

Thiaminase Activity in Alaskan Forage Fishes

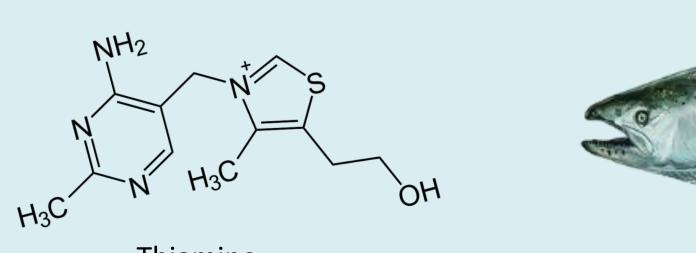
Drew Porter^{1,3}, Kathrine Howard², Cody Pinger³

¹Alaska Sea Grant, ²Alaska Department of Fish and Game, ³Alaska Fisheries Science Center, NOAA/NMFS

Correspondence: drew.porter@noaa.gov

OVERVIEW

- Thiamine (i.e., vitamin B1) is an essential enzyme co-factor for cell metabolism that must be obtained through diet.
- Thiamine deficiency complex has been attributed to early life stage mortality and population declines in salmonids from the Laurentian Great Lakes, California's Central Valley, and the Baltic Sea.
- Thiamine deficiency complex in salmon is hypothesized to be caused by consumption of prey containing thiaminase, an enzyme that destroys thiamine.
- Chinook Salmon (Oncorhynchus tshawytscha) populations in Alaska have undergone significant declines and reduced productivity in recent years.
- Deficient levels of thiamine have been measured in eggs and muscle tissue of Chinook Salmon from both Southeast Alaska (unpublished) and the Yukon River.¹
- To date, thiaminase activity has not been measured in Alaska forage fish that potentially serve as prey for Chinook Salmon.



OBJECTIVE

 Measure thiaminase activity in forage fish species collected from three different Alaska marine ecosystems.

COLLECTION AREAS



Figure 1. Collection areas for Alaska forage fish species. Fish were collected from the Northern Bering Sea (orange), the Arctic (blue), and Southeast Alaska (green).

RESULTS

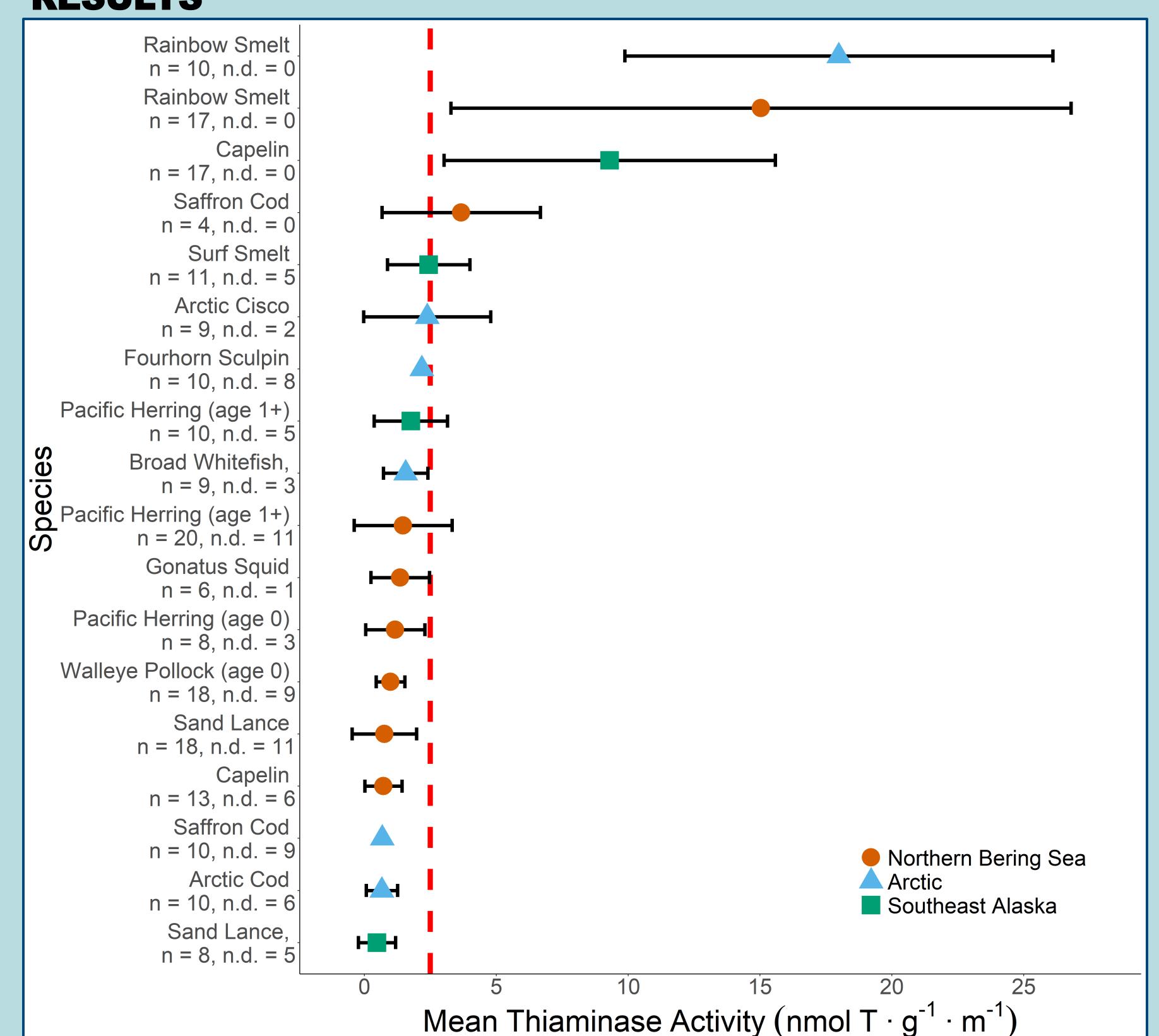


Figure 2. Mean thiaminase activity measured in species collected from Alaska marine ecosystems. Error bars $= \pm 1$ standard deviation, n = sample size, n.d. = samples with nodetectable thiaminase activity, red line = thiaminase activity threshold (>2.5 nmol T·g⁻¹·m⁻¹) observed to cause thiamine deficiency in consumers.²

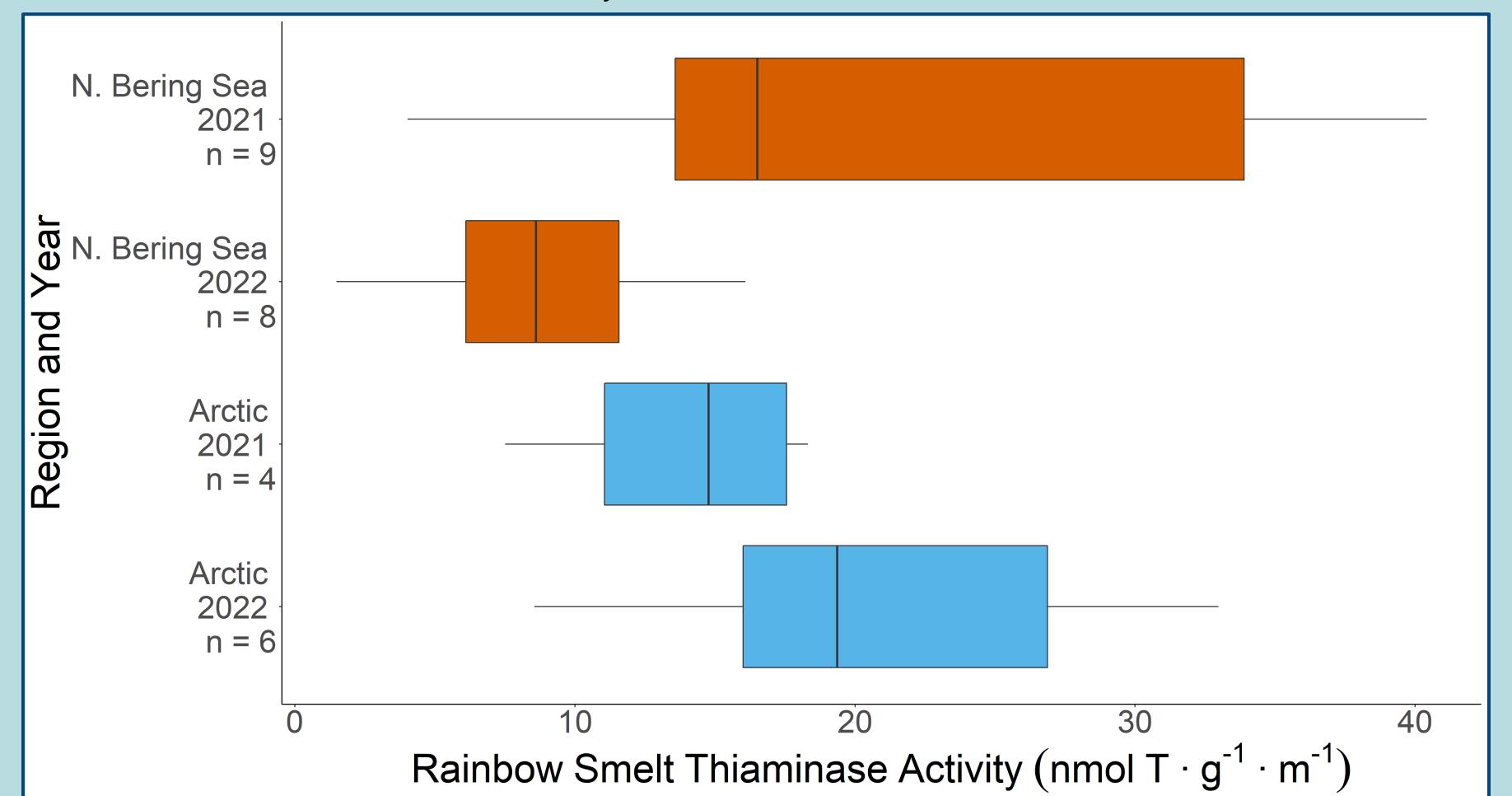
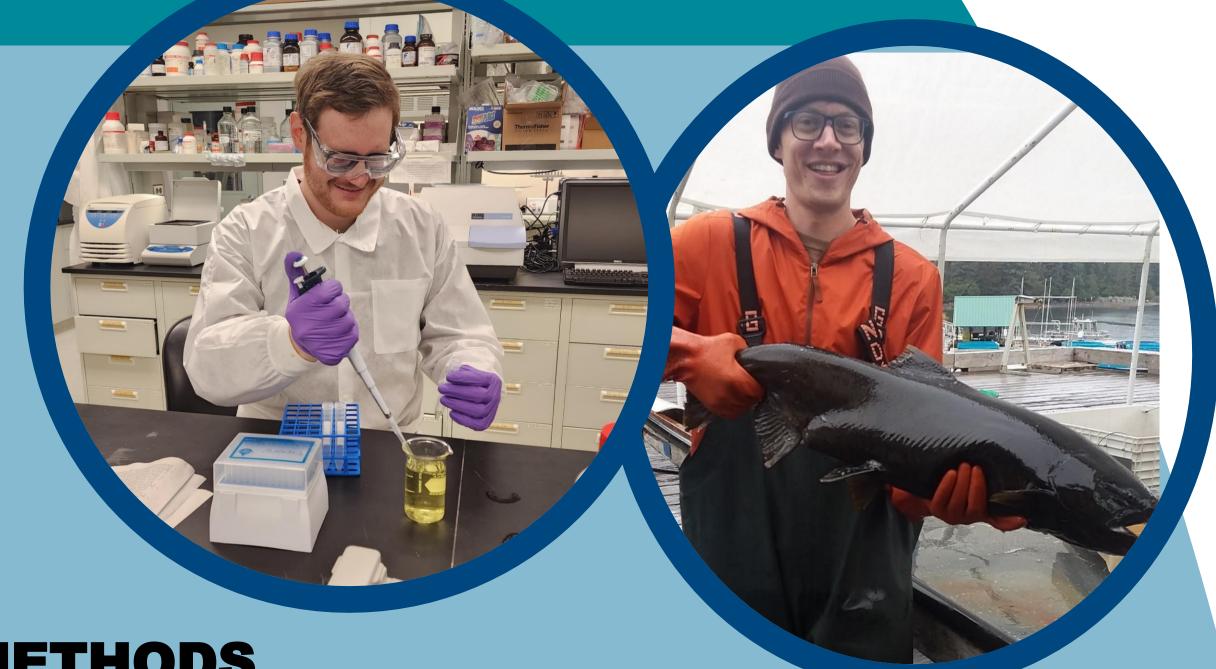


Figure 3. Thiaminase activity measured in Rainbow Smelt collected from two Alaska marine ecosystems over two years.



METHODS

Frozen fish samples were homogenized and analyzed for thiaminase activity (expressed as nanomoles of thiamine degraded per gram per minute) using the spectrophotometric 4-nitrothiophenol assay.3

MAIN FINDINGS

- Rainbow Smelt (Osmerus mordax dentex) displayed the highest thiaminase activity overall and displayed regional and interannual variability, consistent with trends observed elsewhere.4
- Capelin (Mallotus villosus) from Southeast Alaska had much higher thiaminase activity levels than individuals from the Bering Sea did. Notably, these populations are genetically distinct.⁵
- Thiaminase activity varied by species, region, and year. Several purported prey species of Chinook Salmon displayed thiaminase activity above a threshold level known to cause thiamine deficiency in consumers.

NEXT STEPS

- Measure these individuals for total body thiamine and lipid content to gain greater insight into prey quality in Alaska marine environments.
- Further investigate the observed interannual variability of thiaminase activity within species by analyzing archived individuals and individuals collected in subsequent years.
- Assess survey abundance of forage fishes with prey quality data to develop a spatial understanding of how thiaminase positive prey species are distributed in Alaska marine environments.

ACKNOWLEDGEMENTS

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REFERENCES

¹Larson, S., & Howard, K. (2019). Alaska Department of Fish and Game, Fishery Data Series No. 19-22, Anchorage

²Honeyfield, D. C., et al. (2016). North Pacific Anadromous Fish Commission, Bulletin No. 6, 21–31. ³Kraft, C. E., et al. (2014). PLoS ONE, 9(3), e92688.

⁴Tillitt, D. E., et al. (2005). Journal of Aquatic Animal Health, 17(1), 13–25.

⁵Dodson, J. J., et al. (2007). Molecular Ecology, 16(23), 5030–5043.