

Package ‘gJLS’

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Type Package

Title Generalized Joint Location Scale (gJLS) Test

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Description Joint Location Scale (JLS) test to simultaneously test for mean and variance differences between genotype groups.

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LazyData TRUE

RoxygenNote 5.0.1

Imports quantreg, nlme

URL <http://github.com/dsoave/gJLS>

BugReports <http://github.com/dsoave/gJLS/issues>

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gJLS_test	<i>Generalized Joint Location Scale (gJLS) Test</i>
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Description

This function performs the Joint Location Scale (JLS) test (Soave and Sun 2017, Soave et al. 2015) to simultaneously test for mean and variance differences between groups, allowing for correlated errors and group uncertainty. The gJLS test uses Fisher’s combined p-value method to combine evidence from the individual, generalized locaiton (gL) and scale (gS) tests.

Usage

```
gJLS_test(model.loc, model.scale, data, correlation = NULL)
```

Arguments

<code>model.loc</code>	a two-sided linear formula object describing the location test model, with the response (y) on the left and a ~ operator separating the covariates of interest on the right, separated by + operators.
<code>model.scale</code>	a two-sided linear formula object describing the scale test model, with the response (y) on the left and a ~ operator separating the covariates of interest on the right, separated by + operators.
<code>data</code>	a data frame containing the variables named in model and correlation arguments. This is required.
<code>correlation</code>	an optional corStruct object describing the within-group correlation structure. The correlation structure must be called directly from the nlme package using "nlme::" (see examples below). See the documentation of corClasses for a description of the available corStruct classes. If a grouping variable is to be used, it must be specified in the form argument to the corStruct constructor. Defaults to NULL, corresponding to uncorrelated errors.

Details

No missing data are allowed - function will return an "error". Absolute residuals, are estimated using least absolute deviation (LAD) regression. Outcome (phenotype) must be quantitative and covariate (genotype) may be discrete (categorical) or continuous.

Value

a table consisting of test statistics, degrees of freedom and p-values for each of the generalized location (gL), scale (gS) and, joint location-scale (gJLS) tests.

`numDF` the gS test statistic numerator degrees of freedom

`denDF` the gS test statistic denominator degrees of freedom

`gS_p` the gS test p-value

Author(s)

David Soave

References

Soave, D., Corvol, H., Panjwani, N., Gong, J., Li, W., Boelle, P.Y., Durie, P.R., Paterson, A.D., Rommens, J.M., Strug, L.J., and Sun, L. (2015). A Joint Location-Scale Test Improves Power to Detect Associated SNPs, Gene Sets, and Pathways. *American journal of human genetics* 97, 125-138.

Soave, D. and Sun, L. (2017). A Generalized Levene's Scale Test for Variance Heterogeneity in the Presence of Sample Correlation and Group Uncertainty. *Biometrics* (Accepted).

See Also

[gL_test](#), [gS_test](#)

Examples

```
#####
## Example simulating data from model [i] (Soave et al. 2015 AJHG)
#####

n<-2000 ## total sample size
pA<-0.3 ## MAF
pE1<-0.3 ## frequency of exposure E1

## Genotypes (XG)
genocount<-rmultinom(1,size=n,prob=c(pA*pA, 2*pA*(1-pA), (1-pA)*(1-pA)))
XG<-c(rep(0, genocount[1]), rep(1, genocount[2]), rep(2,genocount[3]))
XG<-sample(XG,size=length(XG),replace=FALSE)

## Exposures (E1)
E1<-rbinom(n,1,prob=pE1)

## Phenotype (y)
y<-0.01*XG+0.3*E1+0.5*XG*E1+rnorm(n,0,1)

# Additive model (will work with dosages)
JLS_test(y,XG,XG)
# Genotypic model (will not work with dosages --> factor() will create many groups)
JLS_test(y,factor(XG),factor(XG))

X2=round(cbind(XG==1,XG==2)) #convert to 2 columns

# Genotypic model --> same result as results above using JLS_test(y,factor(X),factor(X))
# This is how genotype probabilities will be analyzed (using a 2 column design matrix)
JLS_test(y,X2,X2)
```

gL_test

Generalized Location (gL) Test

Description

This function performs a generalized location (gL) test (Soave and Sun 2017) using the generalized least squares function, `gls()`, from the `nlme` package (Pinheiro and Bates 2000) to allow for correlated errors.

Usage

```
gL_test(model, data, correlation = NULL)
```

Arguments

<code>model</code>	a two-sided linear formula object describing the model, with the response (y) on the left and a ~ operator separating the covariates of interest on the right, separated by + operators.
<code>data</code>	a data frame containing the variables named in model and correlation arguments. This is required.

correlation an optional corStruct object describing the within-group correlation structure. The correlation structure must be called directly from the nlme package using "nlme::" (see examples below). See the documentation of corClasses for a description of the available corStruct classes. If a grouping variable is to be used, it must be specified in the form argument to the corStruct constructor. Defaults to NULL, corresponding to uncorrelated errors.

Details

No missing data are allowed - function will return an "error". Outcome (phenotype) must be quantitative and covariate (genotype) may be discrete (categorical) or continuous.

Value

gL_F the gL test statistic
 numDF the gL test statistic numerator degrees of freedom
 denDF the gL test statistic denominator degrees of freedom
 gL_p the gL test p-value

Author(s)

David Soave

References

Soave, D., Corvol, H., Panjwani, N., Gong, J., Li, W., Boelle, P.Y., Durie, P.R., Paterson, A.D., Rommens, J.M., Strug, L.J., and Sun, L. (2015). A Joint Location-Scale Test Improves Power to Detect Associated SNPs, Gene Sets, and Pathways. American journal of human genetics 97, 125-138.

Soave, D. and Sun, L. (2017). A Generalized Levene's Scale Test for Variance Heterogeneity in the Presence of Sample Correlation and Group Uncertainty. Biometrics (Accepted).

See Also

[gS_test](#), [gJLS_test](#)

Examples

```
#####
## Example simulating data from model [i] (Soave et al. 2015 AJHG)
#####

n<-2000 ## total sample size
pA<-0.3 ## MAF
pE1<-0.3 ## frequency of exposure E1

## Genotypes (XG)
genocount<-rmultinom(1,size=n,prob=c(pA*pA, 2*pA*(1-pA), (1-pA)*(1-pA)))
XG<-c(rep(0, genocount[1]), rep(1, genocount[2]), rep(2,genocount[3]))
XG<-sample(XG,size=length(XG),replace=FALSE)

## Exposures (E1)
E1<-rbinom(n,1,prob=pE1)
```

```
## Phenotype (y)
y<-0.01*XG+0.3*E1+0.5*XG*E1+rnorm(n,0,1)

# Additive model (will work with dosages)
JLS_test(y,XG,XG)
# Genotypic model (will not work with dosages --> factor() will create many groups)
JLS_test(y,factor(XG),factor(XG))

X2=round(cbind(XG==1,XG==2)) #convert to 2 columns

# Genotypic model --> same result as results above using JLS_test(y,factor(X),factor(X))
# This is how genotype probabilities will be analyzed (using a 2 column design matrix)
JLS_test(y,X2,X2)
```

gS_test

*Generalized Scale (gS) Test***Description**

This function performs the generalized scale (gS) test (Soave and Sun 2017). This extension of Levene's (1960) test allows for correlated errors and group uncertainty.

Usage

```
gS_test(model, data, correlation = NULL)
```

Arguments

model	a two-sided linear formula object describing the model, with the response (y) on the left and a ~ operator separating the covariates of interest on the right, separated by + operators.
data	a data frame containing the variables named in model and correlation arguments. This is required.
correlation	an optional corStruct object describing the within-group correlation structure. The correlation structure must be called directly from the nlme package using "nlme::" (see examples below). See the documentation of corClasses for a description of the available corStruct classes. If a grouping variable is to be used, it must be specified in the form argument to the corStruct constructor. Defaults to NULL, corresponding to uncorrelated errors.

Details

No missing data are allowed - function will return an "error". Absolute residuals, are estimated using least absolute deviation (LAD) regression. Outcome (phenotype) must be quantitative and covariate (genotype) may be discrete (categorical) or continuous.

Value

gS_F the gS test statistic
 numDF the gS test statistic numerator degrees of freedom
 denDF the gS test statistic denominator degrees of freedom
 gS_p the gS test p-value

Author(s)

David Soave

References

Soave, D., Corvol, H., Panjwani, N., Gong, J., Li, W., Boelle, P.Y., Durie, P.R., Paterson, A.D., Rommens, J.M., Strug, L.J., and Sun, L. (2015). A Joint Location-Scale Test Improves Power to Detect Associated SNPs, Gene Sets, and Pathways. *American journal of human genetics* 97, 125-138.

Soave, D. and Sun, L. (2017). A Generalized Levene's Scale Test for Variance Heterogeneity in the Presence of Sample Correlation and Group Uncertainty. *Biometrics* (Accepted).

See Also

[gL_test](#), [gJLS_test](#)

Examples

```
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## Example simulating data from model [i] (Soave et al. 2015 AJHG)
#####

n<-2000 ## total sample size
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genocount<-rmultinom(1,size=n,prob=c(pA*pA, 2*pA*(1-pA), (1-pA)*(1-pA)))
XG<-c(rep(0, genocount[1]), rep(1, genocount[2]), rep(2,genocount[3]))
XG<-sample(XG,size=length(XG),replace=FALSE)

## Exposures (E1)
E1<-rbinom(n,1,prob=pE1)

## Phenotype (y)
y<-0.01*XG+0.3*E1+0.5*XG*E1+rnorm(n,0,1)

# Additive model (will work with dosages)
JLS_test(y,XG,XG)
# Genotypic model (will not work with dosages --> factor() will create many groups)
JLS_test(y,factor(XG),factor(XG))

X2=round(cbind(XG==1,XG==2)) #convert to 2 columns

# Genotypic model --> same result as results above using JLS_test(y,factor(X),factor(X))
# This is how genotype probabilities will be analyzed (using a 2 column design matrix)
JLS_test(y,X2,X2)
```

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