9.1) Unconstrained Bayesian topology inference (RevBayes).

```
##########################
# Reading in the Data #
#####################################
filenames <- v("18sublist.aln.fasta", "16sublist.aln.fasta")
n_data_subsets <- filenames.size()
for (i in 1:n_data_subsets) {
  data[i] = readDiscreteCharacterData(filenames[i])
taxa <- data[1].taxa()
n_species <- data[1].ntaxa()
n_branches <- 2 * n_species - 3
mvi = 0
mni = 0
Substitution Model
# Loop over each data subset #
for (i in 1:n_data_subsets) {
  er_prior[i] <- v(1,1,1,1,1,1)
  er[i] ~ dnDirichlet(er prior[i])
  moves[++mvi] = mvSimplexElementScale(er[i], alpha=10, tune=true, weight=3)
  pi_prior[i] <- v(1,1,1,1)
  pi[i] ~ dnDirichlet(pi_prior[i])
  moves[++mvi] = mvSimplexElementScale(pi[i], alpha=10, tune=true, weight=2)
  Q[i] := fnGTR(er[i],pi[i])
  alpha_prior_mean[i] <- 5.0
  alpha prior sd[i] <- 0.587405
  alpha[i] ~ dnLognormal(alpha prior mean[i], alpha prior sd[i])
  gamma_rates[i] := fnDiscretizeGamma( alpha[i], alpha[i], 4, false )
  moves[++mvi] = mvScale(alpha[i], weight=2)
  pinvar[i] ~ dnBeta(1,1)
  moves[++mvi] = mvScale(pinvar[i], lambda=0.1, tune=true, weight=2.0)
  moves[++mvi] = mvSlide(pinvar[i], delta=0.1, tune=true, weight=2.0)
}
###############
# Tree model #
###############
```

```
topology ~ dnUniformTopology(taxa=taxa)
moves[++mvi] = mvNNI(topology, weight=1.0)
moves[++mvi] = mvSPR(topology, weight=1.0)
for (i in 1:n_branches) {
 br lens[i] ~ dnExponential(10.0)
  moves[++mvi] = mvScale(br lens[i])
}
TL := sum(br lens)
# unite topology and branch length vector into phylogeny object
phylogeny := treeAssembly(topology, br lens)
#####################
# PhyloCTMC Model #
#####################
part rate mult ~ dnDirichlet(rep(1.0, n data subsets))
moves[++mvi] = mvSimplexElementScale(part_rate_mult, alpha=10, tune=true, weight=2)
for (i in 1:n_data_subsets) {
  part rate[i] := part rate mult[i] * n data subsets
for (i in 1:n data subsets) {
  phyloSeq[i] ~ dnPhyloCTMC(tree=phylogeny, Q=Q[i], branchRates=part_rate[i],
siteRates=gamma rates[i], plnv=pinvar[i], type="DNA")
  phyloSeq[i].clamp(data[i])
###########
# Analysis #
###########
mymodel = model(phylogeny)
# add monitors
monitors[++mni] = mnModel(filename="output/gene.log",printgen=10)
monitors[++mni] = mnFile(phylogeny, filename="output/gene.trees", printgen=100)
monitors[++mni] = mnScreen(alpha,printgen=100)
# run the analysis
mymcmc = mcmc(mymodel, moves, monitors)
mvmcmc.burnin(10000.1000)
mymcmc.run(300000)
# summarize output
treetrace = readTreeTrace("output/gene.trees")
treetrace.summarize()
map tree = mapTree(treetrace, "output/gene map.tre")
q()
```

9.2) Constrained Bayesian topology inference (RevBayes).

```
############################
# Reading in the Data #
#####################################
filenames <- v("18sublist.aln.fasta", "16sublist.aln.fasta")
n data subsets <- filenames.size()
for (i in 1:n_data_subsets) {
  data[i] = readDiscreteCharacterData(filenames[i])
# Get some useful variables from the data. We need these later on.
taxa <- data[1].taxa()
n_species <- data[1].ntaxa()
n_branches <- 2 * n_species - 3
mvi = 0
mni = 0
Substitution Model
# Loop over each data subset #
for (i in 1:n_data_subsets) {
  # exchangeability rates for partition i
  er_prior[i] <- v(1,1,1,1,1,1)
  er[i] ~ dnDirichlet(er_prior[i])
  moves[++mvi] = mvSimplexElementScale(er[i], alpha=10, tune=true, weight=3)
  # stationary frequencies for partition i
  pi_prior[i] <- v(1,1,1,1)
  pi[i] ~ dnDirichlet(pi prior[i])
  moves[++mvi] = mvSimplexElementScale(pi[i], alpha=10, tune=true, weight=2)
  # rate matrix for partition i
  Q[i] := fnGTR(er[i],pi[i])
  # +Gamma for partition i
  alpha_prior_mean[i] <- 5.0
  alpha_prior_sd[i] <- 0.587405
  alpha[i] ~ dnLognormal(alpha prior mean[i], alpha prior sd[i])
  gamma_rates[i] := fnDiscretizeGamma( alpha[i], alpha[i], 4, false )
  # add moves for the alpha parameter
  moves[++mvi] = mvScale(alpha[i], weight=2)
  # the probability of a site being invariable
  pinvar[i] ~ dnBeta(1,1)
  moves[++mvi] = mvScale(pinvar[i], lambda=0.1, tune=true, weight=2.0)
```

```
moves[++mvi] = mvSlide(pinvar[i], delta=0.1, tune=true, weight=2.0)
}
###############
# Tree model #
#############
#Constraints on clades
clade_A = clade("Resomia_ornicephala", "Frillagalma_vityazi")
clade B = clade("Cordagalma ordinatum", "Forskalia formosa", "Forskalia asymmetrica",
"Forskalia edwardsii", "Forskalia tholoides")
clade C = clade("Athorybia rosacea", "Agalma clausi", "Agalma elegans", "Agalma okenii",
"Nanomia_bijuga", "Halistemma_rubrum")
clade D = clade("Lychnagalma utricularia", "Physophora gilmeri", "Physophora hydrostatica",
clade C)
clade BD = clade( clade B, clade D)
clade ABC = clade( clade A, clade B, clade D)
clade_E = clade( clade_ABC, "Erenna_richardi", "Erenna_sirena", "Erenna laciniata".
"Stephalia_dilata", "Thermopalia_sp", "Marrus_claudanielis", "Physonect_sp",
"Stephanomia amphytridis")
clade constraints = [ clade A, clade B, clade C, clade D, clade BD, clade ABC, clade E]
topology ~ dnUniformTopology( taxa=taxa, constraints=clade constraints)
moves[++mvi] = mvNNI(topology, weight=1.0)
moves[++mvi] = mvSPR(topology, weight=1.0)
for (i in 1:n branches) {
 br lens[i] ~ dnExponential(10.0)
 moves[++mvi] = mvScale(br lens[i])
TL := sum(br lens)
phylogeny := treeAssembly(topology, br lens)
####################
# PhyloCTMC Model #
#####################
part rate mult ~ dnDirichlet(rep(1.0, n data subsets))
moves[++mvi] = mvSimplexElementScale(part rate mult, alpha=10, tune=true, weight=2)
for (i in 1:n_data_subsets) {
  part rate[i] := part rate mult[i] * n data subsets
for (i in 1:n data subsets) {
  phyloSeq[i] ~ dnPhyloCTMC(tree=phylogeny, Q=Q[i], branchRates=part_rate[i],
siteRates=gamma rates[i], plnv=pinvar[i], type="DNA")
  phyloSeq[i].clamp(data[i])
############
```

```
# Analysis #
############
mymodel = model(phylogeny)
# add monitors
monitors[++mni] = mnModel(filename="output/gene_ct.log",printgen=10)
monitors[++mni] = mnFile(phylogeny, filename="output/gene_ct.trees", printgen=100)
monitors[++mni] = mnScreen(alpha,printgen=100)
# run the analysis
mymcmc = mcmc(mymodel, moves, monitors)
mymcmc.burnin(10000,1000)
mymcmc.run(300000)
# summarize output
treetrace = readTreeTrace("output/gene_ct.trees")
treetrace.summarize()
map_tree = mapTree(treetrace, "output/gene_map_ct.tre")
q()
```

9.3) Bayesian timetree with fixed topology, with relaxed molecular clock.

```
Timetree full model specification and MCMC set up for estimating
   time-calibrated phylogenies under a strict clock model
setOption("useScaling", "true")
### Load the sequence alignment
D <- readDiscreteCharacterData(file="concatenate.fasta")
### initialize an iterator for the moves vector
mi = 1
n_sites <- D.nchar()
# Birth-death, rho=55/186, fixed topology
## read in the tree
backbone <- readTrees("timetree rev constrained/backbone rooted ultram.tre")[1]
n taxa <- backbone.ntips()
taxa <- backbone.taxa()
##### Birth-death process model #####
diversification ~ dnExponential(10.0)
turnover ~ dnBeta(2.0, 2.0)
denom := abs(1.0 - turnover)
birth rate := diversification / denom
death_rate := (turnover * diversification) / denom
### fix rho to 0.2957, since there are ~186 described species and we have sampled 55
rho <- 0.2957
### the root age is an independent stochastic node with a lognormal prior
#arbitrary numbers since we don't have time calibrations from fossils
mean_ra <- 0.5
stdv ra <- 0.001
mu_ra <- ln(mean_ra) - ((stdv_ra*stdv_ra) * 0.5)
root time ~ dnLnorm(mu ra, stdv ra, offset=1)
############################
#Constraints on clades#
############################
```

```
### the time tree is a stochastic node modeled by the constant rate birth-death process
(dnBDP)
tree prior = dnBDP(lambda=birth rate, mu=death rate, rho=rho, rootAge=root time,
samplingStrategy="uniform", condition="nTaxa", taxa=taxa)
timetree ~ dnConstrainedTopology(tree prior, backbone = backbone)
timetree.setValue(backbone)
moves[mi++] = mvScale(diversification,lambda=1.0,tune=true,weight=3.0)
moves[mi++] = mvSlide(turnover,delta=1.0,tune=true,weight=3.0)
###### Tree Moves #######
### add moves on the tree node times, including the root time, which is outside of the timetree
moves[mi++] = mvNodeTimeSlideUniform(timetree, weight=30.0)
### and moves for the tree topology
moves[mi++] = mvNNI(timetree, weight=8.0)
moves[mi++] = mvNarrow(timetree, weight=8.0)
moves[mi++] = mvFNPR(timetree, weight=8.0)
Timetree UCLN relaxed-clock model specification
n branches <- 2 * n taxa - 3
ucln mean ~ dnExponential(2.0)
ucln_sigma ~ dnExponential(3.0)
ucln var := ucln sigma * ucln sigma
ucln mu := In(ucln mean) - (ucln var * 0.5)
moves[mi++] = mvScale(ucln mean, lambda=1.0, tune=true, weight=4.0)
moves[mi++] = mvScale(ucln sigma, lambda=0.5, tune=true, weight=4.0)
for(i in 1:n branches){
  branch rates[i] ~ dnLnorm(ucln mu, ucln sigma)
  moves[mi++] = mvScale(branch rates[i],lambda=1.0,tune=true,weight=2.0)
moves[mi++] = mvVectorScale(branch rates,lambda=1.0,tune=true,weight=2.0)
moves[mi++] = mvVectorSingleElementScale(branch rates,lambda=30.0,tune=true,weight=1.0)
mean rt := mean(branch rates)
n species <- D.ntaxa()
er prior <-v(1,1,1,1,1,1)
er ~ dnDirichlet(er prior)
moves[mi++] = mvSimplexElementScale(er, alpha=10, tune=true, weight=3)
pi prior <- v(1,1,1,1)
pi ~ dnDirichlet(pi prior)
moves[mi++] = mvSimplexElementScale(pi, alpha=10, tune=true, weight=2)
Q := fnGTR(er,pi)
```

```
phySeq ~ dnPhyloCTMC(tree=timetree, Q=Q, branchRates=mean_rt, nSites=n_sites,
type="DNA")
phySeq.clamp(D)
#####################################
#####################################
mymodel = model(er)
### set up the monitors that will output parameter values to file and screen
monitors[1] = mnModel(filename="output/TimeTree_siphs_mcmc.log", printgen=10)
monitors[2] = mnFile(filename="output/TimeTree_siphs_mcmc.trees", printgen=10, timetree)
#monitors[3] = mnScreen(printgen=100, root time)
### workspace mcmc ###
mymcmc = mcmc(mymodel, monitors, moves)
### pre-burnin to tune the proposals ###
mymcmc.burnin(generations=5000,tuningInterval=100)
### run the MCMC ###
mymcmc.run(generations=10000)
### display proposal acceptance rates and tuning ###
mymcmc.operatorSummary()
### summarize the trees ###
tt = readTreeTrace("output/TimeTree siphs mcmc.trees", "clock")
tt.summarize()
### write MAP tree to file
mapTree(tt, "output/TimeTree siphs mcmc MAP.tre")
```

q()

9.4) Backbone tree topology used in 9.3.

```
(((Lensia_conoidea:0.12776145, Sulculeolaria_quadrivalvis:0.12776145):
0.0724035863, Muggiaea_atlantica:0.2001650363):0.01772580604, Chelophyes_appendiculata:
0.2178908423):0.0445064669):0.1207186703,(Sphaeronectes christiansonae:
0.02137209896, Sphaeronectes koellikeri: 0.02137209896): 0.3617438805): 0.06838539141,
(((Chuniphyes moserae:0.1070447673, Chuniphyes multidentata:0.1070447673):
0.1400302115, Kephyes_ovata: 0.2470749788): 0.1096919361, Crystallophyes_amygdalina:
0.3567669149):0.09473445601):0.05336762283, Tottonophyes_enigmatica:0.5048689937):
0.03684738417,((((Hippopodius hippopus:0.1629358231,Voqtia pentacantha:0.1629358231):
0.07817927636, Voqtia qlabra: 0.2411150995): 0.2274944812, Gymnopraia lapislazula:
0.4686095807):0.01821777298,Rosacea_flaccida:0.4868273537):0.02139388915,
(Desmophyes_haematogaster:0.19046821,Lilyopsis_fluoracantha:0.19046821):0.3177530328):
0.03349513508):0.0250076704, Craseoa_lathetica: 0.5667240483):0.02922570111,
((Nectadamas_diomedeae:0.2000425839, Nectopyramis_natans:0.2000425839):
0.03947243462, Athorybia_rosacea: 0.03947243462): 0.01150594871, Agalma_elegans:
0.05097838333):0.00782366888.Agalma clausi:0.05880205221):
0.03987968999, Halistemma_rubrum: 0.0986817422): 0.1363393864, Nanomia_bijuga:
0.2350211286):0.1574236295,((Physophora_gilmeri:0.1996894035,Physophora_hydrostatica:
0.1996894035):0.04503863, Lychnagalma utricularia:0.2447280335):0.1477167245):
0.01217998655,((((Forskalia edwardsii:0.06686802613,Forskalia tholoides:0.06686802613):
0.05276556323,Forskalia_formosa:0.1196335894):0.04922444644,Forskalia_asymmetrica:
0.1688580358):0.2108026519, Cordagalma_ordinatum: 0.3796606877):0.02496405696):
0.002302330604,(Frillagalma_vityazi:0.3758143426,Resomia_ornicephala:0.3758143426):
0.03111273266) : 0.1491307595, \\ (((Erenna\_richardi: 0.02029803167, Erenna\_sirena: 0.03111273266)) : 0.1491307595, \\ (((Erenna\_richardi: 0.02029803167, Erenna\_sirena: 0.03112766)) : 0.1491307596, \\ (((Erenna\_richardi: 0.02029803167, Erenna\_sirena: 0.0311276)) : 0.1491307596, \\ (((Erenna\_richardi: 0.0202980316, Erenna: 0.0202980)) : 0.1491307596, \\ (((Erenna\_richardi: 0.0202980316, Erenna: 0.0202980)) : 0.1491307596, \\ (((Erenna\_richardi: 0.0202980)) : 0.149130, \\ (((Erenna\_richardi: 
0.02029803167):0.009412289298, Erenna laciniata: 0.02971032097):
0.05264891477, Stephanomia amphytridis: 0.08235923574): 0.09325613073,
((Marrus claudanielis:0.08423382499, Physonect sp:0.08423382499):0.05121942472,
(Stephalia_dilata:0.03092743461,Thermopalia_sp:0.03092743461):0.1045258151):
0.04016211677):0.3804424682):0.06347821529):0.01324242897.(Bargmannia amoena:
0.09229156571, Bargmannia elongata: 0.09229156571): 0.5404869133): 0.0277360691,
(Apolemia lanosa: 0.04834004414, Apolemia rubriversa: 0.04834004414): 0.6121745039):
0.1083139728,((Rhizophysa eysenhardtii:0.1600232864,Rhizophysa filiformis:0.1600232864);
0.2114603422, Physalia_physalis: 0.3714836287): 0.3973448922): 0.2311714792,
(((Clytia_hemisphaerica:0.7494295556, Hydra_circumcincta:0.7494295556):
0.1469102, Ectopleura_dumortieri: 0.8963397556): 0.1036602444, ((Porpita_porpita:
0.1556570118. Velella velella: 0.1556570118): 0.4634878517. Staurocladia wellingtoni:
0.6191448635):0.3808551365):-4.884981308e-15);
```