

ORIGINAL RESEARCH PAPERS

Bioactive Polysaccharides from Traditional Chinese Medicine Herbs as Anticancer Adjuvants

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ABSTRACT

Purpose of study: To review the nature, extent, bioactivities, and clinical application of polysaccharides in Traditional Chinese Medicine (TCM) especially as adjuvants in cancer treatment.

Methodology: Literature review.

Findings: Many fungal- and plant-derived bioactive polysaccharides with a broad range of immunomodulatory activities are found in TCM. Some such polysaccharides have been developed into drugs and show clinical efficacy in controlled trials while the majority of such compounds remain as nutraceuticals with only preliminary research. Such polysaccharides are generally nontoxic and possess other bioactivities such as inducing differentiation, stimulating hematopoiesis, antimetastasis, and antiangiogenesis, which make them ideal adjuvants in modern cancer therapy.

Conclusion: Bioactive polysaccharides occur extensively in TCM herbs and are the basis of potentially useful application of TCM as an adjuvant in cancer therapies.

BIOACTIVITY OF POLYSACCHARIDES

As a major class of biomolecules, carbohydrates are the most complex and least appreciated for their bioactivity (Stryer, 1995). Recently, an increasing number of reports describing the isolation and bioactivity of polysaccharide glucans and proteoglucans from plant and other sources highlight the potential role of this class of molecules in cancer therapy as a result of its immunostimulatory as well as other anticancer properties (Wong et al., 1994).

BIOACTIVE POLYSACCHARIDES IN CHINESE HERBS

Known bioactive polysaccharides are found in fungi, lichens, higher plants, marine, as well as animal sources throughout the world, but some of the most well characterized and clinically relevant polysaccharides are found in Traditional Chinese Medicine (TCM) (Ooi and Liu, 2000), especially those herbs from the TCM materia medica classically characterized as tonic in nature or having *Fu-Zhen* (Sun et al., 1981) properties. Many such tonic Chinese herbs

have been found to possess immunomodulatory bioactivity and are potentially useful in cancer therapy (Sun, 1986). As such, the search and characterization of novel, safe, and effective natural compounds from Chinese herbs is a significant goal for anticancer research.

IMMUNOMODULATORY PROPERTY OF POLYSACCHARIDES AND THE β -GLUCAN RECEPTOR

Naturally derived polysaccharides including heteroglycans and proteoglycans of certain molecular weight and structure have specific broad-ranged immunomodulatory properties that have been recognized for several decades. Such immunomodulating activity includes activation of macrophages (Adachi et al., 1990), monocytes (Czop and Austen, 1985a), natural killer (NK) cells (Peter et al., 1988), lymphocyte-activated killer cells (Yamasaki et al., 1989), dendritic cells (Tsujitani et al., 1992), tumor-infiltrating lymphocytes (Kariya et al., 1991), and other lymphocytes (Kumazawa et al., 1985). The stimulated release of various cytokines including interferons (Kandefer-Szerszen and Kawecki, 1973), interleukins (Sakagami et al., 1988), tumor necrosis factor (Abel and Czop, 1992) and colony-stimulating factors (Hashimoto et al., 1990) have also been documented. Such polysaccharides are thus considered multicytokine inducers and this is probably because of induction of gene expression of various immunomodulatory cytokines and cytokine receptors (Liu et al., 1999).

An important feature of the bioactivity of immunomodulatory polysaccharides is the importance of its structure-function relation. Differences in molecular weight, tertiary structure or conformation, and composition all affect polysaccharide bioactivity. In general, polysaccharides in a configuration with β 1-3, 1-4, or 1-6 branch chains are necessary for activity and complex branch-chained polysaccharides with anionic structures and higher molecular weights have greater immunostimulating activities (Cleary et al., 1999). Differences in bioactivity may be caused by differences in receptor affinity or receptor-ligand interaction on the cell surface (Mueller et al., 2000).

The description of a β -glucan receptor on monocytes by Czop and Austen (1985b), served as a basis to understand the immunopotentiating bioactivity of polysaccharides and explains why herbs and materials from different sources with similarly structured polysaccharide content share similar immunomodulatory activity.

POLYSACCHARIDES AS ANTITUMOR ADJUVANTS

The usefulness of bioactive polysaccharides found in TCM with a β 1,3 1,4 or 1,6 in enhancing the immune system and therefore indirectly reducing tumorigenesis as well as tumor growth has been extensively demonstrated in animals while prolonged survival as a result of treatment with polysaccharide-derived nutraceuticals and drugs have been noted in a number of controlled clinical trials carried out in Japan and China.

IMMUNOMODULATORY AND ANTITUMOR POLYSACCHARIDES IN TCM

Immunopotentiating traditional Chinese herbs with proven antitumor activity may be broadly considered as fungals or botanicals. Almost 200 species of such fungi have demonstrable antitumor activity although not all such fungi are in the TCM pharmacopeia (Borchers et al., 1999). Fungals, especially those from the *Basidiomycetes* family have been found to possess bioactive polysaccharides (Wasser and Weis, 1999). According to a survey by Jong and Donovan (1989), 109 antitumor substances from fungi were from *Basidiomycetes* and 51 of these were glucans or polysaccharide compounds from no less than 26 different species. Some of these fungal polysaccharides have been systematically studied as well as developed into nutraceuticals (e.g., *Agaricus blazei* [Itoh et al., 1994], *Cordyceps sinensis* [Kuo et al., 1996], *Ganoderma* spp. [Chang, 1996], *Grifola frondosa* [Hishida et al., 1988]) or drugs KrestinTM (Kureha Chemicals Industry Corp., Tokyo, Japan) from *Coriolus versicolor* (Kondo and Torisu, 1985), Lenti-

TABLE 1. MEDICINAL FUNGI REPORTED TO CONTAIN BIOACTIVE POLYSACCHARIDE

<i>Agaricus blazei</i> (Ohno et al., 2001)
<i>Auricularia auricula</i> (Misaki, 1981)
<i>Flammulina velutipes</i> (Leung et al., 1997)
<i>Hericium erinaceum</i> (Mizuno et al., 1992)
<i>Inonotus</i> spp. (Wasser and Weis, 1999)
<i>Phellinus</i> spp. (Han et al., 1999)
<i>Pleurotus</i> spp. (Chenghua et al., 2000)
<i>Polyporus</i> spp. (Zhang, et al., 1991)
<i>Poria</i> spp. (Kanayama et al., 1986)
<i>Tricholoma</i> spp. (Liu et al., 1996) ^a
<i>Tremella</i> spp. (Xia and Lin, 1989)

^aLiu F, Ooi VE, Liu WK, Chang ST. Immunomodulation and antitumor activity of polysaccharide-protein complex from the culture filtrates of a local edible mushroom, *Tricholoma lobayense*. Gen Pharmacol 1996;27:621–624.

nan from *Lentinus edodes* (Chihara et al., 1987), Schizophyllan from *Schizophyllum communes* (Komatsu et al., 1962), but others have been only studied preliminarily (See Table 1).

As a representative agent, Lentinan from *Lentinus edodes* was identified in the late 1960s by Chihara et al. (1970). It is a branched-chain molecule with a backbone of 1,3 β -D-glucan and side chains of β 1,3 and β 1,6-D-glucose residues. It has been demonstrated to elicit antitumor activity by the stimulation of host-mediated immune responses and thus inhibit the growth of implanted tumors in laboratory animals (Chihara, 1983). Lentinan has also been demonstrated to be active as a parenteral agent in prolonging survival in recurrent and metastatic gastric and colorectal cancer when given in combination with chemotherapy in controlled clinical trial (Wakui et al., 1986).

Another representative agent is Krestin [PSK] (Kureha Chemicals Industry Corp.), which is a protein-bound polysaccharide extracted from *Coriolus versicolor* (Kondo and Torisu, 1985). Unlike lentinan, PSK is a β 1,4 glucan containing 10% protein and is active orally. PSK has also been statistically demonstrated to prolong survival in clinical trials involving gastric (Nakazato et al., 1994), colorectal (Mitomi et al., 1992), esophageal (Ogoshi et al., 1995), nasopharyngeal (Go and Chung, 1989), non-small-cell lung (Hayakawa et al., 1993), and breast cancer.

In addition to fungi, many so-called *Fu-Zhen* (tonifying) traditional Chinese herbs con-

tain bioactive polysaccharides and have been studied for their immunomodulatory and antitumor activity. These botanicals include common herbs such as *Actinidia chinensis* (Zhang and Lin, 1988), *Angelica sinensis* (Choy et al., 1994), *Astragalus membranaceus* (Huang et al., 1982), *Ligustrum lucidum* (Lau et al., 1994), *Panax ginseng* (Lee et al., 1997) as well as others (Table 2).

TCM usually uses herbal formulae and most useful TCM formulas for patients with cancer contain herbs with immunopotentiating activity from its polysaccharide content (Ito and Shimura, 1985a, 1985b). Examples of standard TCM (and Kampo) formulas with published experimental results demonstrating such immunostimulatory properties include *Xiao-Chai-Hu-Tang* [*Sho-saiko-to*] (Nagatsu et al., 1989), *Shi-quan-da-bu-tang* [*Juzen-taiho-to*] (Zee-Cheng, 1992) and *Bu-zhong-yi-qi-tang* [*Hochu-ekki-to*] (Li et al., 1999).

TABLE 2. REPRESENTATIVE TRADITIONAL CHINESE/KAMPO HERBS REPORTED TO CONTAIN BIOACTIVE POLYSACCHARIDES

<i>Acanthopanax Giralddii</i> Harms (Wang et al., 1992)
<i>Achyranthes bidentata</i> (Li and Li, 1997)
<i>Aloe</i> spp. (Zhang and Tizard, 1996)
<i>Atractylodes</i> (Inagaki et al., 2001)
<i>Beniscasa cerifera</i> (Kumazawa et al., 1985)
<i>Cinnamomum cortex</i> (Haranaka et al., 1985) ^a
<i>Curcuma zedoaria</i> (Kim et al., 2000)
<i>Codonopsis pilosula</i> (Wang et al., 1996)
<i>Dipsacus asperoides</i> (Zhang et al., 1997)
<i>Epimedium sagittatum</i> (Liu et al., 1991)
<i>Imperata cylindrica</i> (Pinilla and Luu, 1999)
<i>Isatis indigotica</i> (Xu and Lu, 1991)
<i>Malva verticillata</i> (Gonda et al., 1990)
<i>Panax notoginseng</i> (Gao et al., 1996) ^b
<i>Pseudostellaria heterophylla</i> (Wong et al., 1992)
<i>Radix bupleuri</i> (Geng and Chen, 1989)
<i>Radix glycyrrhizia</i> (Nose et al., 1998)
<i>Radix hadysari</i> (Lan et al., 1987) ^c
<i>Radix pseuoginseng</i> (Lin, 1988)
<i>Radix Rehmannia</i> (Xu, 1992)
<i>Salvia miltiorrhiza</i> (Hromakova et al., 1999)
<i>Zizyphi fructus</i> (Yamaoka et al., 1996)

^aHaranaka K, Satomi N, Sakurai A, Haranka R, Okada N, Kobayashi M. Antitumor activities and tumor necrosis factor producibility of traditional Chinese medicines and crude drugs. Cancer Immol Immunother 1985;20 (1):1–5.

^bGao H, Wang F, Lien EJ, Trousdale MD. Immunostimulating polysaccharides from *Panax notoginseng*. Pharm Res 1996;13(8):1196–1200.

^cLan ZF, Zhang ZL, Cheng GQ, Wang FL, Xi SF. Effects of radix hadysari polysaccharide on immunological function and transplanted tumors in mice [in Chinese]. Zhongguo Yao Li Xue Bao 1987;8(3):275–277.

CLINICAL OBSERVATIONS ON POLYSACCHARIDES AS ANTICANCER ADJUVANTS

It is important to realize that although TCM herbs that contain bioactive polysaccharides may derive some of their anticancer efficacy via immunopotentiality, many such herbs contain other complementary antineoplastic substances.

Conversely, such bioactive polysaccharides may have other antitumor actions beyond immunopotentiality. Such antitumor mechanisms include induction of cellular differentiation (Chen et al, 1997), antiangiogenesis (Kano, 1994), antimetastasis (Kobayashi et al., 1995). Furthermore, the polysaccharides have other applications beyond antitumor in patients with cancer. Such agents may also be useful in enhancing hematopoiesis (Liu et al., 1991), ameliorating side-effects of chemotherapy and radiation as well as generally improving the well-being of patients with cancer.

Most clinical trials of bioactive polysaccharides in cancer have used the agents with conventional treatments such as chemotherapy and radiation. It is important to note that such polysaccharides have been found to be clinically useful across a spectrum of solid tumors including colorectal, gastric, lung, and breast cancers, with the overall result of enhancing survival.

Regarding the issue of side-effects, in addition to the report of a low incidence of allergic reactions to individual herbs or polysaccharide drugs, major complications and/or organ toxicity has not yet been reported with this family of agents.

Because there is always concern regarding potential adverse interaction with conventional therapy with herbal or nutraceutical products, it is important to note that there have not been studies to suggest negative interactions with polysaccharide-derived agents and chemotherapy or radiation.

Not all polysaccharides are comparable and it is not prudent to rely entirely on *in vitro* data on one aspect of a polysaccharide's effectiveness (e.g., NK cell stimulation) as a basis of comparing various different polysaccharide-derived agents. While clinical trial data may not be available for many such agents, clinical

decision making should be guided by trial data, if available, or by the extent and quality of available medical literature on each agent. Furthermore, it is important to consider the choice of polysaccharide agent carefully because there can be significant differences in bioactivity secondary to differences in species, cultivation, method of extraction, formulation, as well as route and amount of dosage. Practicality, availability of agent, cost, and potential efficacy are the main clinical considerations when choosing a suitable polysaccharide to apply in a patient.

FUTURE DIRECTIONS IN THE DEVELOPMENT OF POLYSACCHARIDES AS CANCER ADJUVANTS

From existing laboratory and clinical evidence, it is certain that bioactive polysaccharides in TCM herbs are multifaceted and useful adjuncts in cancer care. However, lack of standardization and pharmacokinetic data among a spectrum of popular polysaccharide-based nutraceuticals, limited controlled trial data in the West on such agents, and relative lack of knowledge about these herbal agents among conventional cancer care professionals hamper the wide application of this unique class of agents. It is hoped that standardization as well as further clinical studies will be the basis for advancement in our knowledge and use of such agents.

REFERENCES

- Abel G, Czop JK. Stimulation of human monocyte beta-glucan receptors by glucan particles induces production of TNF-alpha and IL-1 beta. *Int J Immunopharmacol* 1992;14:1363-1373.
- Adachi Y, Ohno N, Ohsawa M, Oikawa S, Yadomae T. Macrophage activation in vitro by chemically cross-linked (1 → 3)-beta-D-glucans. *Chem Pharm Bull (Tokyo)* 1990;38:988-992.
- Borchers AT, Stern JS, Hackman RM, Keen CL, Gershwin ME. Mushrooms, tumors, and immunity. *Proc Soc Exp Biol Med* 1999;221:281-293.
- Chang RY. Potential application of Ganoderma polysaccharides in the immune surveillance and chemoprevention of cancer. In: Royse DJ, ed. *Mushroom Biology and Mushroom Products*. University Park, PA, Pennsylvania State University Press, 1996:153-159.

- Chen YJ, Shiao MS, Lee SS, Wang SY. Effect of *Cordyceps sinensis* on the proliferation and differentiation of human leukemic U937 cells. *Life Sci* 1997;60:2349–2359.
- Chenghua D, Xingliang Y, Xiaoman G, Yan W, Jingyan Z, Huibi X. A beta-D-glucan from the sclerotia of *pleurotus tuber-regium* (Fr.) Sing. *Carbohydr Res* 2000;328:629–633.
- Chihara G, Hamuro J, Maeda Y, Arai Y, Fukuoka F. Fractionation and purification of the polysaccharides with marked antitumor activity, especially lentinan, from *Lentinus edodes* (Berk.) Sing. (an edible mushroom). *Cancer Res* 1970;30:2776–2781.
- Chihara G. Preclinical evaluation of lentinan in animal models. *Adv Exp Med Biol* 1983;166:189–197.
- Chihara G, Hamuro J, Maeda YY, Shiio T, Sugita T, Takasuka N, Sasaki T. Antitumor and metastasis-inhibitory activities of lentinan as an immunomodulator: An overview. *Cancer Detect Prev Suppl* 1987;1:423–443.
- Choy YM, Leung KN, Cho CS, Wong CK, Pang PK. Immunopharmacological studies of low molecular weight polysaccharide from *Angelica sinensis*. *Am J Chin Med* 1994;22:137–145.
- Cleary A, Kelly GE, Husband AJ. The effect of molecular weight and beta 1,6 linkages on priming of macrophage function in mice by (1,3)-beta-D-glucan. *Immunol Cell Biol* 1999;77:395–403.
- Czop JK, Austen KF. Generation of leukotrienes by human monocytes upon stimulation of their beta-glucan receptor during phagocytosis. *Proc Natl Acad Sci* 1985a;82:2751–2755.
- Czop JK, Austen KF. A β -glucan inhibitable receptor on human monocytes. *J Immunol* 1985b;134:2588–2593.
- Geng JX, Chen SR. Isolation and identification of polysaccharides from radix *Bupleuri* [in Chinese]. *Zhongguo Zhong Yao Za Zhi* 1989;14:37–40.
- Go P, Chung CH. Adjuvant PSK immunotherapy in patients with carcinoma of the nasopharynx. *J Int Med Res* 1989;17:141–149.
- Gonda R, Tomoda M, Kanari M, Shimizu N, Yamada H. Constituents of the seed of *Malva verticillata*. VI. Characterization and immunological activities of a novel acidic polysaccharide. *Chem Pharm Bull (Tokyo)* 1990;38:2771–2774.
- Han SB, Lee CW, Jeon YJ, Hong ND, Yoo ID, Yang KH, Kim HM. The inhibitory effect of polysaccharides isolated from *Phellinus linteus* on tumor growth and metastasis. *Immunopharmacology* 1999;41:157–164.
- Hashimoto K, Suzuki I, Ohsawa M, Oikawa S, Yadomae T. Enhancement of hematopoietic response of mice by intraperitoneal administration of a beta-glucan, SSG, obtained from *Sclerotinia sclerotiorum*. *J Pharmacobiodyn* 1990;13:512–517.
- Hayakawa K, Mitsunashi N, Saito Y, Takahashi M, Katano S, Shiojima K, Furuta M, Niibe H. Effect of Krestin (PSK) as adjuvant treatment on the prognosis after radical radiotherapy in patients with non-small cell lung cancer. *Anticancer Res* 1993;13:1815–1820.
- Hishida I, Nanba H, Kuroda H. Antitumor activity exhibited by orally administered extract from fruit body of *Grifola frondosa* (Maitake). *Chem Pharm Bull (Tokyo)* 1988;36:1819–1827.
- Hromadkova Z, Ebringerova A, Valachovic P. Comparison of classical and ultrasound-assisted extraction of polysaccharides from *Salvia officinalis* L. *Ultrason Sonochem* 1999;5:163–168.
- Huang ZS, Lu GB, Guo JH. Studies on the polysaccharides of Huang Qi (*Astragalus*). *Acta Pharm Sin* 1982;17:200–204.
- Inagaki N, Komatsu Y, Sasaki H, Kiyohara H, Yamada H, Ishibashi H, Tansho S, Yamaguchi H, Abe S. Acidic polysaccharides from rhizomes of *Atractylodes lancea* as protective principle in *Candida*-infected mice. *Planta Med* 2001;67:428–431.
- Itoh H, Itoh H, Amano H, Noda H. Inhibitory action of a (1 \rightarrow 6)-beta-D-glucan-protein complex (FIII-2-b) isolated from *Agaricus blazei* Murill (“himematsutake”) on Meth A fibrosarcoma-bearing mice and its antitumor mechanism. *Jpn J Pharmacol* 1994;66:265–271.
- Ito H, Shimura K. Studies on the antitumor activity of traditional Chinese medicines (I) [in Japanese]. *Gan To Kagaku Ryoho* 1985a;12:2145–2148.
- Ito H, Shimura K. Studies on the antitumor activity of traditional Chinese medicines (II). The antitumor mechanism of traditional Chinese medicines [in Japanese]. *Gan To Kagaku Ryoho* 1985b;12:2149–2154.
- Jong SC, Donovick R. Antitumor and antiviral substances from fungi. *Adv Appl Microbiol* 1989;34:183–262.
- Kanayama H, Togami M, Adachi N, Fukai Y, Okumoto T. Studies of the antitumor active polysaccharides from the mycelia of *Poria cocos* Wolf. III. Antitumor activity against mouse tumors [in Japanese]. *Yakugaku Zasshi* 1986;106:307–312.
- Kandefer-Szerszen M, Kaweck Z. Water extracts of fungi as interferon inducers. *Acta Microbiol. Pol Acad* 1973;5:163–168.
- Kanoh T, Matsunaga K, Saito K, Fujii T. Suppression of in vivo tumor-induced angiogenesis by the protein-bound polysaccharide PSK. *In Vivo* 1994;8:247–250.
- Kariya K, Okamoto N, Fujimoto T, Inoue N, Kihara T, Sugie K, Yagita M, Kanzaki H, Mori T, Uchida A. Lysis of fresh human tumor cells by autologous peripheral blood lymphocytes and tumor-infiltrating lymphocytes activated by PSK. *Jpn J Cancer Res* 1991;82:1044–1050.
- Kim KI, Kim JW, Hong BS, Shin DH, Cho HY, Kim HK, Yang HC. Antitumor, genotoxicity and anticlastogenic activities of polysaccharide from *Curcuma zedoaria*. *Mol Cells* 2000;10:392–398.
- Kobayashi H, Matsunaga K, Oguchi Y. Antimetastatic effects of PSK (Krestin), a protein-bound polysaccharide obtained from basidiomycetes: An overview. *Cancer Epidemiol Biomarkers Prev* 1995;4:275–281.
- Komatsu N, Okubo S, Kikumoto S, Kimura K, Saito G. Host mediated antitumor action of *Schizophyllum commune*. *Gann* 1969;60:137–144.
- Kondo M, Torisu M. Evaluation of anticancer activity of a protein-bound polysaccharide PS-K (Krestin). In: Torisu M, Yoshida T, eds. *Basic Mechanisms and Clinical Treatment of Tumor Metastasis*. Academic Press, New York, 1985:623–636.

- Kumazawa Y, Nakatsuru Y, Fujisawa H, Nishimura C, Mizunoe K, Otsuka Y, Nomoto K. Lymphocyte activation by a polysaccharide fraction separated from hot water extracts of *Angelica acutiloba* Kitagawa. *J Pharmacobiodyn* 1985;8:417–424.
- Kuo YC, Tsai WJ, Shiao MS, Chen CF, Lin CY. *Cordyceps sinensis* as an immunomodulatory agent. *Am J Chin Med* 1996;24:111–125.
- Lau BH, Ruckle HC, Botolazzo T, Lui PD. Chinese medicinal herbs inhibit growth of murine renal cell carcinoma. *Cancer Biother* 1994;9:153–161.
- Lee YS, Chung IS, Lee IR, Kim KH, Hong WS, Yun YS. Activation of multiple effector pathways of immune system by the antineoplastic immunostimulator acidic polysaccharide ginsan isolated from *Panax ginseng*. *Anticancer Res* 1997;17:323–331.
- Leung MY, Fung KP, Choy YM. The isolation and characterization of an immunomodulatory and anti-tumor polysaccharide preparation from *Flammulina velutipes*. *Immunopharmacology* 1997;35:255–263.
- Li T, Tamada K, Abe K, Tada H, Onoe Y, Tatsugami K, Harada M, Kubo C, Nomoto K. The restoration of the antitumor T cell response from stress-induced suppression using a traditional Chinese herbal medicine Hochu-ekki-to (TJ-41:Bu-Zhong-Yi-Qi-Tang). *Immunopharmacology* 1999;43:11–21.
- Li ZK, Li DD. The immunomodulatory effect of *Achyranthes bidentata* polysaccharides [in Chinese]. *Yao Xue Xue Bao* 1997;32:881–887.
- Lin PF. Antitumor effect of *Actinidia chinensis* polysaccharide on murine tumor [in Chinese]. *Zhonghua Zhong Liu Za Zhi* 1988;10:441–444.
- Liu F, Ding G, Li J. Effects of *Epimedium sagittatum* Maxim. Polysaccharides on DNA synthesis of bone marrow cells of “yang deficiency” animal model caused by hydroxyurea [in Chinese]. *Zhongguo Zhong Yao Za Zhi* 1991;16:620–622.
- Liu F, Ooi VE, Fung MC. Analysis of immunomodulating cytokine mRNAs in the mouse induced by mushroom polysaccharides. *Life Sci* 1999;64:1005–1011.
- Misaki A, Kakuta M, Sasaki TM, Tanaka M, Miyahi H. Studies on interrelation of structure and antitumor effects of polysaccharides: Antitumor action of periodate-modified, branched (1,3)- β -D-glucan of *Auricularia auricula-judae* and other polysaccharides containing (1,3)-glycoside. *Carbohydr Res* 1981;92:115–129.
- Mitomi T, Tsuchiya S, Iijima N, Aso K, Suzuki K, Nishiyama K, Amano T, Takahashi T, Murayama N, Oka H, et al. Randomized, controlled study on adjuvant immunochemotherapy with PSK in curatively resected colorectal cancer: The Cooperative Study Group of Surgical Adjuvant Immunochemotherapy for Cancer of Colon and Rectum (Kanagawa). *Dis Colon Rectum* 1992;35:123–130.
- Mizuno T, Wasa T, Ito H, Suzuki C, Ukai N. Antitumor-active polysaccharides isolated from the fruiting body of *Hericium erinaceum*, an edible and medicinal mushroom called yamabushitake or houtou. *Biosci Biotechnol Biochem* 1992;56:347–348.
- Mueller A, Raptis J, Rice PJ, Kalbfleisch JH, Stout RD, Ensley HE, Browder W, Williams DL. The influence of glucan polymer structure and solution conformation on binding to (1 \rightarrow 3)- β -D-glucan receptors in a human monocyte-like cell line. *Glycobiology* 2000;10:339–346.
- Nagatsu Y, Inoue M, Ogihara Y. Modification of macrophage functions by Shosaikoto (kampo medicine) leads to enhancement of immune response. *Chem Pharm Bull (Tokyo)* 1989;27:1540–1542.
- Nakazato H, Koike A, Saji S, Ogawa N, Sakamoto J. Efficacy of immunochemotherapy as adjuvant treatment after curative resection of gastric cancer. *Lancet* 1994;343:1122–1126.
- Nose M, Terawaki K, Oguri K, Ogihara Y, Yoshimatsu K, Shimomura K. Activation of macrophages by crude polysaccharide fractions obtained from shoots of *Glycyrrhiza glabra* and hairy roots of *Glycyrrhiza uralensis* in vitro. *Biol Pharm Bull* 1998;21:1110–1112.
- Ogoshi K, Satou H, Isono K, Mitomi T, Endoh M, Sugita M. Immunotherapy for esophageal cancer: A randomized trial in combination with radiotherapy and radiochemotherapy. Cooperative Study Group for Esophageal Cancer in Japan. *Am J Clin Oncol* 1995;18:216–222.
- Ohno N, Furukawa M, Miura NN, Adachi Y, Motoi M, Yadomae T. Antitumor neta glucan from the cultured fruit body of *Agaricus blazei*. *Biol Pharm Bull* 2001;24:820–828.
- Ooi VE, Liu F. Immunomodulation and anti-cancer activity of polysaccharide-protein complexes. *Curr Med Chem* 2000;7:715–729.
- Peter G, Karoly V, Imre B, Janos F, Kaneko Y. Effects of lentinan on cytotoxicity of human lymphocytes. *Immunopharmacol Immunotoxicol* 1988;10:157–163.
- Pinilla V, Luu B. Isolation and partial characterization of immunostimulating polysaccharides from *Imperata cylindrica*. *Planta Med* 1999;65:549–552.
- Sakagami Y, Mizoguchi Y, Shin T, Seki S, Kobayashi K, Morisawa S, Yamamoto S. Effects of an anti-tumor polysaccharide, schizophyllan, on interferon-gamma and interleukin 2 production by peripheral blood mononuclear cells. *Biochem Biophys Res Commun* 1988;155:650–655.
- Stryer L. Carbohydrates. In: Stryer L, ed. *Biochemistry*. WH Freeman and Co., New York, 1995:447–477.
- Sun Y. Chinese medicinal herbs as biological response modifiers. In: Lotzova E, Herberman R, eds.: *Natural Immunity, Cancer, and Biological Response Modification*. Karger, Basel, 1986:206–211.
- Sun Y, Zhang YH, Yu GQ, et al. Effect of Fu-zheng therapy in the management of malignant diseases. *Chin Med J* 1981;61:97–101.
- Toritsu M, Hayashi Y, Ishimitsu T, Fujimura T, Iwasaki K, Katano N, Yamamoto H, Kimura Y, Takesue M, Kondo M, et al. Significant prolongation of disease-free period gained by oral polysaccharide K (PSK) administration after curative surgical operation of colorectal cancer. *Cancer Immunol Immunother* 1990;31:261–268.
- Tsujitani S, Kakeji Y, Orita H, Watanabe A, Kohnoe S, Baba H, Anai H, Maehara Y, Sugimachi K. Postoperative adjuvant immunochemotherapy and infiltration of

- dendritic cells for patients with advanced gastric cancer. *Anticancer Res* 1992;12:645–648.
- Wakui A, Kasai M, Konno K, Abe R, Kanamaru R, Takahashi K, Nakai Y, Yoshida Y, Koic H, Masuda H, et al. Randomized study of lentinan on patients with advanced gastric and colorectal cancer: Tohoku Lentinan Study Group [in Japanese]. *Gan To Kagaku Ryoho* 1986;13(4pt1):1050–1059.
- Wang JZ, Tsumura H, Shimura K, Ito H. Antitumor activity of polysaccharide from Chinese medicinal herb, *Acanthopanax giraldii* Harms. *Cancer Lett* 1992;65:79–84.
- Wasser SP, Weis AL. Therapeutic effects of substances occurring in higher Basidiomycetes mushrooms: A modern perspective. *Crit Rev Immunol* 1999;19:65–96.
- Wang ZT, Ng TB, Yeung HW, Xu G. Immunomodulatory effect of a polysaccharide-enriched preparation of *Codonopsis pilosula* roots. *Pharmacol* 1996;27:1347–50.
- Wong CK, Leung KN, Fung KP, Choy YM. Immunomodulatory and anti-tumor polysaccharides from medicinal plants. *J Int Med Res* 1994;22:229–312.
- Xia D, Lin ZB. Effects of Tremella polysaccharides on immune function in mice. *Zhongguo Yao Li Xue Bao* 1989;10:453–457.
- Xu JP. Research on liu wei Rehmannia oral liquid against side-effect of drugs of anti-tumor chemotherapy [in Chinese]. *Zhongguo Zhong Si Yi Jie He Za Zhi* 1992;12:734–737, 709–710.
- Xu YM, Lu PC. Experimental studies on immunostimulatory effects of *Isatis indigotica* polysaccharides [in Chinese]. *Zhong Xi Yi Jie He Za Zhi* 1991;6:357–359.
- Yamaoka Y, Kawakita T, Kaneko M, Nomoto K. A polysaccharide fraction of Ziziphi fructus in augmenting natural killer activity by oral administration. *Biol Pharm Bull* 1996;19:936–939.
- Yamasaki K, Sone S, Yamashita T, Ogura T. Synergistic induction of lymphokine-activated killer activity by IL-2 and the polysaccharide lentinan and therapy of spontaneous pulmonary metastases. *Cancer Immunol Immunother* 1989;29:871–892.
- Zee-cheng RK. Shi-quan-da-bu-tang, SQT. A potent Chinese biological response modifier in cancer immunotherapy, potentiation and detoxification of anticancer drugs. *Methods Find Exp Clin Pharmacol* 1992;14:725–736.
- Zhang L, Tizard IT. Activation of a mouse macrophage cell line by acemannan: the major carbohydrate fraction from *Aloe vera* gel. *Immunopharmacolgy* 1996;35:119–128.
- Zhang Y, Kiyohara H, Matsumoto T, Yamada H. Fractionation and chemical properties of immunomodulating polysaccharides from roots of *Dipsacus asperoides*. *Planta Med* 1997;63:393–399.
- Zhang YH, Liu YL, Yan SC. Effect of *Polyporus umbellatus* polysaccharide on function of macrophages in the peritoneal cavities of mice with liver lesions [in Chinese] *Zhong Xi Yi Jie He Za Zhi* 1991;11:225–226.

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