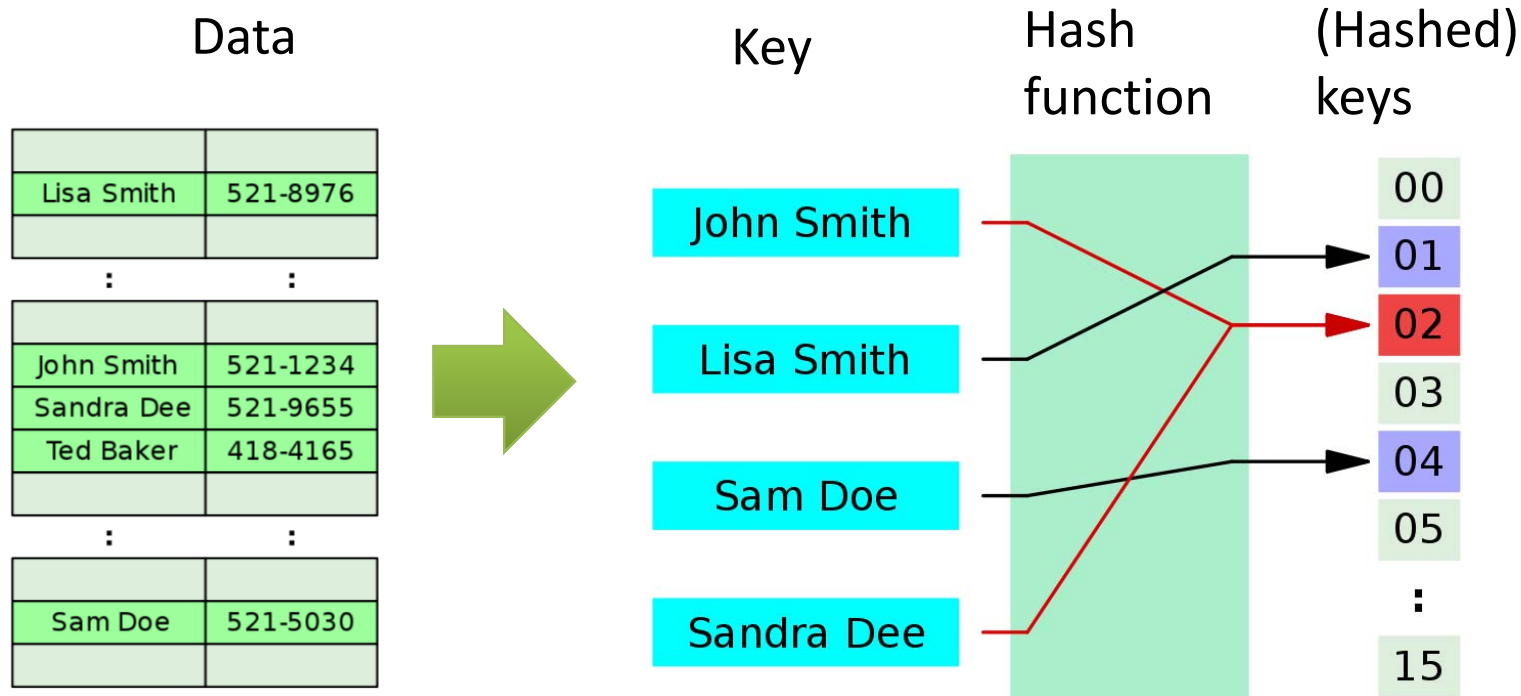
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Hash

Hash function maps (partial) data into (hashed) keys for following-up indexing



HBS: A naive hash function

Let's assume: A = 1, C = 2, G = 4, T = 8, then: $HBS(S) = \sum_i HBS(S_i)$, e.g:

$$HBS(AAAAA) = 1 + 1 + 1 + 1 + 1 = 5$$

$$HBS(GTACG) = 4 + 8 + 1 + 2 + 4 = 19$$

...

123456789012345678901234567890
TAACCCTAACCCTAA...AACCCTAACCC

CCTAA

HBS

$$2+2+8+4+4=20$$

Reference Genome

Index
Table

...

20

...

Address
Table

(CCTAA11)

...

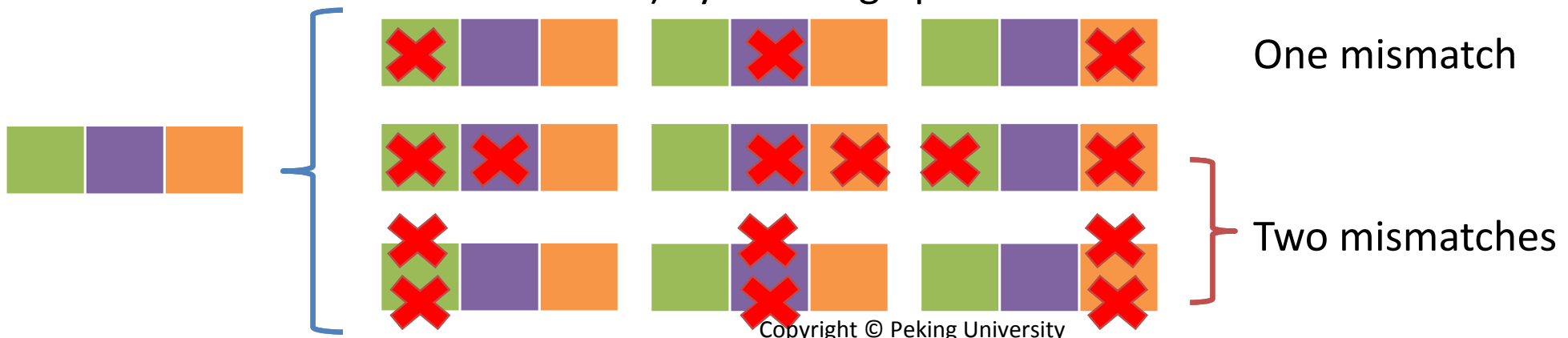
Pigeonhole principle (抽屉原理)

“In mathematics, the pigeonhole principle states that if n items are put into m pigeonholes with $n > m$, then at least one pigeonhole must contain more than one item.”



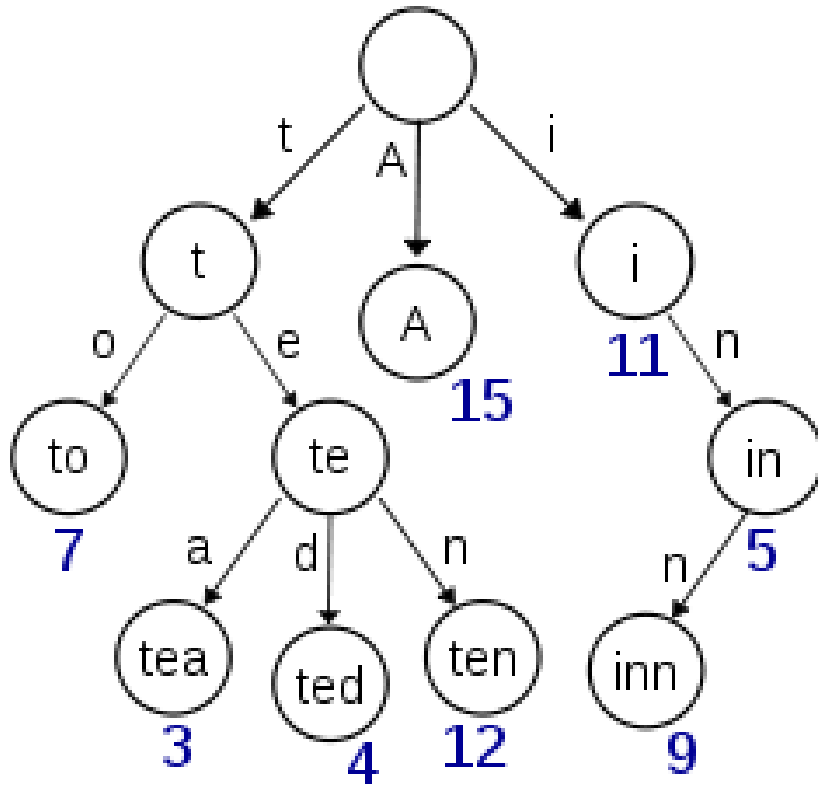
http://en.wikipedia.org/wiki/Pigeonhole_principle

After splitting the read into n (non-overlapped) blocks, there will be **at least $n-m$ perfectly-matched blocks** (i.e. without any mismatch within the block) by allowing up-to- m mismatches.



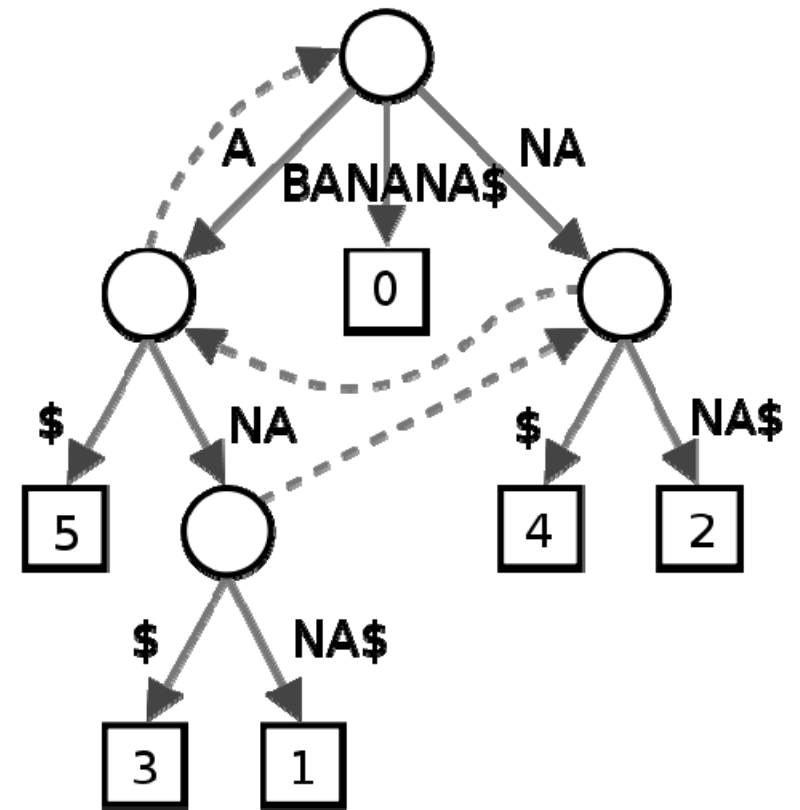
ELAND
MAQ
SOAP1
...

Prefix Tree



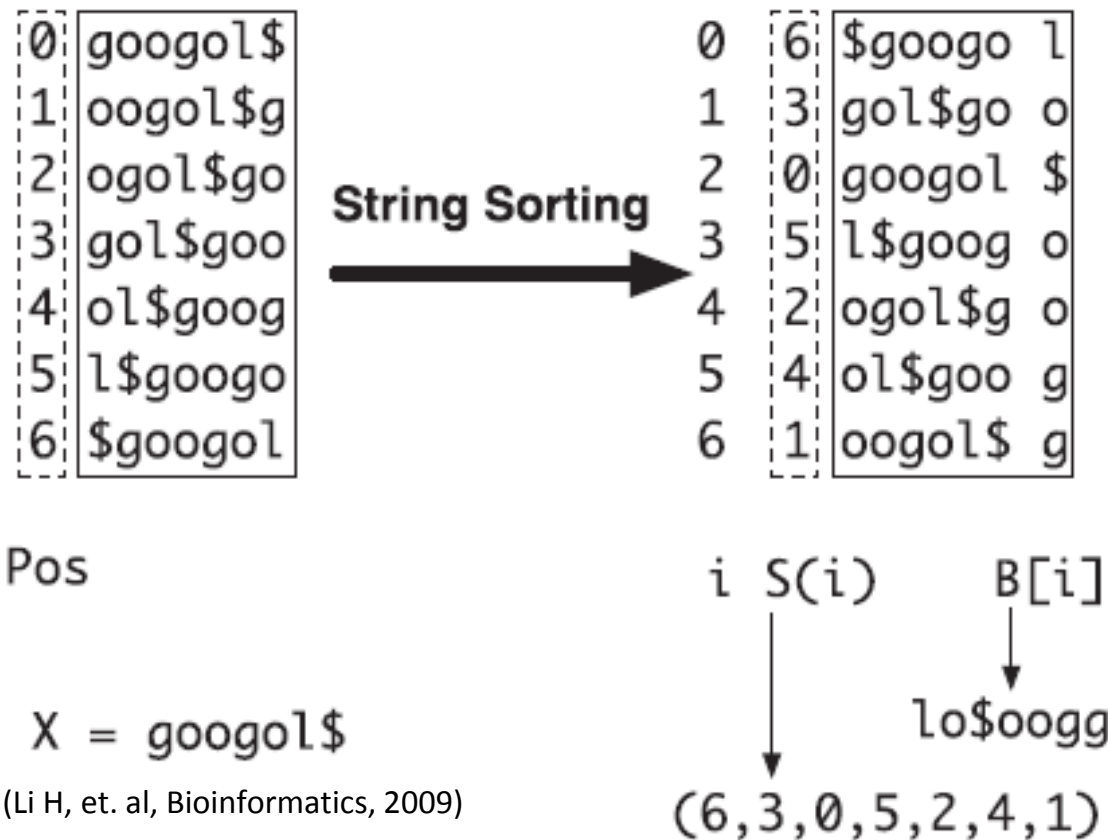
<http://en.wikipedia.org/wiki/Trie>

Suffix Tree



http://en.wikipedia.org/wiki/Suffix_tree

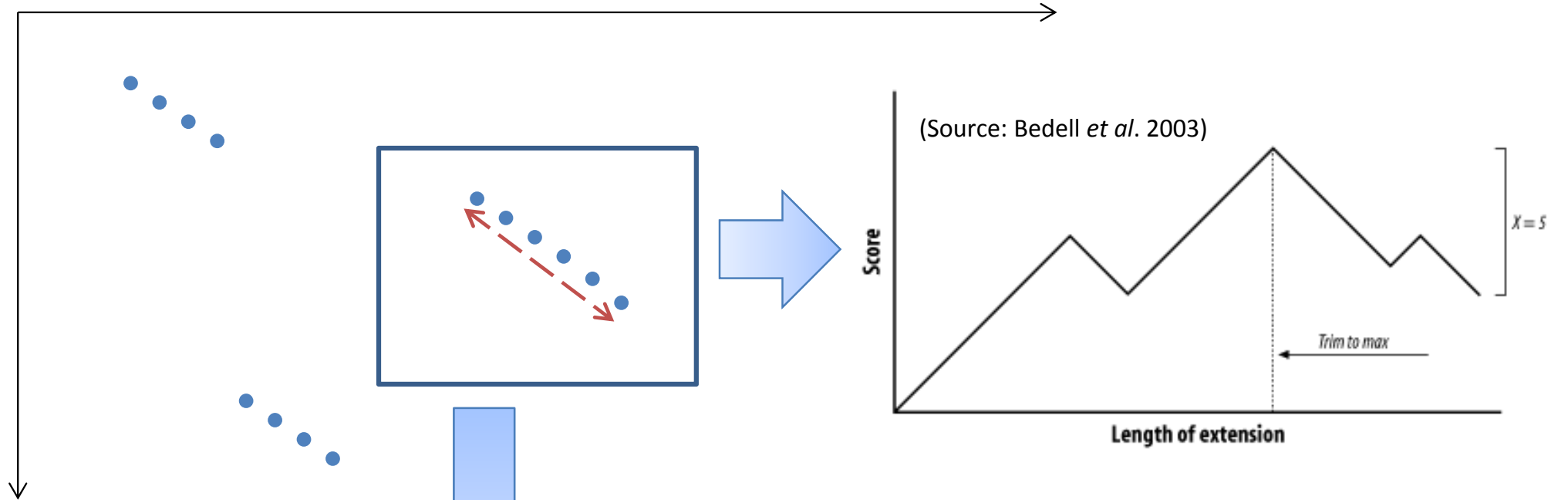
Burrows–Wheeler transform (BWT)



BOWTIE
BWA
SOAP3
...

One of candidate sequence

Query sequence

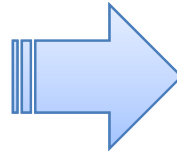


$$F(0, 0) = 0$$

$$F(i, j) = \max \begin{cases} F(i-1, j-1) + s(x_i, y_j) \\ F(i-1, j) + d \\ F(i, j-1) + d \\ 0 \end{cases}$$

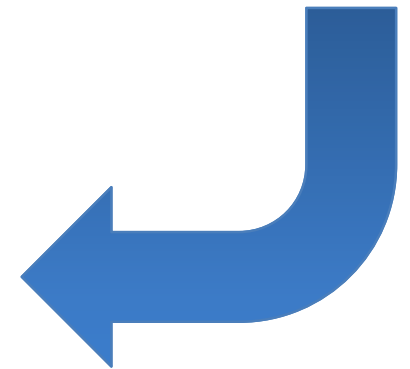
Quality: Given p = the probability of a base calling is *wrong*, its Quality Score can be written as

$$Q = -10 * \log_{10}(p)$$



p	Q
0.1	10
0.01	20
0.001	30
0.0001	40

0	10	20	30	40
!	"#\$%&' () * + , - . / 0 1 2 3 4 5 6 7 8 9 : ; < = > ? @ A B C D E F G H I			
0	10	20	30	40



Mapping Quality

Given reference sequence z (length L), a read sequence x (length l), u is the alignment position of x on z , the probability that z actually coming from the position u is $p(z|x, u)$

(Genome Res. 2008 Nov;18(11):1851.)

$$p(z | x, u) = \prod_{\text{mismatch}} p(z_i) \quad SQ(u) = \log(p(z | x, u)) = \sum_{\text{mismatch}} p(z_i) = \sum_{\text{mismatch}} Q(z_i)$$

Read: ACGT (Quality: 30 30 25 20)

Ref: ACGTACGGA

ACGT	0 +	0 +	0 +	0	SQ (0)
ACGT	30 +	30 +	25 +	20	SQ (1)
ACGT	30 +	30 +	25 +	20	SQ (2)
ACGT	30 +	30 +	25 +	20	SQ (3)
ACGT	0 +	0 +	0 +	20	SQ (4)
ACGT	30 +	30 +	0 +	20	SQ (5)

Mapping Quality

If we assume that a **uniform NULL model**, i.e. the read can randomly come from all possible positions with equal probability, then the error of mapping to a specified position u could be written as

$$E(u) = \frac{SQ(u)}{\sum_i SQ(i)}$$

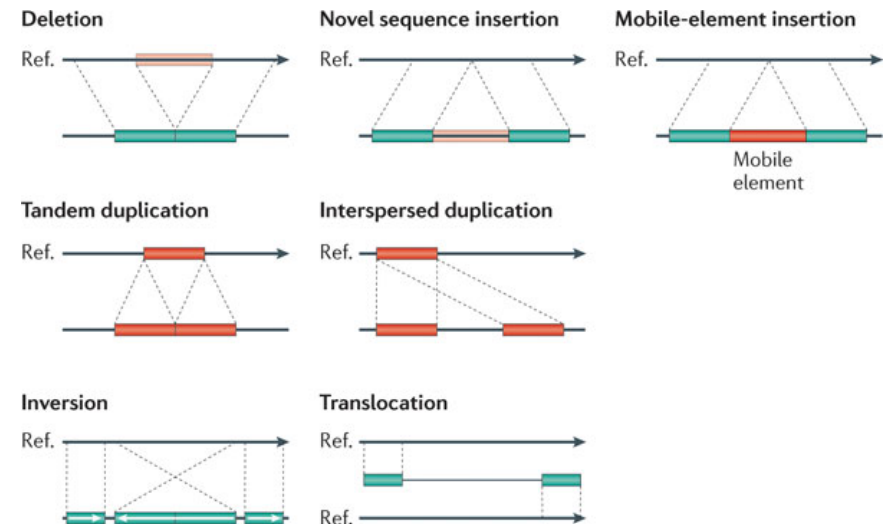
(Genome Res. 2008 Nov;18(11):1851.)

Read: ACGT (Quality: 30 30 25 20)

Ref:	ACGTACGGA	<u>SQ(u)</u>	<u>E(u)</u>
	ACGT	0 + 0 + 0 + 0	0 / 415
	ACGT	30 + 30 + 25 + 20	105 / 415
	ACGT	30 + 30 + 25 + 20	105 / 415
	ACGT	30 + 30 + 25 + 20	105 / 415
	ACGT	0 + 0 + 0 + 20	20 / 415
	ACGT	30 + 30 + 0 + 20	80 / 415

Genetic Variants

- SNV: Single Nucleotide Variant
 - Substitution (SNP)
 - Indel: insertion/deletion
- Structural Variation (SV)
 - Large-scale insertion/deletion
 - Inversion
 - Translocation
 - Copy Number Variation (CNV)



SNP Calling is NOT Genotyping

- “**SNP calling** aims to determine in which **positions** there are polymorphisms or in which **positions** at least one of the bases differs from a reference sequence”
- “**Genotype calling** is the process of determining the **genotype** for each individual and **is typically only done for positions in which a SNP or a 'variant' has already been called.**”

(Source: *Nature Reviews Genetics* 12, 443-451)

Counting: an intuitive (and naïve) approach

...CCATAG TAT CGCCC CGGAATT CGGTATAC...
...CCAT CTATAT CG TCGGAATT CGGTATAC
...CCAT GGCTATAT CGC CTATCGGAAA GCGGTATA
...CCA AGGCTATAT CGC CCTATCGGA TTGCGGTA C...
...CCA AGGCTATAT GCCCTATCG TTTGCGGT C...
...CC AGGCTATAT GCCCTATCG AAATTGTC ATAC...
...CC TAGGCTATA CGCCCTA AAATTGTC GTATAC...
...CCATAGGCTATATCGGCCCTATCGGCAATTGCGGTATAC...

Genetic variants

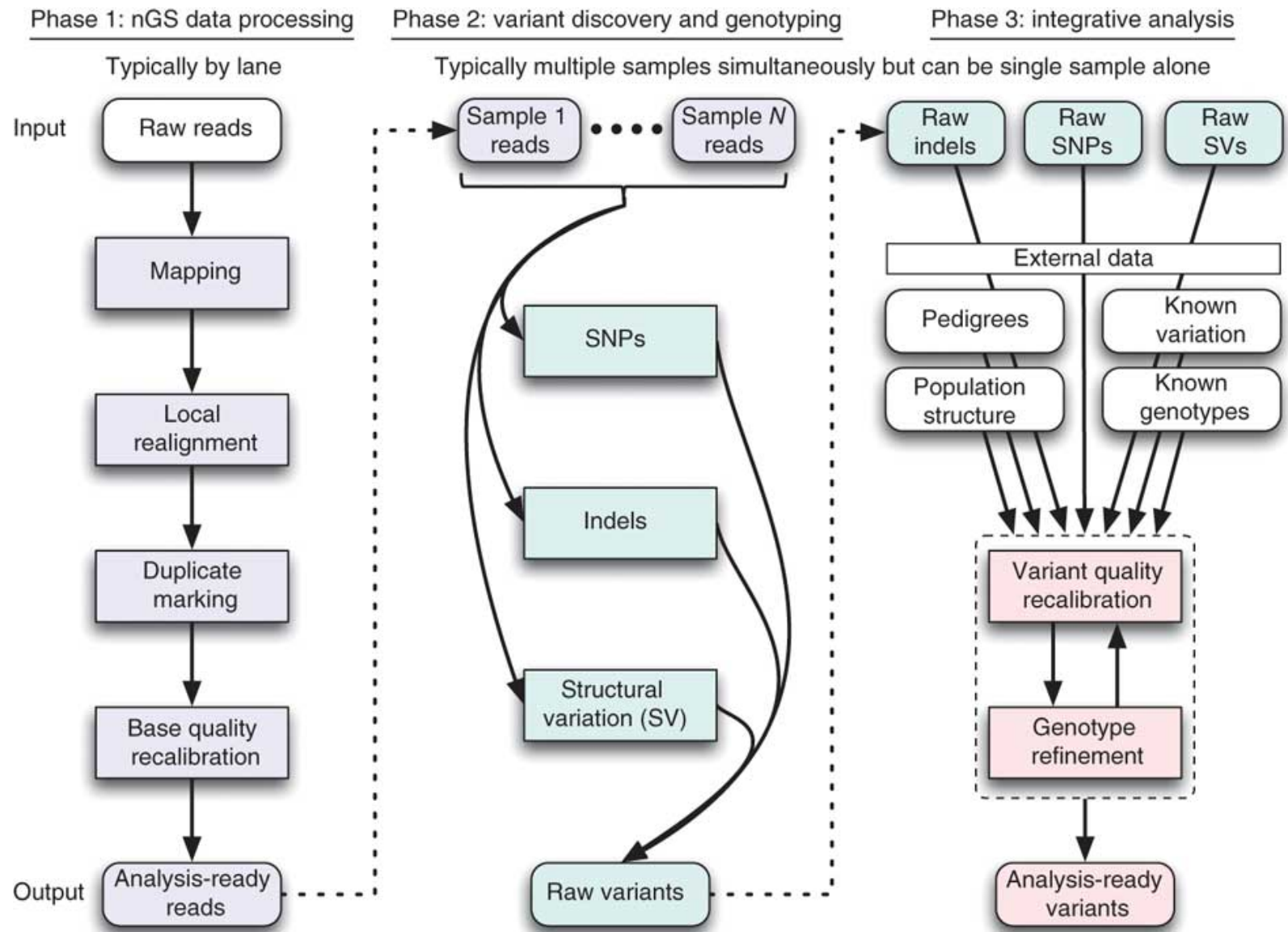
- Counting **high-confident** , **non-reference** allele (i.e. Quality >= 20)
 - Freq <20% or > 80%: **homozygous** genotype
 - Otherwise: **heterozygous**
- Works well for “**deeply sequenced regions**” (DSR), i.e. depth > 25x
 - But suffer from under-calling of heterozygous genotypes for low-coverage regions
 - And can't give an objective measurement for **reliability**

(Source: *Nature Reviews Genetics* 12, 443-451)
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A Simple Probabilistic Model for Genotyping

1. For a diploid genome, there will be at most two different alleles (A and a) observed at a given site:
 - 3 possible genotypes: $\langle A, A \rangle$, $\langle A, a \rangle$, $\langle a, a \rangle$
 - Number of A: k; Number of a: n-k
2. Then, the probability for each genotypes is
 - $P(D | \langle A, A \rangle) = \prod_{n-k} P(x_i)$
 - Similarly, we can see the $P(D | \langle a, a \rangle) = \prod_k P(x_i)$
 - $P(D | \langle A, a \rangle) = 1 - (P(D | \langle A, A \rangle) + P(D | \langle a, a \rangle))$
3. Bayes Formula can be further employed to calculate posterior probabilities, i.e. $P(\langle A, A \rangle | D)$, $P(\langle a, a \rangle | D)$, and $P(\langle A, a \rangle | D)$ if we can estimate the prior probabilities $P(\langle A, A \rangle)$, $P(\langle a, a \rangle)$ and $P(\langle A, a \rangle)$

Genome Analysis ToolKit (GATK)



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