

# Genetic and Functional Drivers of Diffuse Large B Cell Lymphoma<sup>1</sup>

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Presented by Dominik Yang and Diana Lin

# Diffuse Large B Cell Lymphoma (DLBCL)

- Most common type of non-Hodgkin lymphoma (NHL)<sup>2</sup>
- Under a microscope, lymphoma cells appear larger than normal lymphocytes<sup>2</sup>
- Most of the time, starts in the lymph nodes but it can also start in organs and tissues outside the lymph nodes<sup>2</sup>
- 30-40% of cases are diagnosed in stage 1 or 2 (localized)<sup>2</sup>
- Rest of the cases have spread to lymph nodes above and below the diaphragm or to liver, spleen, or bone marrow<sup>2</sup>
- Treated with R-CHOP chemotherapy<sup>3</sup>
  - Rituximab
  - Cyclophosphamide
  - Hydroxydaunomycin
  - Oncovin
  - Prednisolone

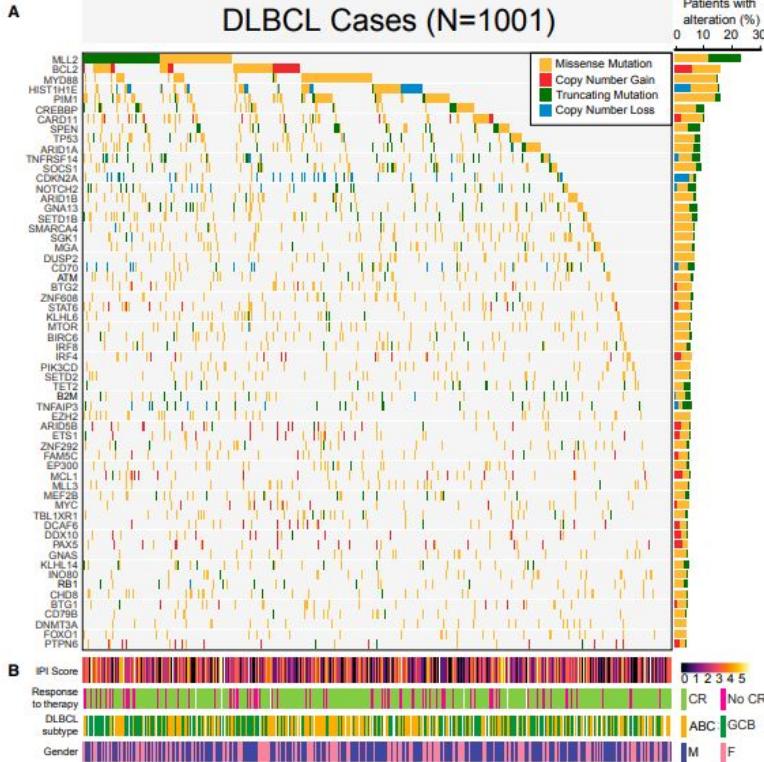
# Background

- Most common hematologic malignancy (annual incidence of over 100,000 cases worldwide)
- **Striking molecular and clinical heterogeneity makes it difficult to study this disease**
- Requires a large sample size to discover rare but important mutations

# Goals

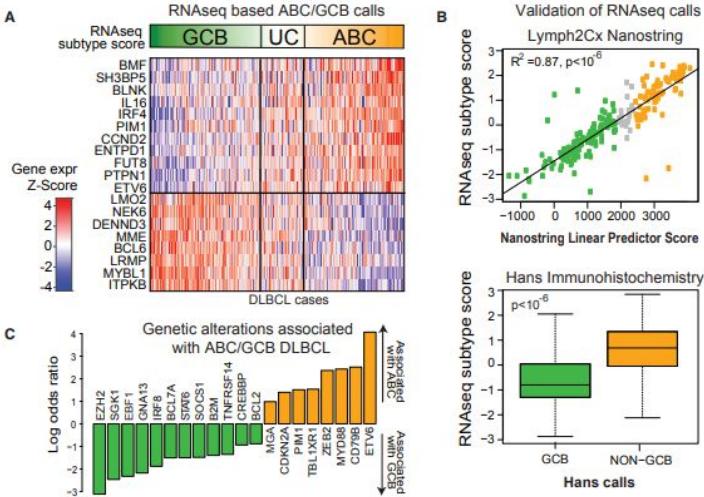
- Define the link between the functional impact of these genetic mutations to the growth of lymphoma cells
- Ultimately be able to create therapeutics that target specific genetic drivers of DLBCL

# Discovery of Genetic Drivers



- Whole-exome sequencing of 1001 DLBCLs
- 150 possible driver genes identified
- Mean of 7.75 mutations per DLBCL patient
- Driver genes express common mutation patterns:
- Missense mutation and copy number gains - oncogene (BCL2, CARD11, IRF4)
- Truncating mutations and copy number losses - tumor suppressor (SPEN, CDKN2A, TNFAIP3)

# Cell-of-Origin Effects



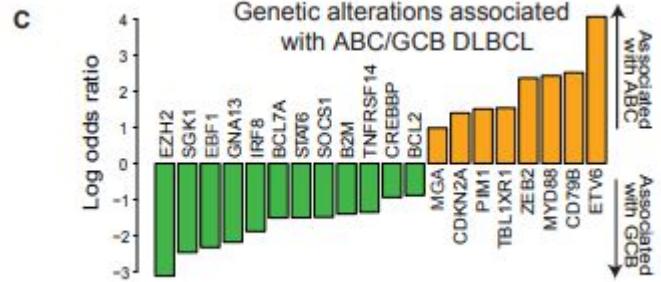
- RNA sequencing of gene expression in 775 patients to determine tumor cell of origin (A)
  - 313 activated B cell-like (ABC)
  - 331 germinal Center B cell-like (GCB)
  - 131 unclassified
- Validated by 2 complementary methods (B)
  - Nanostring assays
  - Immunohistochemistry-based Hans algorithm
- ABC DLBCLs had significantly worse overall survival than GCB DLBCLs

# Cell-of-Origin Effects

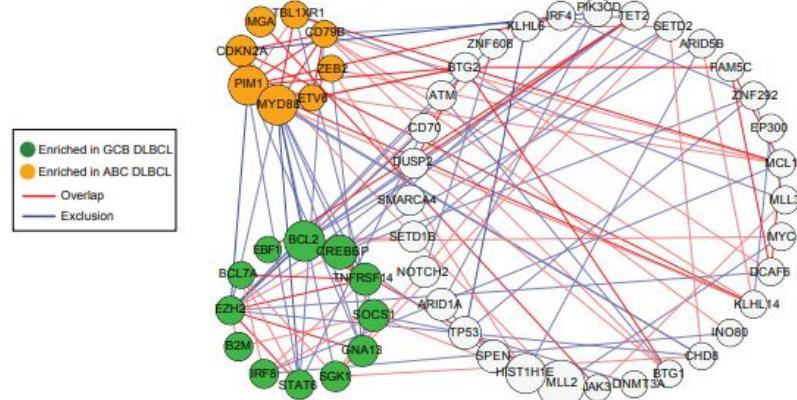
## Question:

Why do ABC DLBCLs have worse survival than GCB DLBCLs in response to standard therapy?

# Connectivity of Driver Genes



- ABC and GCB DLBCLs share most driver genes
- 20 genes had significantly different mutations (C)
- Of the 150 driver genes, 61 were significantly related to other driver genes (D)
  - MLL2 mutations mostly exclusive with MYC
  - TP53 mutations mutually exclusive with KLHL6



# Functional Genomics via CRISPR Screening

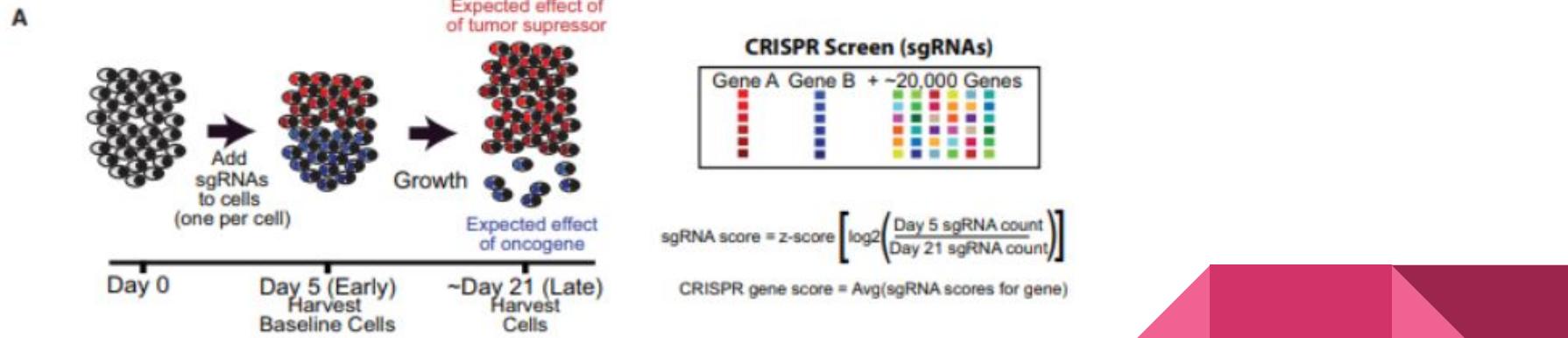
- 3 ABC, 2 GCB DLBCL, 1 BJAB (similar to GCB DLBCL)
- Selectively knock-out each gene in the DLBCLs to observe the effect on cell growth

QUESTION:

What did the authors expect to see from the effect of oncogenes vs tumor-suppressor genes?

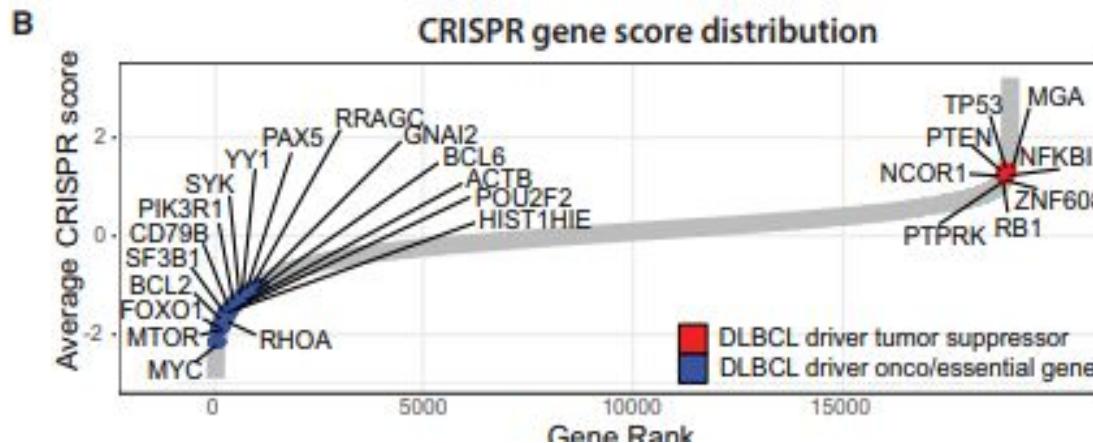
# Functional Genomics via CRISPR Screening

- Expected outcome: cells with oncogene-targeting sgRNAs would be depleted, cells expressing tumor-suppressor targeting sgRNA would be enriched



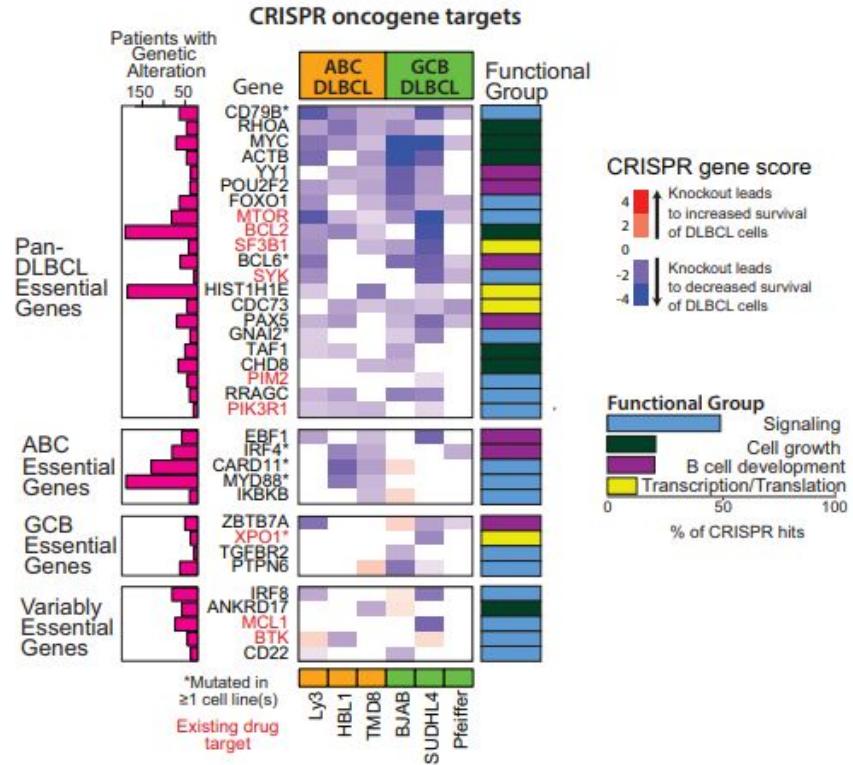
# Functional Genomics via CRISPR Screening

- 1956 essential genes with significant role on cell fitness
- CRISPR score: degree of alteration in each gene as a function of altered abundance of sgRNAs targeting the gene



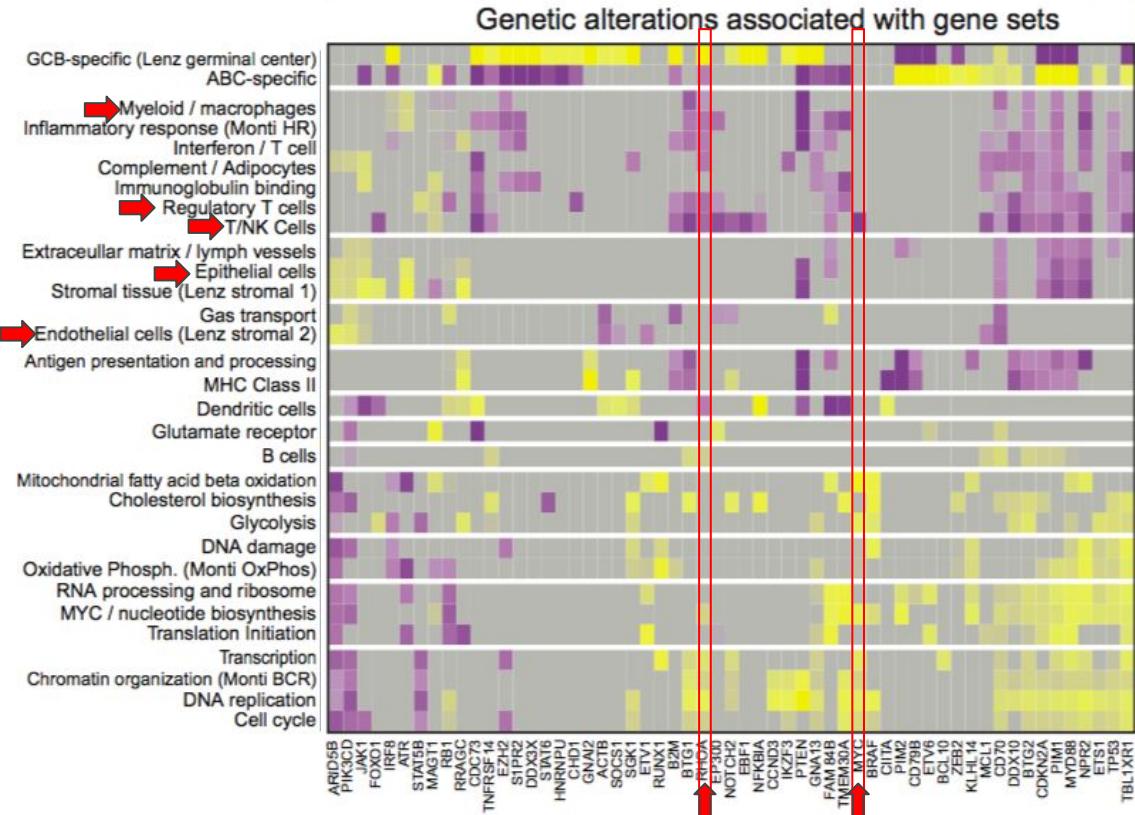
# Functional Genomics via CRISPR Screening

C



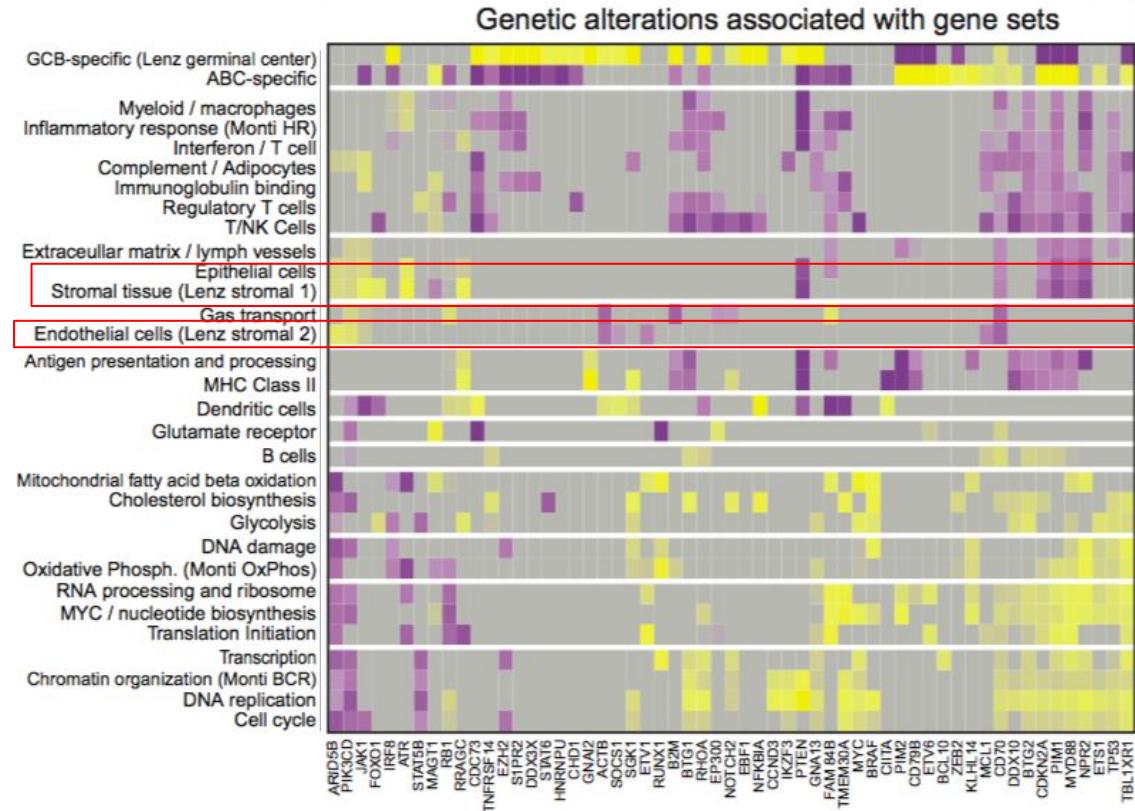
- 35 driver genes identified as oncogenes
  - 9 sub-type specific ABC or GCB
- Four functional groups:
  - Signalling
  - Cell growth
  - B cell development
  - Transcription and translation
- Lethal effects of gene knockdown also affects non-mutated cells

# Gene Expression Signature Associations with Genetic Alterations and Outcome



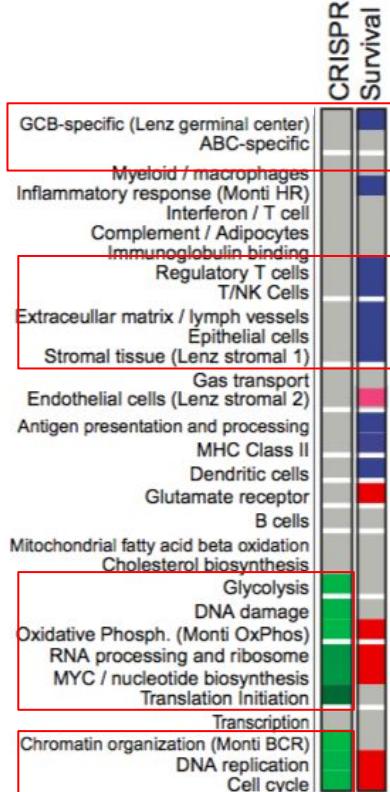
- Gene expression signatures with significant correlation fall into two classes:
  - Immune cell types (regulatory T cells, myeloid cells, NK cells)
  - Stromal connective tissue (epithelial and endothelial cells)
- *RHOA* and *MYC* mutations associated with proliferation-related signatures

# Gene Expression Signature Associations with Genetic Alterations and Outcome



- No reliable association between signalling pathways and mutations in those pathways
  - Non-malignant cells may have affected effective measurements
- No association between stromal signatures and mutations (insignificant)

# Gene Expression Signature Associations with Genetic Alterations and Outcome

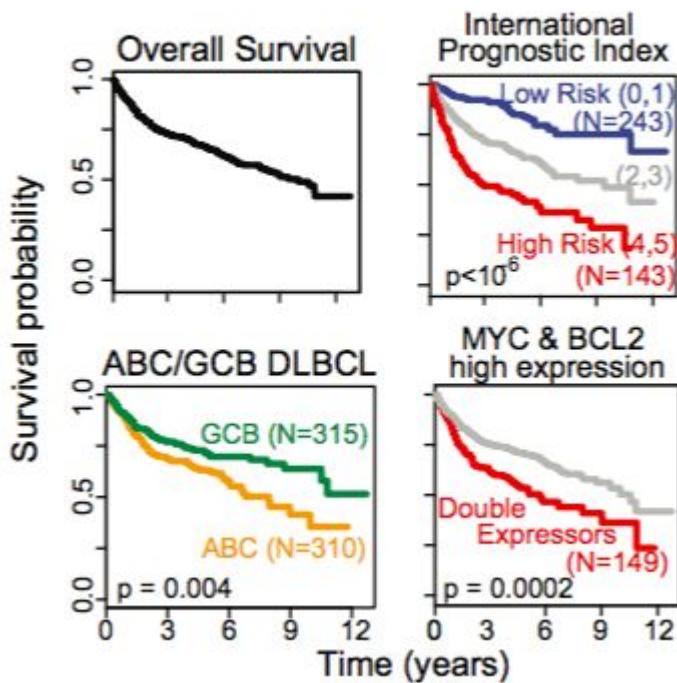


- Gene sets related to cancer-related processes contained significantly more CRISPR-identified driver genes
  - DNA damage, oxidative phosphorylation, DNA replication, cell-cycle, RNA processing
- GCB DLBCL associated with better survival rate
- Stromal and immune signatures also associated with better survival rate
  - Regulatory T cells
- Proliferation signatures associated with worse survival rates
  - MYC/Nucleotide biosynthesis



# Clinical Characteristics of DLBCL Driver Genes

A



- GCB DLBCL patients have a better survival probability than ABC DLBCL
- High expression of *MYC* & *BCL2* are associated with a worse prognosis and lower survival probability

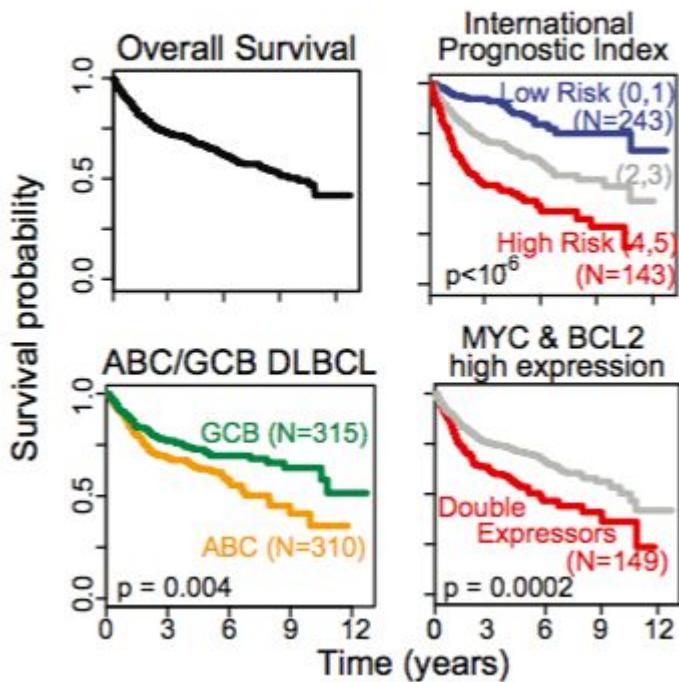
## Question:

What are the proteins that *MYC* & *BCL2* encode involved in?

B

# Clinical Characteristics of DLBCL Driver Genes

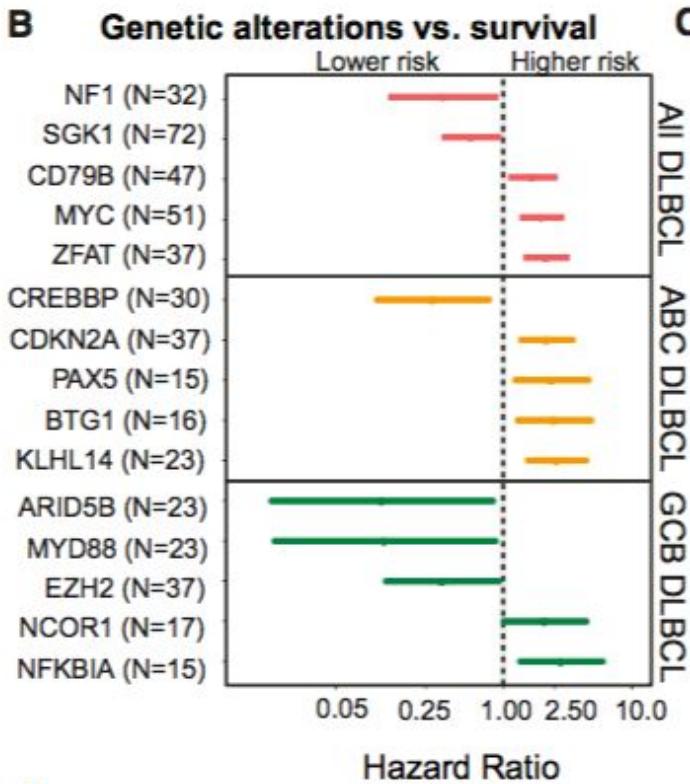
A



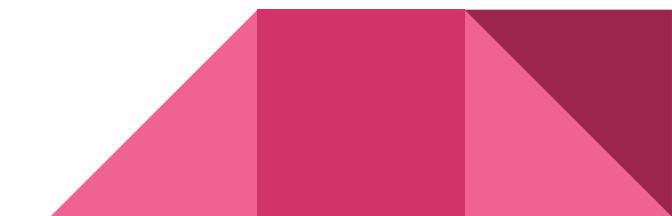
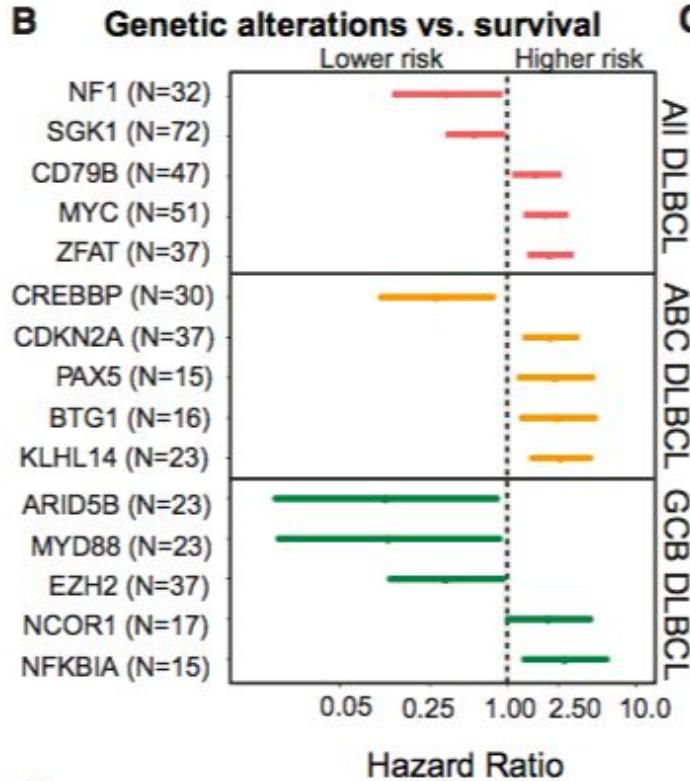
- GCB DLBCL patients have a better survival probability than ABC DLBCL
- High expression of *MYC* & *BCL2* are associated with a worse prognosis and lower survival probability
  - *MYC*: codes for a transcription factor involved in processes of the cell cycle, **apoptosis**, etc.<sup>4</sup>
  - *BCL2*: codes Bcl-2 family of regulator proteins involved in the regulation of **apoptosis**<sup>4</sup>

B

# Clinical Characteristics of DLBCL Driver Genes



# Clinical Characteristics of DLBCL Driver Genes



# Clinical Characteristics of DLBCL Driver Genes

**Question:**

What do a lot of these genes have in common?

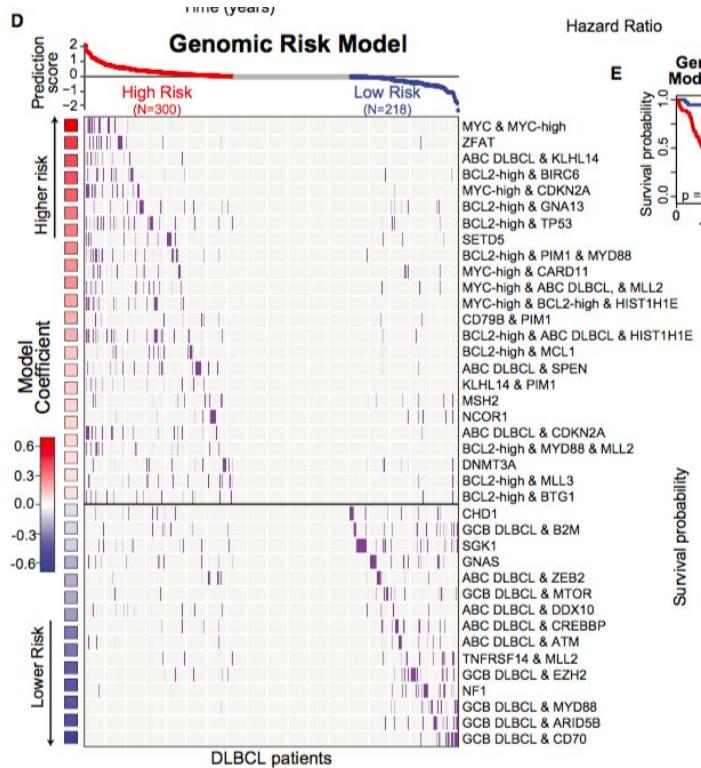
# Clinical Characteristics of DLBCL Driver Genes

- *NF1*: codes for protein neurofibromin which acts as a **tumour suppressor** protein<sup>5</sup>
- *SGK1*: codes for Serine/Threonine Kinase, involved in regulating **cell proliferation** and **apoptosis**<sup>6</sup>
- *CD79B*: codes for Igβ protein of **B cell** antigen<sup>7</sup>
- *ZFAT*: codes a zinc finger protein<sup>8</sup>
- *KLHL14*: codes for adapter protein for certain **ubiquitin** ligases<sup>9</sup>
- *BTG1*: codes for **anti-proliferative** protein involved in regulating cell growth and **proliferation**<sup>10</sup>
- *PAX5*: codes for **B-cell** lineage specific activator protein<sup>11</sup>
- *CDKN2A*: codes for a protein that stabilizes **tumour suppressor** protein p53<sup>12</sup>
- *CREBBP*: encodes CREB binding protein that is involved in growth control and **chromatin remodeling**
- *NCOR1*: codes for protein that promotes **chromatin condensation**<sup>13</sup>
- *NFKBIA*: codes for **NFκB** inhibitor<sup>14</sup>
- *EZH2*: codes for histone methyltransferase involved in **transcriptional repression**<sup>15</sup>
- *MYD88*: codes for adaptor protein involved in **immune** cell signaling
- *ARID5B*: encodes a DNA binding protein that is involved in **transcription regulation**<sup>16</sup>

# Comparison of Clinical vs Genomic Risk Model

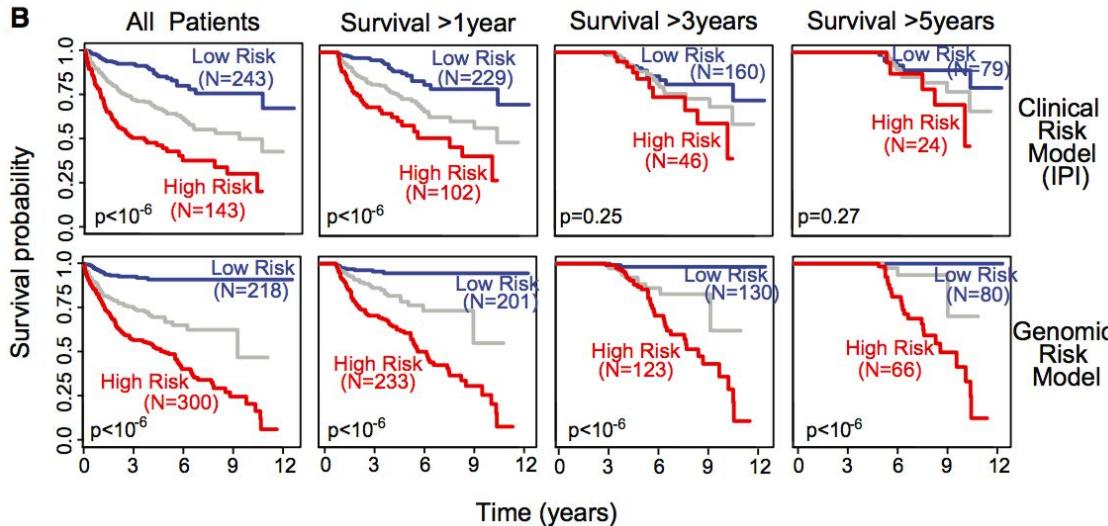
- Clinical Risk Model
  - Based on the International Prognostic Index (IPI) score<sup>18</sup>
    - Age > 60 (1 pt)
    - Stage III or IV disease (1pt)
    - Elevated serum LDH (1 pt)
      - High levels of LDH (lactate dehydrogenase) indicate tissue damage and cellular destruction
    - WHO score of 2, 3 or 4 (1 pt)<sup>18</sup>
      - 2: symptomatic; <50% in bed during the day
      - 3: symptomatic; >50% in bed during the day but not bedbound
      - 4: bedbound
    - > 1 extranodal site (1 pt)

# Comparison of Clinical vs Genomic Risk Model



- Genomic Risk Model
  - Based on genetic and molecular features
    - Known expression subgroups
      - ABC, GCB subgroups
      - MYC, BCL2 high expression
    - Mutations and copy-number events in the 150 DLBCL associated genes identified in this study

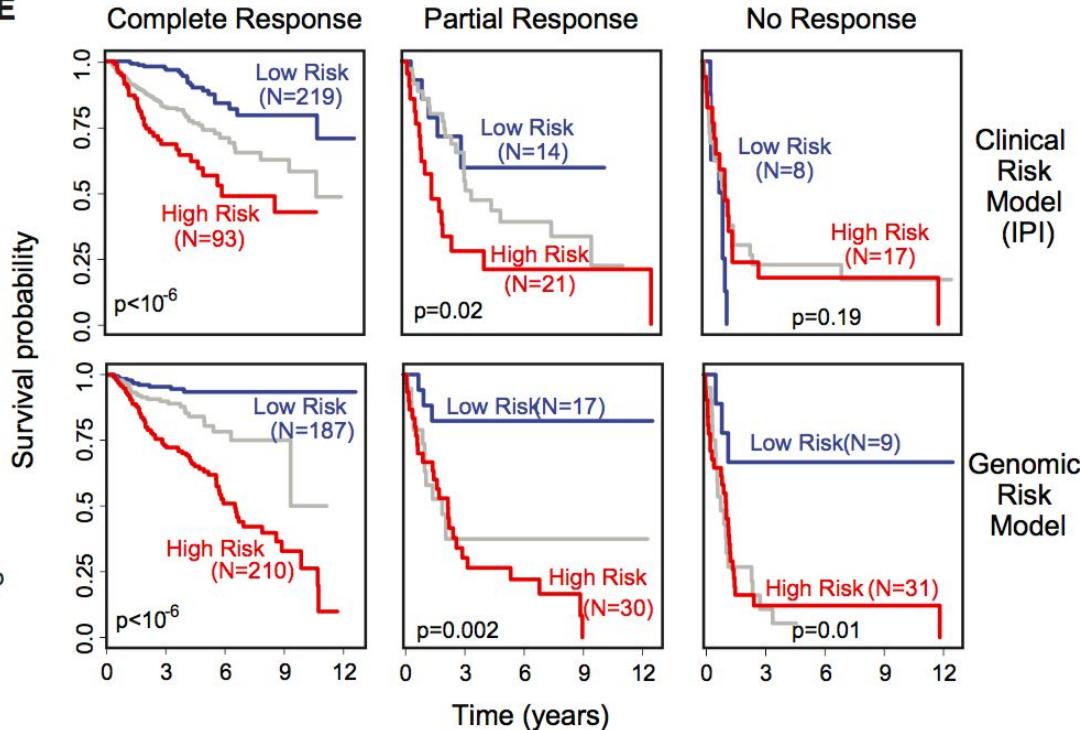
# Comparison of Clinical vs Genomic Risk Model



- Clinical Risk Model is highly prognostic in early stages (early mortality)
- Genomic Risk Model **better** at predicting long-term mortality (early and late mortality)

# Comparison of Clinical vs Genomic Risk Model

E



- Early mortality of those that completely respond to treatment can be predicted by the Genomic Risk Model
- Indicates that genomic profile of a patient strongly influences their prognostic risk

# Summary

- DLBCL is the most prevalent blood cancer in the world
- Diverse number of mutations causing DLBCL make it difficult to understand the impact of the mutations for therapeutic reasons
- 150 driver genes were identified, including 27 newly identified genes
- Functional screening showed many possible therapeutically targetable genes
- Can now distinguish which DLBCL patients would likely not benefit well from standard therapy, and can pursue new targeted therapies
- Genomic risk model can be used clinically: measure cell of origin, BCL2 and MYC expression, targeted sequencing of driver genes

# Future Outlook

- Better understanding of the combined effects of the gene combinations and potential to aid in developing new therapeutic approaches
- Apply genomic methodologies to large cohorts and uncover patterns in a variety of other cancers with genetic heterogeneity that would not be discovered in small sample sizes
- Clinical and functional drivers of DLBCL identified → first step in improving outcomes for DLBCL

# Strengths and limitations

- Pros:
  - Large sample size
  - Lots of statistical verification for their results
  - Provides novel and valuable information that can be translated into a clinical setting
- Cons:
  - A lot of their starting data is reliant on other researchers' data
  - There were many cases where the authors did not explain why they chose to do certain things
  - Requires a lot of statistical educational background to fully understand some tests they conducted
  - Requires a deep understanding of biological methodology and techniques to fully understand the paper on another level

# Questions?



# References

1. Reddy A, Zhang J, Davis NS, et al. Genetic and Functional Drivers of Diffuse Large B+Cell Lymphoma. *CELL Cell.* 2017;171(2):481-494.e415.
2. Dearden C, Matutes E. Non-Hodgkin's lymphoma. *MPMED Medicine.* 2004;32(6):78-84.
3. Schmitz N, Zeynalova S, Nickelsen M, et al. CNS International Prognostic Index: A Risk Model for CNS Relapse in Patients With Diffuse Large B-Cell Lymphoma Treated With R-CHOP. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology.* 2016;34(26):3150-3156.
4. Miao Y, Hu S, Lu X, et al. Double-hit follicular lymphoma with MYC and BCL2 translocations: a study of 7 cases with a review of literature. *YHUPA Human Pathology.* 2016;58:72-77.
5. Susanne AMT, Lauren F, Margaret RW. Review Article : NF1 Mutations and Molecular Testing. *Journal of Child Neurology.* 2002;17(8):555-561.
6. Di Cristofano A. SGK1 : The Dark Side of PI3K Signaling. *BS:CTDB Current Topics in Developmental Biology.* 2017;123:49-71.
7. Chu PG, Arber DA. CD79: A Review. *Applied Immunohistochemistry & Molecular Morphology.* 2001;9(2):97-106.
8. Tsunoda T, Shirasawa S. Roles of ZFAT in haematopoiesis, angiogenesis and cancer development. *Anticancer research.* 2013;33(7):2833-2837.
9. Braybrook C, Warry G, Howell G, et al. Identification and characterization of KLHL4, a novel human homologue of the Drosophila Kelch gene that maps within the X-linked cleft palate and Ankyloglossia (CPX) critical region. *Genomics.* 2001;72(2):128-136.
10. Matsuda S, Rouault J, Magaud J, Berthet C. In search of a function for the TIS21/PC3/BTG1/TOB family. *FEBS letters.* 2001;497(2-3):2-3.
11. Medvedovic J, Ebert A, Tagoh H, Busslinger M. Pax5 : A Master Regulator of B Cell Development and Leukemogenesis. *BS:AI.* 2011;111:179-206.
12. Kannengiesser C, Avril MF, Spatz A, Laud K, Lenoir GM, Bressac-de-Paillerets B. CDKN2A as a uveal and cutaneous melanoma susceptibility gene. *Genes, chromosomes & cancer.* 2003;38(3):265-268.
13. Wang W, Song X-W, Bu X-M, Zhang N, Zhao C-H. PDCD2 and NCoR1 as putative tumor suppressors in gastric gastrointestinal stromal tumors. *Cell Oncol Cellular Oncology : The official journal of the International Society for Cellular Oncology.* 2016;39(2):129-137.
14. Geng P, Ou J, Li J, et al. Genetic Association Between NFKBIA -881A>G Polymorphism and Cancer Susceptibility. *Medicine.* 2015;94(31).
15. Crea F, Fornaro L, Bocci G, et al. EZH2 inhibition: targeting the crossroad of tumor invasion and angiogenesis. *Cancer Metastasis Rev Cancer and Metastasis Reviews.* 2012;31(3-4):753-761.
16. Gutiérrez-Camino, LÚpez-LÚpez E, Martín-Guerrero I, et al. Intron 3 of the ARID5B gene: a hot spot for acute lymphoblastic leukemia susceptibility. *Journal of cancer research and clinical oncology.* 2013;139(11):1879-1886.
17. Wilder RB, Rodriguez MA, Medeiros LJ, et al. International prognostic index-based outcomes for diffuse large B-cell lymphomas. *CNCR Cancer.* 2002;94(12):3083-3088.
18. Oken MM, Creech RH, Tormey DC, et al. Toxicity and response criteria of the Eastern Cooperative Oncology Group. *AMERICAN JOURNAL OF CLINICAL ONCOLOGY.* 1982;5(6):649-656.