

Lab Meeting - Data

Diya

Project Overview


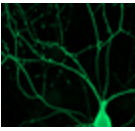

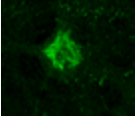

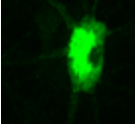
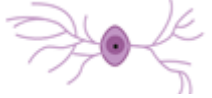
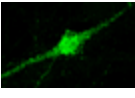

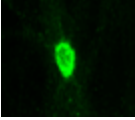
Previous project: Compare the organization of Munc13 and PSD-95 at Ex→Ex synapses vs Ex→PV (parvalbumin-expressing) synapses

Overall goal: Investigate the organization of excitatory synapses onto different inhibitory neuron subtypes (in addition to PV)

1. Develop method(s) to identify inhibitory subtypes
 - a. Identify markers for inhibitory subtypes
 - b. Determine the populations of various subtypes, ensuring they are significant to investigate
 - c. Generate an inhibitory specific cell fill

Inhibitory Neuron Subtype Markers

Cell type	Percent CA1 neurons
Axo-axonic (AAC)	0.04%
Parvalbumin-expressing basket cells (PVBCs)	1.5%
Bistratified Cells (BiCs)	0.7%
Cholecystikinin-expressing basket cells (CCKBCs)	1%
Dendrite targeting CCK expressing Interneurons	0.4%
Oriens lacunosum-moleculare interneurons (O-LMs)	0.5%
Neurogliaform (NGFCs)	1%
Ivy cells (IVCs)	2.5%
Interneuron selective interneurons (ISIs)	2.2%

Inhibitory Neuron Type(s)	Antibody/ Neuropeptide Marker	Probable Morphology	Staining
Axo-axonic and PV-expressing basket cells	Anti-Parvalbumin (PV)		
CCK-expressing basket cells/ CCK interneurons	Anti-Cholecystikinin (CCK)		
Oriens lacunosum-moleculare interneurons	Anti-Somatostatin (SST)		
Neurogliaform and ivy cells	Anti-Neuropeptide Y (NPY)		
Interneuron selective interneurons	Anti-Vasoactive Intestinal Peptide (VIP)		

Results: Larger and Denser Munc13 Clusters in Ex→Inh Synapses, but Variable Munc13 to PSD-95 Ratios Among Inhibitory Neuron Subtypes

Table 1 Cell Type Percentages

Subtype Marker	Average Percentage Per Coverslip
GFP	10.74%
CCK	7.55%
PV	4.95%
SST	2.85%
NPY	3.00%

Table 2 Sample Sizes for Synaptic Analysis

Subtype Marker	Total Number of Cells Analyzed	Total Number of Synapses Analyzed
GFP	30	11519
CCK	14	1112
PV	16	2769
SST	47	6251
NPY	26	4015

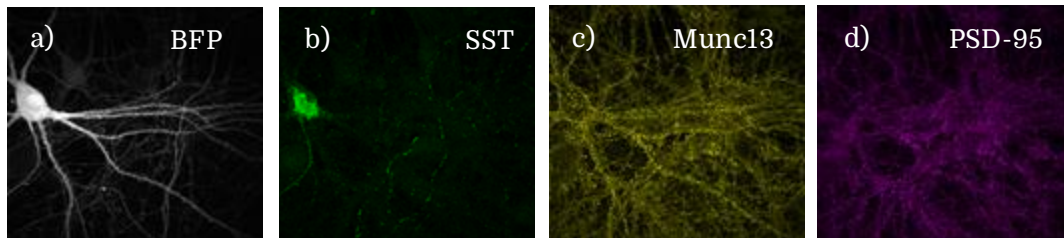
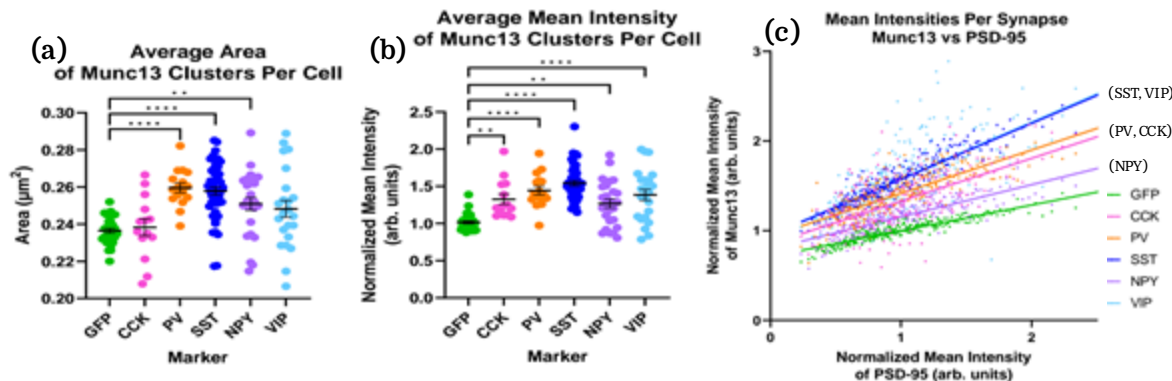


Figure 3. a) The inhibitory specific BFP cell fill visualizes the neuron's dendrites. b) The anti-SST antibody confirms the presence of the SST neuropeptide, identifying the inhibitory subtype (fig 2). c) Synaptic staining shows Munc13 clusters. d) Synaptic staining shows PSD-95 clusters. Taken at 60x.

Figure 4. Average area (a) and intensity (b) of Munc13 clusters collected from confocal images is compared by synapse subtypes shown in figure 2. Asterisk (*) identifies a significant increase compared to Ex→Ex (GFP). c) Munc13 intensity graphed against PSD-95 intensity indicates SST and VIP, PV and CCK, and NPY have statistically different protein ratios.



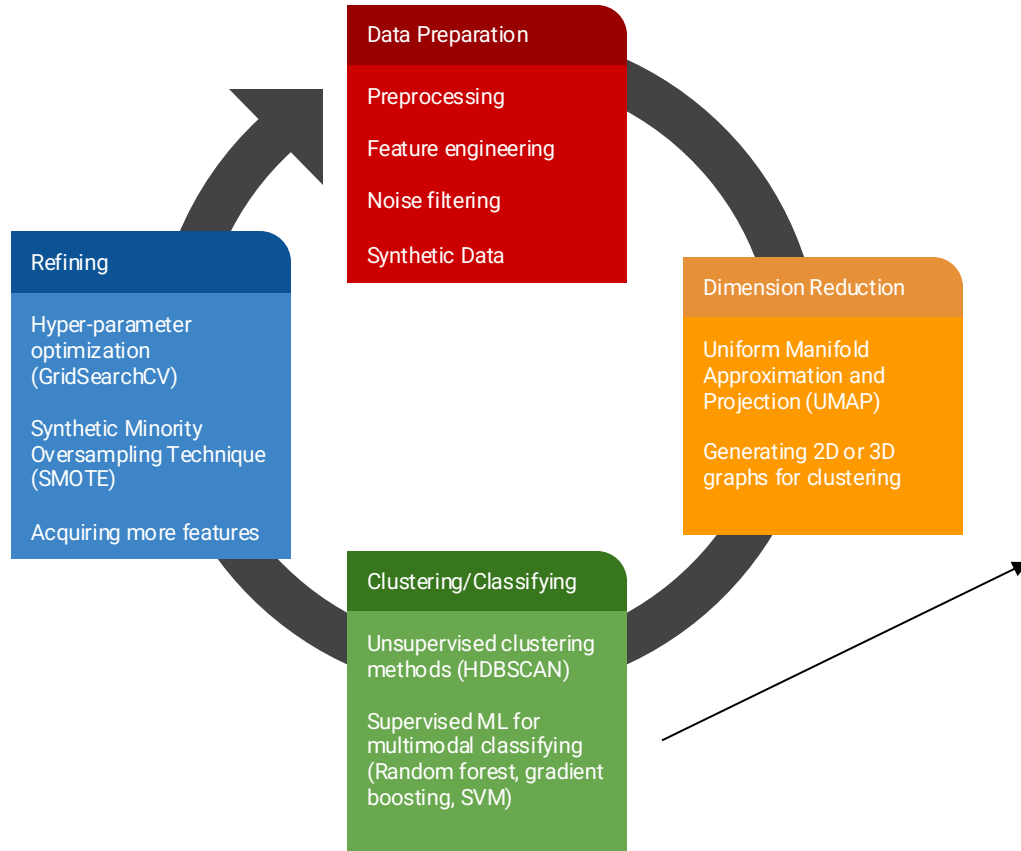
Further Analysis - Summer Project

Overview: Categorize Ex->Inh Synapses and Identify Key Features Using Machine Learning Methods

Goals:

1. Determine synapse types/subtypes based on the organization of Munc13 and PSD95
 - a. Determine how closely those types correlate to postsynaptic cell type
2. Identify synaptic protein organization features we can leverage to distinguish between synapses with different postsynaptic cell types
 - A. Combine multiple methods to create data analysis workflow and test/evaluate already existing tools on our data set
 - B. Compare several algorithms and perform parameter sweeps at each step to define an optimal approach

Analysis Process



Algorithms to Compare

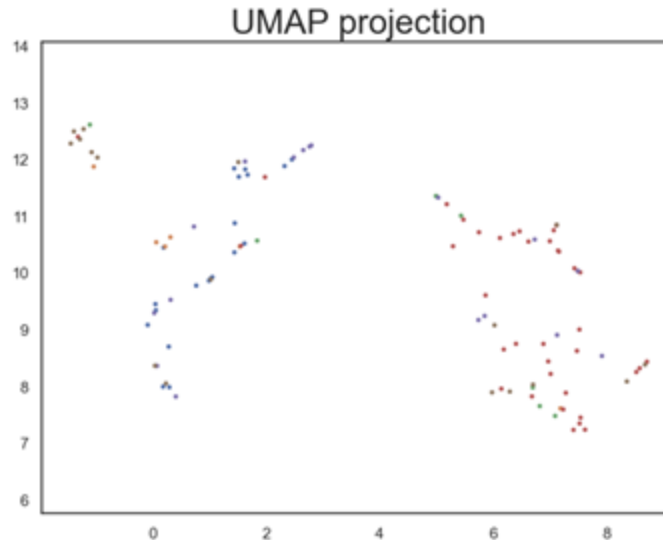
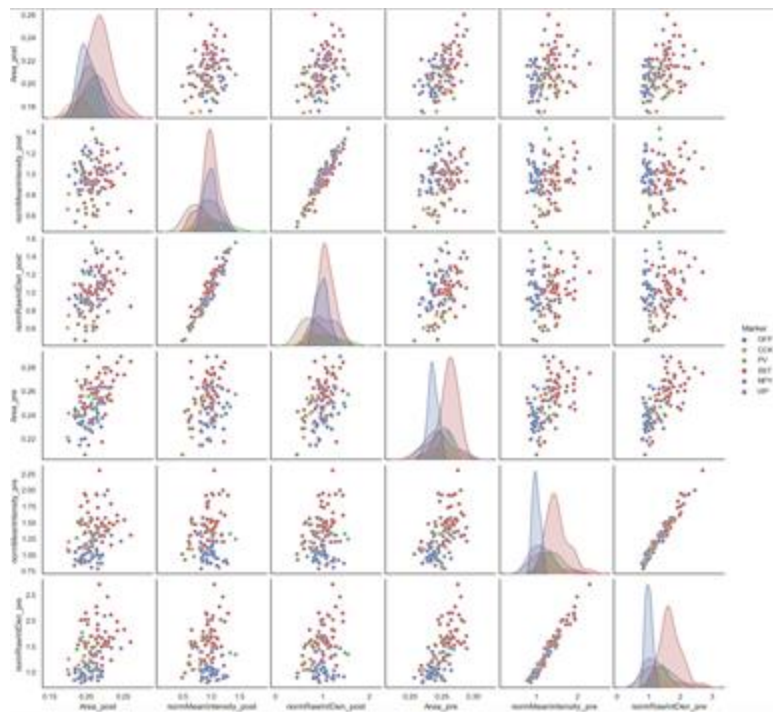
Support Vector Machine: best for high-dimensional data, small to medium-sized datasets, using strategies like one-vs-one or one-vs-all, can handle noisy data with the use of the kernel trick and soft margin approach

Gradient Boosting: handles complex relationships between features, supports multiclass classification, can handle noisy data well by focusing on difficult-to-classify instances

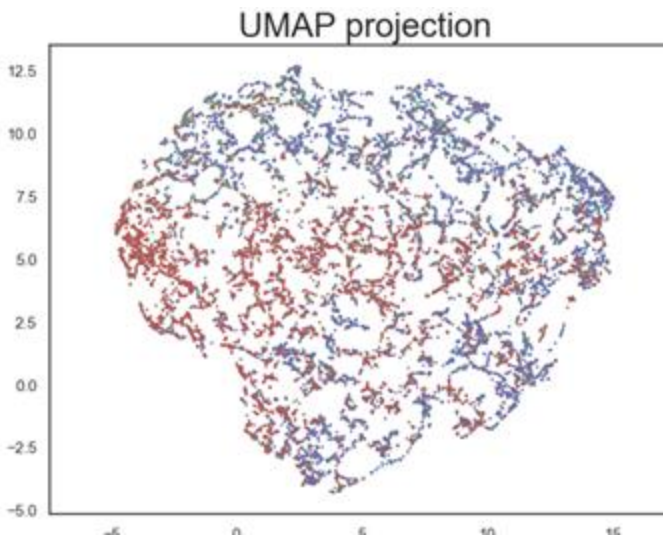
Random Forest Classifier: handles datasets with a large number of features, inherently supports multiclass classification, robust to noise and can handle overlapping data well by averaging predictions from multiple trees

Uniform Manifold Approximation and Projection

6 dimensions to 2 dimensions



Means
per Cell

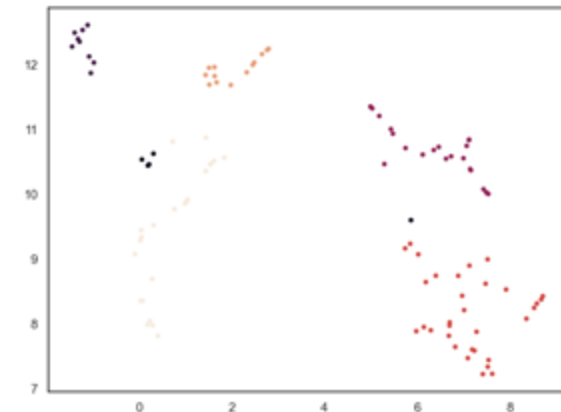


Synaptic

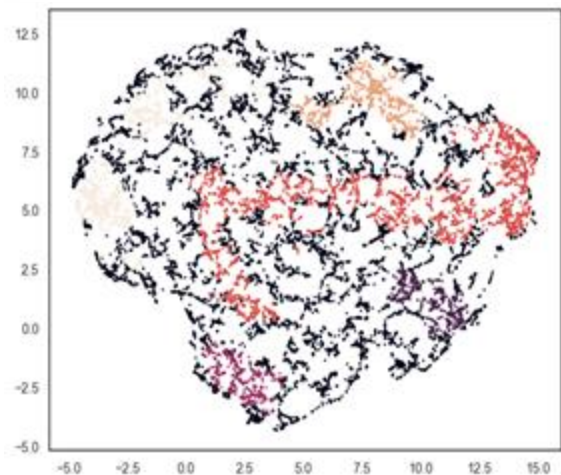
Clustering Algorithm - HDBSCAN

Min_cluster_size = 5

Mean Per Cell



Synaptic

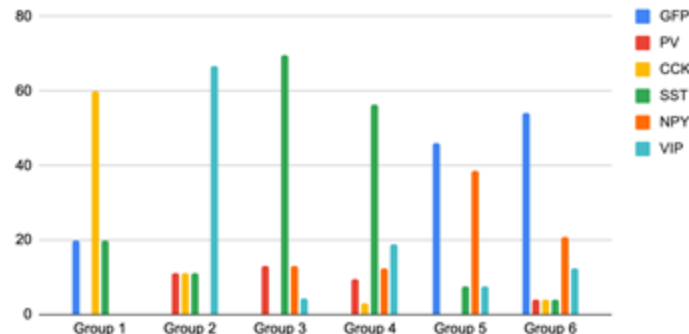


Min_cluster_size = 300

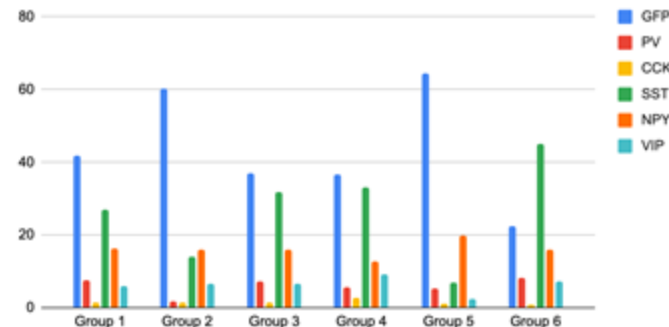
Percentage of each subtype per group

Group	%GFP	%PV	%CCK	%SST	%NPY	%VIP
Group 0	20.0	0.0	60.0	20.0	0.0	0.0
Group 1	0.0	11.11	11.11	11.11	0.0	66.67
Group 2	0.0	13.04	0.0	69.57	13.04	4.35
Group 3	0.0	9.38	3.12	56.25	12.5	18.75
Group 4	46.15	0.0	0.0	7.69	38.46	7.69
Group 5	54.17	4.17	4.17	4.17	20.83	12.5

Percentages per Group - HDBSCAN, Means per Cell



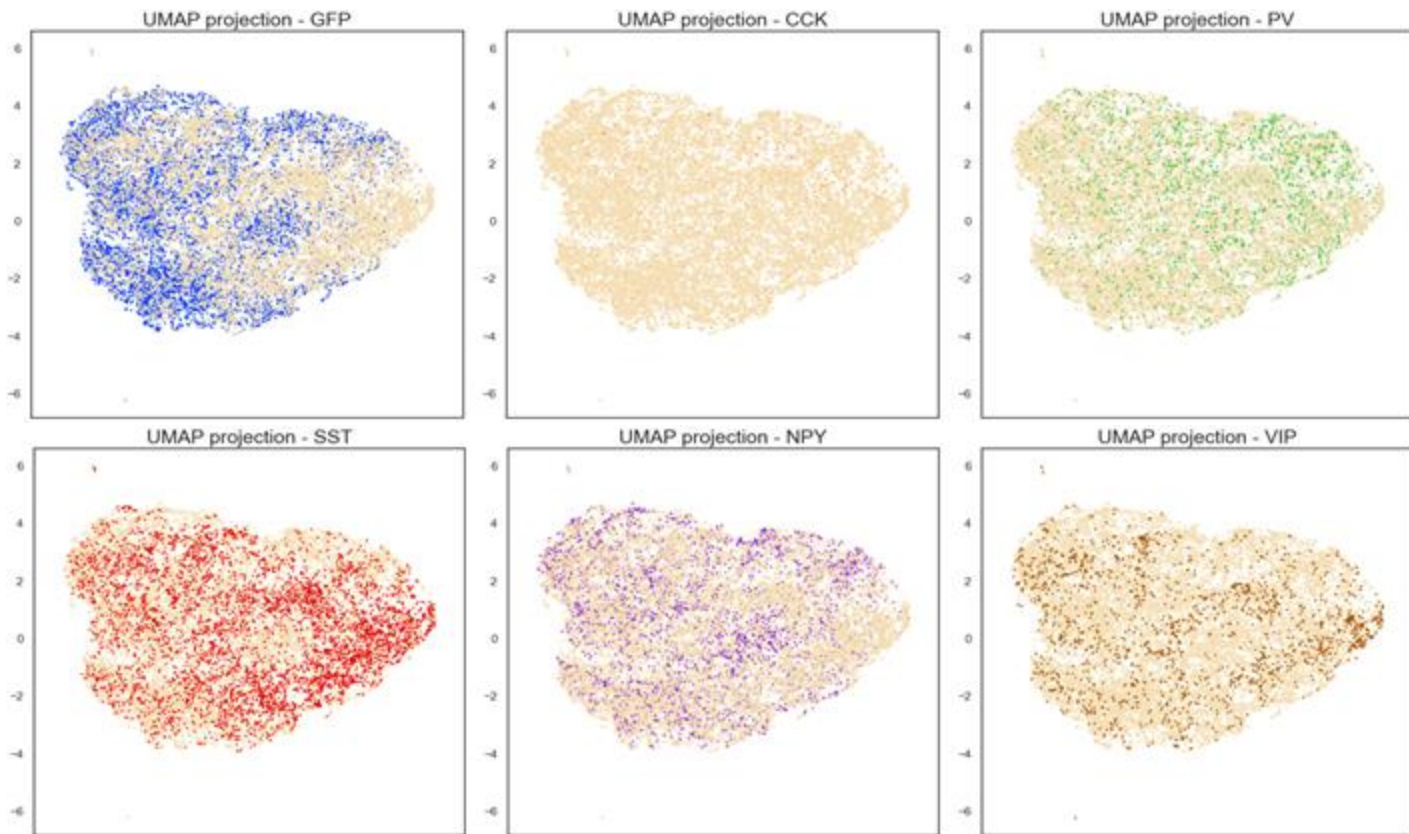
Percentages per Group - HDBSCAN, Synaptic



Acquiring additional features

Reran acquisition
in ImageJ to
include additional
measurements

32 features for
Munc13 and PSD-
95 (64 total)

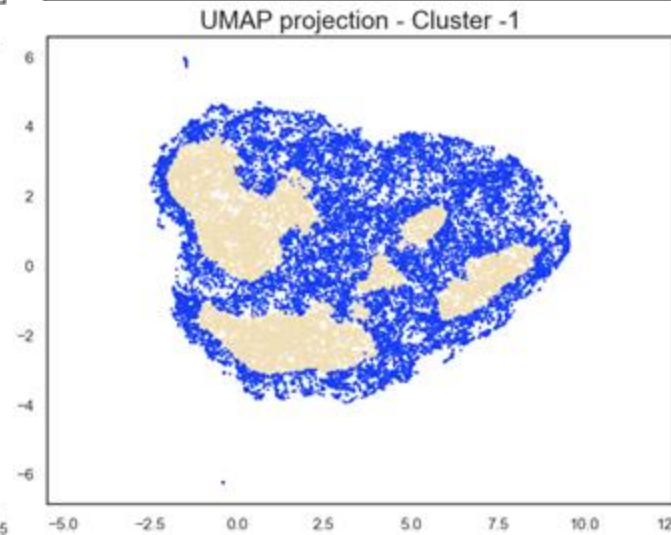
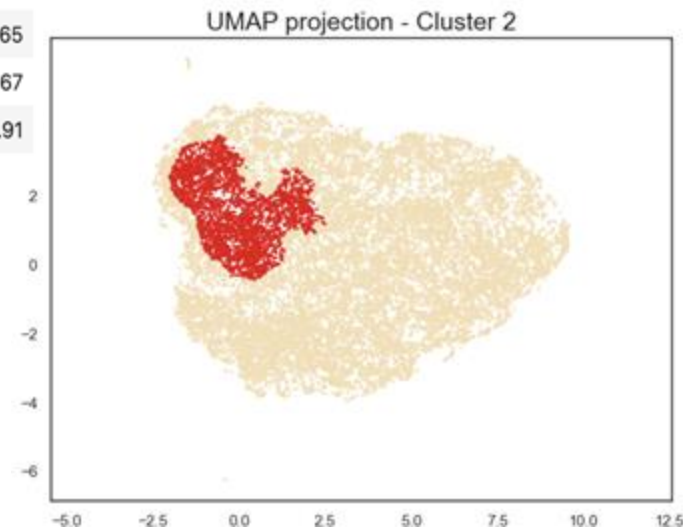
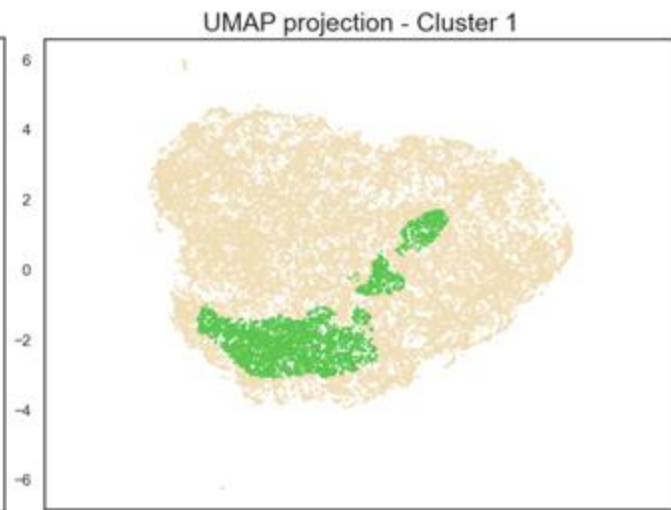
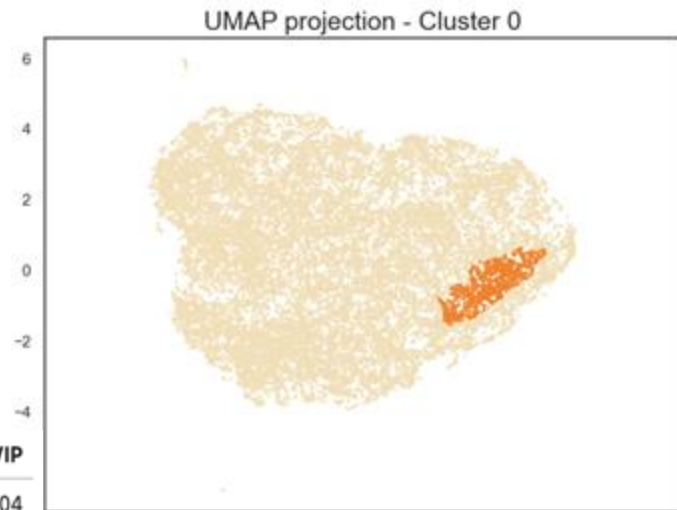


HDBSCAN

Orange - mostly SST
Red - mostly GFP

Group	%GFP	%PV	%CCK	%SST	%NPY	%VIP
Group -1	39.98	11.38	4.26	23.15	15.19	6.04
Group 0	8.73	17.53	3.99	48.08	13.02	8.65
Group 1	51.85	6.64	3.00	20.00	13.84	4.67
Group 2	51.32	6.69	4.43	17.35	14.29	5.91

Min_cluster_size=
1000



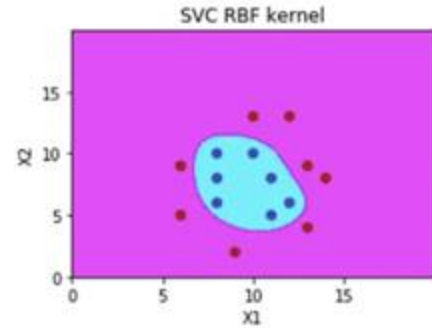
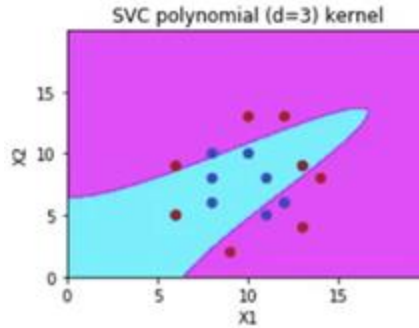
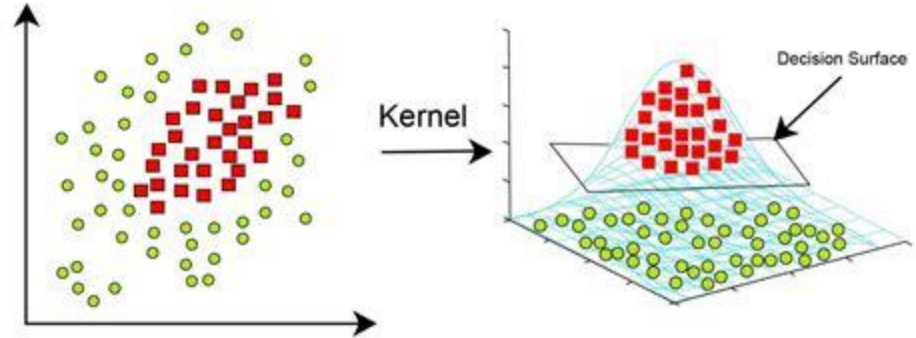
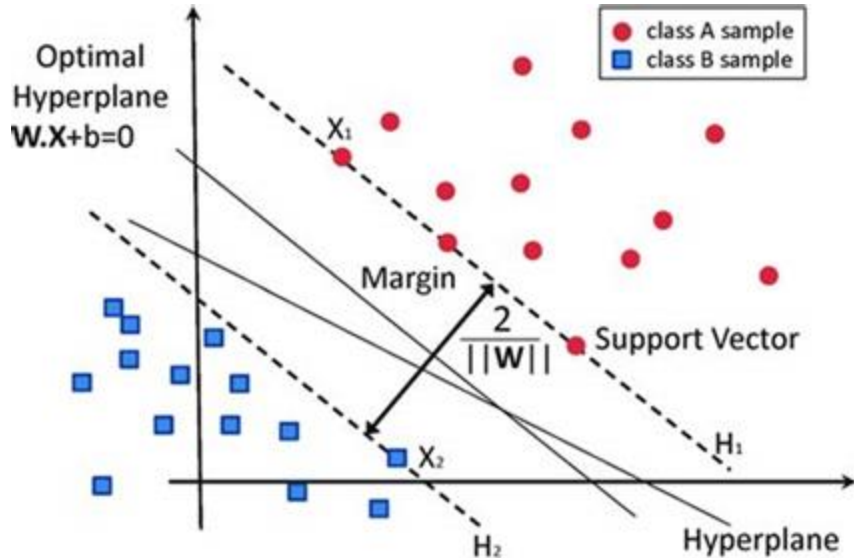
Limitations and Conclusion

1. Co-expression of markers across multiple inhibitory subtypes
 - a. Bistratified cells - SST+ and NPY+
2. Highly variable synaptic staining within certain subtypes
3. Relatively low sample size for ML analysis
4. Features limited to only Munc13 and PSD-95
 - a. Unable to create robust protein profile for each synapse type
5. Limited by confocal data rather than super-resolution

Mainly due to a low sample size and limited features, unsupervised clustering is inconclusive, so we must rely on the classes we identified via staining for further analysis using supervised methods.

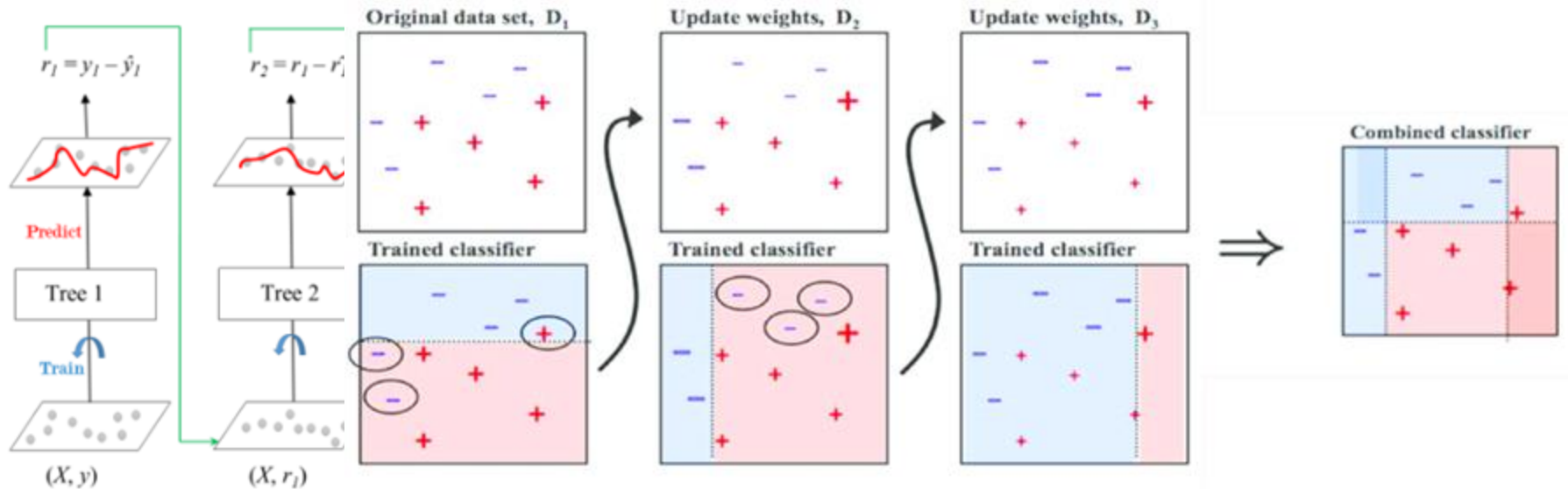
Support Vec

- Establishes hyperplane (decision bound
- Finds optimal equation for the line, with
- Kernels trick - projects data into higher c
- polynomial and Radial Basis Function (F



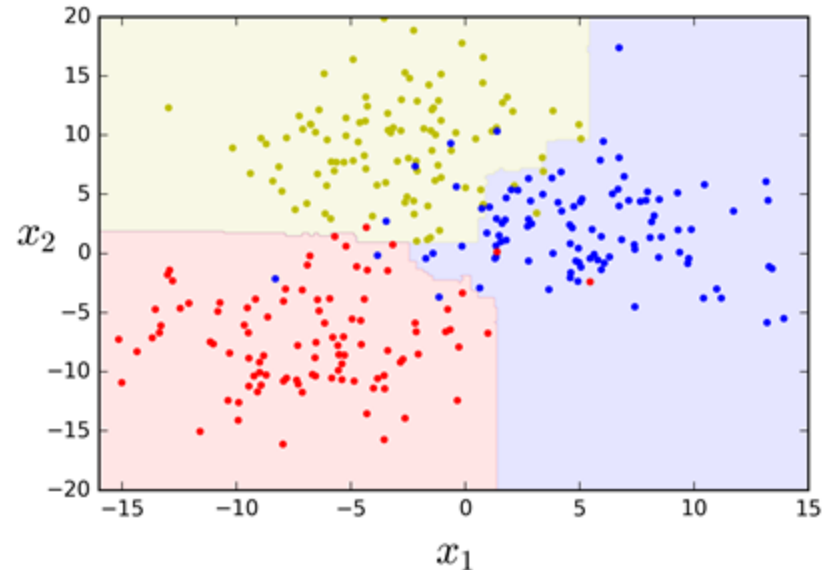
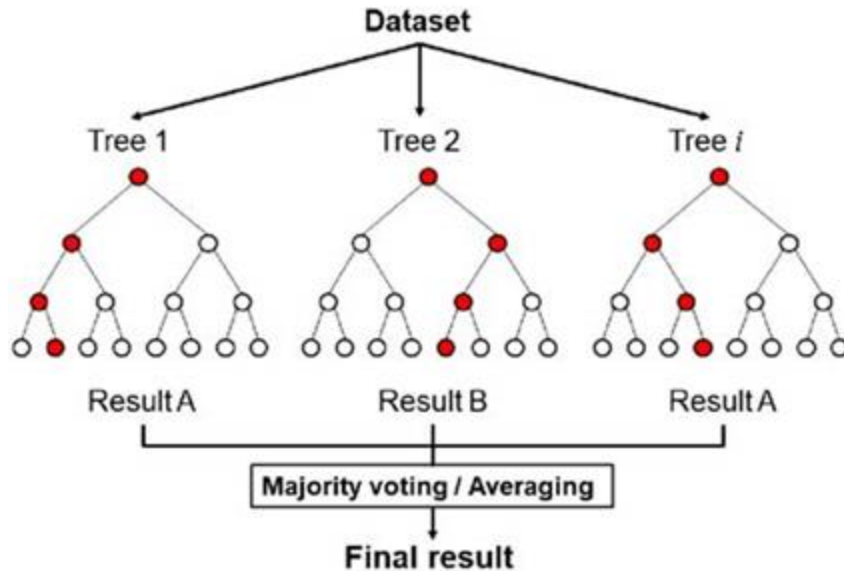
Gradient Boosting

- Ensemble Learning method which trains the model sequentially and each new model tries to correct the previous model
- Computes the gradient of the loss function, trains a new weak model to minimize this gradient, and adds new predictions to ensemble, repeating until a stopping criterion is met



Random Forest Classification

- Also an ensemble method so it combines predictions from other models.
- Each of the smaller models in the random forest ensemble is a decision tree
- Multiple decision trees created using different random subsets of the data and features and most popular prediction among trees chosen

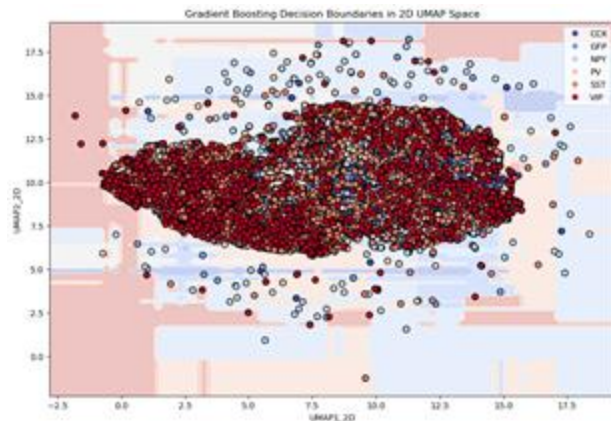
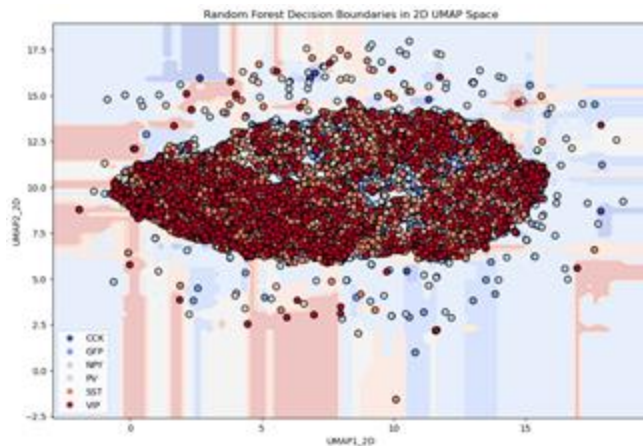
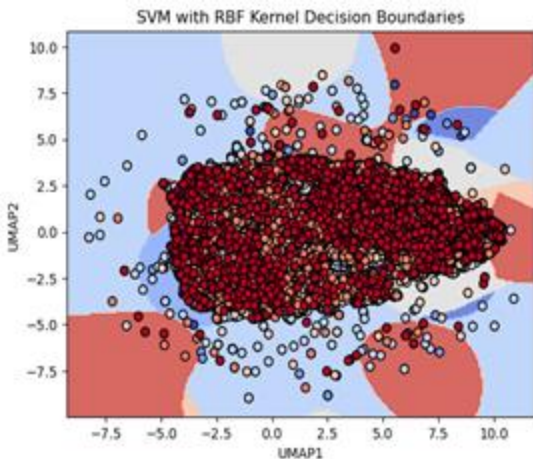


Supervised Learning Algorithms

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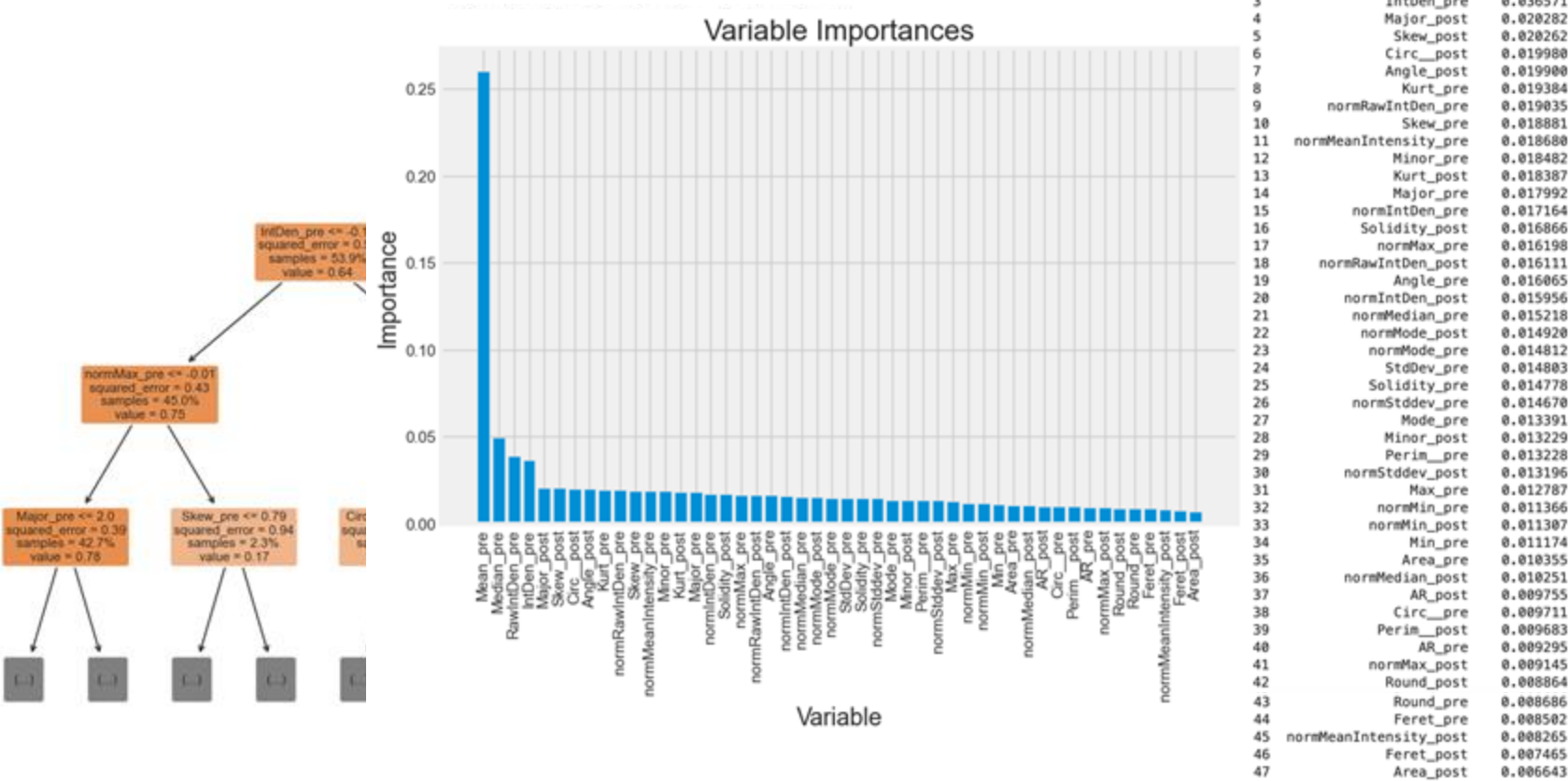
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Random Forest Classifier: handles datasets with a large number of features, inherently supports multiclass classification, robust to noise and can handle overlapping data well by averaging predictions from multiple trees



After discussing with Evelina, a research associate from the Ament lab, I learned some techniques that might be a better approach, focusing specifically on data preprocessing and feature selection.

Next Steps - Feature rank/selection



Next steps - One vs one

- Rather than inputting all the subtypes into one ML algorithm, run a loop that does pairwise comparisons
- Will likely be able to differentiate at least one of the subtypes more successfully than all at once

```
Accuracy: 0.9482273494653911
Confusion Matrix:
[[3356  97]
 [ 179 1699]]
Classification Report:
              precision    recall  f1-score   support

     0       0.95         0.97     0.96       3453
     1       0.95         0.90     0.92       1878

 accuracy          0.95          0.95          0.95          5331
 macro avg         0.95          0.94          0.94          5331
 weighted avg      0.95          0.95          0.95          5331
```

