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On the Road to Sustainable Energy Storage Technologies: Synthesis of Anodes for Na-Ion Batteries from Biowaste

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Abstract: Hard carbon is one of the most promising anode materials for sodium-ion batteries. In this work, new types of biomass-derived hard carbons were obtained through pyrolysis of different kinds of agro-industrial biowaste (corncob, apple pomace, olive mill solid waste, defatted grape seed and dried grape skin). Furthermore, the influence of pretreating the biowaste samples by hydrothermal carbonization and acid hydrolysis was also studied. Except for the olive mill solid waste, discharge capacities typical of biowaste-derived hard carbons were obtained in every case ($\approx 300 \text{ mAh}\cdot\text{g}^{-1}$ at C/15). Furthermore, it seems that hydrothermal carbonization could improve the discharge capacity of biowaste samples derived from different nature at high cycling rates, which are the closest conditions to real applications.

Keywords: sustainable batteries; Na-ion battery; biowaste; hard carbon; anode



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1. Introduction

The European Green Deal is the strategy of the European Union (UE) to make Europe climate neutral to 2050 and, at the same time, a world leader in climate issues. This plan is based on four main pillars: industrial innovation, bioeconomy, energy innovation and circular economy [1]. In short, there is an urgent need to use innovation to change the current status quo into a scenario where: (1) biological raw materials are used as a source of products and energy, (2) energy comes from renewable energy sources that are climate-neutral, and (3) waste is reintroduced into the economy in the form of secondary raw materials. On the road to innovation in renewable energy, it is necessary to advance in electrochemical energy storage technologies, with the aim of overcoming the limitations of intermittent production of solar, wind or wave energy. Innovation in this field currently requires the design of sustainable batteries that, among other aspects, are not based on critical materials such as lithium or that incorporate materials fabricated from biological feedstock [2].

For the past few years, sodium-ion batteries (SIBs) have received increased attention as an alternative to lithium-ion batteries (LIBs) for stationary energy storage, due to the fact that sodium and lithium have similar chemical properties, such as the intercalation chemistry and, contrary to the case of lithium, there are abundant reserves of sodium on earth [3–5]. Even if fundamental principles of SIBs and LIBs are almost the same, Na-ion batteries usually exhibit lower specific capacities, shorter cycle life and poorer rate

capabilities, because of the higher mass and radius of Na ($22.99 \text{ g}\cdot\text{mol}^{-1}$; 1.02 \AA) when compared to those of Li ($6.94 \text{ g}\cdot\text{mol}^{-1}$; 0.76 \AA) [6]. Different kind of materials and mainly layered oxides, polyanionic and Prussian Blue systems have been studied as cathodes for SIBs [7–10].

Regarding anode materials many promising non-carbonaceous (e.g., Ti-, Ge- and P-based compounds) [11–16] and carbonaceous materials such as carbon black, hard carbon, carbon spheres, hollow carbon nanowires, carbon nanofibers, carbon nanotubes, graphite and graphene have been tested for SIBs [17–20]. The carbonaceous materials must facilitate the intercalation/deintercalation of Na^+ and accelerate the kinetics, but most of them suffer from low reversible capacity and severe volume expansion during the electrochemical reaction process. However, hard carbon presents outstanding electrochemical features, such as considerable reversible specific capacity and excellent electrical conductivity, being today the most promising anode material [21,22].

Hard carbon is normally obtained from pyrolysis of organic precursors at temperatures above $1000 \text{ }^\circ\text{C}$ and heating rates between $1\text{--}10 \text{ }^\circ\text{C min}^{-1}$ under inert atmospheres [23]. The organic nature of the hard carbon precursors gives the opportunity to incorporate biological raw materials in the fabrication of battery components; in fact, the use of bio-based precursors is a common approach when preparing hard carbons for SIBs [24]. Mainly, two groups of bio-precursors can be distinguished: on the one hand, reagent grade bio-chemicals, such as sucrose, glucose, cellulose and lignin [25–28]. On the other hand, biowaste coming from forestry and agricultural activities [29–33]. In general, anodes synthetized from reagent grade bio-chemicals show better electrochemical performance compared to those obtained from biowaste. However, taking into account that the use of sugars as raw materials for non-alimentary purposes is controversial and that the utilization of biowaste for manufacturing bio-products is at the core of the bioeconomy and the circular economy, it is worth paying particular attention to production of hard carbons from biowaste.

With the objective of overcoming the lower electrochemical performance of hard carbons obtained from direct pyrolysis of biowaste, this work evaluates the effect of acid hydrolysis (AH) and hydrothermal carbonization (HTC) as pretreatment steps for biowaste prior to pyrolysis. Acid hydrolysis, both in dilute and concentrated forms, is a conventional process to convert cellulose and hemicellulose into fermentable sugars, normally as a previous step to bioethanol production via fermentation [34,35]. On the other hand, HTC is a carbonization process happening in the reaction between biomass and water under mild conditions ($180\text{--}280 \text{ }^\circ\text{C}$, $1\text{--}72 \text{ h}$, autogenous pressure $2\text{--}6 \text{ MPa}$), normally focused on producing a carbonaceous substance (hydrochar) to be used as solid fuel, activated carbon precursor or soil amendment [36,37]. In the end, HTC is also a hydrolysis process caused by the action of temperature and the weak acid nature of water [38], with the advantage of being a water-based environmentally friendly strategy [39,40]. These two processes partially dissolve hemicellulose and cellulose, converting the initial biowaste into a lignin-rich biomass. Considering that lignin is one of the most used biomaterial as precursor for hard carbons due to its high crosslinking and non-crystalline biopolymer [41], the approach of this work is to integrate both pretreatments in the synthesis route of hard carbons from biowaste, which could be connected with better electrochemical performance.

2. Results and Discussion

2.1. Characterization of Biowaste Samples

The different biowaste materials were first subjected to proximate, elemental and constituent analysis in order to observe the differences in composition among them. Table 1 gathers the results obtained from these analyses, where glucans are mainly representative of cellulose (but include also starch) and xylans-arabinans represent hemicellulose.

Table 1. Proximate, elemental and constituent analysis of the raw precursors (wt.%).

	Corn Cob	Apple Pomace	Olive Mill Solid Waste	Deffated Grape Seed	Dry Grape Skin
Proximate analysis ¹	<i>Moisture</i>	8.70	5.80	6.70	13.90
	<i>Volatiles</i>	72.70	73.90	68.40	59.00
	<i>Fixed carbon</i> ³	16.80	18.30	16.10	24.60
	<i>Ashes</i>	1.80	2.00	8.80	2.50
	<i>Carbon</i>	43.50	41.70	44.10	47.40
Elemental analysis ¹	<i>Hydrogen</i>	7.00	7.80	6.30	6.70
	<i>Nitrogen</i>	0.40	0.60	1.60	1.90
	<i>Sulfur</i>	0.20	0.20	0.20	0.20
	<i>Oxygen</i>	42.80	48.10	35.80	38.50
	<i>Others</i> ³	6.10	1.60	12.00	5.30
Constituent analysis ²	<i>Extractives</i>	9.60	44.30	37.60	9.70
	<i>Glucans</i>	33.20	15.50	9.50	4.50
	<i>Xylans</i>	27.60	6.60	11.90	10.50
	<i>Arabinans</i>	2.00	4.20	0.90	0.30
	<i>Lignin</i>	11.50	10.30	26.40	56.50
	<i>Others</i> ³	16.10	19.10	13.70	18.50

¹ Wet basis (as received); ² Dry basis; ³ By difference.

Regarding the tentative analysis, all samples are mainly composed of volatile matter (59–74 wt.%) and fixed carbon (16–25 wt.%), with lower values of moisture (6–14 wt.%) and ash (2–9 wt.%). Considering that the objective is to produce a carbonized product, the ideal sample would be one with a high fixed carbon content (so that the yield to carbonized product is high) and low ash content (so that the carbonized product does not present a relevant quantity of inorganic substances which would influence the electrochemical performance). In this sense, *a priori*, it could be said that the two samples of biowaste from wine production (GSEED and GSINK) would be the most favorable ones for a possible industrial use of these precursors. Elemental analysis exhibits the usual composition in biowaste materials, mostly composed by carbon and oxygen, and with very low quantities of sulphur and nitrogen [42].

Subsequent elemental analysis of corn cob and olive mill solid waste derived carbons is displayed in Table 2. It can be seen that the carbon percentage in both pyrolysed and HTC pretreated samples (CORN_1200, CORNHTC_1200, OLI_1200 and OLIHTC_1200) is similar. On the other hand, samples subjected to a previous acid digestion (CORN_PHOSHTC_1200 and OLI_PHOSHTC_1200) show lower C weight percentages. This can be related to higher amounts of inorganic impurities in the samples or to the presence of a higher content in volatile compounds in the final carbon sample compared to the directly pyrolyzed or only pretreated by HTC.

Table 2. Elemental analysis of corn cob and olive mill solid waste derived hard carbons (wt.%).

	Samples	Carbon	Hydrogen	Nitrogen
CORN	<i>1200</i>	80.08	0.45	1.56
	<i>HTC_1200</i>	89.31	0.25	2.68
	<i>PHOSHTC_1200</i>	51.69	0.27	1.60
OLI	<i>1200</i>	70.68	0.83	1.59
	<i>HTC_1200</i>	70.76	0.39	2.27
	<i>PHOSHTC_1200</i>	49.94	0.13	1.46

At last, it can be said that the bio-precursors chosen for this study are different enough regarding constituents' composition, which could lead to present diverse behavior when subjected to the different treatment and pretreatments described above. The results of

the study of fixed carbon, ashes, glucans (cellulose and starch), hemicellulose (xylans and arabinans) and lignin are described in Figure 1. As it can be seen, the amount of fixed carbon is comparable between each biomass. Nevertheless, in the case of ashes, a huge different quantity is noticed between OLI (8.80 wt.%) and the other samples (<4.10 wt.%). Glucans percentages of the samples seem to be heterogeneous and CORN shows the highest content (33.2 wt.%), probably due to the presence of starch, followed by APP (15.5 wt.%). Notwithstanding, the content of hemicellulose is homogeneous, CORN with the highest hemicellulose content (29.6 wt.%) is far from the second one, GSEED (13.5 wt.%). Concerning lignin, the grape-derived materials show the highest values (56.5 wt.% GSEED and 45.2 wt.% GSkin) followed by OLI (26.4 wt.%), while CORN (11.5 wt.%) and APP (10.3 wt.%) present quite lower lignin quantity.

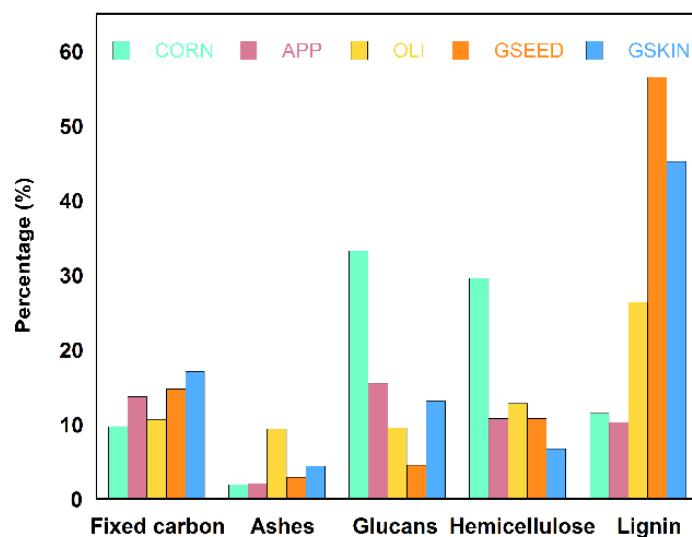


Figure 1. Fixed carbon, ashes, glucans, hemicellulose and lignin analysis comparison of the biowaste samples.

2.2. Characterization of Hard Carbons

Figure 2 shows the diffraction patterns of the produced carbons. XRD is usually used in hard carbons to obtain information about the interlayer spacing between graphene sheets, which is determined based on the position of (002) and (100) diffraction peaks at 20–26° and 43–44°, respectively. This case, all X-ray diffraction diagrams present this typical wide diffraction maxima (002) and (100), which is attributed to the disordered structure of the hard carbons [43]. This indicates that all samples have a turbostratic structure, which implies that the building blocks of the carbon samples are composed of graphite-like microcrystallites that are randomly oriented and distributed throughout the samples.

Apart from these two main peaks, other diffraction peaks are also observed in the diffractograms, probably due to the presence of impurities related to the use of real biowaste as starting material. In this sense, two groups of samples can be distinguished in Figure 2: those with a low quantity of impurities and those with a higher amount of them. In the first group, there are CORN, APP and GSEED (Figure 2a,b,d, respectively). In these cases, the use of HTC and AH + HTC pretreatments reduces the impurities compared to the materials obtained by direct pyrolysis. On the other hand, in OLI and GSkin carbons (Figure 2c,e, respectively) it seems that the impurities decrease with the combined pretreatment AH + HTC, but, on the contrary, they increased with only an HTC pretreatment. These impurities have been related to inorganic compounds, that have been identified as: CaO, MgO, SiO₂, Ca(OH)₂, NaNO₃, K₂O and Ca₂SiO₄ (Powder Diffraction File 01-075-0264, 01-077-2364, 01-083-2468, 01-076-0570, 01-072-0026, 00-027-0431 and 00-029-0369, respectively) [44]. Such substances are consistent with the typical chemical species present in biomass, for example, those coming from the presence of Ca²⁺, Mg²⁺ or Na⁺ salts in the plant [45]. Regarding the

lower carbon content registered in elemental analysis of PHOS-HTC_1200 samples for corn cob and olive mill solid waste, X-ray diffractograms indicate that the amount of inorganic crystalline impurities is the lowest in each set of samples. Thus, the cause of the decreased weight carbon content in PHOS-HTC_1200 samples must be attributed to the significant presence of volatile C-based compounds.

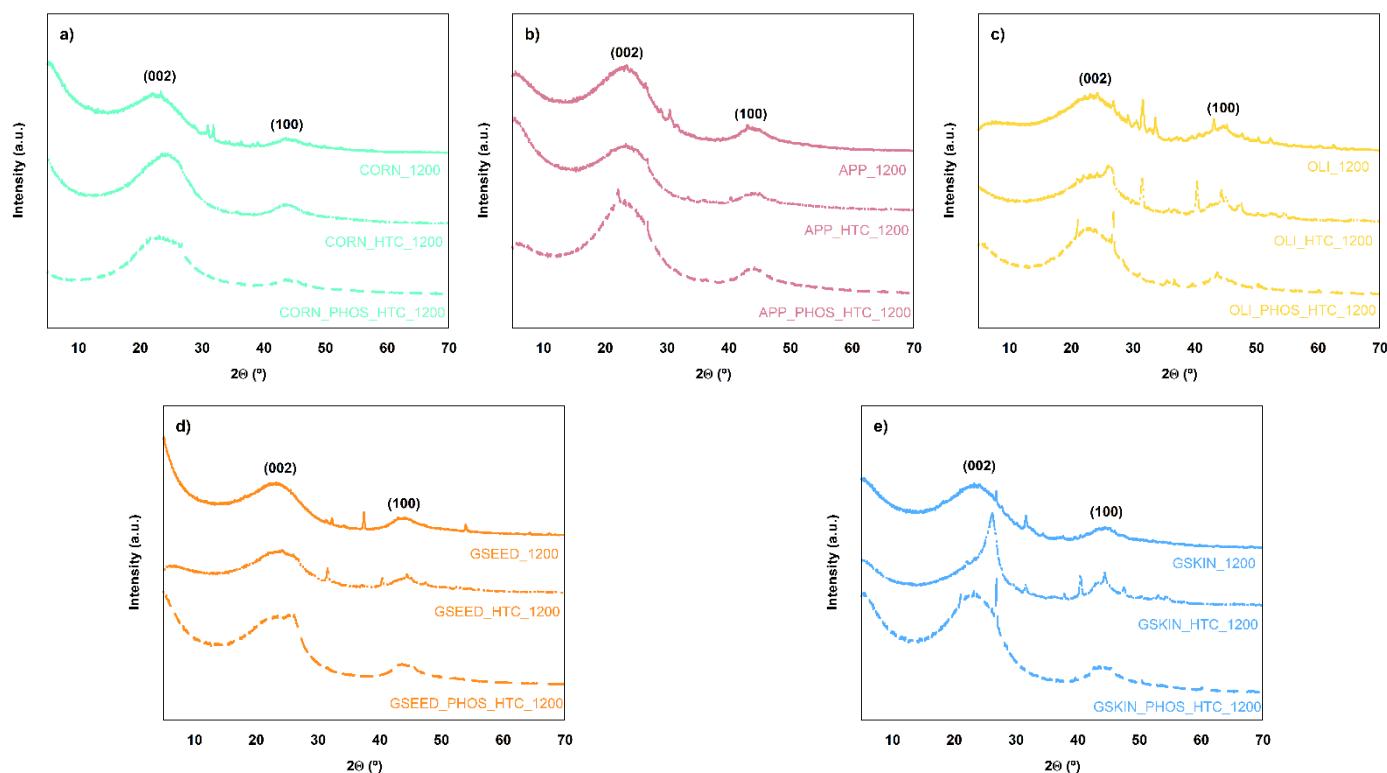


Figure 2. X-ray diffraction diagrams of corn cob (a), apple pomace (b), olive mil solid waste (c), defatted grape seed (d) and dried grape skin (e) samples.

Figure 3 depicts the Raman spectra of the carbon materials obtained from biowaste. As it can be observed, all samples show the typical D ($1300\text{--}1360\text{ cm}^{-1}$) and G ($1580\text{--}1600\text{ cm}^{-1}$) bands of disordered carbons. D band is associated with the vibrational mode of micro-crystalline graphite, which is only Raman-active in the presence of structural defects in graphene planes. On the other hand, G band is linked to stacked graphene sheets defects. This way, the fitting of the Raman spectra allows calculating the ratio between D and G band areas (A_D/A_G), which can be an indicator of the ordering degree of the carbon structure in the samples [43].

Table 3 shows the main parameters calculated from the information obtained from XRD and Raman characterization techniques. These are structural parameters show the interlayer spacing (d_{002} and d_{100}) and microcrystallite dimensions, stack height (L_c), stack width (L_a), the number of stacked graphene sheets (N) and the ratio between D and G Raman-band areas (A_D/A_G). The calculations were determined using Bragg and Scherrer formulas, which rely on the wavelength of the diffractometer (λ), the diffraction angle (2θ) and the full width at half maximum (FWHM), which is the width of the band at half. Therefore, if (002) diffraction angle increases the d_{002} decreases. The same occurs to FWHM; when this parameter increases, L_c and L_a decrease. [46–48].

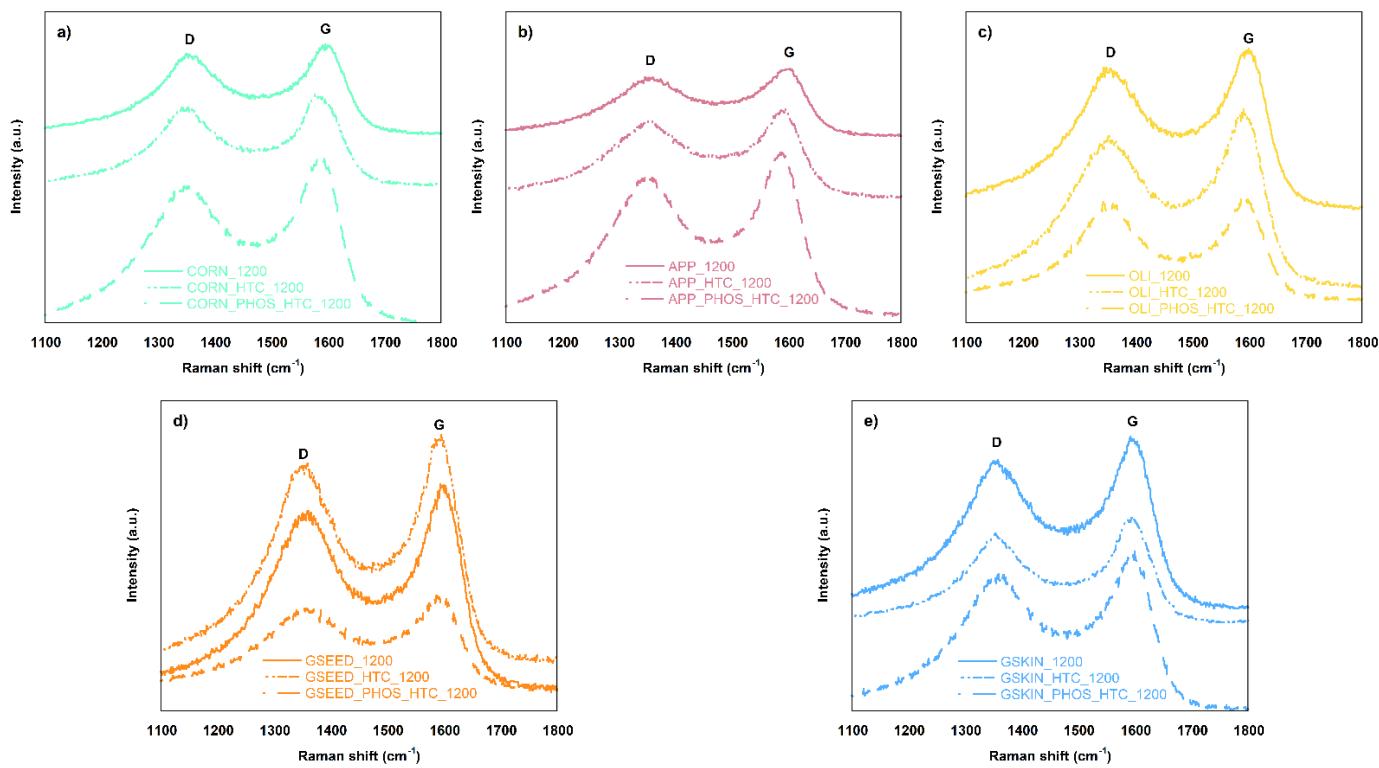


Figure 3. Raman spectra of corn cob (a), apple pomace (b), olive oil mill solid waste (c), grape defatted seed (d) and dried grape skin (e) samples.

Table 3. Physical parameters of the samples calculated from XRD and Raman spectra.

	Samples	d_{002} (Å)	L_a (Å)	L_c (Å)	N	$A_D/A_{G\ 01}$	$A_D/A_{G\ 02}$
CORN	1200	3.98	38.50	10.61	2.67	2.25	2.29
	HTC_1200	3.80	38.47	9.53	2.51	2.66	1.52
	PHOSHTC_1200	3.92	36.39	8.91	2.27	1.58	1.89
APP	1200	3.90	37.06	9.38	2.41	1.91	1.32
	HTC_1200	3.87	37.37	10.23	2.64	2.11	0.87
	PHOSHTC_1200	3.93	36.81	9.42	2.40	1.85	1.77
OLI	1200	3.81	40.52	8.23	2.16	1.70	0.61
	HTC_1200	3.75	35.84	8.63	2.30	1.90	1.71
	PHOSHTC_1200	3.89	45.88	9.41	2.42	1.91	—
GSEED	1200	3.92	39.89	10.01	2.56	1.19	1.16
	HTC_1200	3.85	39.04	9.97	2.59	1.80	—
	PHOSHTC_1200	3.85	41.80	10.03	2.61	2.71	2.00
GSKIN	1200	3.86	35.53	9.51	2.46	2.30	—
	HTC_1200	3.50	45.61	35.67	10.18	1.84	1.77
	PHOSHTC_1200	3.88	35.10	9.46	2.44	2.91	2.98

The interlayer spacing between the graphene sheets (d_{002}) is highly important, given that with the increase of this parameter, the space between the sheets also increases and it is easiest for the sodium-ion insertion in the material, probably leading to a better electrochemical performance. In general, it can be said that all the obtained carbons present d_{002} values equal or higher than 3.44 Å that, together with N values smaller than 500, are typical values for hard carbons [49]. Table 3 shows that HTC pretreatment does not increase the space between the sheets in any case and sometimes produces a significant reduction,

as in the case of CORN and especially GSKIN. The explanation for such a reduction could lie on the fact that sample GSKINHTC_1200 shows higher (002) diffraction angle (θ) than the other materials and, at the same time, much smaller FWHM (see Figure 2). This leads to decrease d_{002} to 3.50, compared to 3.80–3.90 values for the other materials, with the corresponding increase in N up to 10 stacked graphene sheets. This would suggest that the pretreatment of HTC combined with a pyrolysis in GSKIN induce a slightly higher ordering of the material compared to the other samples.

In contrast, pretreatment with AH + HTC increases d_{002} obtained with only HTC in all cases. In the case of APP, OLI and GSKIN samples these d_{002} are greater than those obtained with direct pyrolysis. On the contrary, it seems that the route to acquire carbon materials with higher interlayer spacing in the case of CORN and GSEED is direct pyrolysis, without the use of any kind of pretreatment, nor AH, neither HTC.

The ratios between D and G Raman-band areas (A_D/A_G) are also shown in Table 3. As different spectra were taken in several zones of the analyzed materials, two area ratio values are presented for each material ($A_D/A_G\text{ }_{01}$ and $A_D/A_G\text{ }_{02}$), in order to use them as indicators for the degree of homogeneity of the produced carbons. As it can be seen in Table 3, only five out of twelve samples present some degree of homogeneity (CORN_1200, APP_PHOSHTC_1200, GSEED_1200, GSKINHTC_1200 and GSKINPHOSHTC_1200), while the other carbons show clear signs of heterogeneity. Therefore, when analyzing the influence of the different pretreatments on the resulting carbons, the uncertainty caused by heterogeneity must be taken into account. Furthermore, in order to lead the discussion, it should be mentioned that the typical A_D/A_G values for hard carbons are around 2.65–2.70 [49].

It should be noted that only 3 samples show uniformity in the influence of pretreatments among the different areas analyzed ($A_D/A_G\text{ }_{01}$ and $A_D/A_G\text{ }_{02}$); these are OLI, GSEED and GSKIN. In the case of OLI and GSEED, it appears that the two pretreatments (HTC and AH + HTC) improve the ratio A_D/A_G in the different areas. As a consequence, the GSEEDPHOSHTC_1200 carbon clearly enhances the ratio of those measured after only pyrolysis or after HTC + pyrolysis, reaching the typical values of hard carbons (2.71). On the contrary, HTC carbons present smaller ratio compared to the only pyrolyzed carbons for GSKIN, but this is clearly increased after AH + HTC (2.91). Therefore, it can be concluded that the number of defects in the materials increases notably when using AH combined with HTC pretreatment with GSEED and GSKIN samples. On the contrary, no rigorous conclusions can be drawn from this parameter for the CORN and APP samples, since, depending on the area analyzed, opposing effects can be observed.

SEM micrographs of the raw precursors and the prepared carbons are shown in Figure 4. In the case of CORN, OLI and GSEED, it can be observed that after a pyrolysis step (Figure 4b,j,n, respectively) the integrity of the biostructure they come from (Figure 4a,i,m, respectively) is slightly broken in some points, which allows seeing the internal arrangement of the material. In CORN_1200 it can be appreciated the cell form morphology inside the material with pores of about 8–10 μm size. OLI_1200 and GSEED_1200 show a degraded structure consisting of 100–150 μm wrinkled aggregates (OLI_1200), and >200 μm lumps with pores smaller than 10 microns in them (GSEED_1200). Nevertheless, HTC pretreatment on these three precursors shows deteriorated structures (Figure 4c,k,o, respectively) and, in the case of AH with subsequent HTC, highly deteriorated structure without any chance of distinguishing the initial morphology of each precursor (Figure 4d,l,p, respectively).

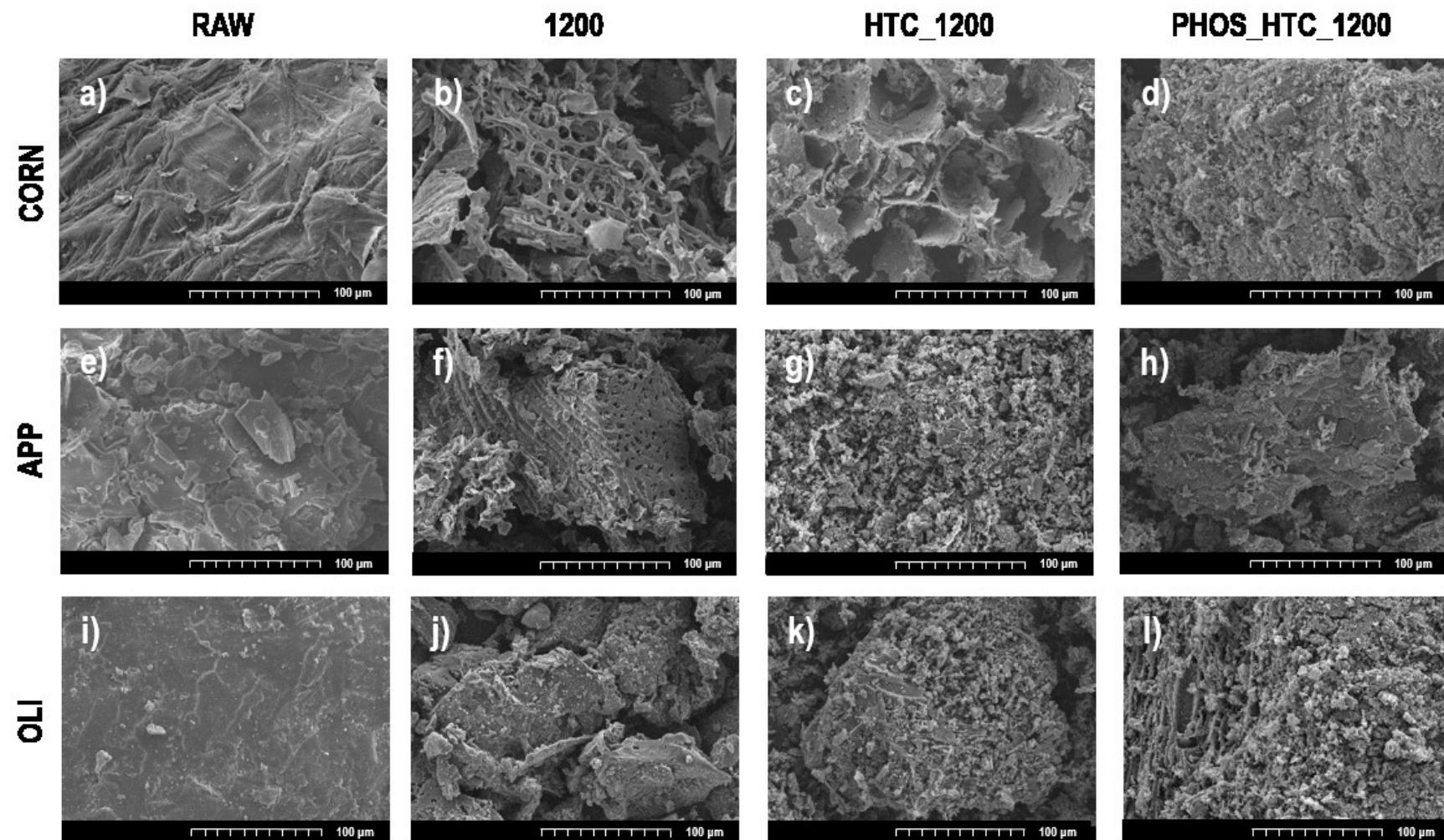


Figure 4. Cont.

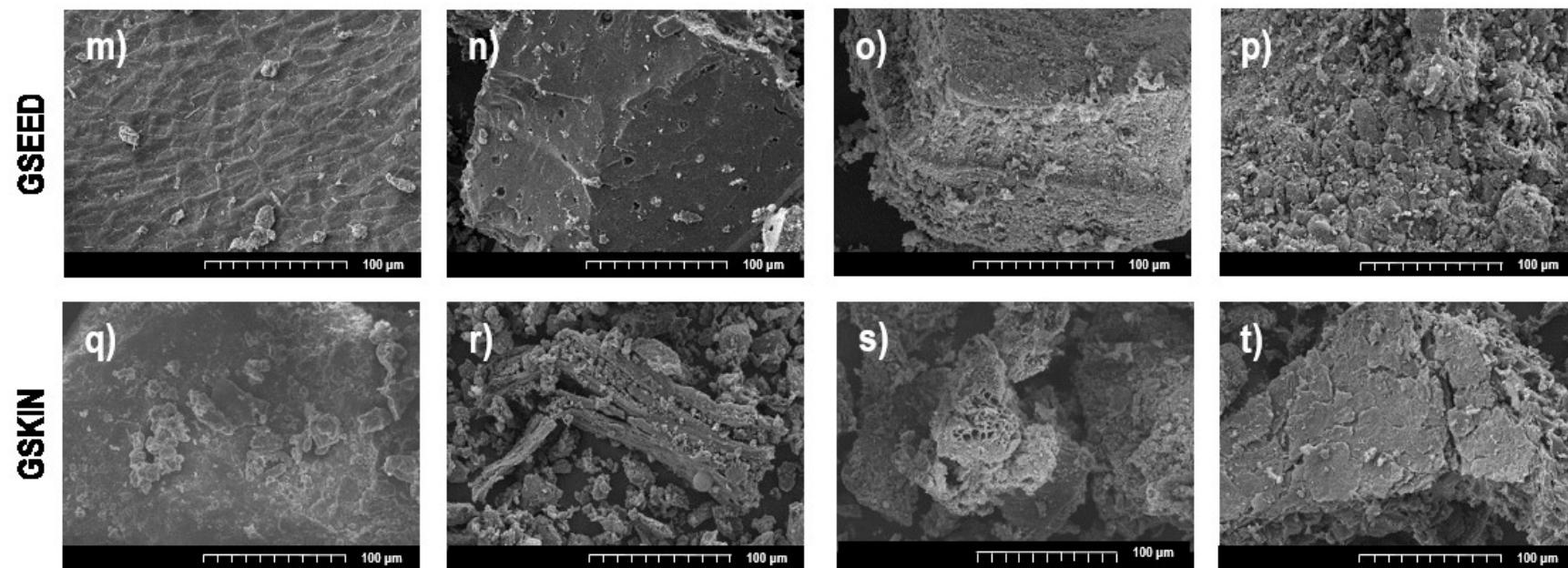


Figure 4. SEM micrographs of the untreated, pyrolyzed (1200), pretreated with HTC (HTC_1200) and pretreated with acid and later with HTC (PHOS-HTC_1200) samples of corn cob (a–d), apple pomace (e–h), olive mil solid waste (i–l), defatted dried grape skin (m–p) and dried grape skin (q–t), respectively.

On the other hand, APP_1200 also exhibits broken aggregates larger than 100 μm with small pores in them after direct pyrolysis (Figure 4f). However, the only use of HTC leads to complete disaggregation of the material, which is reduced to particles smaller than 10 μm (Figure 4g). Then, the combination of AH and HTC on APP precursor (APP_PHOSHTC_1200) cause higher level of biostructure destruction (Figure 4h). Finally, GSKIN presents the highest degree of biostructure damage after pyrolysis (Figure 4r) compared to the materials subjected to HTC pretreatment (Figure 4s) and to combined AH and HTC pretreatment (Figure 4t). Biowaste hard carbons can preserve their natural porous structures derived from the plant vessels for water transfer, which is beneficial for Na^+ adsorption and insertion. However, the use of different treatments, such as pyrolysis, hydrothermal carbonization and acid hydrolysis can lead to modify their initial microstructure, to form micropores and mesopores. In the case of hydrothermal process and also in acid digestion, crystalline inorganic impurities could be significantly removed, which can be linked to an increase in the pore formation. In summary, the morphological analysis of the prepared carbon samples indicates that the use of different pretreatments led to diverse morphologies and textures depending on the starting material.

2.3. Electrochemical Tests

Figure 5 shows the voltage profile of the first cycle of samples derived from corncob as representative of the whole set of samples (Figure 5a) and the first cycle and its subsequent cycles from CORN_1200 sample (Figure 5b).

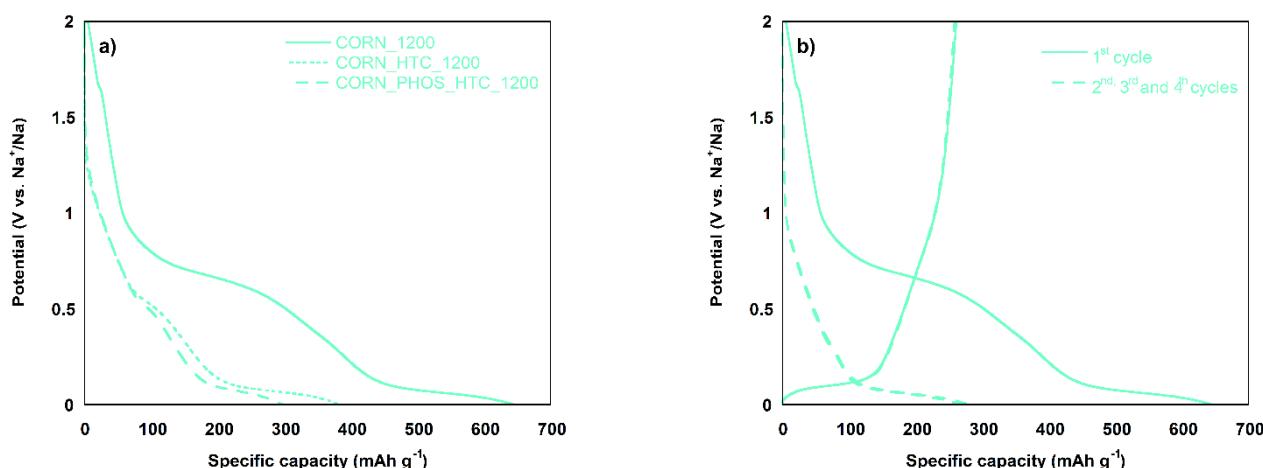


Figure 5. Electrochemical performance of the first discharge cycle of CORN_1200, CORN-HTC_1200 and CORN_PHOSHTC_1200 (a) and the first charge/discharge and subsequent cycles of CORN_1200 (b) at C/15.

Analysis of the voltage profile of CORN materials in the first discharge (Figure 5a) shows that all of them present the typical electrochemical signature of hard carbon vs. sodium metal: a slope region below 1 V vs. Na/Na^+ followed by a plateau close to 0 V vs. Na/Na^+ [50]. Figure 5b shows that the initial discharge capacity, that is, the specific capacity of each material in the first cycle, presents a larger curve than that of the subsequent cycles. This means that the specific capacity (C_{sp}) in the first cycle is higher than in the following ones, due to the Na ion (Na^+) insertion-adsorption and the formation of the Solid Electrolyte Interphase (SEI), that occurs always during the first discharge [51]. The formation of a stable SEI mitigates further parasitic reactions between the anode material and the electrolyte and allows an efficient reversible electrochemical cycling. However, the natural precipitation of a compact, stable and suitable passivation film depends on the complex interplay of many concurrent factors, such as the composition of the electrolyte or the surface chemistry of the hard carbon and its morphology [52].

This phenomenon happens for all the anodic materials as it can be seen in Table 4, where initial discharge capacities, together with the main parameters corresponding to the electrochemical performance of every material are shown.

Table 4. Electrochemical performances in each sample at C/15 (24.8 mA·g⁻¹).

Samples		Initial Discharge Capacity (mAh·g ⁻¹)	Specific Capacity (mAh·g ⁻¹)	Initial Coulombic Efficiency (%)
CORN	1200	644.48	265.17	43.39
	HTC_1200	378.96	268.54	73.49
	PHOS-HTC_1200	297.09	169.66	61.69
APP	1200	644.64	247.19	40.73
	HTC_1200	457.76	222.47	50.41
OLI	PHOS-HTC_1200	381.57	196.63	55.11
	1200	444.09	149.44	38.75
	HTC_1200	468.09	174.16	50.00
GSEED	PHOS-HTC_1200	320.86	182.90	64.81
	1200	289.59	158.42	66.47
	HTC_1200	450.76	244.94	60.25
GSKIN	PHOS-HTC_1200	437.63	162.92	43.65
	1200	446.47	215.73	57.03
	HTC_1200	385.61	149.44	43.96
	PHOS-HTC_1200	437.84	243.82	61.89

The electrochemical performances reported in Table 4 present a wide range of values, which could be related to the precursor used and the effects of the pretreatments on it. However, the majority of values in Table 4 are higher than those reported in other works carried out with biowaste [53]. CORN and APP show quite high initial discharge capacities after direct pyrolysis (644.48 and 644.64 mAh·g⁻¹, respectively). Then, these capacities worsen as samples are pretreated with HTC and AH + HTC. In the case of OLI and GSKIN, apart from showing lower initial discharge capacities (444.09 and 446.47 mAh·g⁻¹, respectively), they behave quite different in relation to the two pretreatments. On the one hand, the initial capacity of OLI is slightly enhanced with HTC pretreatment, but significantly decreases when AH + HTC pretreatment is used. On the other hand, HTC pretreatment diminishes the initial capacity of GSKIN, but it is almost recovered when AH + HTC is applied in the pretreatment step. At last, GSEED is the only sample that clearly improves its capacity thanks to the use of the pretreatment stages, showing a slightly better performance with exclusive HTC pretreatment.

Moreover, Table 4 and Figure 6 present slope-plateau contribution to specific capacity for the first cycle (a) and Initial Coulombic Efficiency (ICE) (b) percentages for each sample. From Figure 6a, it can be observed that most of the samples have higher plateau proportion with direct pyrolysis (OLI, GSEED and GSKIN) or with HTC pretreatment (CORN and APP), but none of them with combined AH and HTC.

Analysis of the ICE (Figure 6b) indicates that the use of HTC pretreatment and the combined pretreatment with AH improves the ICE values compared to the pyrolytic ones, reaching at least 50% of ICE for most of the precursors (CORN, APP, OLI and GSKIN). However, GSEED and GSKIN do not show this trend and, in these cases, ICE values higher than 55% are only reached by using direct pyrolysis and HTC pretreatment (GSEED) and direct pyrolysis and the AH combined with HTC pretreatment (GSKIN). It must be remarked that, except GSEED, the use of HTC pretreatment with or without previous acid digestion causes an increase in the ICE, which is a parameter that must be maintained as high as possible in order to obtain carbon materials to be used as anodes in commercial Na-ion batteries. Thus, CORN-HTC_1200 presenting the highest ICE value among the tested samples (73.49%), would be the best behaving material in terms of Coulombic efficiency.

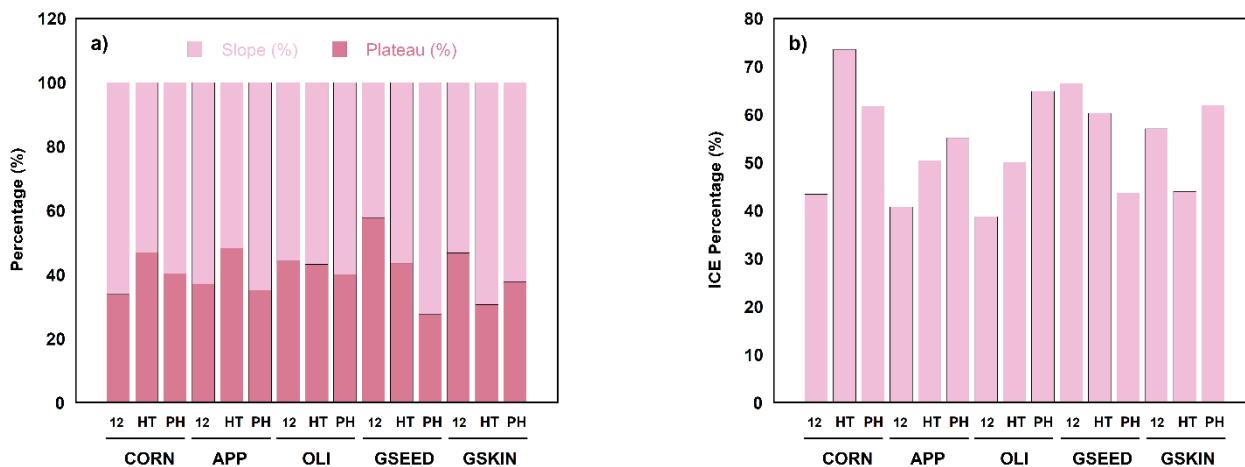


Figure 6. Slope-plateau proportion (a) and Initial Coulombic Efficiency (ICE) (b) in each material.

Figure 7 displays the discharge rate capability of each one of the carbonaceous materials prepared in this work, that is, the specific capacity (C_{sp}) at different cycling speeds. Among the used cycling rates, C/15 and C/10 are considered moderate/low, whereas C is taken as high rate. A final set of 5 cycles at C/15 is carried out on every material in order to observe if the electroactive material recovers its initial C_{sp} or has been degraded during continuous cycling at various rates. In all cases, it can be appreciated that the use of higher cycling rates induces a decrease in C_{sp} , which is due to kinetic limitations of electrochemical reactions. On the other hand, C_{sp} values in the range of the previous ones registered in the initial cycles at C/15 were observed, so it can be considered that there was no significant material degradation with cycling and the observed specific capacity values for each rate can be attributed only to the material performance.

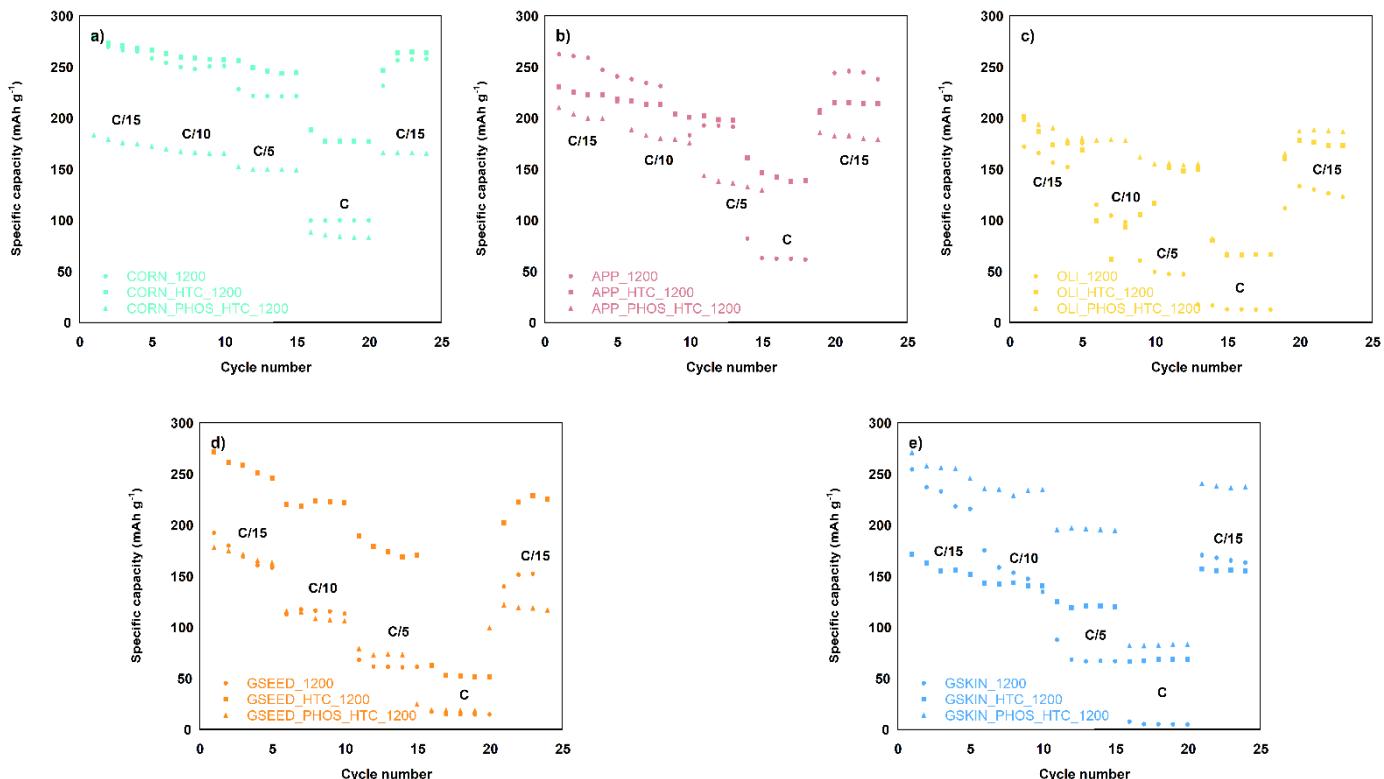


Figure 7. Discharge rate capability of corncob (a), apple pomace (b), olive mill solid waste (c), defatted grape seed (d) and dried grape skin (e) samples.

In CORN samples (Figure 7a), the use of HTC pretreatment improves slightly the C_{sp} , especially at high rates (56% at C-rate), whereas by using a previous AH no improvement in the electrochemical performance is observed. In the case of APP materials (Figure 7b), direct pyrolysis provides the best electrochemical results compared to the other synthesis routes, except at C-rate (high rate), where it can be observed a substantial improvement in the C_{sp} of the HTC pretreated sample (44% at C-rate). Figure 7c shows the rate performance of OLI derived carbons, where it can be seen that the carbon sample with the best electrochemical results is the one pretreated with AH and HTC. Taking into account that OLI is a biowaste with a significant quantity of ash (8.8 wt.%), perhaps the dissolution of part of this inorganic content due to pretreatment with phosphoric acid has improved its electrochemical performance. In addition, HTC has been also previously reported as an effective strategy to remove inorganic impurities [54]. Despite the improvement observed with the pretreatments, this sample is the one with the worst electrochemical performance.

Finally, contrary to expected, GSEED and GSkin (Figure 7d,e) do not show similar response to the different pretreatments. In the case of GSEED (Figure 7d), the use of HTC pretreatment enhances electrochemical performance at all rates compared to the other GSEED samples, this enhancement is of a 67% at C/15, 52% at C/10, 36% at C/5 and 28% at C-rate. In this sample the use of AH seems not to be necessary. In fact, the subsequent use of HTC after acid treatment does not produce a carbon with similar electrochemical performance to GSEED-HTC_1200. On the other hand, carbons obtained from GSkin precursor (Figure 7e) show better results in the case of AH and HTC pretreated material.

The best specific capacities were obtained by using corn cob with a combined treatment of HTC + pyrolysis, probably due to the higher content of glucans (cellulose and starch) and hemicellulose (xylans and arabinans) in the precursor. CORN-HTC_1200 presents the best specific capacity value at high rates, that are considered closer to real charge/discharge conditions.

The comparison of cycling behavior of the synthesized materials with values reported in literature shows that the hard carbons prepared in this work are below the registered capacities obtained from reagent grade chemicals such as magnesium gluconate and glucose, which are close to $500 \text{ mAh}\cdot\text{g}^{-1}$ at C/15 [55]. However, the capacities reported for hard carbons coming from biowaste ($\approx 300 \text{ mAh}\cdot\text{g}^{-1}$ at C/15) are in the range of those obtained in this work as it can be seen in Table 5. Additional characterizations (such as SAXS, BET surface area measurement and post-mortem analysis) are needed in the future to offer better insights.

Table 5. Electrochemical performances in each sample at C/15 ($24.8 \text{ mA}\cdot\text{g}^{-1}$).

	Electrochemical Performance ($\text{mAh}\cdot\text{g}^{-1}$)	Initial Coulombic Efficiency (%)	References
<i>Waste tea bag</i>	282.40 (30.00 $\text{mA}\cdot\text{g}^{-1}$)	69.00	[30]
<i>Maple tree</i>	332.00 (30.00 $\text{mA}\cdot\text{g}^{-1}$)	88.30	[56]
<i>Argan shell</i>	286.00 (25.00 $\text{mA}\cdot\text{g}^{-1}$)	76.90	[57]
<i>Sugarcane bagasse</i>	290.00 (30.00 $\text{mA}\cdot\text{g}^{-1}$)	70.00	[58]
<i>Oatmeal</i>	272.40 (20.00 $\text{mA}\cdot\text{g}^{-1}$)	46.64	[59]
<i>Corn cob</i>	268.54 (24.80 $\text{mA}\cdot\text{g}^{-1}$)	73.49	This work
<i>Grape skin</i>	243.82 (24.80 $\text{mA}\cdot\text{g}^{-1}$)	61.89	This work
<i>Grape seed</i>	244.94 (24.80 $\text{mA}\cdot\text{g}^{-1}$)	60.25	This work
<i>Apple pomace</i>	247.19 (24.80 $\text{mA}\cdot\text{g}^{-1}$)	40.73	This work

3. Materials and Methods

3.1. Biowaste

Five types of biowaste related to agro-industrial activities with a high level of implementation in Spain were selected as bio-precursors for hard carbon production. (1) Corncob (CORN) coming from corn production and supplied by a local farmer;

(2) apple pomace (APP) coming from cider production and supplied by an association that elaborates fresh cider in Basque Country (*Bizkaiko Sagardoa*); (3) olive mill solid waste (OLI) coming from olive oil production and supplied by an olive oil press located in Navarra (*Hacienda Ortigosa*); (4) defatted grape seed (GSEED) and (5) dried grape skin (GSKIN) coming from wine and subsequent liquor production and supplied by a distillery dedicated to the exploitation of the wine production waste (*Agralco S.Coop.*).

The quantity of biowaste generated in olive oil production in Spain is a big concern, since this country is the first world producer of olive oil (around 1.3 Mt/year) and, taking into account the low olive oil/olive ratio (0.2/1), the production of olive mill solid waste represents approximately 1.5 Mt/year [60]. Spain is also the second world producer of wine, which accounts up to 4.6 Mt of wine annually and the corresponding 1.26 Mt of winery derived biowaste [61]. On the other hand, corn is the fourth most produced cereal in Spain (the one with the highest production in the spring season), reaching 3.6 Mt in the 2018 harvest [62]. It is estimated that waste generated in corn production is 40% of the harvest, within which, cobs represent another 40% (0.6 Mt/year) [63]. At last, although cider does not reach the production levels mentioned above, apple pomace has been selected as a possible emerging waste given the increasing cider production observed in recent years [64]. The majority of the above-mentioned biowaste is nowadays used as solid fuel or raw material for composting. However, in order to expand the possibilities of bioeconomy, other alternatives of material recovery must be found.

3.2. Characterization Techniques

The chemical properties of the biowaste samples were determined by proximate analysis, elemental analysis and analysis of constituents. The proximate analysis was carried out in the LECO TGA-701 thermobalance, following the ASTM E871-82, ASTM E872-82 and ASTM E1534 standards. The elemental analysis (C, H, N, S, O) was accomplished in the Eurovector 2000 elemental analyzer. The constituent analysis, including extractive substances, structural carbohydrates (glucans, xylans and arabinans) and lignin, was performed according to UNE-EN-ISO 18134-2:2015 standard, and “Determinations of extractives in Biomass” and “Determinations of structural Carbohydrates and lignin in Biomass” NREL protocols.

The hard carbons produced were characterized by X-ray diffraction and Raman spectroscopy. X-ray diffraction was carried out with the PANalytical Xpert PRO diffractometer with Cu-K α radiation source ($\lambda = 1.5418 \text{ \AA}$) and Raman spectroscopy by using the Renishaw inVia spectrometer (514 nm Ar $^+$ laser, 4 scans by measurement between 150 and 3500 cm $^{-1}$). At last, the morphology of both, the raw materials (biowaste samples) and the resulting hard carbons, was observed using the scanning electron microscope (SEM) Hitachi S-4800.

3.3. Hard Carbon Production

Each sample of biowaste was powdered by milling (Retsch, Mixer Mill 301), decreasing the particle size to 5 μm . GSEED and APP were previously freeze-dried (Heto PowerDry LL3000) to prevent from any kind of biologic degradation. Three types of hard carbons were produced with each biowaste sample: direct pyrolysis hard carbons, HTC + pyrolysis hard carbons and AH + HTC + pyrolysis hard carbons. The pyrolysis was carried out in a tubular furnace (TZF 1200 °C Carbolite), following a thermal program consisting in heating up 1 g of each sample in a N₂ atmosphere from room temperature to 1200 °C at a heating rate of 5 °C/min and maintaining at such temperature for 2 h. The hard carbons produced in this way were named as NAME_1200. When biowaste samples were pretreated by HTC, 5 g of sample were introduced in a 300 mL autoclave (Autoclave Engineers) with 250 mL of water and both were heated up to 250 °C and maintained for 24 h under autogenous pressure (≈ 40 bar). The obtained hydrochars were pyrolyzed under the conditions mentioned above and the resulting carbon samples were named as NAMEHTC_1200. At last, when AH was also used to pretreat the biowaste samples, 25 gr of sample were dispersed in concentrated H₃PO₄ (Honeywell, 85%) and maintained at room temperature for 24 h. After AH and

subsequent washing and filtering until neutral pH, the hydrolyzed biowaste samples were hydrothermally carbonized and later pyrolyzed under the above-quoted conditions. The obtained materials were entitled as NAME_PHOSHTC_1200.

3.4. Electrochemical Tests

The working electrodes consisted of 80 wt.% active material (corn cob, apple pomace, olive mill solid waste, defatted grape seed and dried grape skin derived carbons) and 10 wt.% conducting carbon black (Ketjen black, AkzoNobel) and the rest is composed by polyvinylidene fluoride (PVDF, Solvay) solved homogeneously in N-methyl-2-pyrrolidone (NMP). Al foil was used as current collector. The electrodes were punched out with diameters of 7/16 inches. The CR2032-type coin cells were assembled in an Ar-filled glove Box (Labstar, MBraun). The electrolyte was a solution of 1 mol/L NaPF₆ in ethylene carbonate/dimethyl carbonate (EC/DMC, 1:1 in volume). Metallic sodium was used as the counter electrode and glass microfiber filters (Whatman GF/F) were used as the separator. All electrochemical measurements were carried out on a multichannel potentiostat/galvanostat (BioLogic VPM3) at room temperature. The charge/discharge experiments were performed between 2.0 and 0.002 V at different current rate in which C-rate is defined as discharge to the graphite full capacity ($372 \text{ mAh}\cdot\text{g}^{-1}$). The reproducibility of performance of each hard carbon material has been carried out 3 and 4 times. All the reactants and equipments have been purchased in Spain.

4. Conclusions

The main conclusion of this work is that, depending on the nature of the biowaste, different synthesis routes must be followed in the preparation of pyrolytic hard carbons. However, in general, hydrothermal carbonization improves capacity at high cycling rates, which are the closest conditions to real application. The electrochemical performance of the materials tested in this work is similar to other hard carbons obtained from biowaste. The best capacities were obtained by using corn cob with a combined treatment of HTC + pyrolysis, probably due to the presence of a higher amount of glucans (cellulose and starch) and hemicellulose (xylans and arabinans) in the precursor. Despite the similar specific capacities of CORN_1200 and CORNHTC_1200 at low rates, the second one presents the best electrochemical performance in terms of both initial Coulombic efficiency and specific capacity at high rates, closer to real charge/discharge conditions.

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