

## Quick Guide for the Adept™ Rapid PCR-Plus Protocol

# Element Adept Library Compatibility Workflow

## Introduction

This quick guide provides concise instructions for the Adept PCR-Plus Protocol, which uses the Adept Rapid PCR-Plus Kit. For comprehensive information and detailed instructions, see the *Element Adept Library Compatibility Workflow User Guide for the Rapid PCR-Plus Protocol (MA-00040)*.

## Amplify Library

1. Prepare 0.05–0.5 pmol linear library in 0.1 mM EDTA and 10 mM Tris, pH 8.0.
2. Combine the following reagents, allowing 10–15% overage.

Reagent	Volume per Reaction (μl)
Adept PCR+ Primer Mix	5
Adept PCR+ Master Mix	25

3. Add 20 μl library to a new PCR plate.
4. Add 30 μl master mix.
5. Run the following program:

Temperature	Time	Number of Cycles
Volume set to 50 μl		
Lid set to 105°C		
98°C	1.5 minutes	1
98°C	30 seconds	5
60°C	30 seconds	
72°C	30 seconds	
72°C	1 minute	1
4°C	Hold	1

## Clean Up Library

1. Add 50 μl beads (1x).
2. Mix beads and library:
  - » For a plate, shake at 1500–1800 rpm for 2 minutes.
  - » For tubes, pipette 10 times.

3. Incubate at room temperature for 5 minutes.
4. Place on the magnet and wait until supernatant clears.  
**Keep on the magnet.**
5. Remove the entire volume of supernatant.
6. Wash the content of each well or tube:
  - a. Add 200 μl 80% ethanol and incubate 30–60 seconds.
  - b. Remove and discard ethanol.
  - c. Repeat steps **a–b** one time.
  - d. Remove residual ethanol.
7. Air-dry for 3–5 minutes.
8. Add 32 μl low TE buffer (10 mM Tris, pH 8.0 and 0.1 mM EDTA).
9. Resuspend the beads:
  - » For a plate, shake at 1500–1800 rpm for 2 minutes.
  - » For tubes, pipette 10 times.
10. Incubate at room temperature for 2 minutes.
11. Place on the magnet and wait until supernatant clears.
12. Transfer 30 μl **supernatant** to a DNA LoBind tube.
13. Proceed to [Quantify Library](#) and sequencing or store at -25°C to -15°C for ≤ 15 days.

## Quantify Library

1. Quantify 2 μl library using Qubit.
2. Size 2 μl library using TapeStation.
3. Calculate the molar concentration:

$$nM = (ng/\mu l) / (660 * average\ library\ size) * 1,000,000$$