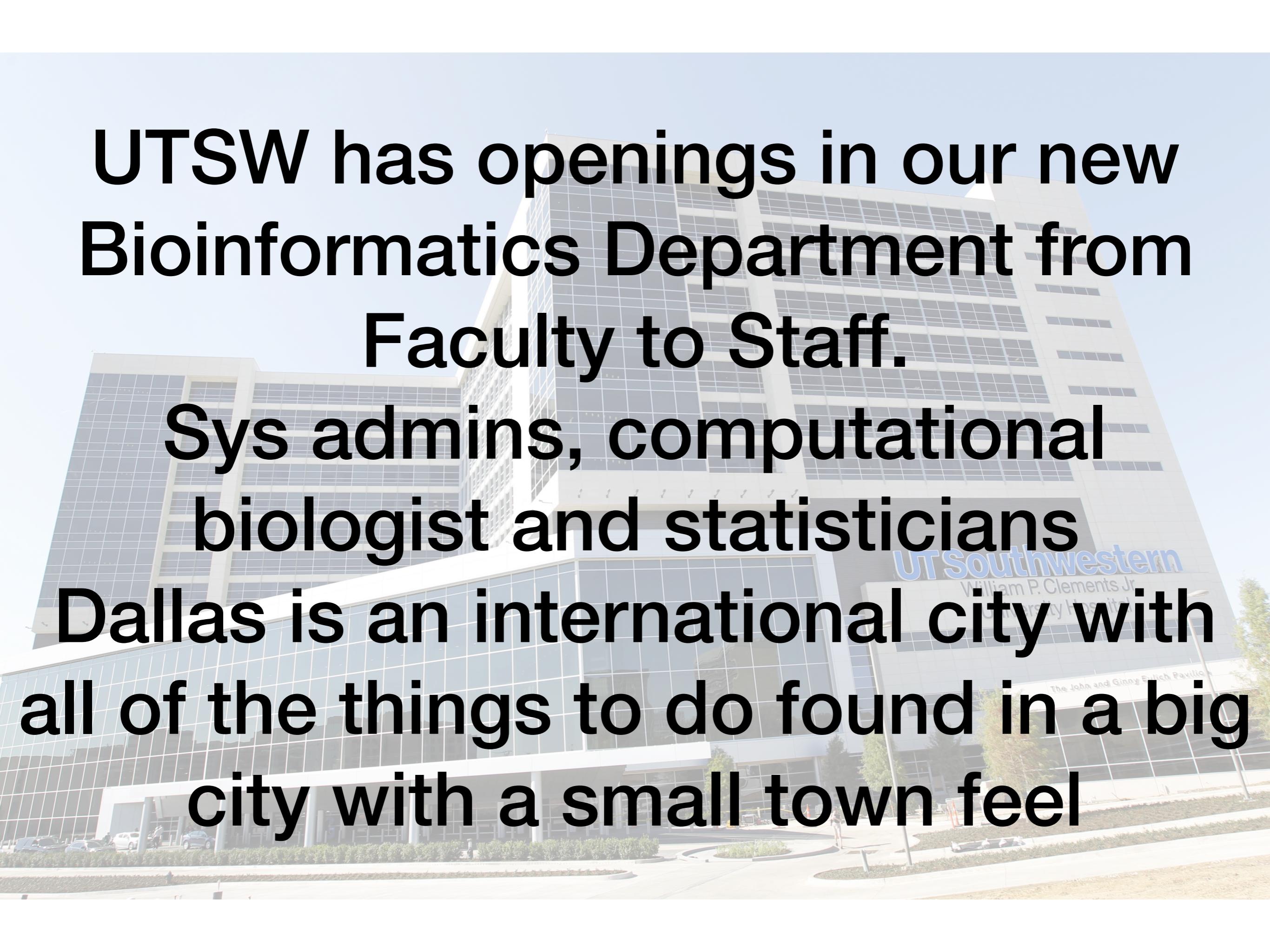


# Sequence Homology and Analysis

*Understanding How FASTA and BLAST work to  
optimize your sequence similarity searches.*



Brandi Cantarel, Ph.D  
UTSW, Department of Bioinformatics  
Programming for Biology 2018



**UTSW has openings in our new  
Bioinformatics Department from  
Faculty to Staff.**

**Sys admins, computational  
biologist and statisticians**

**Dallas is an international city with  
all of the things to do found in a big  
city with a small town feel**

# Take Home Messages

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1. *Homologous* sequences share a common ancestor, but most sequences are *non-homologous*
2. Compare protein sequence for distant comparison and DNA for close comparisons
3. Sequence Homology can be reliably inferred from statistically significant similarity (non-homology cannot from non-similarity)
4. Homologous proteins share common structures, but not necessarily common functions
5. Sequence statistical significance estimates are accurate (verify this yourself)
6. Smaller databases increase search sensitivity
7. Statistical accuracy can be evaluated by examining the “highest scoring unrelated sequence” or by random shuffles

# What is Understanding Homology Important

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- Most gene databases use sequence similarity to infer gene function (with a few exceptions)
  - In the absence of high-throughput biochemistry experimentation, homology is used to predict gene function and pathway assignments.
- Many of these predictions are correct however distinguishing orthologs (deviation from speciation) and paralogs (deviation from gene duplication) is difficult
- E-values are more reliable than percent identity

What is Homology?  
How do we recognize it?

# History of Sequence Similarity

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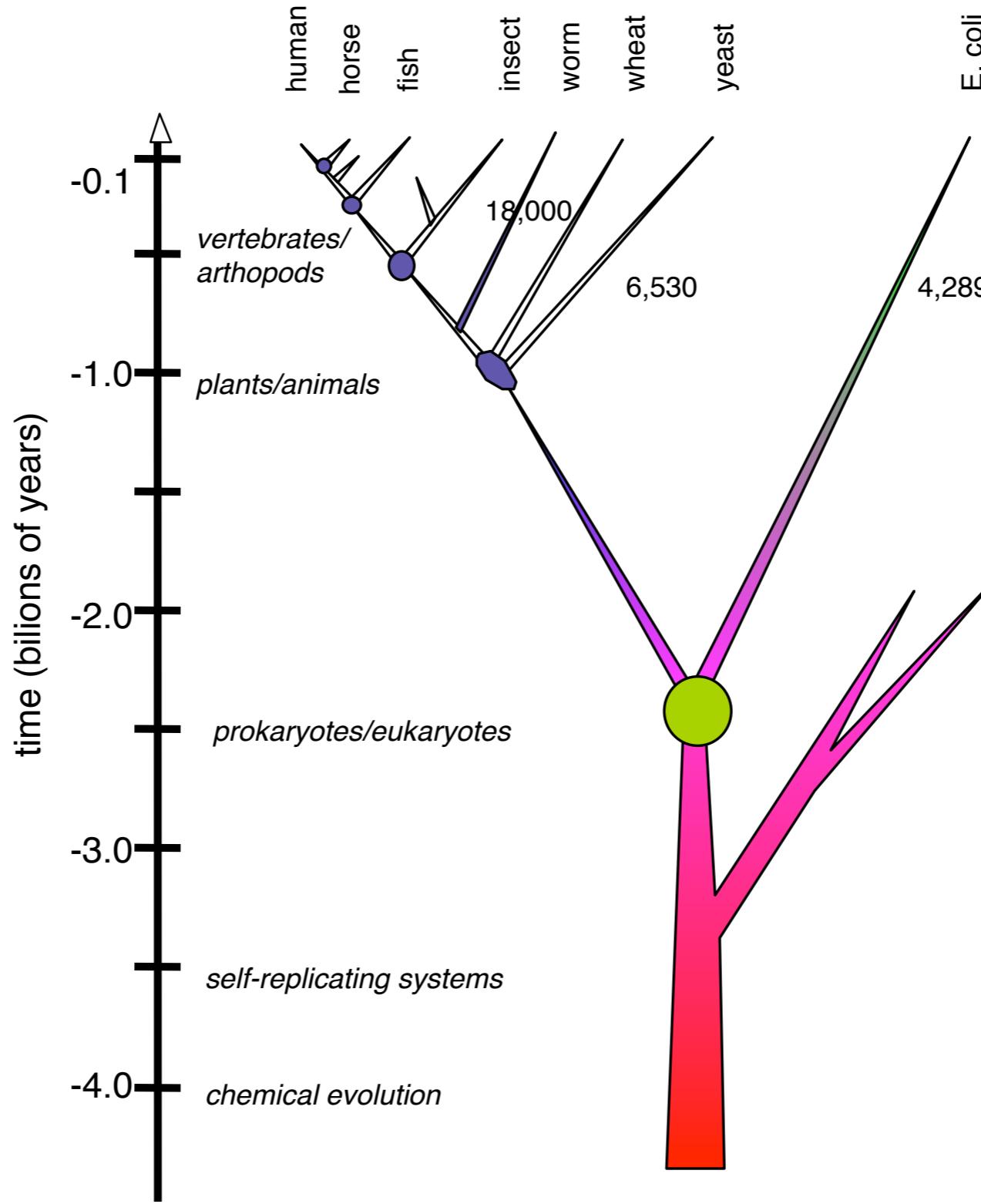
1950s	Fredrick Sanger Determines the Sequence of Insulin
1970	Margaret Dayhoff compiled one of the first protein sequence databases, manually aligns sequences and creates the first protein substitution matrices
1979	Temple Smith and Michael Waterman proposed an optimal alignment algorithm for local alignments
1981	William Pearson and David Lipman implement an optimization of the Smith-Waterman algorithm using short exact matching words (FASTA)
1985	Stephen Altshul, Warren Gish, Webb Miller, Eugene Myers and David Lipman, propose a faster alignment tool to identify high identity matches without GAPS
1990	Randall Smith and Temple Smith implement pattern-induced multiple-sequence alignment (PUMA)
1992	Sean Eddy implements multiple sequence alignments using hidden Markov Models (HMMER)
1995	Warren Gish, branches the development of BLAST
1996	Gapped BLAST and PSI-BLAST
1997	Gapped BLAST and PSI-BLAST

# Establishing homology from statistically significant similarity

---

- For most proteins, homologs are easily found over long evolutionary distances (500 My – 2 By) using standard approaches (BLAST, FASTA)
- Difficult for distant relationships or very short domains
- Most default search parameters are optimized for distant relationships and work well

# Homologous Sequences Share a Common Ancestor

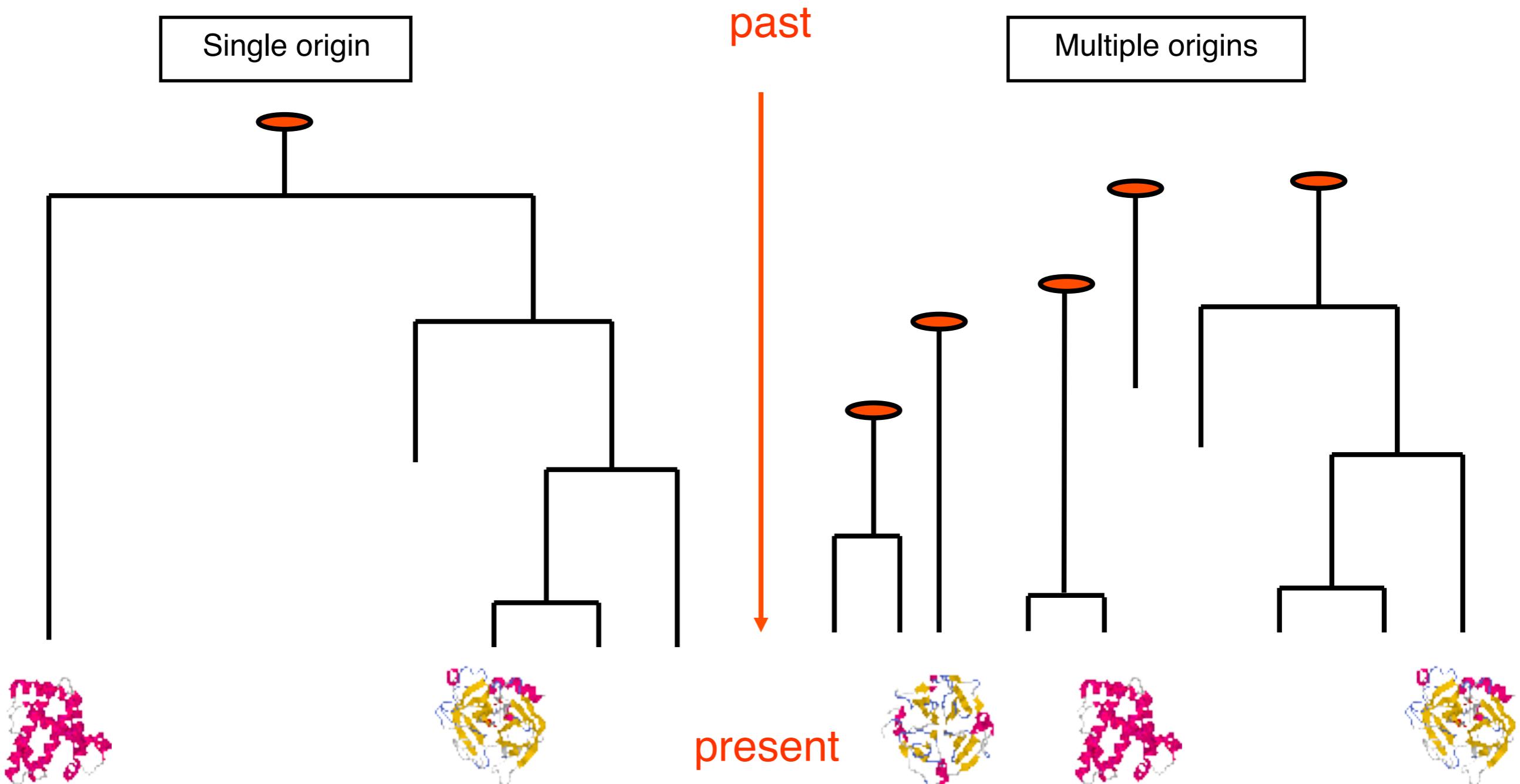


# Homology is Confusing: Ways we have seen it defined

---

- Protein/Genes/DNA that share a common ancestor
- Specific positions/columns in a multiple sequence alignment that have a 1:1 relationship over evolutionary history
  - Is it possible to be 50% homologous?
- Specific morphological/functional characteristics that share a recent divergence (clade)
  - Are all wings homologous (bat, butterfly, eagle)?

# Homology is Confusing: Are all sequences homologous?



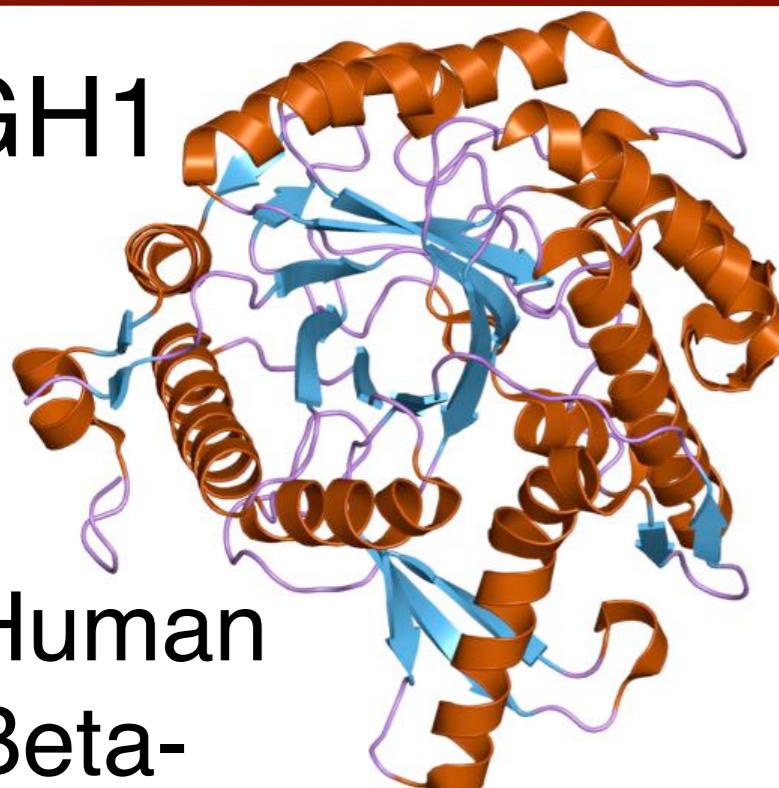
# Homology Using Sequence/Structural Comparisons

---

- Homology is shared ancestry
- Convergence are independent events resulting in the same outcome.
- Sequences are inferred to share a common ancestor based on statistically significant **excess** similarity
- Any evidence of this **excess** similarity can be used to infer homology (sequence or structure)
- Lack of evidence cannot be used to infer non-homology
- One must weight the evidence for each hypothesis (Convergence or Homology)

# When do we infer homology?

GH1



Human  
Beta-  
glucosidase

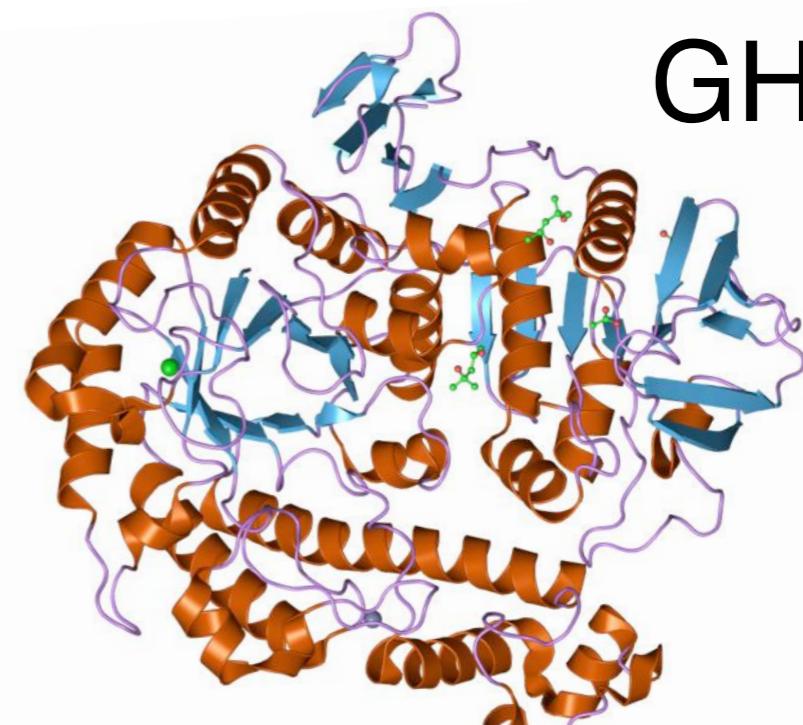
Sequence: Score = 205 bits  
Expect = 3e-57  
%ID = 30%  
Structure: RMSD = 2.63  
Score = 1044  
P-Value = 0

Sequence: Score = 16.2  
Expect = 1.3  
%ID = 26%  
Structure: RMSD = 3.80  
Score = 364.8  
P-Value = 1.97e-02

Lactococcus lactis  
Beta-galactosidase



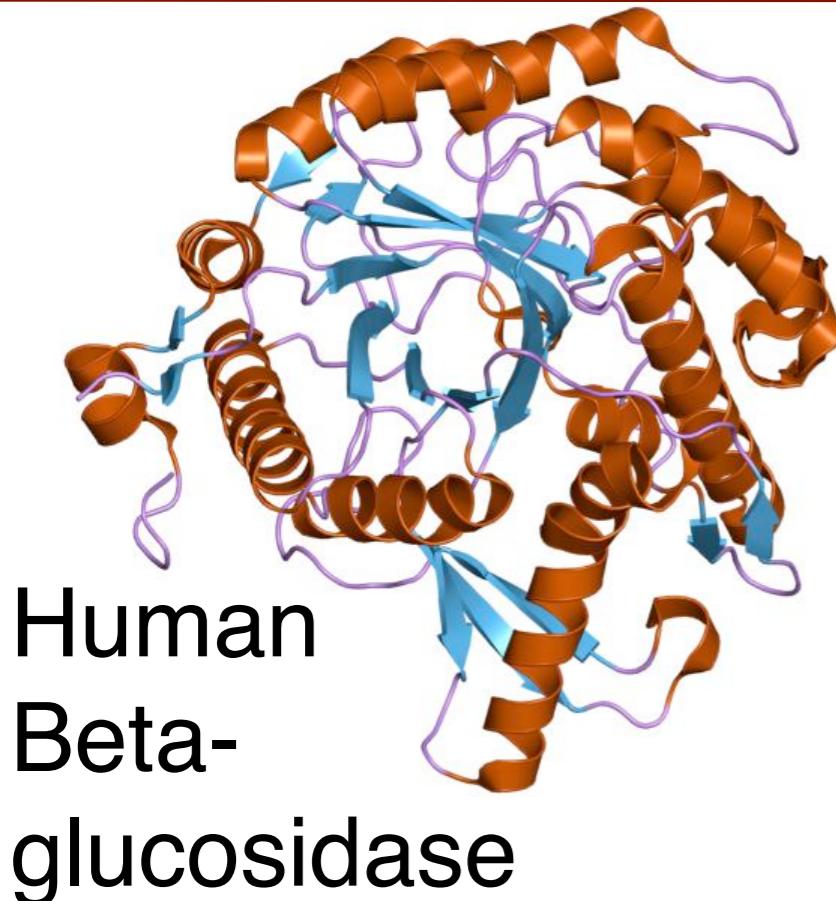
GH42



Human Beta-  
galactosidase

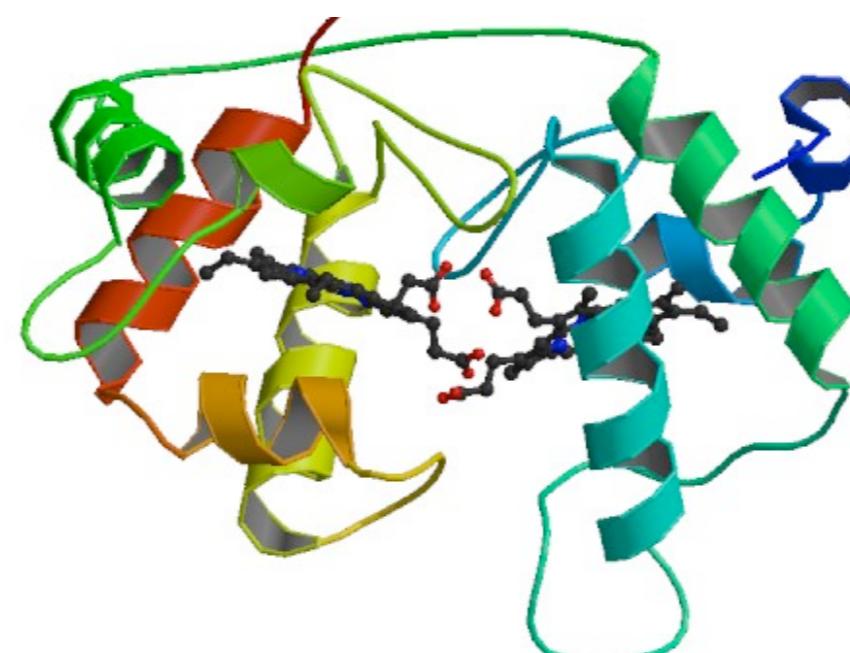
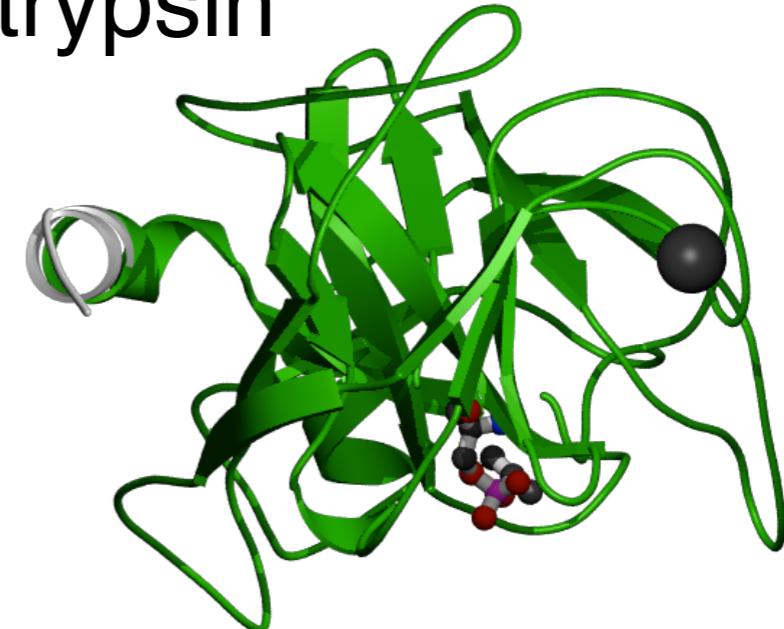
Sequence: Score = 22.3  
Expect = 0.051  
%ID = 25%  
Structure: RMSD = 393.5  
Score = 393.5  
P-Value = 1.65e-07

# When do we infer non-homology?



Sequence: Score = 13.5 bits (23)  
Expect = 6.4  
%ID = 36%  
Structure: P-value: 7.57e-01  
Score: 122.45  
RMSD: 4.74  
%Id: 4.3%

Bovine trypsin



CYTOCHROME  
C4

Non-homologous Proteins have  
different structures

# What BLAST Does

---

?

Similarity <=> Homology

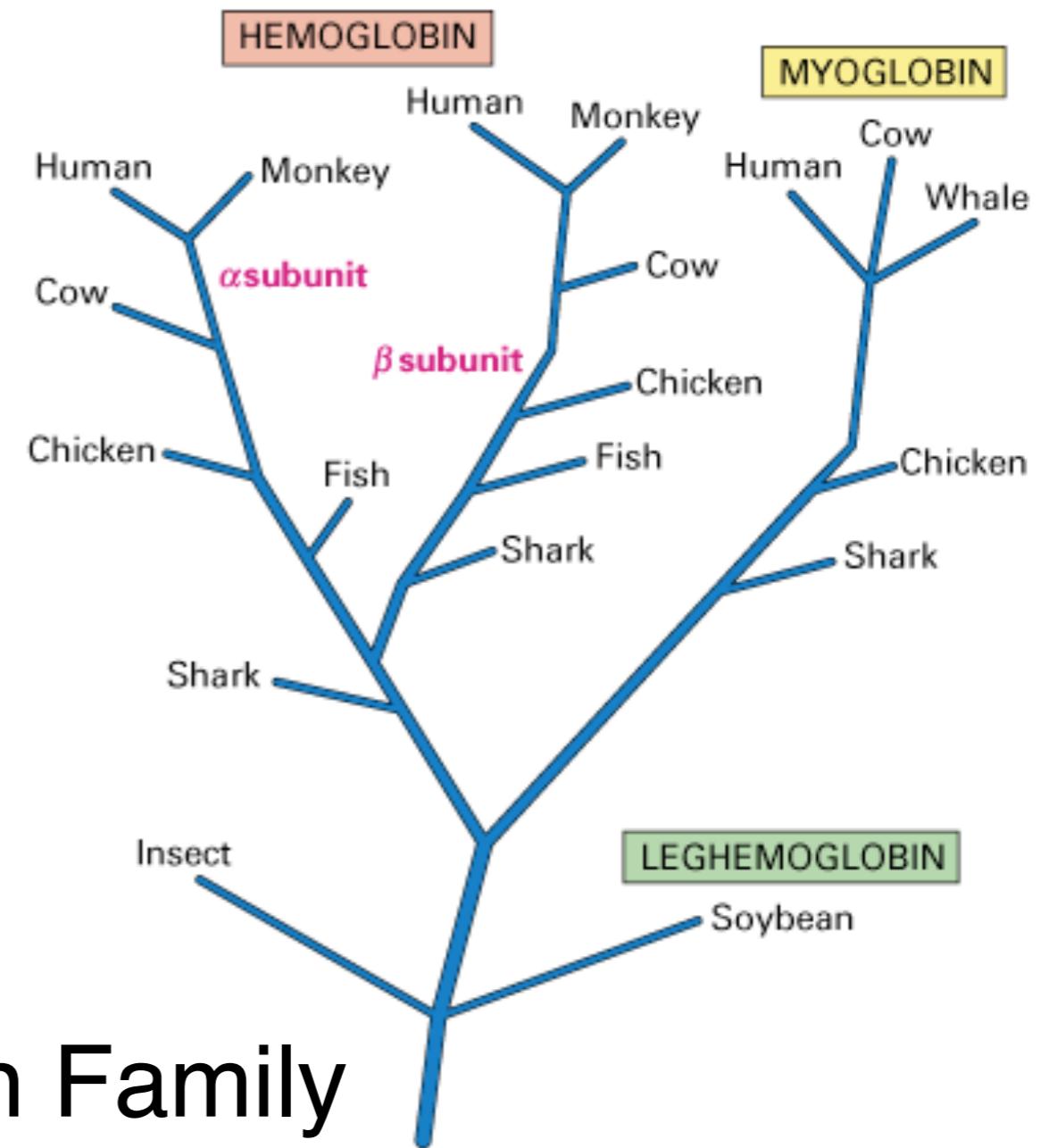
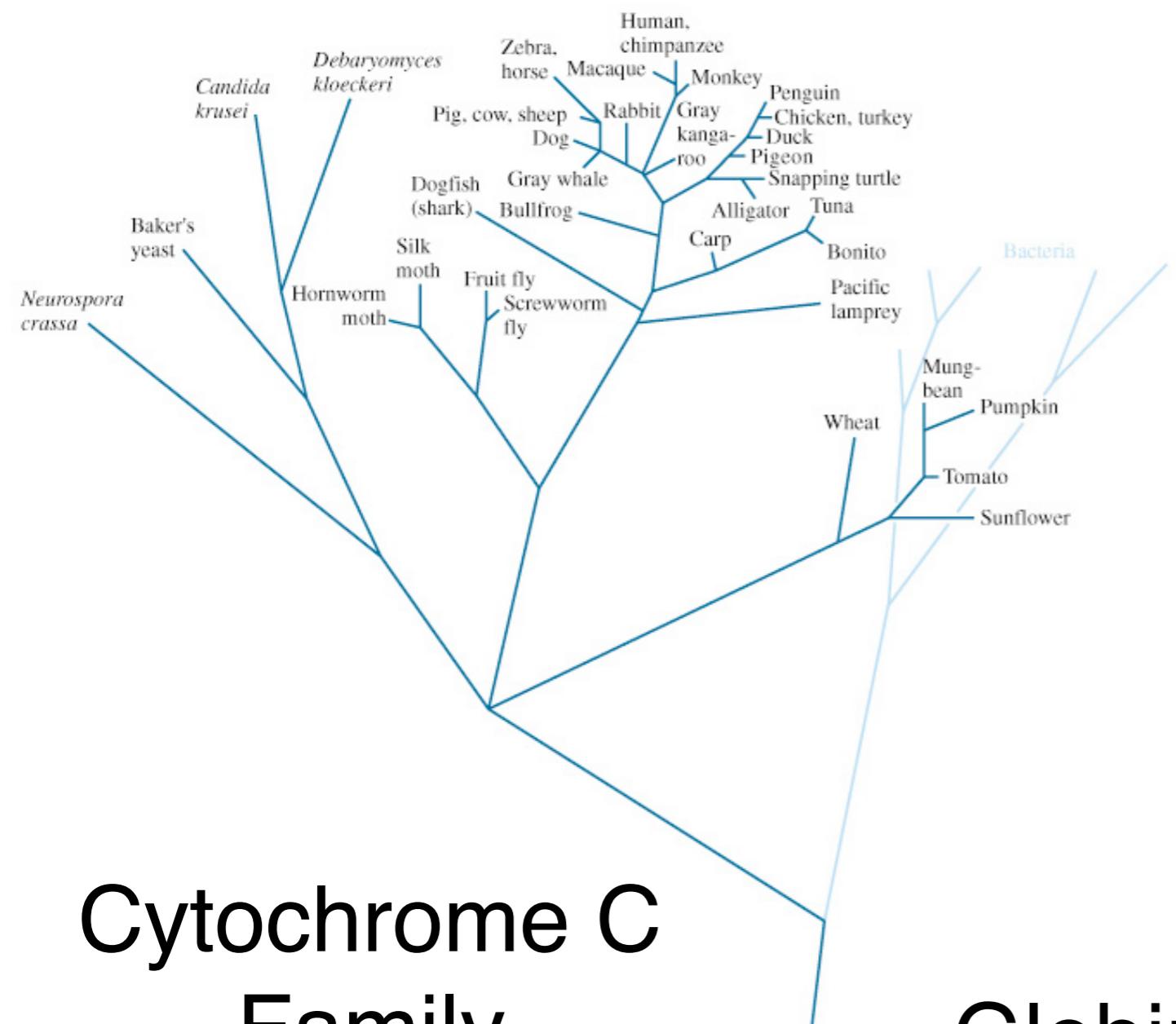
?

Statistical Significance <=> Biological Significance

Divergence OR Convergence

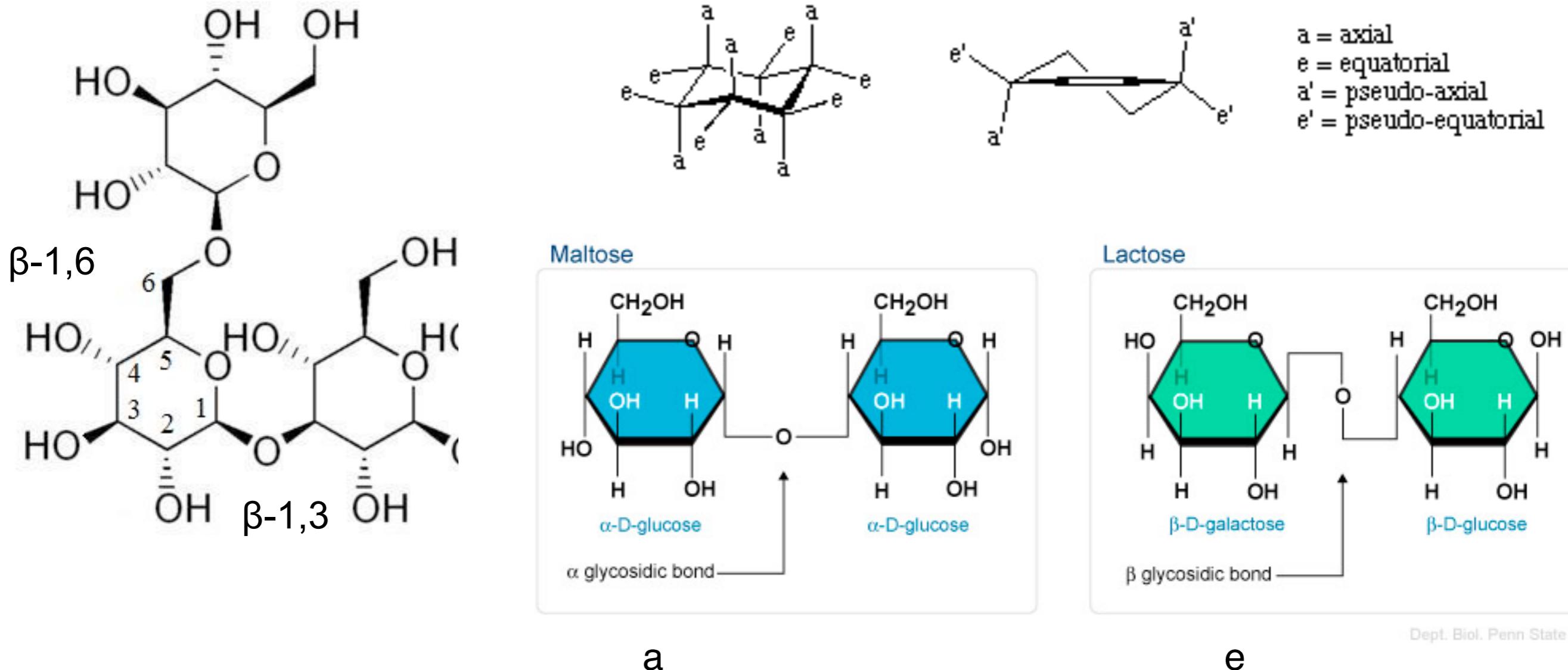
# Orthologs vs Paralogs

## Inferring Function



# Orthologs vs Paralogs

## Inferring Function



Dept. Biol. Penn State ©2008

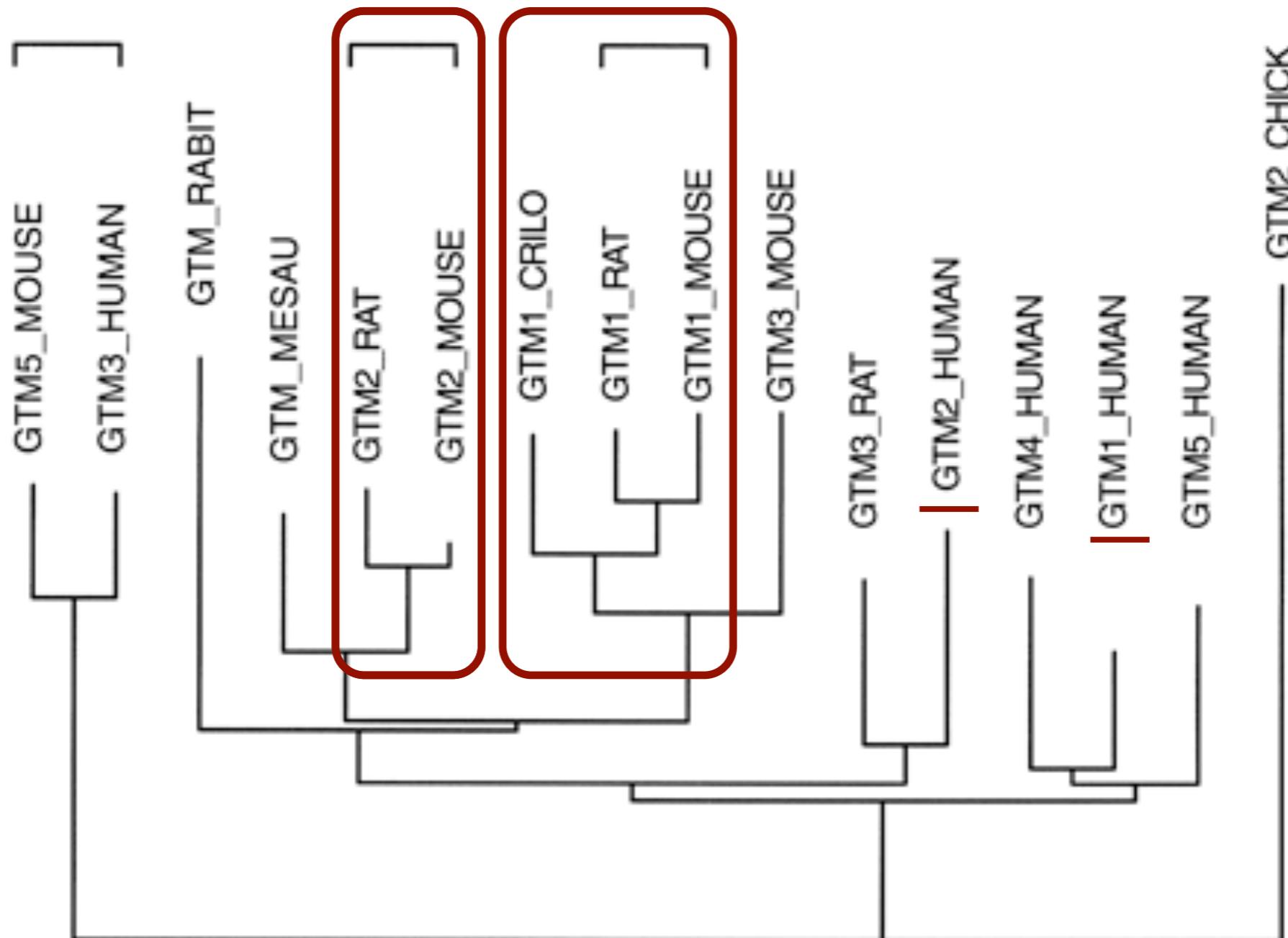
Homologs Often Maintain Similar Chemical Functions

# Orthology can be difficult to infer

---

- Over modest distances(human/mouse) post- speciation duplication is common
- Over large distances (human/fly,bacteria), duplication/ loss/replacement may be common
- Homology inferences have false-negatives, but the false-positive rate can be reliably controlled
- Orthology inferences will have both false positives and false negatives
- Paralogous proteins often have similar chemical functions (may act in different pathways)

# Orthology can be difficult to infer



doi: 10.1101/gr.9.4.373 *Genome Res.* 1999. 9: 373-382

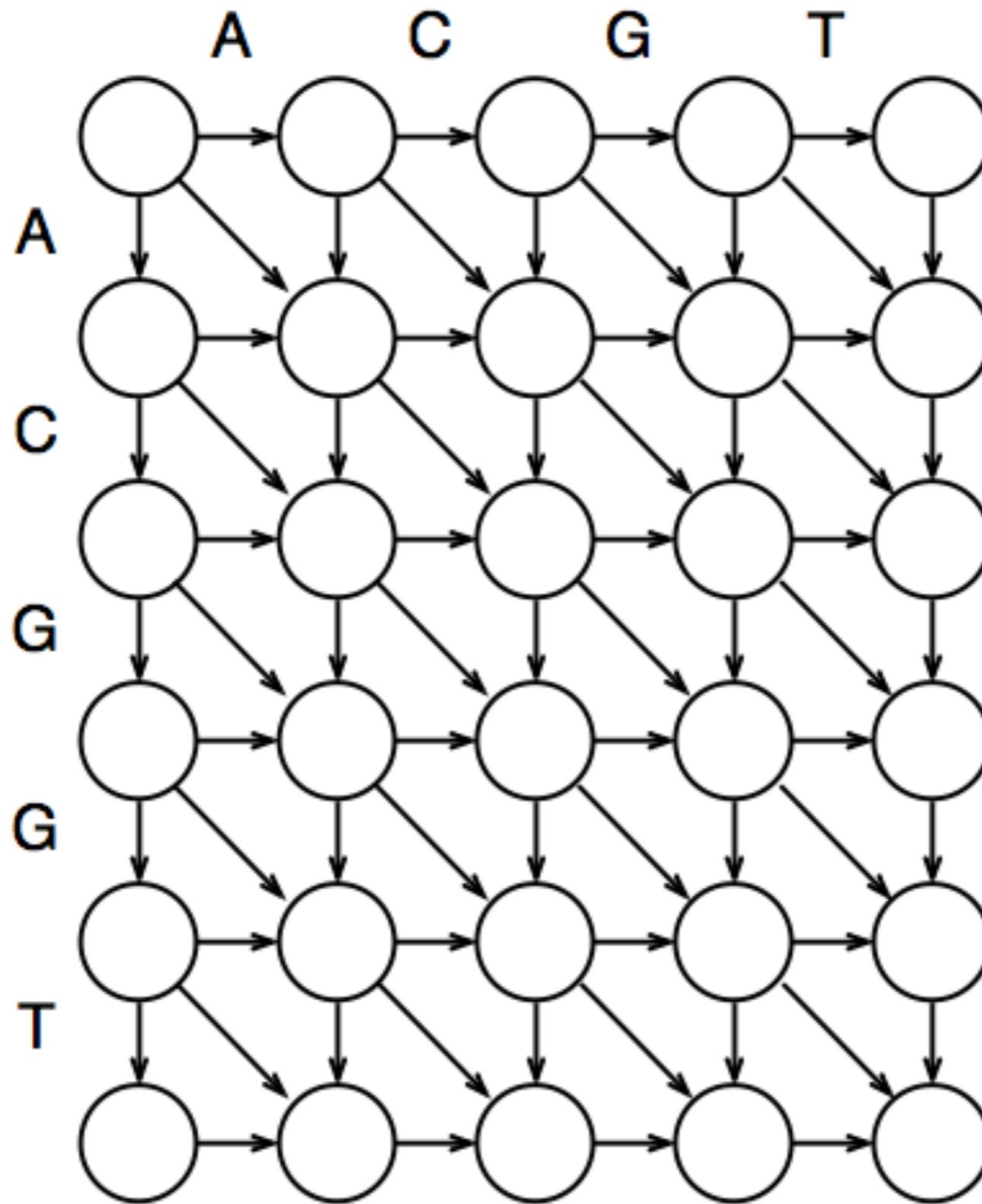
How do we measure  
sequence similarity by  
alignment and scoring  
matrices?

# Simple Alignments

Match: 1

Mismatch: -1

Gap: -2



# Simple Alignments

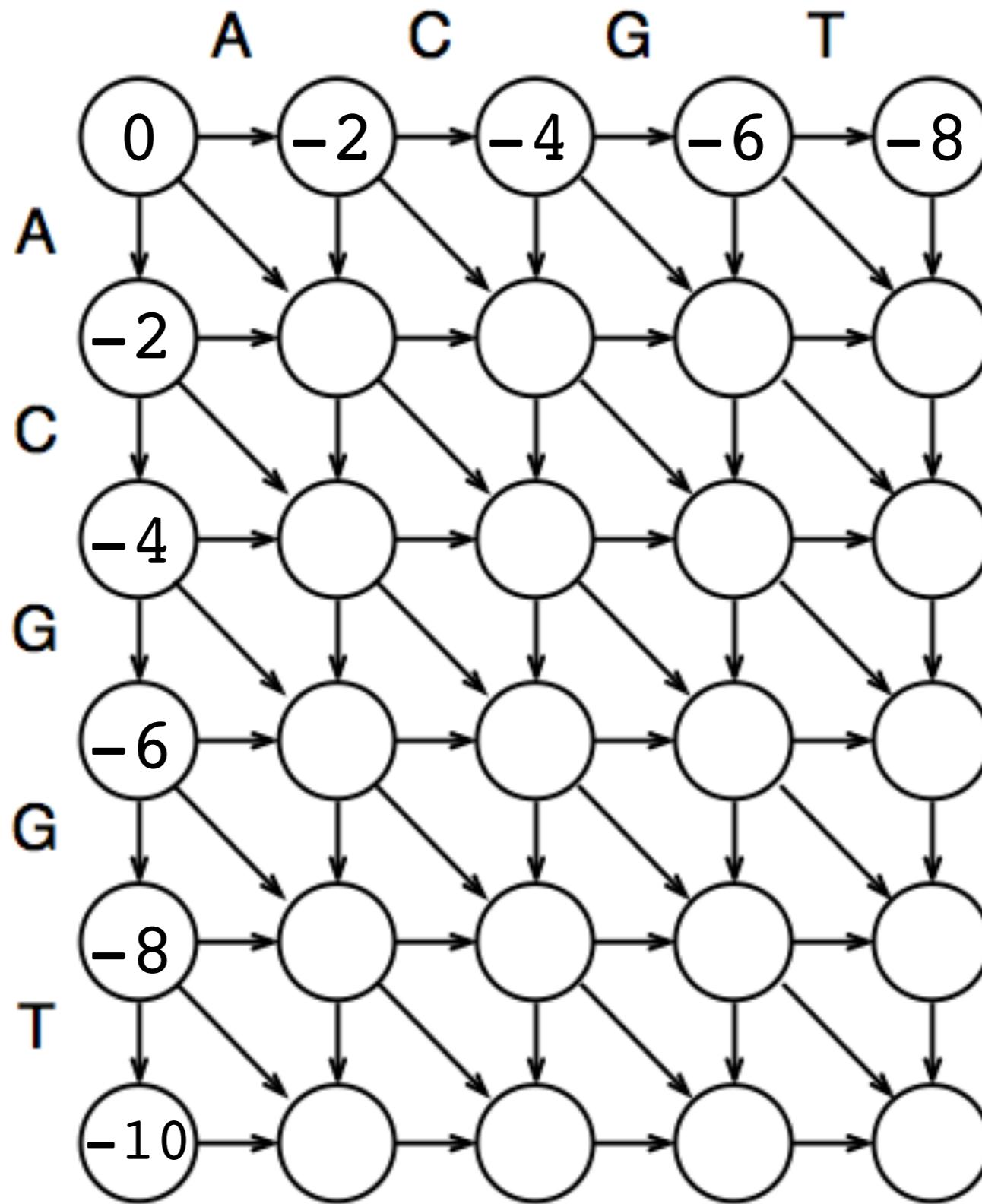
Match: 1

Mismatch: -1

Gap: -2

-----ACGT

ACGGT-----



# Simple Alignments

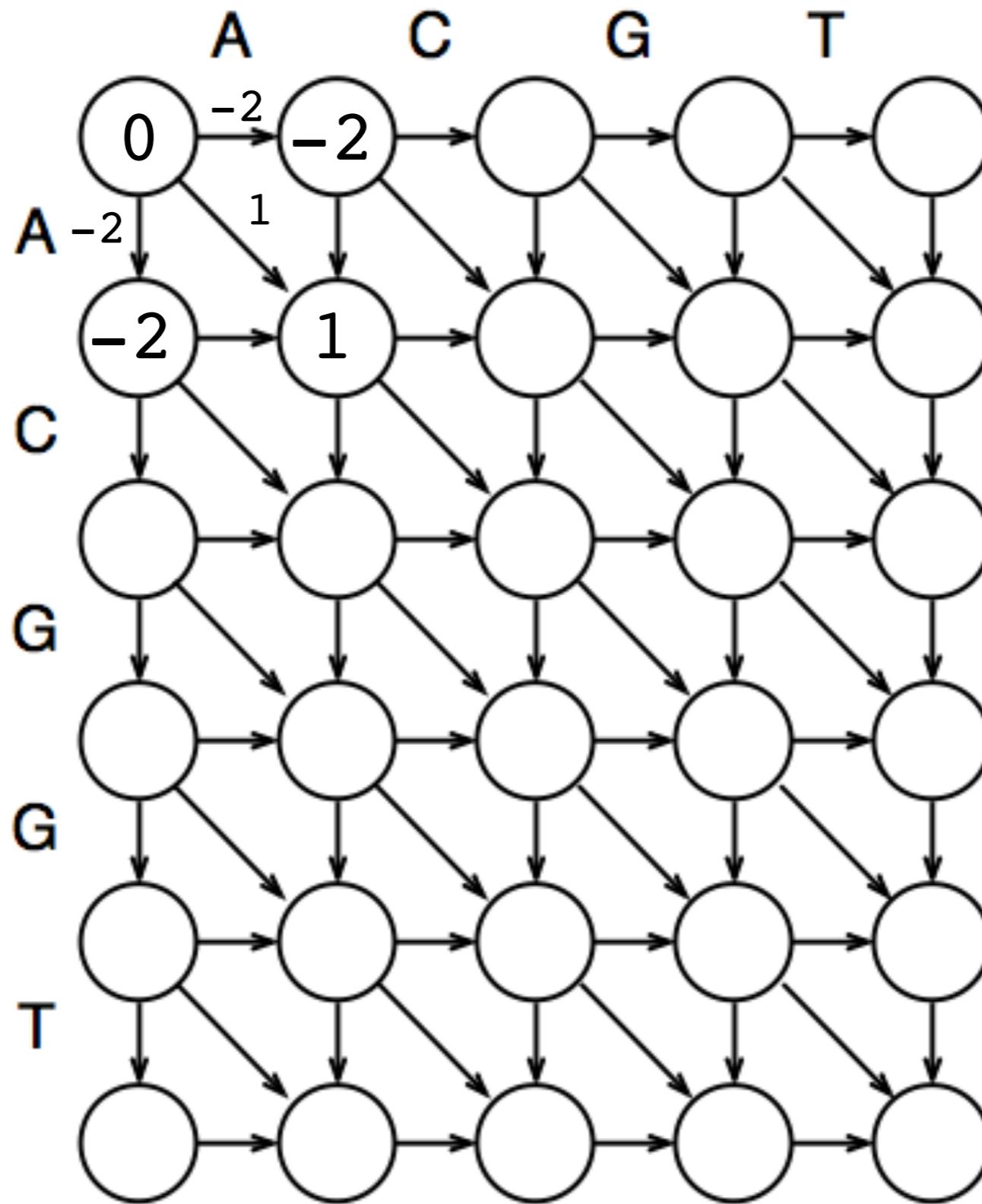
Match: 1

Mismatch: -1

Gap: -2

A

A



# Simple Alignments

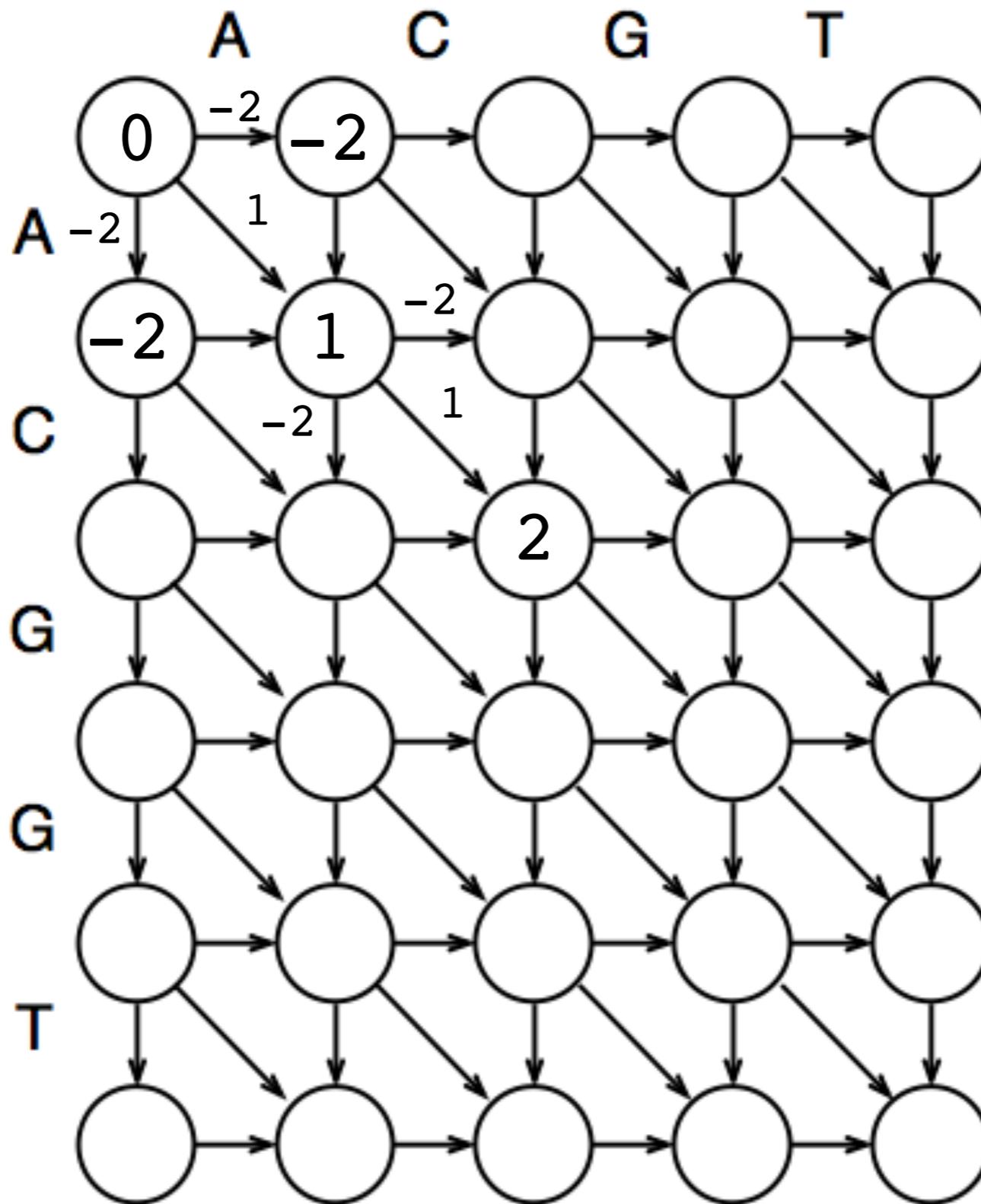
Match: 1

Mismatch: -1

**Gap:** -2

AC

AC



# Simple Alignments

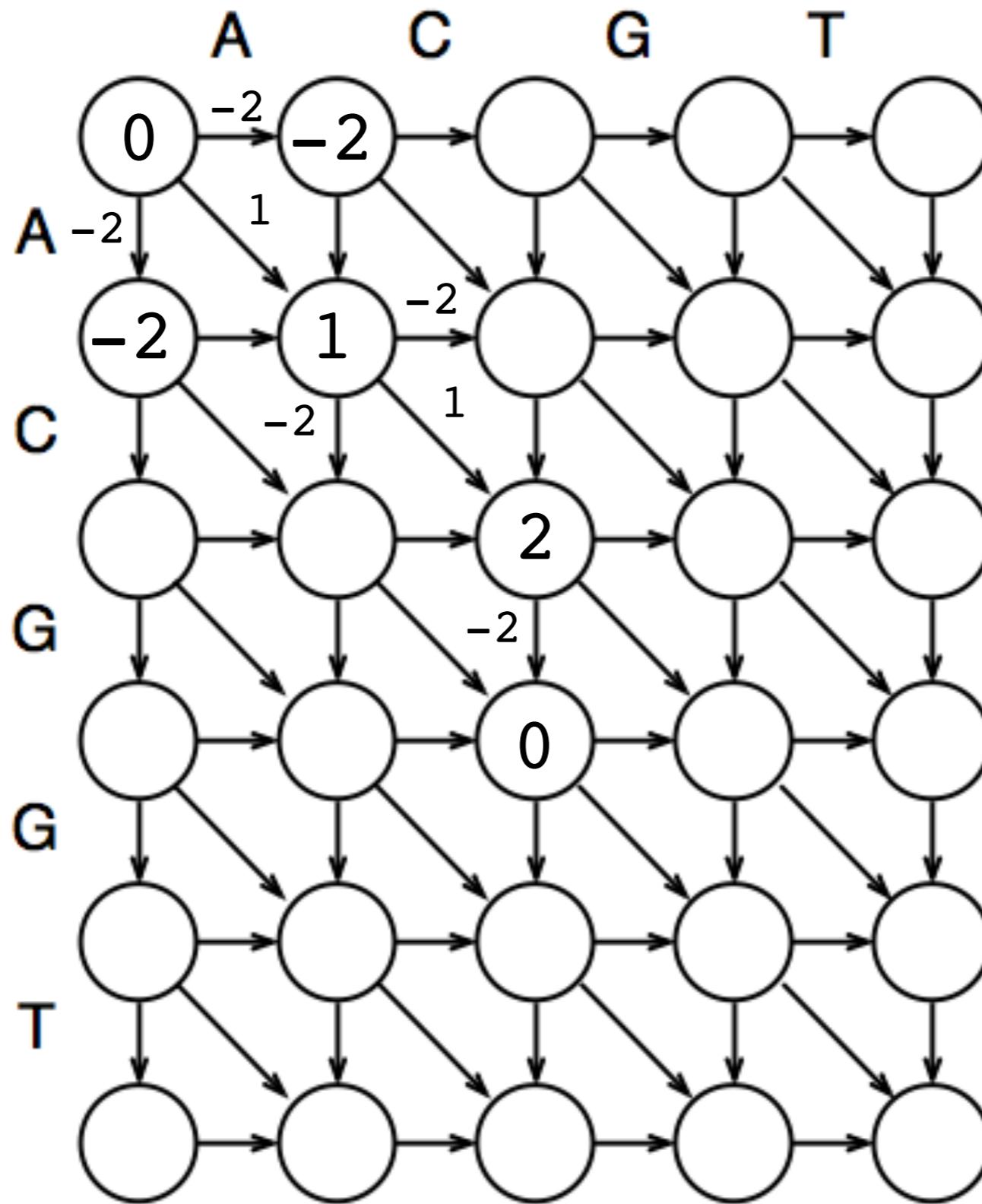
Match: 1

Mismatch: -1

Gap: -2

AC-

ACG



# Simple Alignments

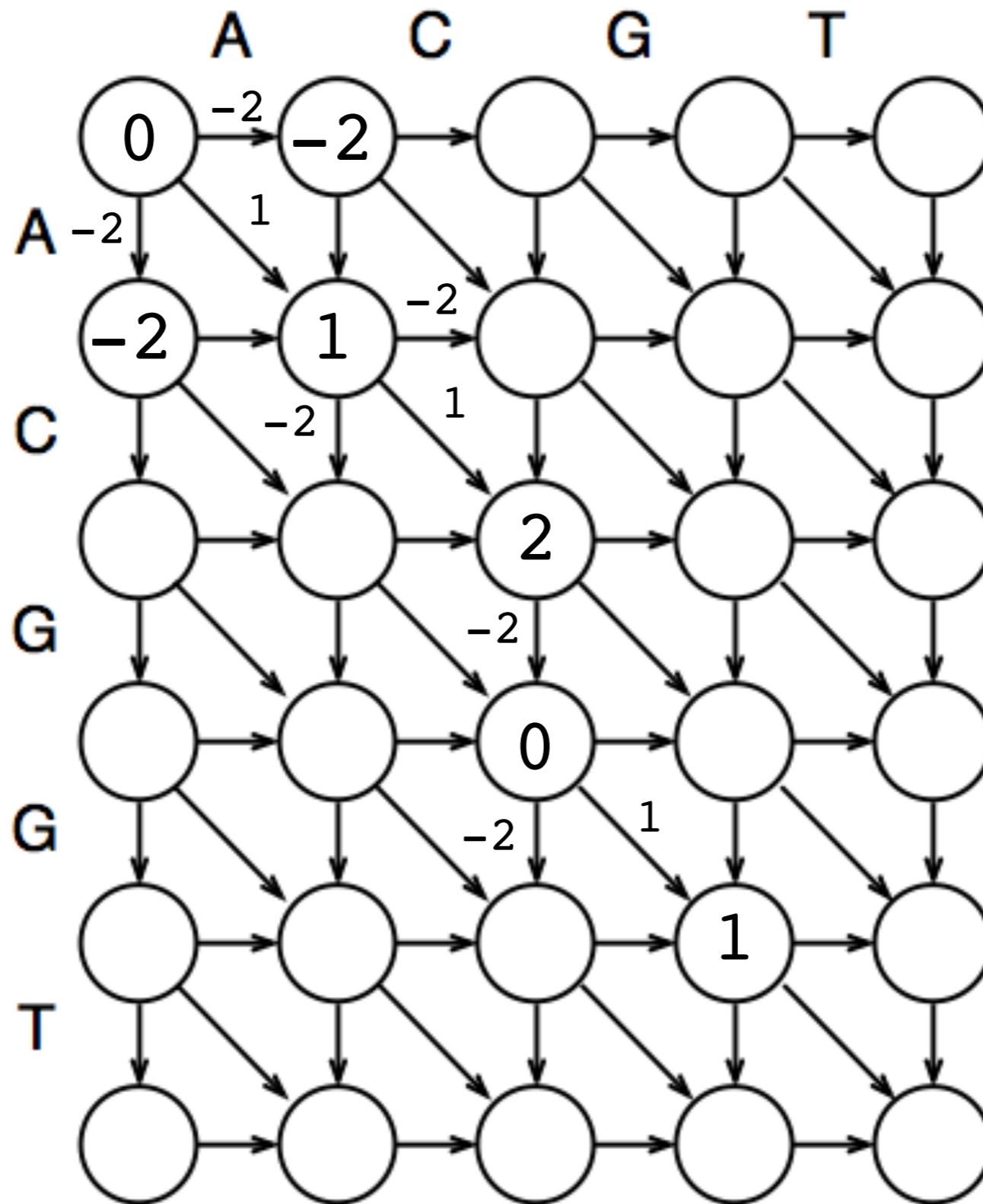
Match: 1

Mismatch: -1

Gap: -2

AC-G

ACGG



# Simple Alignments

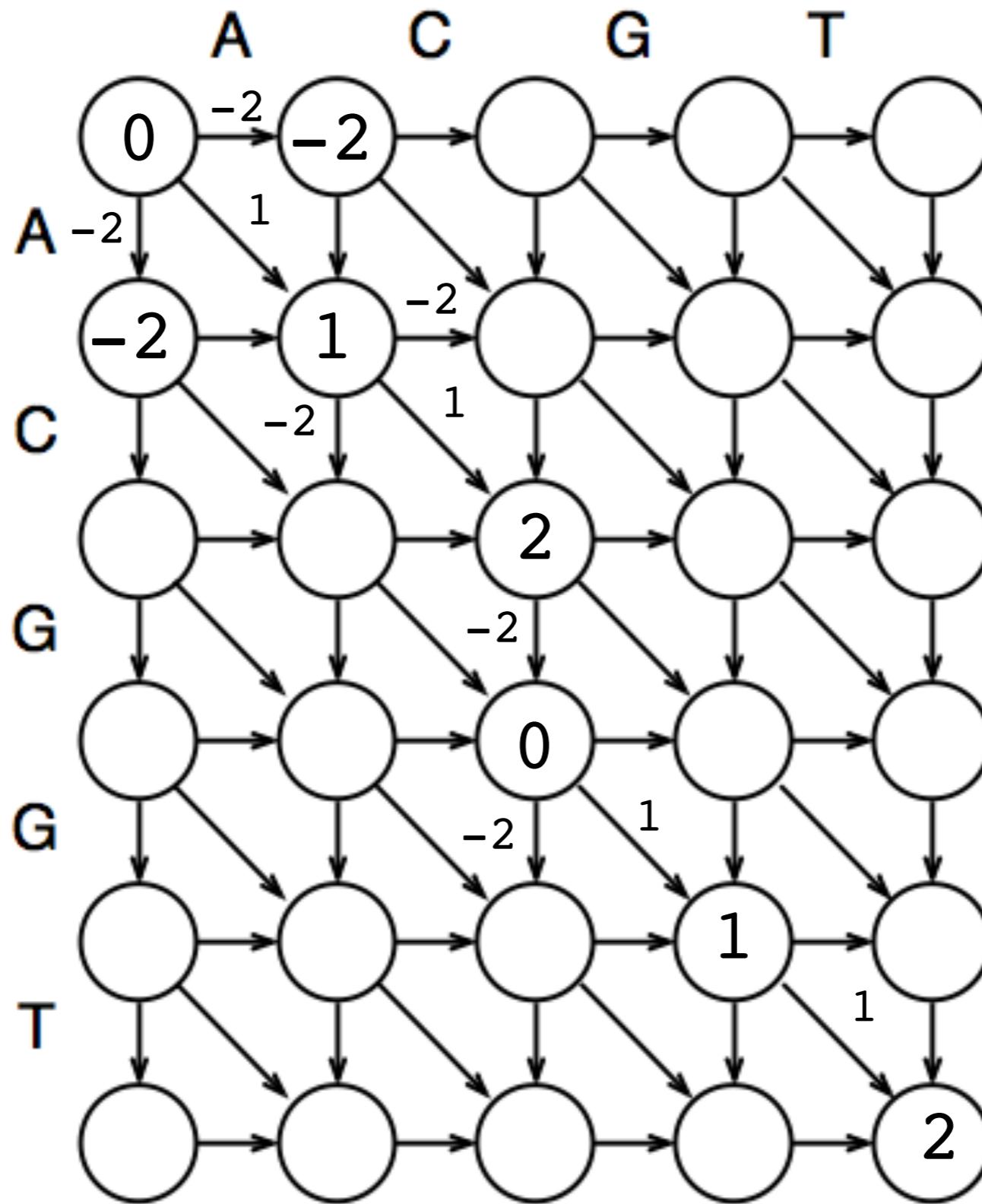
Match: 1

Mismatch: -1

**Gap:** -2

# AC-GT

**ACGGT**



# Simple Alignments

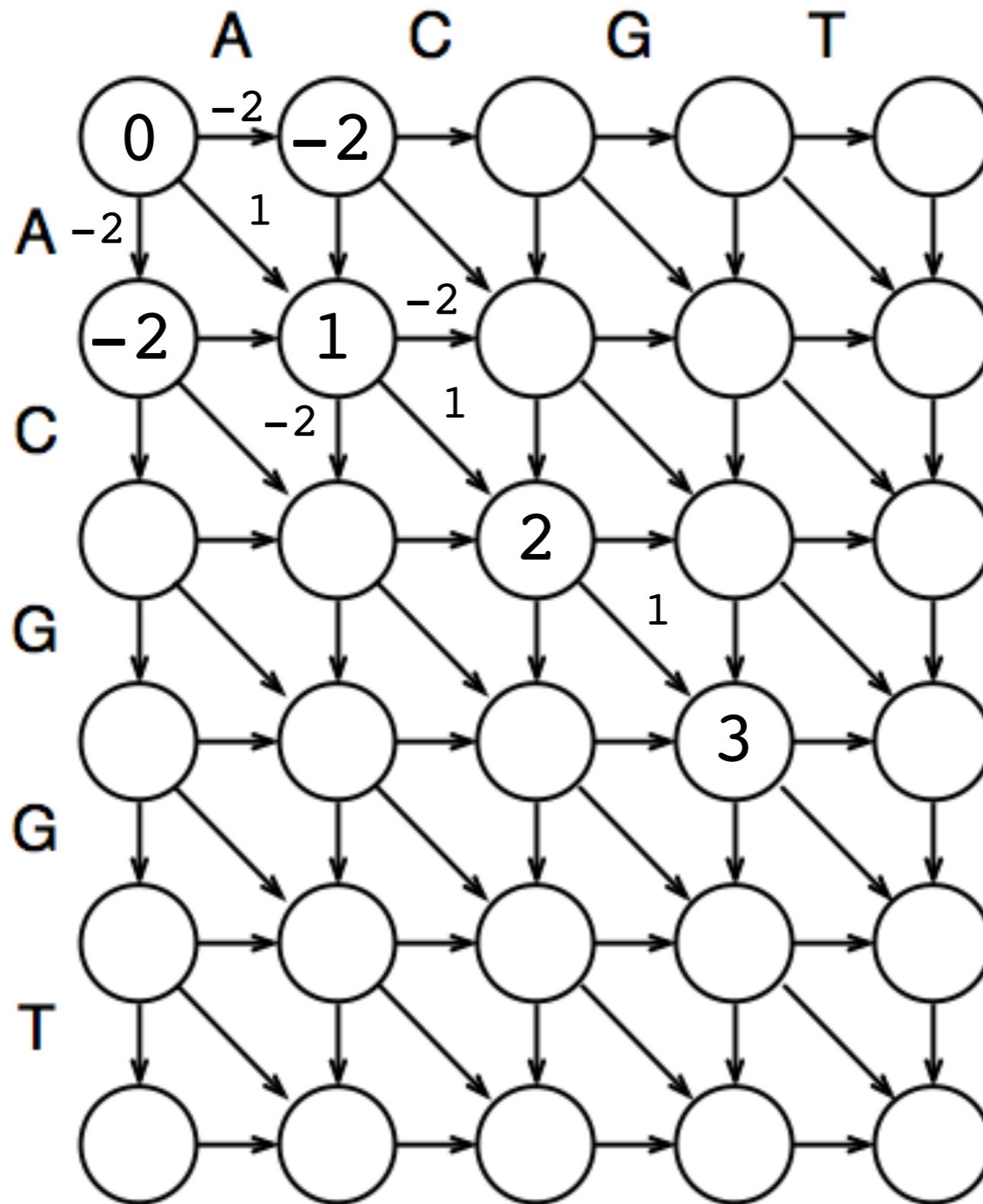
# Match: 1

Mismatch: -1

**Gap:** -2

**ACG**

ACG



# Simple Alignments

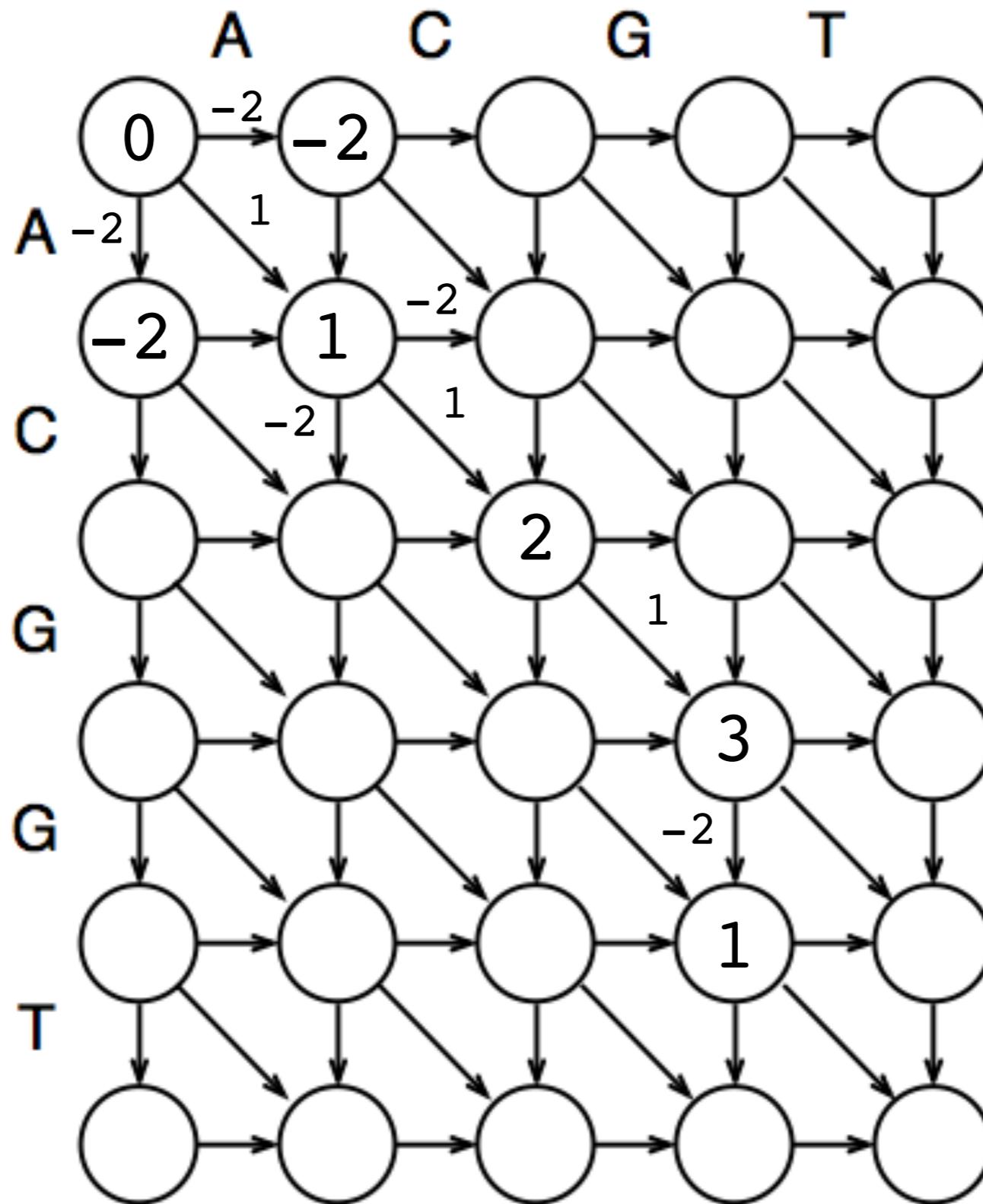
Match: 1

Mismatch: -1

Gap: -2

ACG-

ACGG



# Simple Alignments

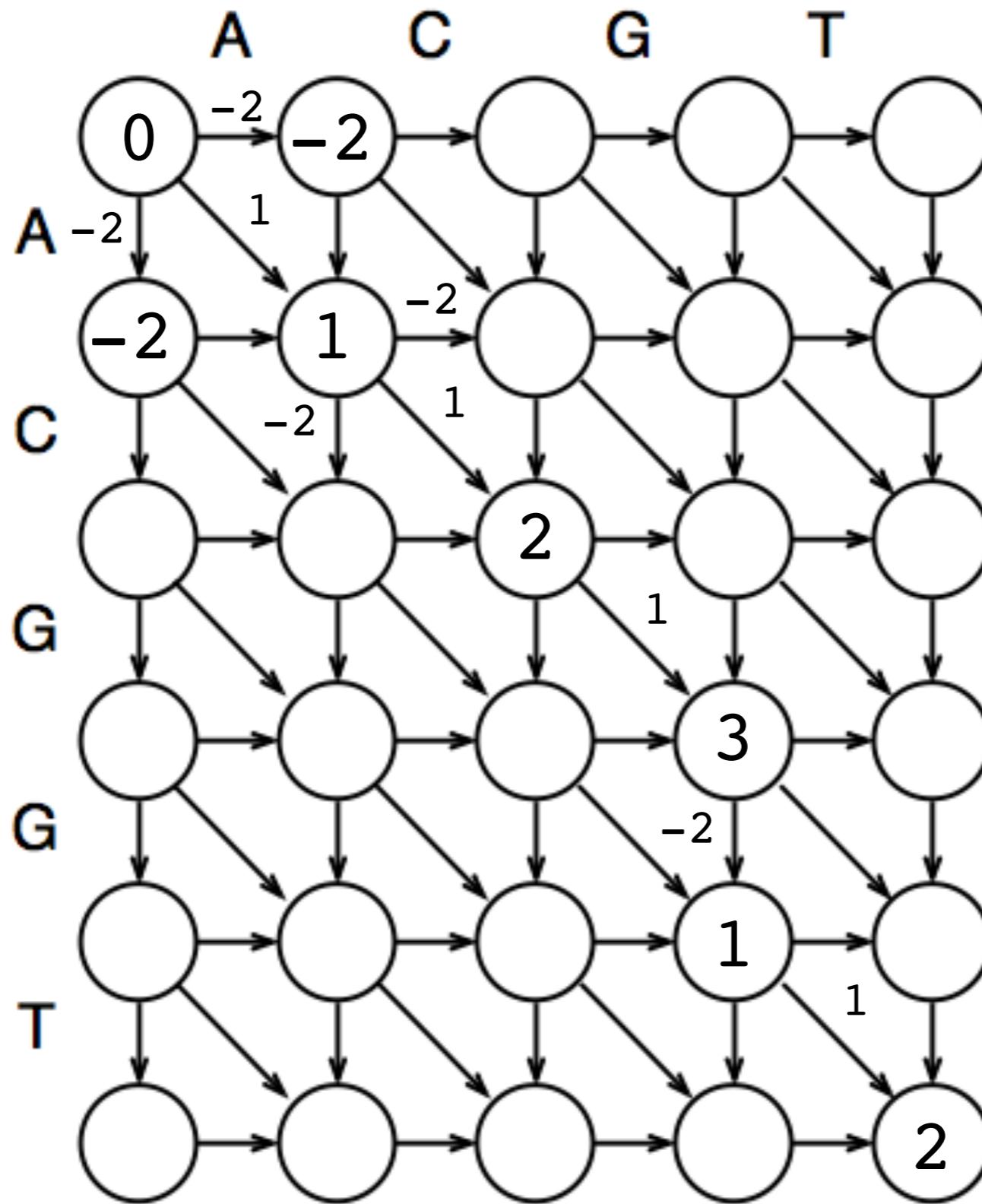
Match: 1

Mismatch: -1

Gap: -2

ACG-T

ACGGT



# Global or Local

	PM	I	L	G	Y	W	N	V	R	G	L
P:											
P:											
Y	.		:	.							
T					..						
I	:	.				:	.				.
V	...				:						.
Y	.		:	.							
F	...		..								.
P:											
V	...			:		.					
R					:						
G						:					

: = Match

. = Similar

- = Gap

## Local Alignment

PM-ILGYWNVRGL  
::: : . : . : : :  
PPYTIV-YFPVRG

## Global Alignment

-PMILGYWNVRGL  
PPYTIVYFPVRG-

# Global Alignments

## Global Alignment

-PMILGYWNVRGL  
  :   .   :   .   :   :  
PPYTIVYFPVRG-

Basis:

$$F_{0j} = d * j$$

$$F_{i0} = d * i$$

Recursion, based on the principle of optimality:

$$F_{ij} = \max(F_{i-1,j-1} + S(A_i, B_j), F_{i,j-1} + d, F_{i-1,j} + d)$$

The pseudo-code for the algorithm to compute the F matrix therefore looks like this:

```
for i=0 to length(A)
    F(i,0) ← d*i
for j=0 to length(B)
    F(0,j) ← d*j
for i=1 to length(A)
    for j=1 to length(B)
    {
        Match ← F(i-1,j-1) + S(Ai, Bj)
        Delete ← F(i-1, j) + d
        Insert ← F(i, j-1) + d
        F(i,j) ← max(Match, Insert, Delete)
    }
```

# Local Alignments

## Local Alignment

AAPMILGYWNVRGLBB  
  •••••  
DDPPYTIVYFPVRGCCCC

A matrix  $H$  is built as follows:

$$H(i, 0) = 0, \quad 0 \leq i \leq m$$

$$H(0, j) = 0, \quad 0 \leq j \leq n$$

if  $a_i = b_j$  then  $w(a_i, b_j) = w(\text{match})$  or if  $a_i \neq b_j$  then  $w(a_i, b_j) = w(\text{mismatch})$

$$H(i, j) = \max \left\{ \begin{array}{ll} 0 & \\ H(i - 1, j - 1) + w(a_i, b_j) & \text{Match/Mismatch} \\ H(i - 1, j) + w(a_i, -) & \text{Deletion} \\ H(i, j - 1) + w(-, b_j) & \text{Insertion} \end{array} \right\}, \quad 1 \leq i \leq m, 1 \leq j \leq n$$

Where:

- $a, b$  = Strings over the Alphabet  $\Sigma$
- $m = \text{length}(a)$
- $n = \text{length}(b)$
- $H(i, j)$  - is the maximum Similarity-Score between a suffix of  $a[1..i]$  and a suffix of  $b[1..j]$
- $w(c, d), c, d \in \Sigma \cup \{'-\}$ , '-' is the gap-scoring scheme

# Search Algorithms

Algorithm	Value Calculated	Scoring Matrix	Gap penalty	Time Requirement	Reference
Needleman-Wunsch	Global similarity	Any	Penalty/Gap	O(n <sup>2</sup> )	Needleman and Wunsch, 1970
Sellers	Global distance	Unity	Penalty/Gap	O(n <sup>2</sup> )	Sellers, 1974
Smith-Waterman	Local Similarity	$S_{ij} < 0.0$	Affine (q+rk)	O(n <sup>2</sup> )	Smith and Waterman, 1981 Gotoh, 1982
SRCHN	Approx. local similarity	diagonal	Penalty/Gap	O(n) – O(n <sup>2</sup> )	Wilbur and Lipman, 1983
FASTP/FASTA	Approx. local similarity	$S_{ij} < 0.0$	Limit Size (q+rk)	O(n <sup>2</sup> )/K	Lipman and Pearson, 1985, Pearson and Lipman, 1988
BLAST	Maximum Segment Score	$S_{ij} < 0.0$	Multiple Segment	O(n <sup>2</sup> )/K	Altschul et al 1990
BLAST2.0	Approx. local similarity	$S_{ij} < 0.0$	(q+rk)	O(n <sup>2</sup> )/K	Altschul et al 1997

# Scoring Matrices For Proteins

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Scoring matrices can set the evolutionary look-back time for a search

- Lower PAM (PAM10/MDM10 ... PAM60) for closer (90% - 50% identity)
- Higher BLOSUM for higher conservation (BLOSUM50 distant, BLOSUM80 conserved)
- Shallow scoring matrices for short domains/short queries (metagenomics)
  - Matrices have “bits/position” (score/position), 40 aa at 0.7 bits/position (BLOSUM62) means 28 bit max score (50 bits significant)
- Deep scoring matrices allow alignments to continue, possibly outside the homologous region

# PAM Matrices

---



- The PAM matrices were introduced by Margret Dayhoff in 1979
- They were based on 1572 observed mutations in 71 families of closely related proteins.
- Each matrix has the twenty standard amino acids in its twenty rows and columns
- The value in a given cell represents the probability of a substitution of one amino acid for another.

# Details on Scoring Matrices

Pam40

	A	R	N	D	E	I	L
A	8						
R	-9	12					
N	-4	-7	11				
D	-4	-13	3	11			
E	-3	-11	-2	4	11		
I	-6	-7	-7	-10	-7	12	
L	-8	-11	-9	-16	-12	-1	10

Pam250

	A	R	N	D	E	I	L
A	2						
R	-2	6					
N	0	0	2				
D	0	-1	2	4			
E	0	-1	1	3	4		
I	-1	-2	-2	-2	-2	5	
L	-2	-3	-3	-4	-3	2	6

$q_{ij}$  : replacement frequency at PAM40, 250

$$q_{R:N(40)} = 0.000435$$

$$p_R = 0.051$$

$$q_{R:N(250)} = 0.002193$$

$$p_N = 0.043$$

$$\text{I}_2 S_{ij} = \lg_2 (q_{ij}/p_i p_j) \quad \text{I}_e S_{ij} = \ln(q_{ij}/p_i p_j) \quad p_R p_N = 0.002193$$

$$\text{I}_2 S_{R:N(40)} = \lg_2 (0.000435/0.00219) = -2.333$$

$$\text{I}_2 = 1/3; S_{R:N(40)} = -2.333/\text{I}_2 = -7$$

$$\text{I}_2 S_{R:N(250)} = \lg_2 (0.002193/0.002193) = 0$$

# PAM Matrices

$$\lambda S = \log\left(\frac{q_{ij}}{p_i p_j}\right)$$

- S is the replacement score of i to j
- $\lambda$  term is used to scale the matrix so that individual scores can be accurately represented with integers
- $q_{ij}$  is Replacement frequency of i to j
- $p_i$  is the expected frequency of i

**Table 1:** Relative mutabilities and the distribution of amino acids in M. Dayhoff's database of observed amino acid changes.

		$mut_i$	$f_i$			$mut_i$	$f_i$
Ala	A	100	0.087	Leu	L	40	0.085
Arg	R	65	0.041	Lys	K	56	0.081
Asn	N	134	0.040	Met	M	94	0.015
Asp	D	106	0.047	Phe	F	41	0.040
Cys	C	20	0.033	Pro	P	56	0.051
Gln	Q	93	0.038	Ser	S	120	0.070
Glu	E	102	0.050	Thr	T	97	0.058
Gly	G	49	0.089	Trp	W	18	0.010
His	H	66	0.034	Tyr	Y	41	0.030
Ile	I	96	0.037	Val	V	20	0.065

- Scoring matrices can be designed for different evolutionary distances (less=shallow; more=deep)
- Deep matrices allow more substitution

PAM1: Predicts one mutation per 100 aa

PAM40: Predicts 40 mutations per 100 aa

PAM250: Predicts 250 mutations per 100 aa

# Details on Scoring Matrices

---

## PAM

- Evolutionary model - extrapolated from PAM1
- PAM20: 20% change (mammals)
- PAM250: 250% change (<20% identity)
- Gap penalties should vary
- shallow matrices (PAM10-40) for short sequences and short distances

## BLOSUM

- Empirically determined, no extrapolation (no model)
- BLOSUM45-50 - distant (1/3 bits)
- BLOSUM80 - very highly conserved (not small change), high info/position
- BLOSUM62 - 1/2 bits

### PAM : BLOSUM

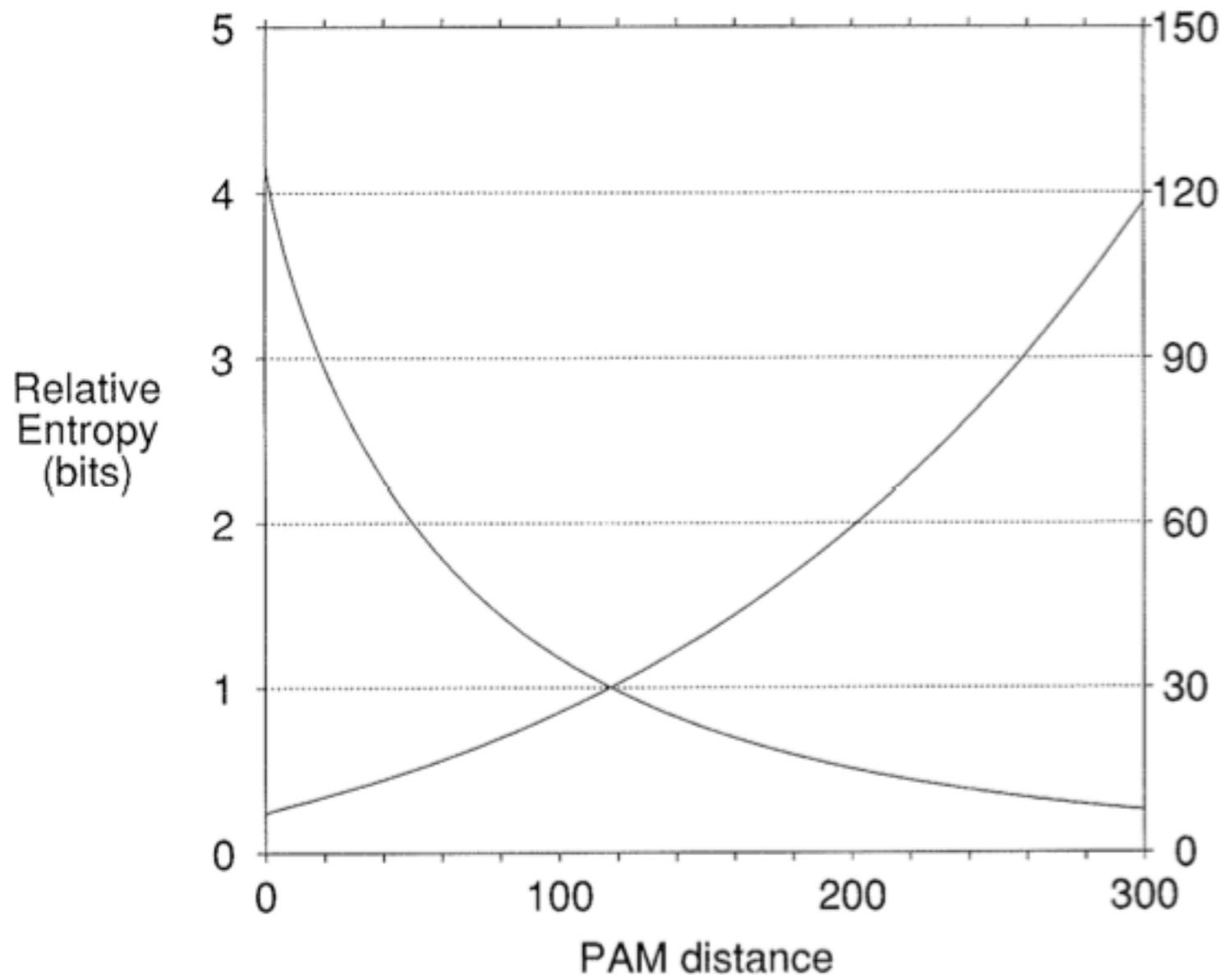
PAM100 :	BLOSUM90
PAM120 :	BLOSUM80
PAM160 :	BLOSUM60
PAM200 :	BLOSUM52
PAM250 :	BLOSUM45

# Scoring Matrices

---

- PAM and BLOSUM matrices greatly improve the sensitivity of protein sequence comparison – low identity with significant similarity
- PAM matrices have an evolutionary model - lower number, less divergence – lower=closer; higher=more distant
- BLOSUM matrices are sampled from conserved regions at different average identity – higher=more conservation
- Short alignments require shallow matrices (closer)
- Shallow matrices set maximum look-back time

# Details on Scoring Matrices

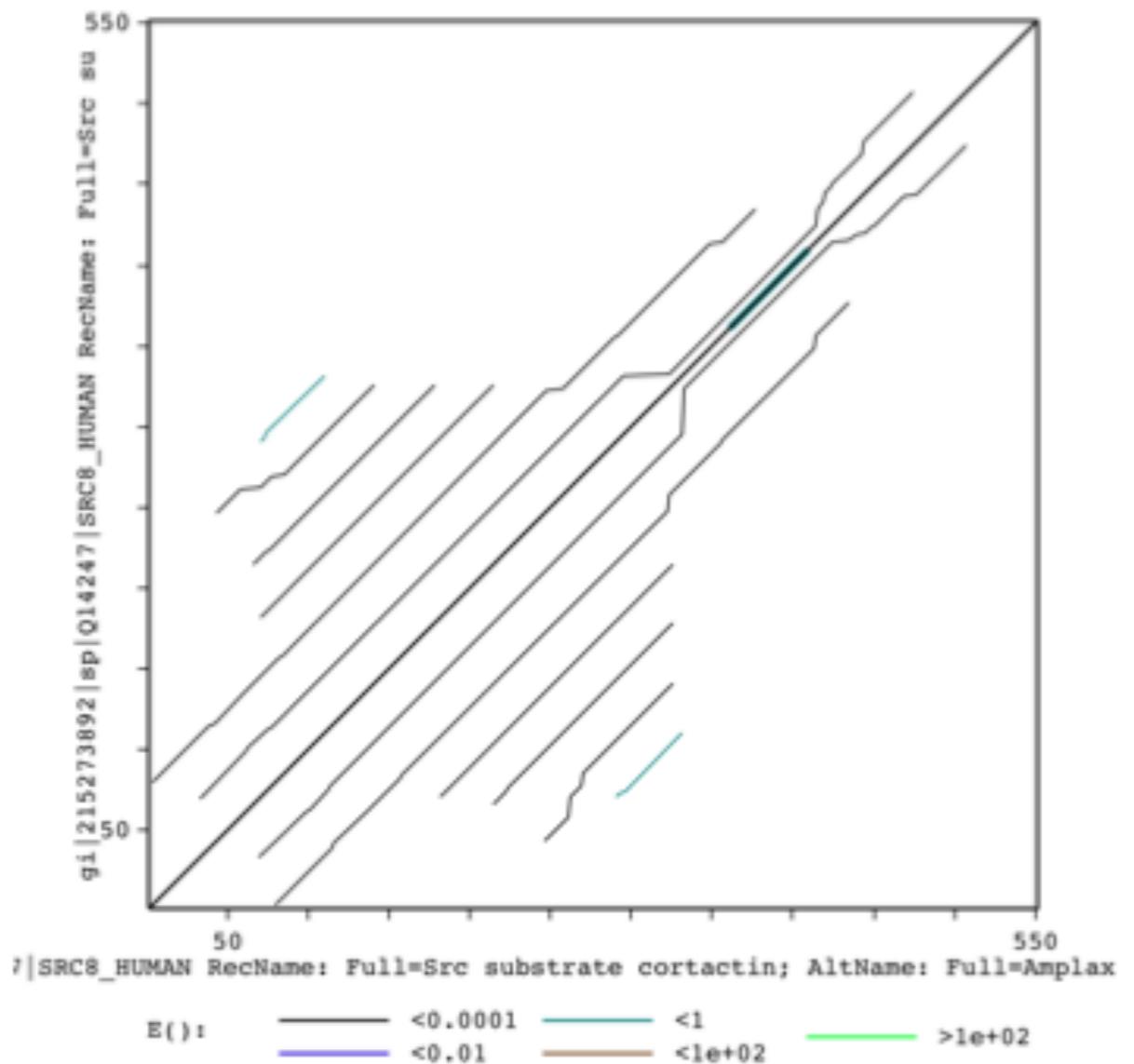


As sequences diverge,  
there is less information  
per position

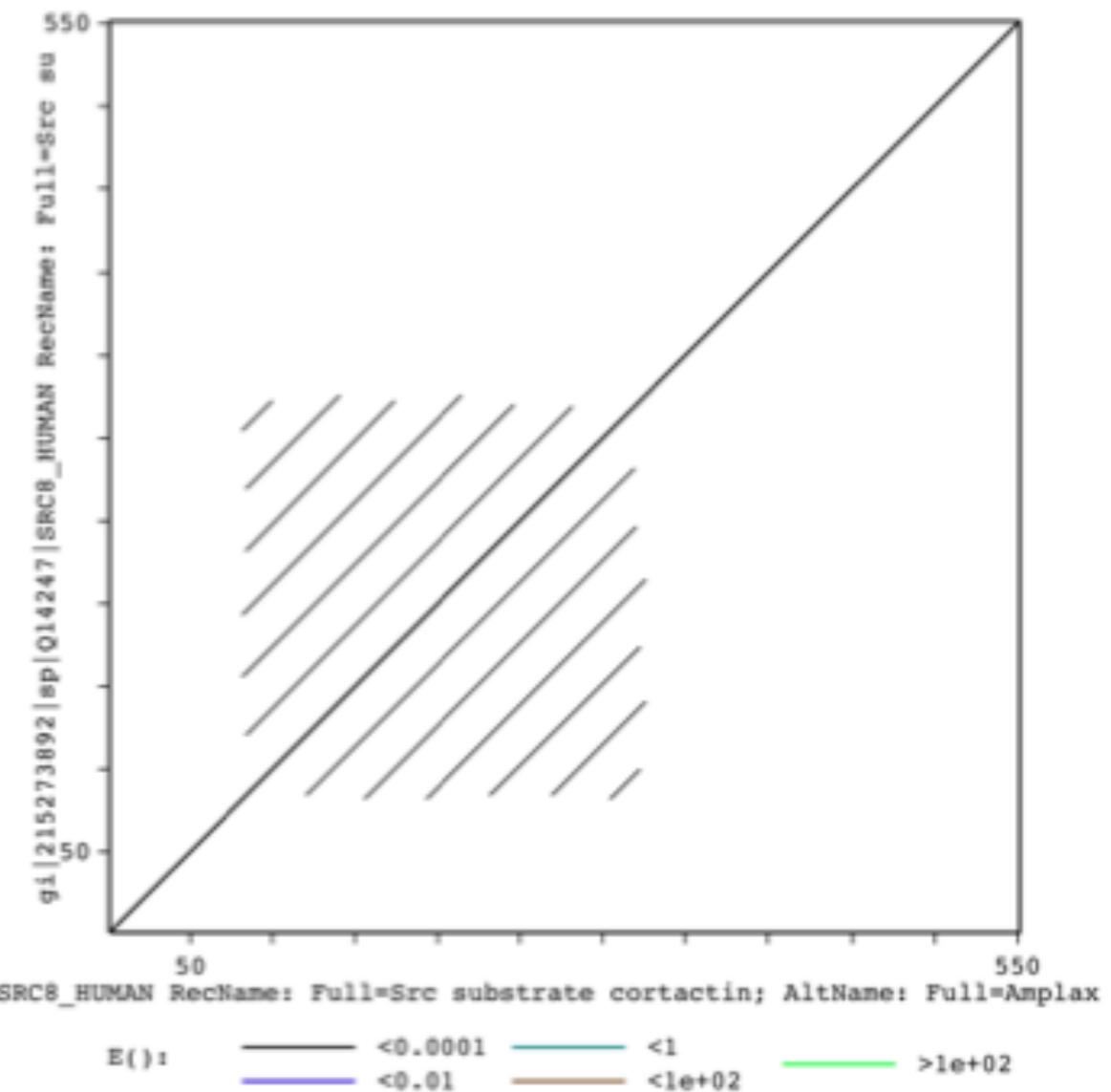
As sequences diverge,  
longer alignments are  
required to contain the  
score threshold

# Stringent Score Leads to Short Alignments

BLOSUM62 -11/-1



MD20 -26/-4



# Scoring Alignments

```
>>sp|P07925|ATP6_MAIZE ATP synthase a chain (ATPase protein 6) (291 aa)
initn: 96 initl: 56 opt: 116 Z-score: 161.2 bits: 37.6 E(13351): 0.0048
Smith-Waterman score: 175; 24.7% identity (57.9% similar) in 247 aa overlap (16-251:31-259)
Entrez Lookup Re-search database General re-search
          10      20      30      40      50      60
gi|231      MKIVLYYFVNMFISGIFQIANVEVGQHFYWSILGFQIHGQLINSWIVILIIGFLSIYTTKNL-
          :: : ..... : .. .... .:...: .:..: ..:..: ..:..: ..:..:
sp|P07 MERNGEivnnngsiiipggggpvTESPLDQFGIHPILDLNIGK-YYVSFTnls1--smllt1glvlllv-f--vvtkkggg
          10      20      30      40      50      60      70
          70      80      90     100     110     120     130     140
gi|231 TLVPANKQIFIELVTEFITDISKTQIGEKEYS---KWPYIGTMFLFIFVSNWSGALIPWKIIELPNGELGAPTNDINTT
          :: : ..::.. .:.. .:..: .. .:..: ..:..: ..:..: ..:..: ..:..:
sp|P07 ksvPNAFQSLVELIYDFVPNLVNEQIGGLSGNVKHKKFFPCISVTFTFSLFRNPQG-MIPFSF-----TVTSHFLIT
          80      90     100     110     120     130      140
          150     160     170     180     190     200     210
gi|231 AGLAILTSLAYFYAGLNKKGLTYFKKYVQPTPILPIN---ILEDFT---KPLSLSFRLFGNILADELVVAVLVSLVPL
          :... .. .:..:..: ..:..: ..:..: ..:..: ..:..: ..:..: ..:..: ..:..: ..:..:
sp|P07 LALSFSIFIGITIVGFQRHGLHFFS-f11pagvp1plapflvlle1ISHCFRALSSGIRLFANMMAGHSSVKILSGFAWT
          150     160     170     180     190     200     210     220
          220     230     240     250
gi|231 IVPVPLIFLGLFTSGIQALIFATLSGSYIGEAMEGHH
          ... : : .. : ..: ..: ..: ..: ..:
sp|P07 MLFLNNIFYFLGDLGPLFIVLA-LTGLELGVAISQAHVSTISICIYLNDATNLHQNESFHNCIKTRSQS
          230     240     250     260     270     280     290
```

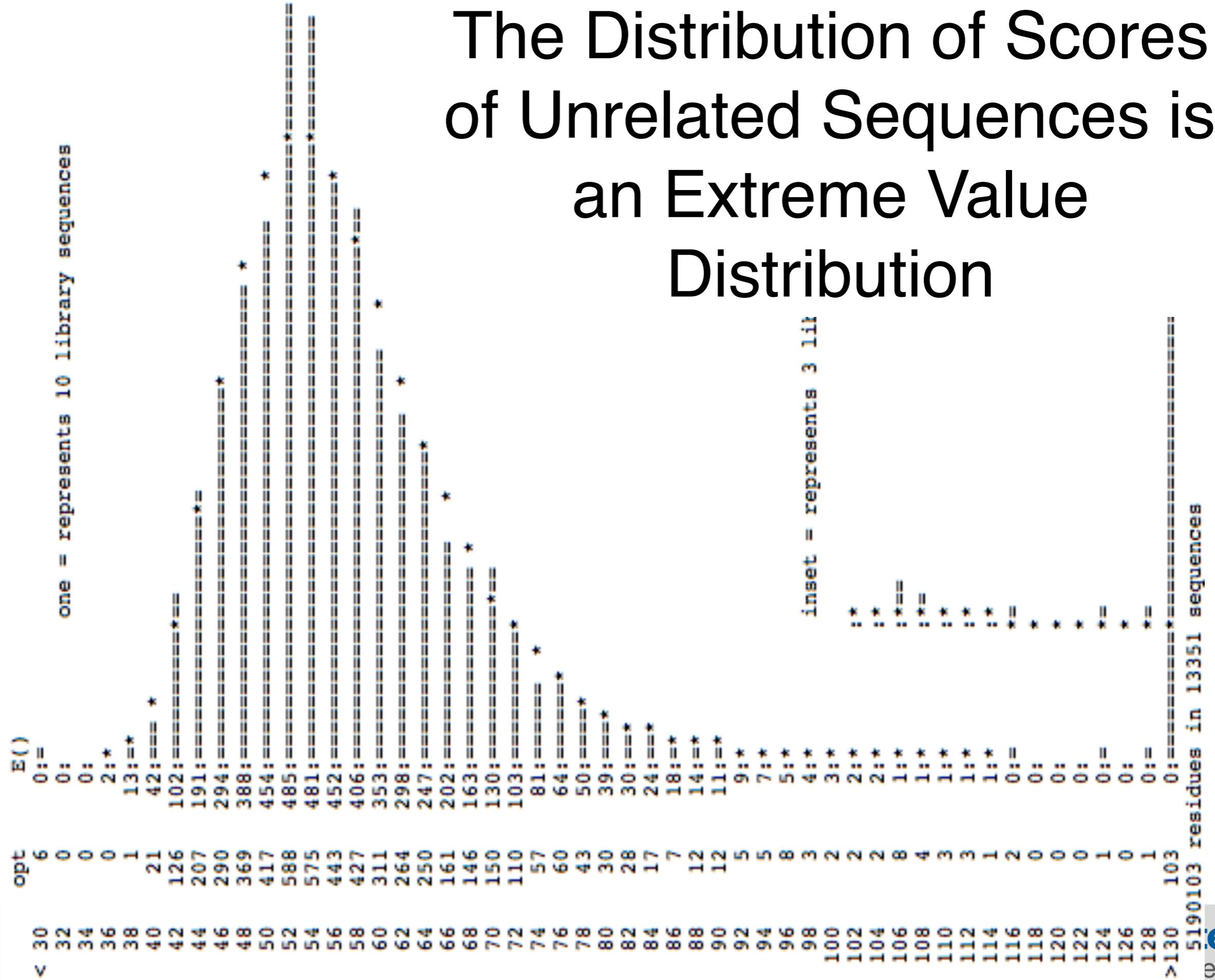
Alignments are scored using the scoring matrix

# Inferring Homology from Statistical Significance

---

- Real *UNRELATED* sequences have similarity scores that are indistinguishable from *RANDOM* sequences
- If a similarity is NOT *RANDOM*, then it must be NOT *UNRELATED*
- Therefore, NOT *RANDOM* (statistically significant) similarity must reflect *RELATED* sequences

# The Distribution of Scores of Unrelated Sequences is an Extreme Value Distribution



# What is an Expectation Value

---

- The Expectation Values is the probability of the score times the number of sequences in your search library
  - The number of times you expect to get that p-value by chance in the search that was performed.

## Library Size

```
>>sp|P07925|ATP6_MAIZE ATP synthase a chain (ATPase protein 6) (291 aa)
initn: 96 initl: 56 opt: 116 Z-score: 161.2 bits: 37.6 E(13351): 0.0048
Smith-Waterman score: 175; 24.7% identity (57.9% similar) in 247 aa overlap (16-251:31-259)
Entrez Lookup Re-search database General re-search
```

# Highest Scoring Unrelated Sequenced E() ~ 1

---

The best scores are:										
				s-w	bits	E(13351)	%_id	%_sim	alen	
sp P26205 BGLT_TRIRP	Cyanogenic beta-glucosidase precur	( 425)	1187	281.9	4.3e-76	0.452	0.763	392	<a href="#">align</a>	
sp P26204 BGLS_TRIRP	Non-cyanogenic beta-glucosidase pr	( 493)	1179	279.9	1.9e-75	0.406	0.704	497	<a href="#">align</a>	
sp P11546 LACG_LACLA	6-phospho-beta-galactosidase (Beta	( 468)	712	171.6	7.5e-43	0.326	0.603	494	<a href="#">align</a>	
sp P12614 BGLS_AGRSA	Beta-glucosidase (Gentiobiase) (Ce	( 459)	699	168.6	5.9e-42	0.302	0.590	483	<a href="#">align</a>	
sp P31835 CDGT2_PAEMA	Cyclomaltodextrin glucanotransfer	( 713)	110	31.7	1.5	0.251	0.561	187	<a href="#">align</a>	
sp P26537 VL1 HPV5B	Major capsid protein L1	( 525)	106	30.9	1.9	0.245	0.504	139	<a href="#">align</a>	
sp P02667 CS2LA_RAT	Alpha-S2-casein-like A precursor (C	( 179)	97	29.2	2.1	0.288	0.652	66	<a href="#">align</a>	
sp Q03763 DSG1_BOVIN	Desmoglein-1 precursor (Desmosomal	(1043)	109	31.3	2.8	0.206	0.497	286	<a href="#">align</a>	
sp P09282 UL32_VZVD	Probable major envelope glycoprotei	( 585)	101	29.7	4.8	0.237	0.568	118	<a href="#">align</a>	
sp Q92040 ANX12_COLLI	Annexin A1 isoform p37 (Annexin I	( 343)	96	28.7	5.5	0.251	0.508	179	<a href="#">align</a>	
sp P16330 CN37_MOUSE	2',3'-cyclic-nucleotide 3'-phospho	( 420)	97	28.9	6.1	0.227	0.529	172	<a href="#">align</a>	
ref NP_276832.1	transcriptional regulator Icc related	( 262)	91	27.7	8.8	0.285	0.455	123	<a href="#">align</a>	

# Highest Scoring Unrelated Protein

The best scores are:

				opt bits	E(13351)	%_id	%_sim	alen	
sp P00846 ATP6_HUMAN	ATP synthase a chain (ATPase prote	( 226)	1124	289.8	4.1e-79	1.000	1.000	226	<a href="#">align</a>
sp P00847 ATP6_BOVIN	ATP synthase a chain (ATPase prote	( 226)	1075	277.5	2e-75	0.779	0.951	226	<a href="#">align</a>
sp P00848 ATP6_MOUSE	ATP synthase a chain (ATPase prote	( 226)	1057	273.0	4.5e-74	0.757	0.916	226	<a href="#">align</a>
sp P00849 ATP6_XENLA	ATP synthase a chain (ATPase prote	( 226)	499	133.4	4.7e-32	0.533	0.847	229	<a href="#">align</a>
sp P00854 ATP6_YEAST	ATP synthase a chain precursor (AT	( 259)	357	97.9	2.7e-21	0.353	0.694	232	<a href="#">align</a>
sp P00851 ATP6_DROYA	ATP synthase a chain (ATPase prote	( 224)	323	89.4	8.3e-19	0.378	0.721	222	<a href="#">align</a>
ref NP_008281.1 ATP6_10704	ATP synthase F0 subunit 6 [D	( 224)	321	88.9	1.2e-18	0.375	0.710	224	<a href="#">align</a>
sp P00852 ATP6_EMENI	ATP synthase a chain precursor (AT	( 256)	266	75.1	1.9e-14	0.304	0.691	230	<a href="#">align</a>
sp P14862 ATP6_COCHE	ATP synthase a chain (ATPase prote	( 257)	221	63.8	4.7e-11	0.313	0.650	214	<a href="#">align</a>
sp P68526 ATP6_TRITI	ATP synthase a chain (ATPase prote	( 386)	204	59.5	1.5e-09	0.289	0.651	235	<a href="#">align</a>
sp P05499 ATP6_TOBAC	ATP synthase a chain (ATPase prote	( 395)	185	54.7	4e-08	0.283	0.635	233	<a href="#">align</a>
sp P07925 ATP6_MAIZE	ATP synthase a chain (ATPase prote	( 291)	182	54.0	4.7e-08	0.311	0.667	180	<a href="#">align</a>
sp P0AB98 ATP6_ECOLI	ATP synthase a chain (ATPase prote	( 271)	166	50.1	7e-07	0.233	0.585	236	<a href="#">align</a>
sp P15993 AROP_ECOLI	Aromatic amino acid transport prot	( 457)	103	34.2	0.072	0.234	0.622	111	<a href="#">align</a>
sp P27178 ATP6_SYNYY3	ATP synthase a chain (ATPase prote	( 276)	92	31.5	0.27	0.265	0.571	170	<a href="#">align</a>
sp P00329 ADH1_MOUSE	Alcohol dehydrogenase 1 (Alcohol d	( 375)	89	30.7	0.64	0.344	0.607	61	<a href="#">align</a>
sp P06757 ADH1_RAT	Alcohol dehydrogenase 1 (Alcohol deh	( 376)	85	29.7	1.3	0.339	0.629	62	<a href="#">align</a>
sp P00161 CYB_EMENI	Cytochrome b	( 387)	83	29.2	1.9	0.308	0.593	91	<a href="#">align</a>
sp P29631 CYB_POMTE	Cytochrome b	( 308)	81	28.8	2	0.274	0.584	113	<a href="#">align</a>
sp P00328 ADH1S_HORSE	Alcohol dehydrogenase S chain	( 374)	82	29.0	2.2	0.328	0.590	61	<a href="#">align</a>
sp P00327 ADH1E_HORSE	Alcohol dehydrogenase E chain	( 375)	82	29.0	2.2	0.328	0.590	61	<a href="#">align</a>
sp P11599 HLYB_PROVU	Alpha-hemolysin translocation ATP-	( 707)	86	29.8	2.3	0.277	0.625	112	<a href="#">align</a>
sp P03880 ANI1_EMENI	Intron-encoded DNA endonuclease I-	( 488)	83	29.1	2.5	0.389	0.630	54	<a href="#">align</a>
sp P07327 ADH1A_HUMAN	Alcohol dehydrogenase 1A (Alcohol	( 375)	79	28.2	3.6	0.265	0.556	117	<a href="#">align</a>
sp P41680 ADH1_PERMA	Alcohol dehydrogenase 1 (Alcohol d	( 375)	79	28.2	3.6	0.241	0.583	108	<a href="#">align</a>
sp P24956 CYB_EQUGR	Cytochrome b	( 379)	79	28.2	3.7	0.315	0.576	92	<a href="#">align</a>
sp P10724 ALR_BACST	Alanine racemase	( 388)	79	28.2	3.8	0.233	0.535	86	<a href="#">align</a>
sp P03046 CIM_BPMU	Cim protein (Kil protein)	( 74)	66	25.4	5.1	0.208	0.623	53	<a href="#">align</a>
sp P72588 DNLJ_SYNYY3	DNA ligase (Polydeoxyribonucleotid	( 669)	81	28.6	5.1	0.250	0.570	128	<a href="#">align</a>

# Unrelated or Too Distance

The best scores are:

			opt bits	E(13351)	%_id	%_sim	alen	
sp P0AB98 ATP6_ECOLI	ATP synthase a chain (ATPase prote	( 271)	1650	428.4	1.1e-120	1.000	1.000	271 <a href="#">align</a>
sp P06451 ATPI_SPIOL	Chloroplast ATP synthase a chain p	( 247)	161	49.1	1.5e-06	0.270	0.616	211 <a href="#">align</a>
sp P06289 ATPI_MARPO	Chloroplast ATP synthase a chain p	( 248)	161	49.1	1.5e-06	0.261	0.621	211 <a href="#">align</a>
sp P06452 ATPI_PEA	Chloroplast ATP synthase a chain pre	( 247)	158	48.3	2.6e-06	0.274	0.614	223 <a href="#">align</a>
sp P69371 ATPI_ATRBE	Chloroplast ATP synthase a chain p	( 247)	156	47.8	3.7e-06	0.270	0.607	211 <a href="#">align</a>
sp P00848 ATP6_MOUSE	ATP synthase a chain (ATPase prote	( 226)	149	46.0	1.2e-05	0.259	0.617	193 <a href="#">align</a>
sp P00846 ATP6_HUMAN	ATP synthase a chain (ATPase prote	( 226)	148	45.7	1.4e-05	0.237	0.589	236 <a href="#">align</a>
sp P30391 ATPI_EUGGR	Chloroplast ATP synthase a chain p	( 251)	139	43.4	7.6e-05	0.298	0.596	225 <a href="#">align</a>
sp P00847 ATP6_BOVIN	ATP synthase a chain (ATPase prote	( 226)	138	43.2	8.1e-05	0.233	0.581	236 <a href="#">align</a>
sp P0C2Y5 ATPI_ORYSA	Chloroplast ATP synthase a chain p	( 247)	132	41.7	0.00026	0.259	0.603	239 <a href="#">align</a>
sp P68526 ATP6_TRITI	ATP synthase a chain (ATPase prote	( 386)	121	38.9	0.0028	0.259	0.603	239 <a href="#">align</a>
sp P27178 ATP6_SYN3	ATP synthase a chain (ATPase prote	( 276)	116	37.6	0.0048	0.264	0.578	258 <a href="#">align</a>
sp P00854 ATP6_YEAST	ATP synthase a chain precursor (AT	( 259)	113	36.8	0.0077	0.235	0.578	277 <a href="#">align</a>
sp P08444 ATP6_SYN6	ATP synthase a chain (ATPase prote	( 261)	113	36.8	0.0077	0.267	0.600	240 <a href="#">align</a>
sp P00852 ATP6_EMENI	ATP synthase a chain precursor (AT	( 256)	111	36.3	0.011	0.209	0.590	244 <a href="#">align</a>
sp P07925 ATP6_MAIZE	ATP synthase a chain (ATPase prote	( 291)	109	35.8	0.017	0.259	0.578	232 <a href="#">align</a>
sp P00851 ATP6_DROYA	ATP synthase a chain (ATPase prote	( 224)	98	33.0	0.094	0.225	0.549	253 <a href="#">align</a>
sp P14862 ATP6_COCHE	ATP synthase a chain (ATPase prote	( 257)	91	31.2	0.37	0.204	0.608	265 <a href="#">align</a>
ref NP_008281.1 ATP6_10704	ATP synthase F0 subunit 6 [D	( 224)	90	31.0	0.39	0.230	0.576	165 <a href="#">align</a>
sp P09716 US17_HCMVA	Hypothetical protein HVLF1	( 293)	91	31.2	0.42	0.260	0.565	131 <a href="#">align</a>
sp P12446 MAT_INCJJ	Polyprotein p42 [Contains: Protein	( 374)	85	29.7	1.5	0.247	0.559	93 <a href="#">align</a>
sp P00849 ATP6_XENLA	ATP synthase a chain (ATPase prote	( 226)	79	28.2	2.7	0.261	0.630	165 <a href="#">align</a>
sp P06974 FLIM_ECOLI	Flagellar motor switch protein fli	( 334)	81	28.7	2.8	0.308	0.673	52 <a href="#">align</a>
sp P05499 ATP6_TOBAC	ATP synthase a chain (ATPase prote	( 395)	81	28.7	3.3	0.220	0.582	268 <a href="#">align</a>

Proteins Sequences Are  
Better for Comparing  
Divergent or Not Well  
Conserved Genes.

# Question you can ask using sequence similarity

---

Is there an homologous protein?

- Does that homologous protein have a similar domain?
- Does XXX genome have YYY (kinase, GPCR, ...)?

Questions not to ask:

- Does this DNA sequence have a similar regulatory element (too short – never significant)?
- Does (non-significant) protein have the same function/modification/antigenic site?

# DNA or Protein

---

- DNA is better when comparing genomes with few variants (populations within a species) or highly conserved genes or RNA genes.
- Otherwise use Protein Sequences

**fRNAdb**  
**Functional**

A comprehensive non-coding RNA sequence database ver. 3.4

fRNAdb is [Web Service \(SOAP, REST\)](#) Ready.

Total: 510,055 entries

 Catalog     Blast     Download     Help

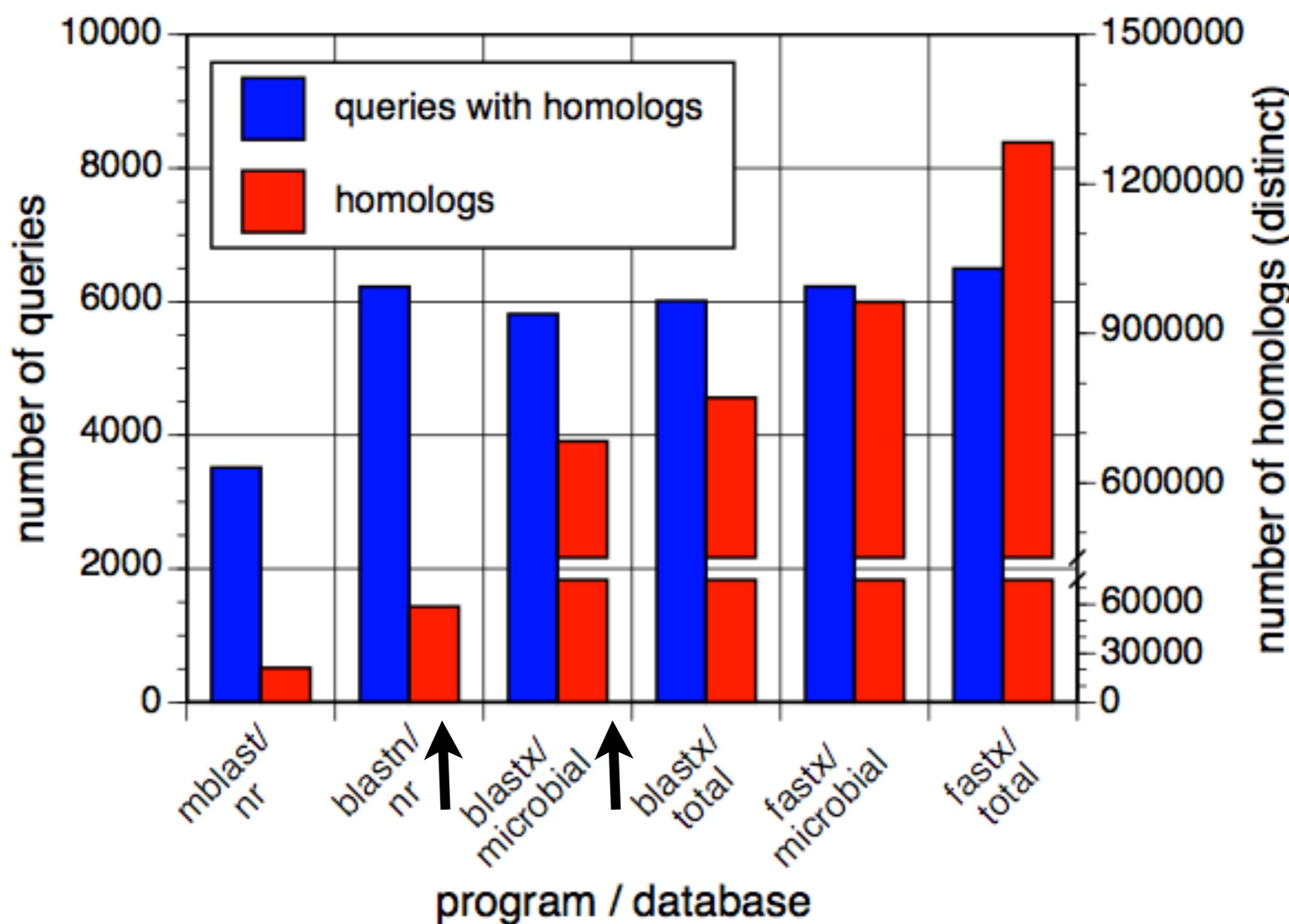


# Protein has a longer look back

The best scores are:

		DNA E(188,018)	tblastx3 E(187,524)	prot. E(331,956)
DMGST	D.melanogaster GST1-1	1.3e-164	4.1e-109	1.0e-109
MDGST1	M.domestica GST-1 gene	2e-77	3.0e-95	1.9e-76
LUCGLTR	Lucilia cuprina GST	1.5e-72	5.2e-91	3.3e-73
MDGST2A	M.domesticus GST-2 mRNA	9.3e-53	1.4e-77	1.6e-62
MDNF1	M.domestica nf1 gene. 10	4.6e-51	2.8e-77	2.2e-62
MDNF6	M.domestica nf6 gene. 10	2.8e-51	4.2e-77	3.1e-62
MDNF7	M.domestica nf7 gene. 10	6.1e-47	9.2e-77	6.7e-62
AGGST15	A.gambiae GST mRNA	3.1e-58	4.2e-76	4.3e-61
CVU87958	Culicoides GST	1.8e-41	4.0e-73	3.6e-58
AGG3GST11	A.gambiae GST1-1 mRNA	1.5e-46	2.8e-55	1.1e-43
BMO6502	Bombyx mori GST mRNA	1.1e-23	8.8e-50	5.7e-40
AGSUGST12	A.gambiae GST1-1 gene	2.3e-16	4.5e-46	5.1e-37
MOTGLUSTR	Manduca sexta GST	5.7e-07	2.5e-30	8.0e-25
RLGSTARGN	R.leguminosarum <i>gstA</i>	0.0029	3.2e-13	1.4e-10
HUMGSTT2A	H. sapiens GSTT2	0.32	3.3e-10	2.0e-09
HSGSTT1	H.sapiens GSTT1 mRNA	7.2	8.4e-13	3.6e-10
ECAE000319	E. coli hypothet. prot.	—	4.7e-10	1.1e-09
MYMDCMA	Methyl. dichlorometh. DH	—	1.1e-09	6.9e-07
BCU19883	Burkholderia maleylacetate red.	—	1.2e-09	1.1e-08
NFU43126	Naegleria fowleri GST	—	3.2e-07	0.0056
SP505GST	Sphingomonas paucim	—	1.8e-06	0.0002
EN1838	H. sapiens maleylaceto. iso.	—	2.1e-06	5.9e-06
HSU86529	Human GSTZ1	—	3.0e-06	8.0e-06
SYCCPNC	Synechocystis GST	—	1.2e-05	9.5e-06
HSEF1GMR	H.sapiens EF1g mRNA	—	9.0e-05	0.00065

# BlastX vs BlastN



# What program do I use?

---

- What is your query sequence?
  - protein: BLAST (NCBI), SSEARCH (EBI)
  - DNA vs Protein: BLASTX (NCBI), FASTX (EBI)
- DNA (structural RNA, repeat family)
  - BLASTN (NCBI), FASTA (EBI)
- Does XXX genome have YYY (protein)?
  - TBLASTN YYY vs XXX genome
  - TFASTX YYY vs XXX genome
- Is Sequence X homologous to Y?
  - BL2SEQ (NCBI), LALIGN, PRSS
- Does my protein contain repeated domains?
  - LALIGN

# Sequence Alignment Via the Web

BLAST®

Home Rec

BLAST finds regions of similarity between biological sequences. [more...](#)

## BLAST Assembled Genomes

Find Genomic BLAST pages:

Enter organism name or id—completions will be suggested

GO

- [Human](#)
- [Mouse](#)
- [Rat](#)
- [Cow](#)
- [Pig](#)
- [Dog](#)
- [Rabbit](#)
- [Chimp](#)
- [Guinea pig](#)
- [Fruit fly](#)
- [Honey bee](#)
- [Chicken](#)
- [Zebrafish](#)
- [Clawed frog](#)
- [Arabidopsis](#)
- [Rice](#)
- [Yeast](#)
- [Microbes](#)

## Basic BLAST

Choose a BLAST program to run.

### [nucleotide blast](#)

Search a nucleotide database using a nucleotide query  
*Algorithms:* blastn, megablast, discontiguous megablast

### [protein blast](#)

Search protein database using a protein query  
*Algorithms:* blastp, psi-blast, phi-blast, delta-blast

### [blastx](#)

Search protein database using a translated nucleotide query

### [tblastn](#)

Search translated nucleotide database using a protein query

### [tblastx](#)

Search translated nucleotide database using a translated nucleotide query

# Sequence Alignment Via the Web

**BLAST®** » blastp suite

Home Recent Results Saved Searches

### Standard Protein BLAST

[blastn](#) [blastp](#) [blastx](#) [tblastn](#) [tblastx](#)

[Enter Query Sequence](#)

BLASTP programs search protein databases using a protein query. [more...](#) [Reset page](#)

Enter accession number(s), gi(s), or FASTA sequence(s) [?](#) [Clear](#)

Query subrange [?](#)

From

To

Or, upload file [Choose File](#) No file chosen [?](#)

Job Title

Enter a descriptive title for your BLAST search [?](#)

Align two or more sequences [?](#)

[Choose Search Set](#)

Database [Protein Data Bank proteins\(pdb\)](#) [?](#)

Organism [Optional](#) Enter organism name or id—completions will be suggested   Exclude [+](#) Enter organism common name, binomial, or tax id. Only 20 top taxa will be shown. [?](#)

Exclude [Optional](#)  Models (XM/XP)  Uncultured/environmental sample sequences

Entrez Query [Optional](#) [YouTube](#) [Create custom database](#) Enter an Entrez query to limit search [?](#)

# Sequence Alignment Via the Web

## FASTA

### FASTA ?

FASTA is another commonly used sequence similarity search tool which uses heuristics for fast **local** alignment searching.

 Protein  Nucleotide  Genomes  Whole Genome Shotgun

### SSEARCH ?

SSEARCH is an optimal (as opposed to heuristics-based) **local** alignment search tool using the Smith-Waterman algorithm. Optimal searches guarantee you find the best alignment score for your given parameters.

 Protein  Nucleotide  Genomes  Whole Genome Shotgun

### PSI-Search ?

PSI-Search combines the sensitivity of the Smith-Waterman search algorithm (SSEARCH) with the PSI-BLAST profile construction strategy to find distantly related protein sequences.

 Protein

### GGSEARCH ?

GGSEARCH performs optimal **global-global** alignment searches using the Needleman-Wunsch algorithm.

 Protein  Nucleotide

## BLAST

### NCBI BLAST ?

NCBI BLAST is the most commonly used sequence similarity search tool. It uses heuristics to perform fast **local** alignment searches.

 Protein  Nucleotide  Vectors

### PSI-BLAST ?

PSI-BLAST allows users to construct and perform a BLAST search with a custom, position-specific, scoring matrix which can help find distant evolutionary relationships. PHI-BLAST functionality is also available to restrict results using patterns.

 Protein

<http://www.ebi.ac.uk/Tools/ss/>

# Sequence Alignment Via the Web

## STEP 1 - Select your databases

### PROTEIN DATABASES

1 Databank Selected

 Clear Selection

- UniProt Knowledgebase
- UniProtKB/Swiss-Prot
- UniProtKB/Swiss-Prot isoforms
- UniProtKB/TrEMBL
- ▶ **UniProtKB Taxonomic Subsets**
- ▶ **UniProt Clusters**
- ▶ **Patents**
- ▶ **Structure**
- ▼ **Other Protein Databases**
  - UniProt Archive
  - IntAct
  - IMGT/HLA
  - IPD-KIR
  - IPD-MHC
  - MACiE Annot Pub

## STEP 2 - Enter your input sequence

Enter or paste a PROTEIN  sequence in any supported format:

<http://www.ebi.ac.uk/Tools/ss/>

or Upload a file:  No file chosen

## STEP 3 - Set your parameters

### PROGRAM

# Sequence Alignment Via the Web

## UVa FASTA Server

New: Annotation features available for SwissProt/PIR1 library searches.

### About

- Getting started
- [fasta\\_guide.pdf](#)

### Other FASTA Servers

- EMBL-EBI
- KEGG (Japan)

### References

- FASTA
- FASTX/FASTY
- Statistics
- FASTS/FASTF

### Software

- FASTA v36 ChangeLog
- Downloads
- Sequence Libraries
- Developer Mailing list

### Other resources

- CHAPS - Convert HMMs and Profiles
- Near optimal alignments
- FASTA Exercises
- NCBI BLAST server
- EMBL-EBI Server

The **FASTA** programs find regions of local or global similarity between Protein or DNA sequences, either by searching Protein or DNA databases, or by identifying local duplications within a sequence. Other programs provide information on the statistical significance of an alignment. Like **BLAST**, **FASTA** can be used to infer functional and evolutionary relationships between sequences as well as help identify members of gene families.

### Protein

- Protein-protein **FASTA**
- Protein-protein Smith-Waterman ([sssearch](#))
- Global Protein-protein (Needleman-Wunsch) ([ggsearch](#))
- Global/Local protein-protein ([glsearch](#))
- Protein-protein with unordered peptides ([fasts](#))
- Protein-protein with mixed peptide sequences ([fastf](#))

### Nucleotide

- Nucleotide-Nucleotide (DNA/RNA [fasta](#))
- Ordered Nucleotides vs Nucleotide ([fastm](#))
- Un-ordered Nucleotides vs Nucleotide ([fasts](#))

[fasta.bioch.virginia.edu](http://fasta.bioch.virginia.edu)

### Translated

- Translated DNA (with frameshifts, e.g. ESTs) vs Proteins ([fastx/fasty](#))
- Protein vs Translated DNA (with frameshifts) ([tfastx/tfasty](#))
- Peptides vs Translated DNA ([tfasts](#))

### Statistical Significance

- Protein vs Protein shuffle ([prss](#))
- DNA vs DNA shuffle ([prss](#))
- Translated DNA vs Protein shuffle ([prfx](#))

### Local Duplications

- Local Protein alignments ([lalign](#))
- Plot Protein alignment "dot-plot" ([plalign](#))
- Local DNA alignments ([lalign](#))
- Plot DNA alignment "dot-plot" ([plalign](#))

# What Database to Search?

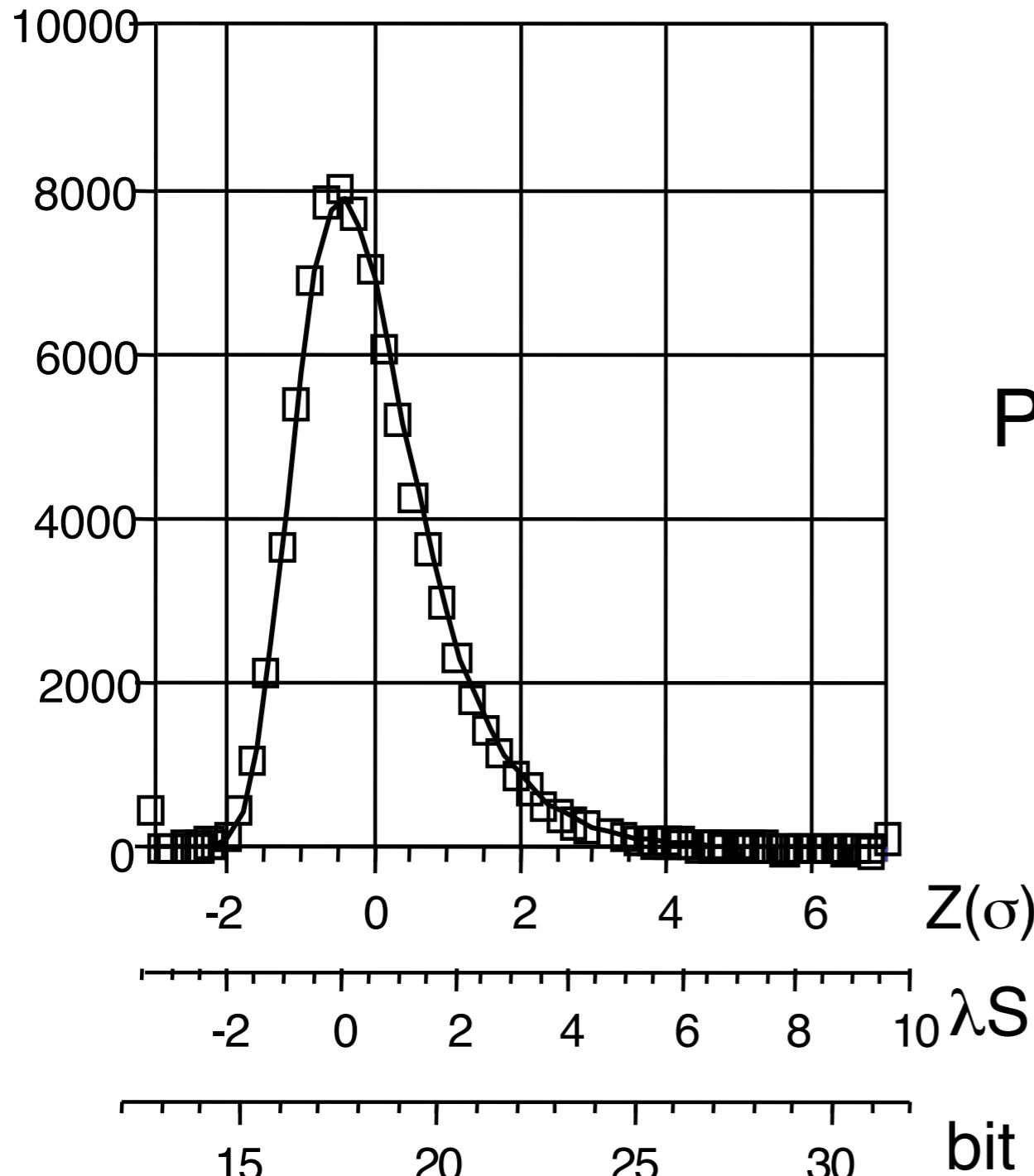
---

- Search the smallest comprehensive database likely to contain your protein of interest
  - vertebrates – human proteins (40,000)
  - fungi – *S. cerevisiae* (6,000)
  - bacteria – *E. coli*, gram positive, etc. (<100,000)
- Search a richly annotated protein set (SwissProt, 450,000)
- Always search NR (> 12 million) *LAST*

Never Search “GenBank” (DNA)

# DB Size Matters

## Smaller is Better



$$S' = \lambda S_{\text{raw}} - \ln K m n$$

$$S_{\text{bit}} = (\lambda S_{\text{raw}} - \ln K) / \ln(2)$$

$$P(S' > x) = 1 - \exp(-e^{-x})$$

$$P(S_{\text{bit}} > x) = 1 - \exp(-mn2^{-x})$$

$$E(S' > x | D) = P D$$

$$P(B \text{ bits}) = m n 2^{-B}$$

$$P(40 \text{ bits}) = 1.5 \times 10^{-7}$$

$$E(40 | D=4000) = 6 \times 10^{-4}$$

$$E(40 | D=12E6) = 1.8$$

# DB Size Matters

## Smaller is Better

gi|114443|sp|P00846.1|ATP6\_HUMAN ATP synthase subunit a; F-ATPase - 226 aa  
vs

gi|16131606|ref|NP\_418194.1| F0 sector of membrane-bound ATP synthase, subunit a [Escherichia coli str. K-12 subst - 271 aa

initn: 159 init1: 104 opt: 148 z-score: 212.5 bits: 46.8  
Smith-Waterman score: 178; 23.7% identity (58.9% similar) in 236 aa overlap (45-264:8-222)

Database	Entries	Length	E()	Time (s)
E.Coli	4237	1350094	3.8E-07	<0.5
Human Ref	38000	17401176	1.9E-05	1
SwissProt	445410165796297		0.0015	10
RefSeq	711441261324908		NS	16

# How Can I Choose my DB?

**BLAST®** » blastp suite

Home Recent Results Saved Searches

### Standard Protein BLAST

[blastn](#) [blastp](#) [blastx](#) [tblastn](#) [tblastx](#)

[Enter Query Sequence](#) [Reset page](#)

BLASTP programs search protein databases using a protein query. [more...](#)

Enter accession number(s), gi(s), or FASTA sequence(s) [?](#) [Clear](#)

Query subrange [?](#)

From

To

Or, upload file [Choose File](#) No file chosen [?](#)

Job Title

Enter a descriptive title for your BLAST search [?](#)

Align two or more sequences [?](#)

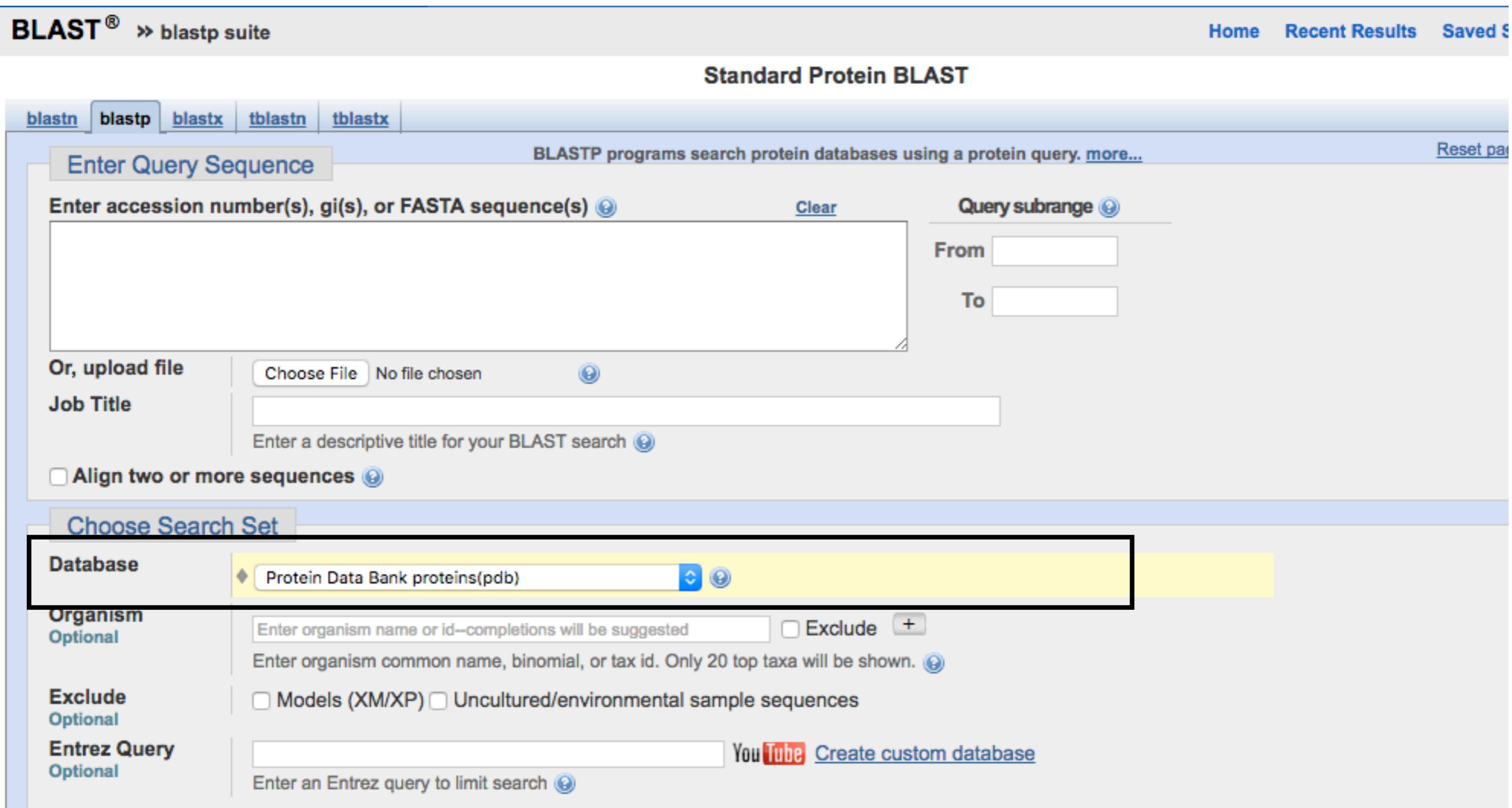
[Choose Search Set](#)

**Database** [Protein Data Bank proteins\(pdb\)](#) [?](#)

**Organism** [Optional](#) Enter organism name or id—completions will be suggested   Exclude [+](#) Enter organism common name, binomial, or tax id. Only 20 top taxa will be shown. [?](#)

**Exclude** [Optional](#)  Models (XM/XP)  Uncultured/environmental sample sequences

**Entrez Query** [Optional](#)  [YouTube](#) [Create custom database](#) Enter an Entrez query to limit search [?](#)



# How can you tell what is the Highest Scoring Unrelated Hit?

```
Query: TMP.q
1>>>gi|28200469|gb|AA031759.1| endo-beta-1,4-mannanase 5A [Cellvibrio - 430 aa
Library: Swissprot (NCBI)
165796297 residues in 445410 sequences

Statistics: Expectation_n fit: rho(ln(x))= 7.6630+/-0.000201; mu= 3.3292+/- 0.012
mean_var=63.4892+/-13.027, 0's: 51 Z-trim(131.3): 79 B-trim: 0 in 0/68
Lambda= 0.160962
statistics sampled from 60000 (180148) to 445316 sequences
Algorithm: Smith-Waterman (SSE2, Michael Farrar 2006) (7.2 Nov 2010)
Parameters: BL50 matrix (15:-5)xS, open/ext: -10/-2
Scan time: 29.700

The best scores are:
          s-w bits E(445410) %_id %_sim alen align
sp|P51529.2|MANA_STRLI Mannan endo-1,4-beta-mannosidase ( 383) 1225 291.3 1.5e-77 0.520 0.789 375 align
sp|P22533.2|MANB_CALSA Beta-mannanase/endoglucanase A; (1331) 896 214.5 7.1e-54 0.403 0.686 382 align
sp|P14768.2|XYNA_CELJU Endo-1,4-beta-xylanase A; Xylan ( 611) 226 59.1 1.9e-07 0.330 0.614 176 align
sp|P10476.2|GUNA_CELJU Endoglucanase A; EGA; Cellulase ( 962) 227 59.2 2.8e-07 0.350 0.657 137 align
sp|P27033.2|GUNC_CELJU Endoglucanase C; Cellodextrinase ( 747) 223 58.4 3.9e-07 0.286 0.636 206 align
sp|P18126.1|GUNB_CELJU Endoglucanase B; EGB; Cellulase ( 511) 201 53.4 8.3e-06 0.327 0.619 202 align
sp|Q74706.1|EGLB_ASPNG Endo-beta-1,4-glucanase B; Endo ( 331) 190 51.0 2.9e-05 0.275 0.558 233 align
sp|Q12647.1|GUNB_NEOPA Endoglucanase B; Cellulase B; En ( 473) 183 49.2 0.00014 0.229 0.469 414 align
sp|O96W08.1|EGLB_ASPKA Probable endo-beta-1,4-glucanase ( 332) 179 48.4 0.00017 0.278 0.543 234 align
sp|P23661.1|GUNB_RUMAL Endoglucanase B; Cellulase B; En ( 409) 166 45.3 0.0018 0.227 0.508 299 align
sp|P54937.1|GUNA_CLOLO Endoglucanase A; Cellulase A; En ( 517) 166 45.3 0.0024 0.209 0.520 406 align
sp|P54937.1|GUNA_CLOLO Endoglucanase A; Cellulase A; En ( 517) 166 45.3 0.0024 0.209 0.520 406 align
```

# Perform a search with your “suspect”

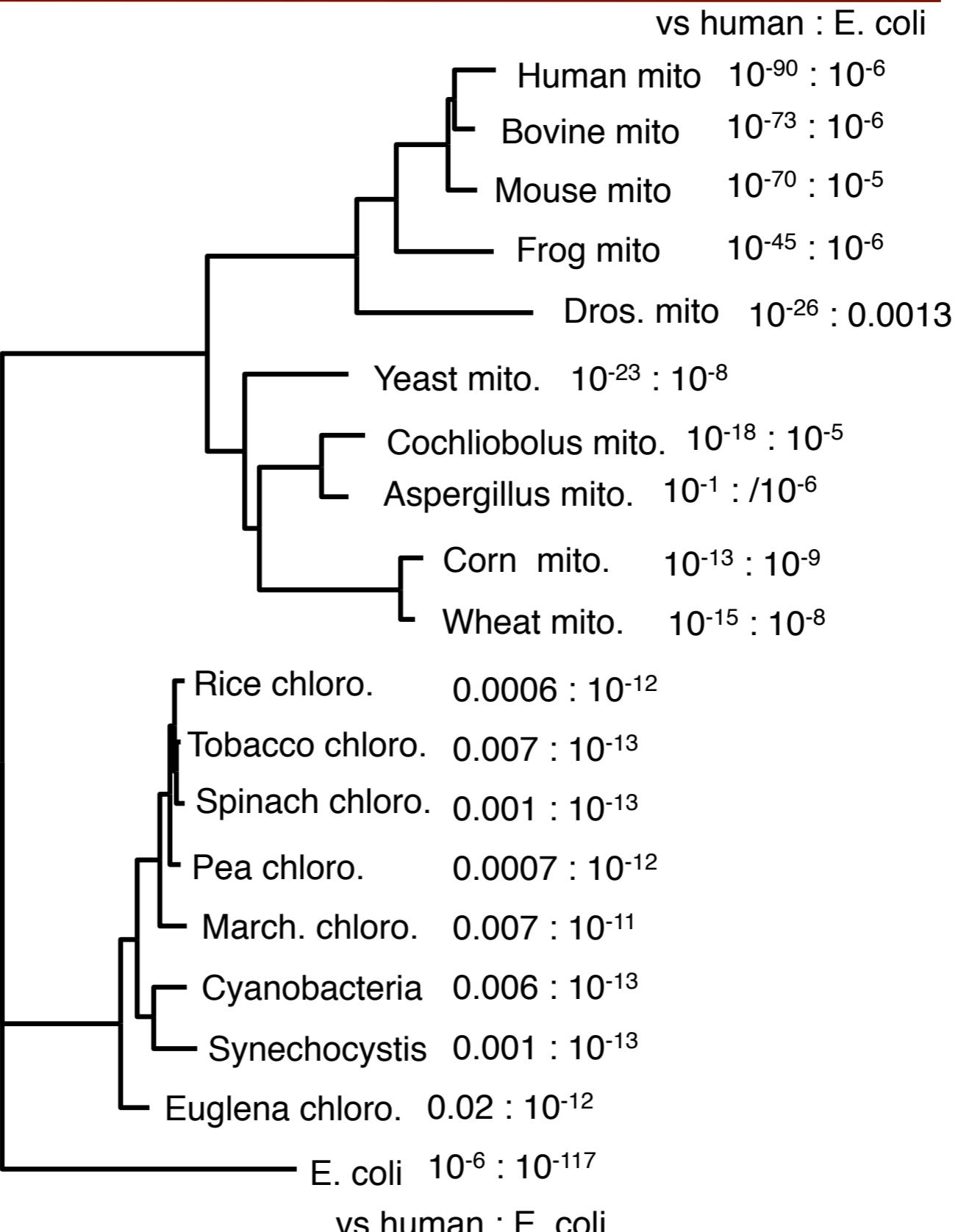
```
The best scores are:
sp|P23661.1|GUNB_RUMAL Endoglucanase B; Cellulase B; En ( 409) 2549 597.9 7.6e-170 1.000 1.000 409 align
sp|P16216.1|GUN1_RUMAL Endoglucanase 1; Cellulase; Endo ( 406) 2186 513.7 1.7e-144 0.806 0.934 407 align
sp|P23660.1|GUNA_RUMAL Endoglucanase A; Cellulase A; En ( 364) 992 236.7 3.7e-61 0.461 0.723 343 align
sp|P54937.1|GUNA_CLOLO Endoglucanase A; Cellulase A; En ( 517) 984 234.7 2.1e-60 0.431 0.727 355 align
sp|Q12647.1|GUNB_NEOPA Endoglucanase B; Cellulase B; En ( 473) 895 214.1 3.1e-54 0.433 0.693 342 align
sp|P10477.1|GUNE_CLOTM Endoglucanase E; Cellulase E; En ( 814) 898 214.5 3.9e-54 0.368 0.679 408 align
sp|P28623.2|GUND_CLOC7 Endoglucanase D; Cellulase D; En ( 515) 894 213.8 4e-54 0.413 0.707 334 align
sp|P17901.1|GUNA_CLOCE Endoglucanase A; Cellulase A; EG ( 475) 875 209.4 7.7e-53 0.403 0.679 380 align
sp|P20847.1|GUN1_BUTFI Endoglucanase 1; Cellulase 1; En ( 547) 855 204.7 2.3e-51 0.389 0.664 378 align
sp|P28621.1|GUNB_CLOC7 Endoglucanase B; Cellulase B; En ( 440) 853 204.4 2.4e-51 0.388 0.703 340 align
sp|P23550.1|GUNB_PAELA Endoglucanase B; Cellulase B; En ( 566) 601 145.8 1.3e-33 0.314 0.638 354 align
sp|P25472.1|GUND_CLOCE Endoglucanase D; Cellulase D; EG ( 584) 570 138.6 2e-31 0.334 0.638 329 align
sp|O08342.1|GUNA_PAEBB Endoglucanase A; Cellulase A; En ( 400) 538 131.3 2.1e-29 0.303 0.612 356 align
sp|P16218.1|GUNH_CLOTH Endoglucanase H; Cellulase H; En ( 900) 507 123.8 9e-27 0.317 0.609 363 align
sp|P19570.1|GUN3_BACS4 Endoglucanase C; Cellulase C; En ( 825) 208 54.4 6.2e-06 0.217 0.506 397 align
sp|Q04469.1|GUN1_CRYFL Endoglucanase 1; Carboxymethyl-c ( 341) 185 49.5 7.8e-05 0.232 0.547 254 align
sp|P07982.1|GUN2_TRIRE Endoglucanase EG-II; EGLII; Cel ( 418) 185 49.4 0.0001 0.224 0.568 340 align
sp|Q2UPQ4.1|EGLB_ASPOR Probable endo-beta-1,4-glucanase ( 333) 181 48.6 0.00014 0.209 0.538 273 align
sp|P06564.1|GUN_BACS1 Endoglucanase; Alkaline cellulase ( 800) 188 49.8 0.00015 0.256 0.555 211 align
sp|P19424.1|GUN_BACS6 Endoglucanase; Alkaline cellulase ( 941) 186 49.3 0.00025 0.263 0.577 194 align
sp|P54583.1|GUN1_ACIC1 Endoglucanase E1; Cellulase E1; ( 562) 176 47.2 0.00064 0.251 0.498 307 align
```

Is a hit from your original search in the re-search?

# Homology through Transitivity

ATP-synt\_A

How do you pick the right sequence homologous to both?



# Unrelated ≠ Random

## low complexity sequence

The best scores are:							s-w	bits	E(13351)	%_id	%_sim	alen
sp P17343 GBB1_CAEEL	Guanine nucleotide-binding protein	( 340)	251	45.2	8.4e-05	0.227	0.531	277	<a href="#">align</a>			
sp P16520 GBB3_HUMAN	Guanine nucleotide-binding protein	( 340)	250	45.0	9.2e-05	0.236	0.528	288	<a href="#">align</a>			
sp P26308 GBB1_DROME	Guanine nucleotide-binding protein	( 340)	249	44.9	0.0001	0.219	0.559	288	<a href="#">align</a>			
sp P62871 GBB1_BOVIN	Guanine nucleotide-binding protein	( 340)	248	44.8	0.00011	0.243	0.558	267	<a href="#">align</a>			
sp P29387 GBB4_MOUSE	Guanine nucleotide-binding protein	( 340)	241	43.8	0.00022	0.234	0.543	265	<a href="#">align</a>			
sp P11017 GBB2_BOVIN	Guanine nucleotide-binding protein	( 326)	240	43.7	0.00023	0.230	0.543	265	<a href="#">align</a>			
sp P04280 PRP1_HUMAN	Basic salivary proline-rich protein	( 392)	242	43.9	0.00023	0.268	0.423	291	<a href="#">align</a>			
sp P62879 GBB2_HUMAN	Guanine nucleotide-binding protein	( 340)	240	43.7	0.00024	0.230	0.543	265	<a href="#">align</a>			
sp P04258 CO3A1_BOVIN	Collagen alpha-1(III) chain	(1049)	246	44.4	0.00044	0.288	0.454	302				
+-			197	37.7	0.046	0.267	0.470	285				
+-			182	35.6	0.19	0.246	0.460	313	<a href="#">align</a>			
sp P29829 GBB2_DROME	Guanine nucleotide-binding protein	( 346)	232	42.6	0.00052	0.233	0.574	258	<a href="#">align</a>			
sp P04474 PRP3_RAT	Acidic proline-rich protein PRP33 pr	( 206)	224	41.5	0.00064	0.300	0.511	190	<a href="#">align</a>			
sp P23232 GBB_LOLFO	Guanine nucleotide-binding protein	( 341)	220	40.9	0.0016	0.215	0.548	279	<a href="#">align</a>			
ref NP_203699.1	alpha 5 type IV collagen isoform 2, pr	(1691)	225	41.5	0.0054	0.256	0.445	308				
+-			208	39.1	0.027	0.256	0.465	301				
+-			202	38.3	0.048	0.280	0.467	321				
+-			183	35.7	0.29	0.251	0.438	347	<a href="#">align</a>			
....	....	....	....	....	....	....	....	....	....	....	....	....

## Filter Low Complexity (SEG)

sp P62871 GBB1_BOVIN	Guanine nucleotide-binding protein	( 340)	225	52.9	4e-07	0.243	0.558	267	<a href="#">align</a>			
sp P23232 GBB_LOLFO	Guanine nucleotide-binding protein	( 341)	220	51.9	8.1e-07	0.215	0.548	279	<a href="#">align</a>			
sp P13712 MSI1_YEAST	Chromatin assembly factor 1 subunit	( 422)	147	37.2	0.026	0.207	0.515	309	<a href="#">align</a>			
sp P53622 COPA_YEAST	Coatomer subunit alpha (Alpha-coat)	(1201)	142	35.8	0.2	0.201	0.479	234	<a href="#">align</a>			
sp P11269 GAG_MLVRD	Gag polyprotein (Core polyprotein)	( 537)	134	34.5	0.22	0.252	0.482	226	<a href="#">align</a>			
sp P29674 LHX1_XENLA	LIM/homeobox protein Lhx1 (LIM hom)	( 403)	129	33.6	0.3	0.299	0.538	117	<a href="#">align</a>			
sp P09256 VGLC_VZVD	Glycoprotein GPV	( 560)	132	34.1	0.3	0.248	0.482	141	<a href="#">align</a>			
sp O13528 YA11A_YEAST	Transposon Ty1-A/Ty1-PR1 Gag poly	( 440)	127	33.2	0.44	0.246	0.508	183	<a href="#">align</a>			
sp P53621 COPA_HUMAN	Coatomer subunit alpha (Alpha-coat)	(1224)	134	34.1	0.63	0.199	0.534	146	<a href="#">align</a>			

# SEG Remove Low Complexity

>gi|122065196|sp|P16371.3|GROU\_DROME Protein groucho; Enhancer of split m9/10 protein; E(spl)m9/10

paaggpppqgp	1-8 9-19 20-122	MYPSPVRH IKFTIADTLERIKEENFLQAQYHSIKLEC EKLSNEKTEMQRHYVEMYEMSYGLNVEMHK QTEIAKRLNTLINQLLPFLQADHQQQVLQA VERAKQVTMQELN
liighqqqhgicqqllqqihaqqvpqggppqp mg	123-154 155-292	
rppsrsgssssrstsps	293-308	
akartptpnaaapapgvnpk qmmpqgpppagypgapyqrpa	309-321 322-341 342-362 363-730	LTKDMEKPGTPG DPYQRPPSDPAYGRPPPMPYDPHAHVRTNG IPHPSALTGGKPAYSFHMNGEGLQPVPFP PDALVGVGIPRHARQINTLSHGEVVCAVTI SNPTKYVYTGGKGCVKVWDISQPGNKNPVS QLDCLQRDNYIRSVKLLPDGRTLIVGGEAS NLSIWDLASPPTPRIKAELTSAAPACYALAI SPDSKVCFSCCSDGNIAVWDLHNEILVRQF QGHTDGASCIDISPDSRLWTGGLDNTVRS WDLREGRQLQQHDFSSQIFSLGYCPTGDWL AVGMENSHVVEVLHASKPDKYQLHLHESCVL SLRFAACGKWFVSTGKDNLNAWRTPYGAS IFQSKE <del>TSSVLS</del> CDISTDDKYIVTGSGDKK ATVYEVIY

# SEG Remove Low Complexity

**Scoring Parameters**

Matrix: BLOSUM62

Gap Costs: Existence: 11 Extension: 1

Compositional adjustments: Conditional compositional score matrix adjustment

**Filters and Masking**

Filter:  Low complexity regions

Mask:  Mask for lookup table only  
 Mask lower case letters

**(A) Program:** FASTA: protein:protein

**(B) Query sequence:** FASTA format

Subset range: \_\_\_\_\_  Use Subset range

Compare your own sequences:  
[Compare sequences](#)

Entrez protein sequence browser  
Entrez DNA sequence browser

Or upload query from file:  [Browse...](#)

Protein  DNA (both-strands)  DNA (forward only)  DNA (rev-comp only)

**(C) Database:**

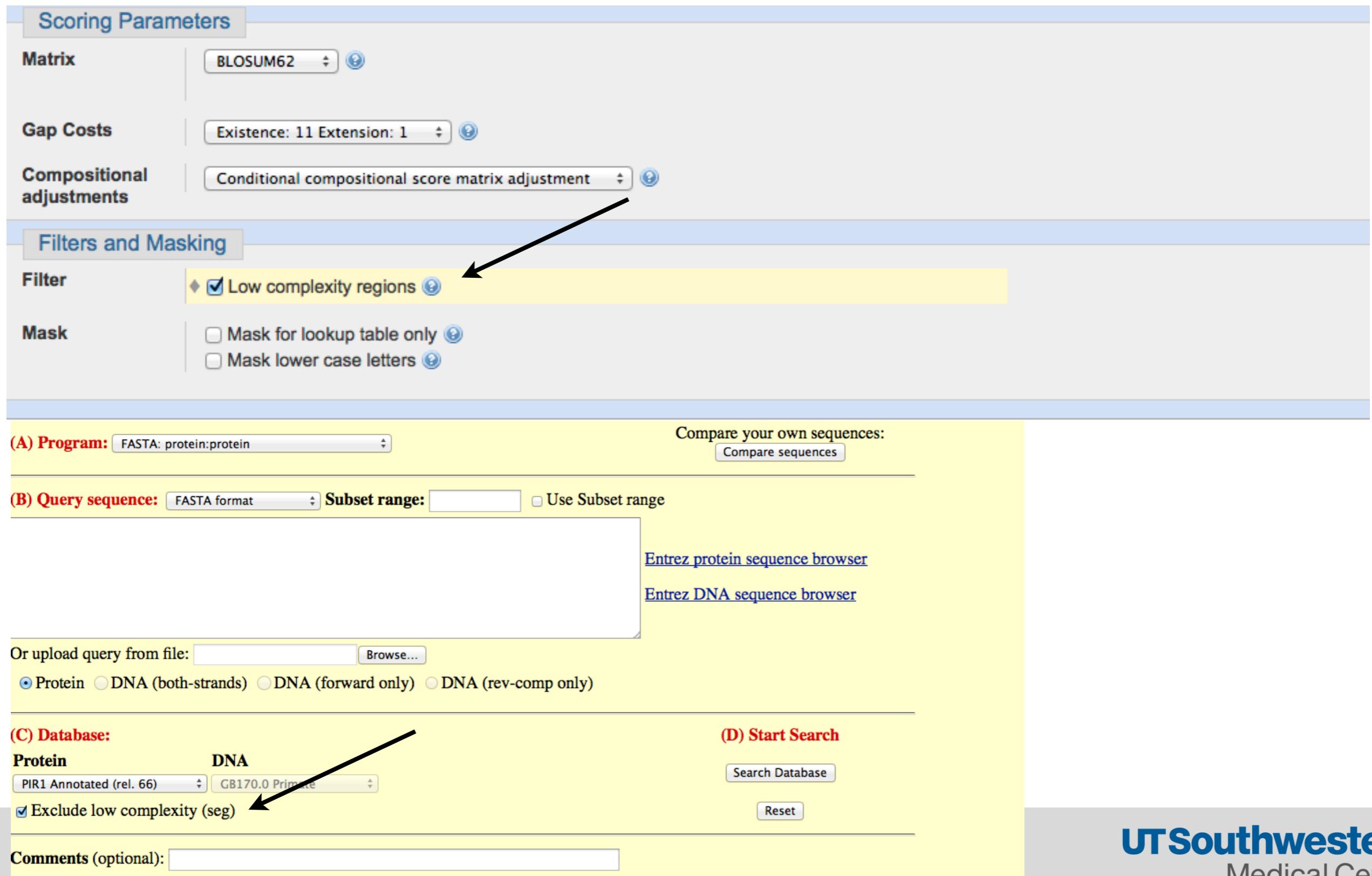
Protein: PIR1 Annotated (rel. 66)  
DNA: GB170.0 Primate

Exclude low complexity (seg)

**(D) Start Search**

[Search Database](#) [Reset](#)

Comments (optional):



# Validating Stats

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- In general, BLASTP statistical estimates are accurate  
The most common errors occur because of low- complexity regions, or biased amino-acid composition  
To confirm statistical accuracy, find the highest scoring non homolog
  - No need to test every hit, test hits that are surprising
  - Confirm homology/non-homology by searching against a different comprehensive database, e.g. SwissProt, or refseq.
  - Non-homologs will find many significant members of other families, but not the family you are testing for
  - Statistical estimates can be confirmed with shuffles

# Validating Stats

Choose: (A) program and (B, C) sequences to compare:

(A) Program: PRSS: protein:protein

(B) Number of shuffles: 200

Uniform  Window

(B.1) Enter first (query) sequence: FASTA format

Subset range:

Annotate Query Sequence (SwissProt accessions)

No annotation

Upload annotation file: Choose File No file chosen

Entrez protein / Entrez DNA sequence browser

Uniprot sequence browser

(B.2) Or upload sequence from file: Choose File No file chosen

Protein  DNA (both-strands)  DNA (forward only)  DNA (rev-comp only)

Use PSSM:

(C.1) Enter the second sequence:

FASTA format

Subset range:

Annotate Target Sequence (SwissProt accessions)

No annotation

Upload annotation file: Choose File No file chosen

Shuffle Sequence

Reset Form

(C.2) Or choose file of sequences/accessions: Choose File No file chosen

Other comparison options:

Scoring matrix: open: ext: Ktup:

Blosum50 (25%) -10 -2 ktup = 2

# Alignment Summary

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- Compare Protein Sequences for long distances, DNA for close relationships.
- Sequence statistical significance estimates are accurate (verify this yourself)  $10^{-6} < E() < 10^{-3}$  is statistically significant
- Local sequence alignments find the best region (so that extending the region reduces the score). Global alignments go from end-to-end.
- The Smith-Waterman algorithm produces local alignments with affine gaps in time  $O(nm)$  and space  $O(n)$ .
- BLAST and FASTA try to approximate Smith-Waterman scores for homologous sequences
- Smaller databases increase search sensitivity
- Statistical accuracy can be evaluated by examining the “highest scoring unrelated sequence” or by random shuffles

# Workshop Time

[https://bcantarel.github.io/cshl\\_homology\\_workshop1](https://bcantarel.github.io/cshl_homology_workshop1)