

# A batch correction method for differential gene network analyses

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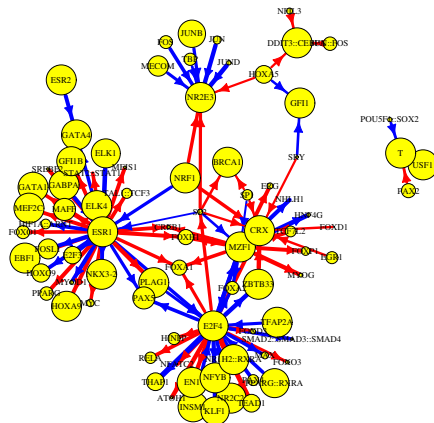
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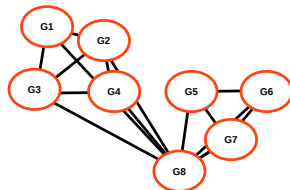
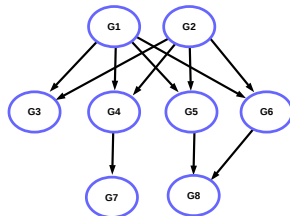
# Inferring gene expression networks



# Types of Network Inference Approaches

Typically, network inference methods fall into two categories:

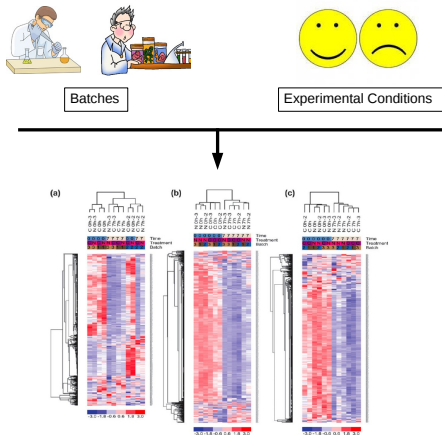
- 1 Gene Regulatory Networks (GRNs)
  - Directed graph
  - Imply a sort of physical interaction
- 2 Gene Coexpression Networks (GCNs)
  - Undirected graph
  - Imply a more general common pathway or process



# Measuring association

- Pearson Correlation
  - Linearity, outliers, etc.
- Spearman Correlation
  - More robust, less sensitive to outliers
- Euclidean Distance
- Mutual Information
  - Non-linear
- Partial Correlation
  - Direct effects

# Batch Effect



Johnson et al. (Biostatistics 2007)

- Laboratory Conditions
- Circadian Rhythm / cell cycle
- Reagents
- Atmospheric Ozone
- Etc. etc.

# Methods for Controlling Batch Effect

- COMBAT - Empirical Bayes approach for location/scale adjustment
- Surrogate Variable Analysis (SVA) - SVD approach for estimating
- Reference based (RATIO-G) methods - Scales sample measurements by the geometric mean of a group of reference measurements
- Distance Weighted Discrimination (DWD) - Based on SVM
- Many more...

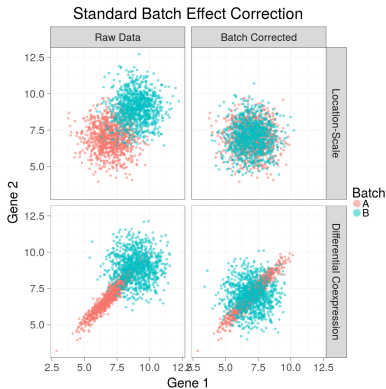
# Limitations of existing batch effect correction methods

- Perfect confounding
- Location/scale assumptions
- Independent effects
- Batches must be known or
- Batches must be estimated (SVA)
- *Differential coexpression*

Batch effect removal methods typically return a corrected gene expression matrix.



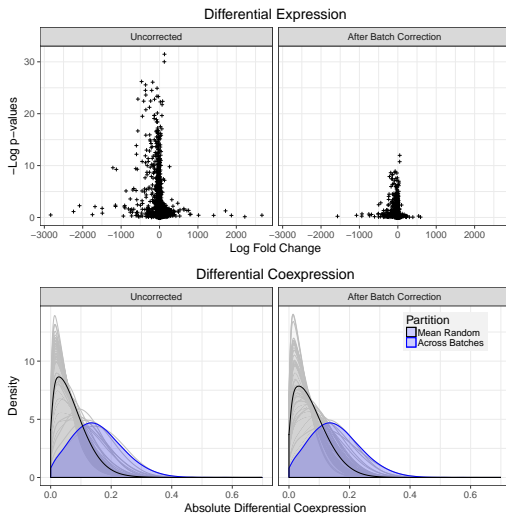
# Limitations to existing batch effect correction methods



- Protocol induced coexpression?
- Differential biological variation

$$f[Gene1|BatchA] = f[Gene1|BatchB]$$

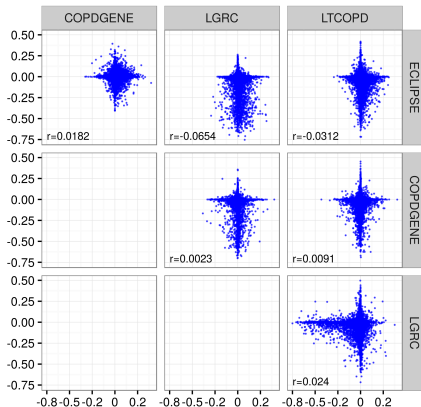
# Limitations to existing batch effect correction methods



ENCODE Project:  
50k genes  
126 samples (63 patients RNA-  
seq'ed at 2 centers)

# Challenges with batch effect on differential coexpression

WGCNA edge weight differences between pairs of studies



- Ultra-high dimensionality
- Differential coexpression
- Modularity

# Estimating the conditional coexpression matrix

Motivating concepts:

- 1.) Provide a regression framework for for the coexpression matrix.
- 2.) Estimate a reduced number of parameters.
- 3.) Exploit modular nature of gene expression patterns.

Our proposal:

Define our parameters as functions of components of variation.

Estimate the eigenvalue contribution of each eigenvector.

# Model

Consider a set of  $N$  samples with  $q$  covariates measuring gene expression across  $p$  genes. Let  $\mathbf{x}_i = (x_{i1}, \dots, x_{iq})$  denote the covariates for sample  $i$  and let  $\mathbf{g}_i = (g_{i1}, \dots, g_{ip})^T$  denote the gene expression values for sample  $i$  for the  $p$  genes.

We can express a model for the gene expression as

$$\mathbf{g}_i = \beta^T \mathbf{x}_i + \epsilon_i \text{ for } i = 1, \dots, N$$

where  $\epsilon_i \sim MVN_p(\mathbf{0}, \Sigma_i)$ . Notably, the covariance of  $\epsilon_i$  differ according to  $i$ .

$$\Sigma_i = \mathbf{Q} \mathbf{D}_i \mathbf{Q}^T$$

where  $\mathbf{D}_i$  is a diagonal matrix with diagonal defined as  $\mathbf{X}_i \Psi_{q \times p}$

# Likelihood Function

$$\mathcal{L}(\mu, \Sigma) = \prod_{i=1}^N \frac{1}{(2\pi)^{\frac{p}{2}} |\Sigma_i|^{\frac{1}{2}}} e^{-\frac{1}{2}(\mathbf{G}_i - \mu)^T \Sigma_i^{-1} (\mathbf{G}_i - \mu)}$$

Where we define  $\Sigma_i$

$$\Sigma_i = \mathbf{Q} \mathbf{D}_i \mathbf{Q}^T$$

Where  $\mathbf{Q}$  is a matrix with columns defined as the eigenvectors of the estimated coexpression matrix,  $\mathbf{G}^* \mathbf{G}^{*T} / N$ .

# Least Squares Estimator

$$\mathbf{0}_q = \sum_{i=1}^N \mathbf{x}_i^T \left[ \mathbf{Q}_h^T \left[ \mathbf{G}_i \mathbf{G}_i^T - \mathbf{Q}_h \mathbf{x}_i \hat{\psi}_h \mathbf{Q}_h^T \right] \mathbf{Q}_h \right] \quad (1)$$

$$\mathbf{0}_q = \sum_{i=1}^N \left[ \mathbf{x}_i^T \mathbf{Q}_h^T \mathbf{G}_i \mathbf{G}_i^T \mathbf{Q}_h - \mathbf{x}_i^T \mathbf{Q}_h^T \mathbf{Q}_h \mathbf{x}_i \hat{\psi}_h \mathbf{Q}_h^T \mathbf{Q}_h \right]$$

$$\mathbf{0}_q = \sum_{i=1}^N \left[ \mathbf{x}_i^T \mathbf{Q}_h^T \mathbf{G}_i \mathbf{G}_i^T \mathbf{Q}_h - \mathbf{x}_i^T \mathbf{x}_i \hat{\psi}_h \right]$$

$$\sum_{i=1}^N \left[ \mathbf{x}_i^T \mathbf{x}_i \right] \hat{\psi}_h = \sum_{i=1}^N \left[ \mathbf{x}_i^T \mathbf{Q}_h^T \mathbf{G}_i \mathbf{G}_i^T \mathbf{Q}_h \right]$$

$$\mathbf{x}^T \mathbf{x} \hat{\psi}_h = \sum_{i=1}^N \left[ \mathbf{x}_i^T \mathbf{Q}_h^T \mathbf{G}_i \mathbf{G}_i^T \mathbf{Q}_h \right]$$

$$\hat{\psi}_h = (\mathbf{x}^T \mathbf{x})^{-1} \sum_{i=1}^N \left[ \mathbf{x}_i^T \mathbf{Q}_h^T \mathbf{G}_i \mathbf{G}_i^T \mathbf{Q}_h \right] \quad (2)$$

$$\hat{\psi} = (\mathbf{x}^T \mathbf{x})^{-1} \sum_{i=1}^N \left[ \mathbf{x}_i^T \mathbf{Q}^T \mathbf{G}_i \mathbf{G}_i^T \mathbf{Q} \right]$$

# The Corrected Coexpression Matrix

With the estimates obtained with our method, it is straightforward to see how fitted values for the coexpression matrix for each sample or experimental condition can be obtained. Given an estimate for  $\Psi$ ,  $\hat{\Psi}$ , we can now estimate the batch-independent coexpression structure as

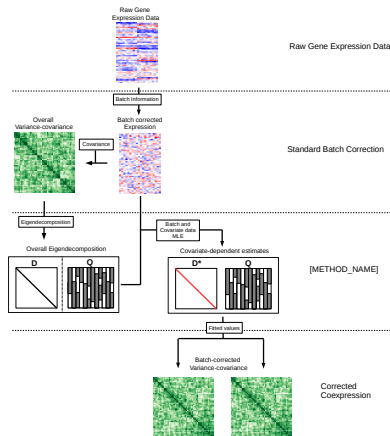
$$\hat{\mathbf{S}} = \mathbf{Q} \text{diag} \left( \bar{\mathbf{X}} \hat{\Psi} \right) \mathbf{Q}^T \text{ or } \hat{\mathbf{S}} = \sum_{i=1}^p \bar{\mathbf{X}} \hat{\Psi}_i \mathbf{Q}_i \mathbf{Q}_i^T$$

The differential coexpression matrix between two conditions, defined in binary as column 2 of  $\mathbf{X}$ , is computed

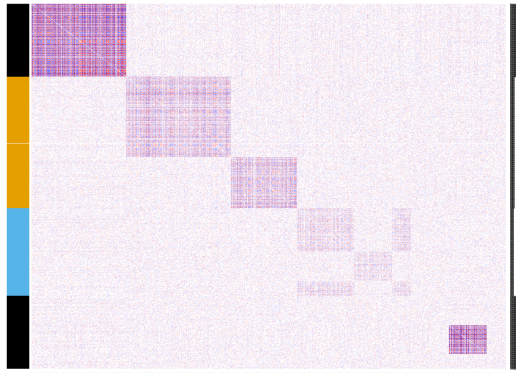
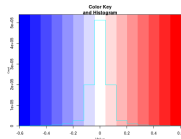
$$\hat{\mathbf{W}} = \mathbf{Q} \text{diag} \left( \hat{\Psi}_{2,\cdot} \right) \mathbf{Q}^T$$



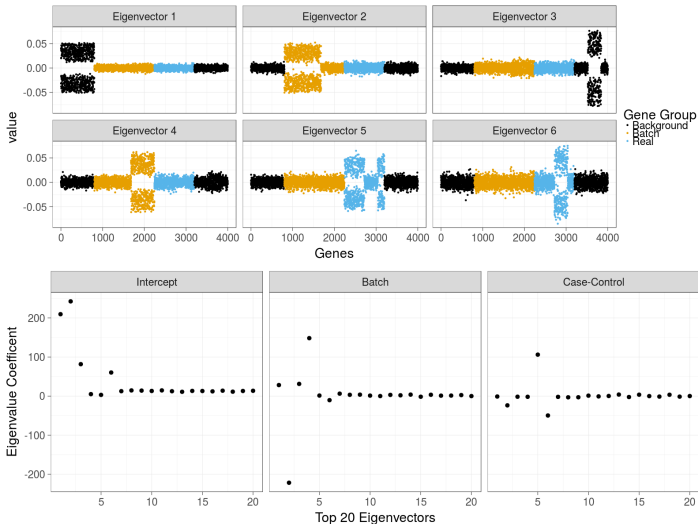
# Example Workflow



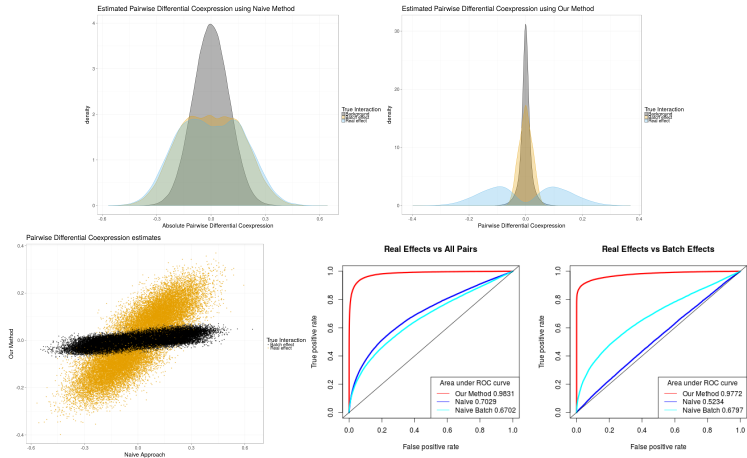
# Simulations



# Simulations



# Simulations



# Immuno-navigator

Soon...

# Acknowledgements

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- Joe Paulson