**Nitrogen demand, availability, and acquisition strategy control plant responses to elevated CO2**

**Running head:** Nitrogen demand and availability control plant responses to elevated CO2

Evan A. Perkowski1,\*, Ezinwanne Ezekannagha1, Nicholas G. Smith1

1Department of Biological Sciences, Texas Tech University, Lubbock, TX

**\***Corresponding author:

2901 Main St., Lubbock, TX, 79409

Email: [evan.a.perkowski@ttu.edu](mailto:evan.a.perkowski@ttu.edu)

**ORCIDs**

Evan A. Perkowski (0000-0002-9523-8892)

Ezinwanne Ezekannagha (0000-0001-7469-949X)

Nicholas G. Smith (0000-0001-7048-4387)

Total word count: 8034 (5350 words excluding Methods)

* Introduction: 1512
* Methods: 2684
* Results: 1088
* Discussion: 2750

Tables: 3

Figures: 4

Supporting Information: 9 tables, 7 figures

**Highlight**

Leaf nitrogen demand drove photosynthetic responses to elevated CO2 independent of nitrogen fertilization. Nitrogen fertilization enhanced whole-plant responses to elevated CO2. Symbiotic nitrogen fixation did not modify plant responses to elevated CO2.

**Abstract**

Plants respond to increasing atmospheric CO2 concentrations by reducing leaf nitrogen content and photosynthetic capacity – patterns that correspond with increased net photosynthesis and growth. Despite the longstanding notion that nitrogen availability regulates these responses, eco-evolutionary optimality theory posits that leaf-level responses to elevated CO2 are driven by leaf nitrogen demand for building and maintaining photosynthetic enzymes and are independent of nitrogen availability. In this study, we examined leaf and whole-plant responses of *Glycine max* L. (Merr) subjected to full-factorial combinations of two CO2, two inoculation, and nine nitrogen fertilization treatments. Nitrogen fertilization and inoculation did not alter leaf photosynthetic responses to elevated CO2. Instead, elevated CO2 decreased the maximum rate of Rubisco carboxylation more strongly than it decreased the maximum rate of electron transport for RuBP regeneration, increasing net photosynthesis by allowing rate-limiting steps to approach optimal coordination. Increasing fertilization enhanced positive whole-plant responses to elevated CO2 due to increased belowground carbon allocation and nitrogen uptake. Inoculation with nitrogen-fixing bacteria did not influence plant responses to elevated CO2. These results reconcile the role of nitrogen availability on plant responses to elevated CO2, showing that leaf photosynthetic responses are regulated by leaf nitrogen demand while whole-plant responses are constrained by nitrogen availability.

**Keywords**

acclimation, biomass, eco-evolutionary optimality, growth chamber, least-cost theory, optimal coordination, photosynthesis, plant functional ecology, resource optimization

**Introduction**

Complex carbon and nitrogen cycles regulate terrestrial ecosystems. Terrestrial biosphere models that incorporate coupled carbon and nitrogen cycles must accurately represent the processes and interactions governing these cycles across different environmental scenarios to simulate carbon and nitrogen fluxes reliably (Hungate *et al.*, 2003; Prentice *et al.*, 2015; Davies-Barnard *et al.*, 2020; Kou-Giesbrecht *et al.*, 2023). However, uncertainties remain regarding how nitrogen availability and acquisition strategy influences leaf- and whole-plant responses to increasing atmospheric CO2 concentrations, leading to divergent predictions of future carbon and nitrogen pools and fluxes across models (Arora *et al.*, 2020; Davies-Barnard *et al.*, 2020, 2022; Meyerholt *et al.*, 2020; Stocker *et al.*, 2025).

Research spanning several decades has documented consistent trends in leaf and whole-plant responses to elevated CO2. At the leaf level, C3 plants commonly exhibit increased net photosynthesis rates that correspond with reduced leaf nitrogen content, stomatal conductance, and photosynthetic capacity when grown under elevated CO2 compared to ambient conditions (Curtis, 1996; Drake *et al.*, 1997; Nakano *et al.*, 1997; Medlyn *et al.*, 1999; Ainsworth *et al.*, 2002; Ainsworth and Long, 2005; Bernacchi *et al.*, 2005; Ainsworth and Rogers, 2007; Crous *et al.*, 2010; Lee *et al.*, 2011; Pastore *et al.*, 2019; Poorter *et al.*, 2022; Cui *et al.*, 2023; Stocker *et al.*, 2025). At the whole-plant level, CO2 enrichment increases total leaf area, promoting greater primary productivity and biomass accumulation (Coleman *et al.*, 1993; Makino *et al.*, 1997; Ainsworth *et al.*, 2002; Ainsworth and Rogers, 2007; Finzi *et al.*, 2007; Poorter *et al.*, 2022). Some studies suggest that elevated CO2 increases belowground carbon allocation and root:shoot ratios (Iversen *et al.*, 2008; Iversen, 2010; Nie *et al.*, 2013; Stocker *et al.*, 2025), although these responses are not consistently observed (Luo *et al.*, 1994; Poorter *et al.*, 2022) and are highly variable across experiments (Stocker *et al.*, 2025).

Two hypotheses – the nitrogen limitation hypothesis and the eco-evolutionary hypothesis – offer contrasting views on how nitrogen availability shapes plant responses to elevated CO2. The nitrogen limitation hypothesis posits that nitrogen availability constrains plant responses to elevated CO2, as nitrogen availability often limits net primary productivity and influences the magnitude of the terrestrial carbon sink (Vitousek and Howarth, 1991; LeBauer and Treseder, 2008; Sigurdsson *et al.*, 2013; Wieder *et al.*, 2015). Elevated CO2 increases whole-plant nitrogen demand for building new tissues, which may lead to greater nitrogen limitation of net primary productivity without additional ecosystem nitrogen inputs (Luo *et al.*, 2004). Thus, increased nitrogen availability should amplify the positive effects of elevated CO2 on net primary productivity and biomass accumulation, provided that nitrogen availability exceeds whole-plant demand. Free-air CO2 enrichment studies offer mixed support for this hypothesis, with some studies supporting its predictions (Reich *et al.*, 2006; Norby *et al.*, 2010) and others not (Finzi *et al.*, 2006; Moore *et al.*, 2006; Liang *et al.*, 2016). The hypothesis also implies that reductions in leaf nitrogen content and photosynthetic capacity under elevated CO2 are linked to ecosystem nitrogen limitation, as positive correlations between soil nitrogen availability, leaf nitrogen content, and photosynthetic capacity are commonly observed (Field and Mooney, 1986; Evans, 1989). However, evidence shows that reductions in leaf nitrogen content and photosynthetic capacity under elevated CO2 are often decoupled from changes in nitrogen availability (Crous *et al.*, 2010; Lee *et al.*, 2011; Pastore *et al.*, 2019), indicating that other factors, such as demand for building and maintaining photosynthetic tissues, might play a more important role in determining leaf-level responses.

Conversely, the eco-evolutionary optimality hypothesis asserts that leaf-level demand to build and maintain photosynthetic enzymes drives leaf-level photosynthetic responses to elevated CO2 and that nitrogen availability does not modify these responses (Harrison *et al.*, 2021). The hypothesis combines photosynthetic least-cost (Wright *et al.*, 2003; Prentice *et al.*, 2014) and optimal coordination (Chen *et al.*, 1993; Maire *et al.*, 2012) theories, suggesting that elevated CO2 downregulates the maximum rate of Ribulose-1,5-bisphosphate (RuBP) carboxylase/ oxygenase (Rubisco) carboxylation (*V*cmax) more strongly than the maximum rate of electron transport for RuBP regeneration (*J*max). The downregulation in *V*cmax is attributed to increased CO2 availability under elevated CO2, which enhances Rubisco affinity for carboxylation relative to oxygenation and reduces nitrogen demand for building and maintaining additional Rubisco enzymes (Bazzaz, 1990; Dong *et al.*, 2022). The eco-evolutionary optimality hypothesis predicts that plants optimize leaf nitrogen allocation to photosynthetic capacity to use available light efficiently while avoiding over-investment in Rubisco, which has high nitrogen and energetic costs to build and maintain (Evans, 1989; Sage, 1994; Evans and Clarke, 2019). This strategy enhances photosynthetic nitrogen-use efficiency and allows increased net photosynthesis rates to be achieved by increasing the co-limitation of net photosynthesis rates by Rubisco carboxylation and electron transport for RuBP regeneration (Chen *et al.*, 1993; Maire *et al.*, 2012; Wang *et al.*, 2017; Smith *et al.*, 2019). Empirical evidence supports this hypothesis (Crous *et al.*, 2010; Lee *et al.*, 2011; Smith and Keenan, 2020; Harrison *et al.*, 2021; Dong *et al.*, 2022; Cui *et al.*, 2023), though few studies have connected these patterns with concurrently measured whole-plant responses.

While the eco-evolutionary optimality hypothesis predicts that leaf-level photosynthetic responses are independent of nitrogen availability, it acknowledges that nitrogen availability likely regulates whole-plant responses to elevated CO2. The hypothesis suggests that the optimal whole-plant response to elevated CO2 involves allocating surplus nitrogen not needed to satisfy leaf-level demand to build and maintain photosynthetic enzymes toward constructing additional optimally coordinated leaves and other plant organs. Furthermore, the hypothesis implies that optimal resource allocation to photosynthetic capacity leads to nitrogen-savings at the leaf-level, which maximizes resource allocation to support whole-plant growth (Smith *et al.*, 2024). Thus, the extent to which plant responses to elevated CO2 align with the nitrogen limitation or eco-evolutionary optimality hypothesis may be a question of scale, with leaf-level responses influenced by leaf-level demand to build and maintain photosynthetic enzymes and whole-plant responses regulated by nitrogen availability.

Nitrogen acquisition strategy complicates the role of nitrogen availability on plant responses to elevated CO2. Plants use a variety of strategies to acquire nitrogen, including direct uptake from the soil or through symbiotic relationships with mycorrhizal fungi and nitrogen-fixing bacteria (Barber, 1962; Gutschick, 1981; Smith and Read, 2008). The carbon costs associated with nitrogen acquisition vary among species with different acquisition strategies and depend on environmental factors such as atmospheric CO2, temperature, light availability, and nutrient availability (Fisher *et al.*, 2010; Brzostek *et al.*, 2014; Terrer *et al.*, 2018; Allen *et al.*, 2020; Perkowski *et al.*, 2021, 2024; Lu *et al.*, 2022; Peng *et al.*, 2023). Carbon costs to acquire nitrogen can influence nitrogen uptake and, in turn, affect nitrogen allocation to different plant organs, investment in photosynthetic tissues, and biomass accumulation (Terrer *et al.*, 2018; Perkowski *et al.*, 2021, 2024; Waring *et al.*, 2023). Therefore, considering nitrogen acquisition strategy is important when examining plant responses to elevated CO2 across nitrogen availability gradients, especially because whole-plant responses to elevated CO2 are often positively correlated with nitrogen uptake (Feng *et al.*, 2015; Stocker *et al.*, 2025). However, few studies account for acquisition strategy when considering the role of nitrogen availability on plant responses to elevated CO2 (Terrer *et al.*, 2016, 2018; Smith and Keenan, 2020). Despite this, emerging evidence suggests that acquisition strategies with lower carbon costs for nitrogen acquisition may mitigate nitrogen limitation at the whole-plant level, though leaf-level responses remain less clear (Terrer *et al.*, 2018; Smith and Keenan, 2020).

Here, we examined whether plant responses to elevated CO2 align with the nitrogen limitation or eco-evolutionary optimality hypothesis and assessed how nitrogen acquisition strategy modifies these responses. Using a growth chamber experiment, we grew *Glycine max* L. (Merr.) seedlings under two CO2 concentrations (420, 1000 ppm CO2), two nitrogen acquisition strategies (with and without *Bradyrhizobium japonicum*), and nine soil nitrogen fertilization treatments (ranging from 0 to 630 ppm N) in a full-factorial design. Inoculation with *B. japonicum* simulated whether plants could acquire nitrogen through associations with symbiotic nitrogen-fixing bacteria. We used this experimental setup to test the following hypotheses:

1. Leaf photosynthetic responses to elevated CO2 will be independent of nitrogen fertilization and inoculation treatment. Instead, elevated CO2 will decrease *V*cmax more than *J*max, increasing the ratio of *J*max to *V*cmax. This response will increase net photosynthesis rates under growth CO2 conditions by allowing rate-limiting steps to approach optimal coordination while enhancing photosynthetic nitrogen-use efficiency.
2. Following the nitrogen limitation hypothesis, increasing nitrogen fertilization will enhance the positive effects of elevated CO2 on total leaf area and total biomass. This response will be due to increased belowground carbon allocation and nitrogen uptake and with increasing nitrogen fertilization that will be stronger under elevated CO2. Biomass responses to elevated CO2 will be driven by a greater increase in belowground biomass than aboveground biomass, as plants will invest in resource acquisition strategies to meet the increased whole-plant nitrogen demand for building new tissues.
3. Following the nitrogen limitation hypothesis, inoculation with nitrogen-fixing bacteria will enhance positive whole-plant responses to elevated CO2. These responses will be strongest under low nitrogen availability, where inoculated plants will invest in nitrogen uptake through symbiotic nitrogen fixation over more costly direct uptake pathways. However, these patterns will diminish with increasing nitrogen fertilization as plants acquire more nitrogen through increasingly less costly direct uptake pathways.

**Materials and methods**

*Seed treatments and experimental design*

*Glycine max* L. (Merr) seeds (Territorial Seed Co., Cottage Grove, OR, USA) were planted in 144 6-liter surface sterilized pots (NS-600, Nursery Supplies, Orange, CA, USA) containing a steam-sterilized 70:30 volume:volume mix of *Sphagnum* peat moss (Premier Horticulture, Quakertown, PA, USA) to sand (Pavestone, Atlanta, GA, USA). Before planting, all *G. max* seeds were surface sterilized in 2% sodium hypochlorite for 3 minutes, followed by three 3-minute washes with ultrapure water (MilliQ 7000; MilliporeSigma, Burlington, MA USA). Subsets of surface-sterilized seeds were inoculated with *Bradyrhizobium japonicum* (Verdesian N-Dure™ Soybean, Cary, NC, USA) in a slurry following manufacturer recommendations (3.12 g inoculant and 241 g ultrapure water per 1 kg seed).

Seventy-two pots were randomly planted using surface-sterilized seeds inoculated with *B.* *japonicum*, while the remaining 72 pots were planted using surface-sterilized uninoculated seeds. Thirty-six pots in each inoculation treatment were placed in one of two atmospheric CO2 treatments (420, 1000 μmol mol-1 CO2). CO2 treatments were decided based on current ambient CO2 concentrations and projections from the Intergovernmental Panel on Climate Change indicating that CO2 concentrations could surpass 1000 ppm by 2100 under the Shared Socioeconomic Pathway 5-8.5 (IPCC, 2021). Plants in each unique inoculation-by-CO2 treatment combination received one of nine nitrogen fertilization treatments equivalent to 0 (0 mM), 35 (2.5 mM), 70 (5 mM), 105 (7.5 mM), 140 (10 mM), 210 (15 mM), 280 (20 mM), 350 (25 mM), or 630 ppm (45 mM) N. This experimental setup resulted in 4 replicates per unique inoculation-by-CO2-by-nitrogen fertilization treatment combination. Nitrogen fertilization treatments were created using a modified Hoagland’s solution (Hoagland and Arnon, 1950) designed to keep concentrations of all other macronutrients and micronutrients equivalent across treatments (Table S1). Plants received the same nitrogen fertilization treatment twice per week in 150 mL doses as topical agents to the soil surface. Plants were well-watered between fertilization doses to ensure that physiology and growth was not limited by water availability.

*Growth chamber conditions*

Plants were randomly placed in one of six calibrated Percival LED-41L2 growth chambers (Percival Scientific Inc., Perry, IA, USA) over two experimental iterations due to chamber space limitation. The first iteration included all plants grown under elevated CO2, while the second included all plants grown under ambient CO2. Average (± SD) CO2 concentrations across chambers throughout the experiment were 439±5 μmol mol-1 CO2 for the ambient treatment and 989±4 μmol mol-1 CO2 for the elevated treatment. Each experimental iteration lasted seven weeks, which was sufficient for plants to grow through the majority of their vegetative growth phase without evidence of reproduction.

Daytime growth conditions were simulated using a 16-hour photoperiod, with incoming light radiation set to chamber maximum (mean±SD: 1230±12 μmol m-2 s-1 across chambers), air temperature set to 25°C, and relative humidity set to 50%. This daylength allowed plants to maximize vegetative growth across the seven-week experiment while minimizing the onset of reproduction. The remaining 8-hour period simulated nighttime growing conditions, with incoming light radiation set to 0 μmol m-2 s-1, chamber temperature set to 17°C, and relative humidity set to 50%. Transitions between daytime and nighttime growing conditions were simulated by ramping incoming light radiation in 45-minute increments and temperature in 90-minute increments over 3 hours (Table S2).

Plants grew under average (± SD) daytime light intensity of 1049±27 μmol m-2 s-1, including ramping periods. In the elevated CO2 iteration, plants grew under 24.0±0.2°C during the day, 16.4±0.8°C during the night, and 51.6±0.4% relative humidity. In the ambient CO2 iteration, plants grew under 23.9±0.2°C during the day, 16.0±1.4°C during the night, and 50.3±0.2% relative humidity. Any differences in climate conditions across the six chambers were accounted for by shuffling the same group of plants throughout the growth chambers. This process was done by iteratively moving the group of plants on the top rack of a chamber to the bottom rack of the same chamber while simultaneously moving the group of plants on the bottom rack of a chamber to the top rack of the adjacent chamber. Plants were moved within and across chambers daily during each experiment iteration.

*Leaf gas exchange measurements*

Leaf gas exchange measurements were collected in all plants (n = 144 individuals) on the seventh week of development, before the onset of reproduction. All gas exchange measurements were collected on the center leaflet of the most recent fully expanded trifoliate leaflet set using LI-6800 portable photosynthesis machines configured with a 6800-01A fluorometer head and 6 cm2 aperture (LI-COR Biosciences, Lincoln, NE, USA). Specifically, net photosynthesis rates (*A*net; μmol m-2 s-1), stomatal conductance rates (*g*sw; mol m-2 s-1), and intercellular CO2 concentrations (*C*i; μmol mol-1) were measured across a range of atmospheric CO2 concentrations (i.e., an *A*net/*C*i curve) using the Dynamic Assimilation™ Technique. The Dynamic Assimilation™ Technique corresponds well with traditional steady-state *A*net/*C*i curves in *G. max* (Saathoff and Welles, 2021; Tejera-Nieves *et al.*, 2024). *A*net/*C*i curves were generated along a reference CO2 ramp down from 420 µmol mol-1 CO2 to 20 µmol mol-1 CO2, followed by a ramp up from 420 µmol mol-1 CO2 to 1620 µmol mol-1 CO2 after a 90-second wait period at 420 µmol mol-1 CO2. The ramp rate for each curve was set to 200 μmol mol-1 min-1, logging every five seconds, generating 96 data points per response curve. All *A*net/*C*i curves were conducted after *A*net and *g*sw stabilized in an LI-6800 cuvette set to a 500 mol s-1 flow rate, 10000 rpm mixing fan speed, 1.5 kPa vapor pressure deficit, 25°C leaf temperature, 2000 μmol m-2 s-1 incoming light radiation, and initial reference CO2 concentration set to 420 µmol mol-1.

Snapshot *A*net measurements were extracted from each *A*net/*C*i curve, both at a common CO2 concentration, 420 µmol mol-1 CO2 (*A*net,420; μmol m-2 s-1), and growth CO2 concentration, 420 and 1000 µmol mol-1 CO2 (*A*net,gc; μmol m-2 s-1). We quantified *A*net,420 to gauge relative investment in photosynthetic tissues between treatment combinations and *A*net,gc to quantify photosynthetic performance between treatment combinations. Dark respiration (*R*d; μmol m-2 s-1) measurements were collected on the same leaflet used to generate *A*net/*C*i curves following at least a 30-minute period of darkness. Dark respiration measurements were collected on a 5-second log interval for 60 seconds after the leaf stabilized in an LI-6800 cuvette set to a 500 mol s-1 flow rate, 10000 rpm mixing fan speed, 1.5 kPa vapor pressure deficit, 25°C leaf temperature, and 420 µmol mol-1 reference CO2 concentration (regardless of CO2 treatment), with incoming light radiation set to 0 μmol m-2 s-1. A single dark respiration value was determined for each leaflet by calculating the mean dark respiration value across the logging interval.

*A*/*C*i *curve-fitting and parameter estimation*

*A*net/*C*i curves were fit using the ‘fitaci’ function in the ‘plantecophys’ R package (Duursma, 2015). This function estimates the apparent maximum rate of Rubisco carboxylation (*V*cmax; µmol m-2 s-1) and apparent maximum rate of electron transport for RuBP regeneration (*J*max; µmol m-2 s-1) based on the Farquhar et al. (1980) biochemical model of C3 photosynthesis. Triose phosphate utilization (TPU) limitation was included as an additional rate-limiting step after visually observing clear TPU limitation for most curves. All curve fits included measured dark respiration values. As *A*net/*C*i curves were generated using a common leaf temperature (25°C), curves were fit using Michaelis-Menten coefficients for Rubisco affinity to CO2 (*K*c; μmol mol-1) and O2 (*K*o; mmol mol-1), and the CO2 compensation point *(Γ*\*; μmol mol-1) reported in Bernacchi et al. (2001). Specifically, *K*c was set to 404.9 μmol mol-1, *K*o was set to 278.4 μmol mol-1, and *Γ*\* was set to 42.75 μmol mol-1. *V*cmax, *J*max, and *R*d estimates are referenced throughout the rest of the paper as *V*cmax25, *J*max25, and *R*d25.

*Leaf trait measurements*

The leaflet used for *A*net/*C*i curves and dark respiration measurements was harvested immediately following gas exchange measurements. Images of each focal leaflet were curated using a flat-bed scanner to determine fresh leaf area using the ‘LeafArea’ R package (Katabuchi, 2015), which automates leaf area calculations using ImageJ software (Schneider *et al.*, 2012). Post-processed images were visually assessed to check against errors in the automation process. Each focal leaflet was dried at 65C for at least 48 hours, weighed, and ground until homogenized. Leaf mass per area (*M*area; g m-2) was calculated as the ratio of dry leaflet biomass to fresh leaflet area. Leaf nitrogen content (*N*mass; gN g-1) was quantified using a subsample of ground and homogenized leaflet tissue through elemental combustion (Costech-4010, Costech, Inc., Valencia, CA, USA). Leaf nitrogen content per unit leaf area (*N*area; gN m-2) was calculated by multiplying *N*mass and *M*area. Photosynthetic nitrogen-use efficiency (*PNUE*gc; µmol CO2 g-1 Ns-1) was estimated as the ratio of *A*net,gc to *N*area.

Chlorophyll content was extracted from a second leaflet in the same trifoliate leaf set as the leaf used to generate *A*net/*C*i curves. A cork borer was used to punch 3-5 0.6 cm2 disks from the leaflet. Images of each set of leaflet disks were curated using a flat-bed scanner to determine wet leaf area using the 'LeafArea' R package (Katabuchi, 2015). Leaflet disks were shuttled into a test tube containing 10 mL dimethyl sulfoxide, vortexed, and incubated at 65°C for 120 minutes (Barnes *et al.*, 1992). Incubated test tubes were vortexed again before being loaded in 150 μL triplicate aliquots to a 96-well plate. Dimethyl sulfoxide was loaded in each plate as a single 150 μL triplicate aliquot and used as a blank. Absorbance measurements at 649 nm (*A*649) and 665 nm (*A*665) were recorded using a plate reader (Biotek Synergy H1; Biotek Instruments, Winooski, VT USA), with triplicate measurements averaged and corrected by the mean of the blank absorbance value. Blank-corrected absorbance values were used to estimate chlorophyll *a* (*Chl*a; μg mL-1) and chlorophyll *b* (*Chl*b; μg mL-1) following equations from Wellburn (1994):

(1)

and

(2)

*Chl*a and *Chl*b were converted to mmol mL-1 using the molar masses of chlorophyll *a* (893.51 g mol-1) and chlorophyll *b* (907.47 g mol-1), then added together to calculate the total chlorophyll content in dimethyl sulfoxide extractant (mmol mL-1). Total chlorophyll content (mmol) was determined by multiplying the total chlorophyll content in dimethyl sulfoxide by the volume of dimethyl sulfoxide extractant (10 mL). Area-based chlorophyll content (*Chl*area; mmol m-2) was calculated by dividing the total chlorophyll content by the total area of the leaflet disks.

*Whole-plant measurements*

All individuals were harvested, and biomass of major organ types (leaves, stems, roots, and nodules when present) were separated immediately following gas exchange measurements on the seventh week of development. Fresh leaf area of all harvested leaflets was measured using an LI-3100C (LI-COR Biosciences, Lincoln, Nebraska, USA). Total fresh leaf area (cm2) was calculated as the sum of all leaflet areas, including those used for gas exchange and chlorophyll extractions. Harvested material was separately dried in an oven set to 65°C for at least 48 hours to a constant mass, weighed, and then ground to homogeneity. Leaves and root nodules were ground using a mortar and pestle, while stems and roots were ground using an E3300 Single Speed Mini Cutting Mill (Eberbach Corp., MI, USA). Total biomass (g) was calculated as the sum of dry leaf, stem, root, and root nodule biomass. Carbon and nitrogen content was measured for each organ type through elemental combustion (Costech-4010, Costech, Inc., Valencia, CA, USA) using ground and homogenized organ tissue subsamples. The ratio of root nodule biomass to root biomass was calculated as an indicator of plant investment toward nitrogen fixation relative to other uptake pathways (e.g., direct uptake). The root:shoot ratio (unitless) was calculated as the ratio of belowground biomass (root and root nodule biomass) to shoot biomass (leaf and stem biomass). Leaf, stem, and root mass fractions were calculated as the dry biomass of each respective organ per unit total biomass (g g-1 in all cases).

Belowground biomass carbon costs to acquire nitrogen were quantified as the ratio of belowground biomass carbon to whole-plant nitrogen biomass (g C g N-1) (Perkowski *et al.*, 2021). Belowground biomass carbon (g C) was calculated as the sum of root and root nodule carbon biomass. Root carbon biomass and root nodule carbon biomass were calculated as the product of the organ biomass and respective organ carbon content. Whole-plant nitrogen biomass (g N) was calculated as the sum of total leaf, stem, root, and root nodule nitrogen biomass. Leaf, stem, root, and root nodule nitrogen biomass was calculated as the product of the organ biomass and respective organ nitrogen content. This calculation does not account for additional carbon costs associated with respiration, root exudation, or root turnover and may underestimate carbon costs to acquire nitrogen (Perkowski *et al.*, 2021).

*Statistical analyses*

Uninoculated plants with substantial root nodule formation (root nodule biomass: root biomass values greater than 0.05 g g-1) were removed from analyses following the assumption that plants were incompletely sterilized or contaminated. This decision resulted in the removal of sixteen plants from the analysis: two plants in the elevated CO2 treatment that received 35 ppm N, three plants in the elevated CO2 treatment that received 70 ppm N, one plant in the elevated CO2 treatment that received 210 ppm N, two plants in the elevated CO2 treatment that received 280 ppm N, two plants in the ambient CO2 treatment that received 0 ppm N, three plants in the ambient CO2 treatment that received 70 ppm N, two plants in the ambient CO2 treatment that received 105 ppm N, and one plant in the ambient CO2 treatment that received 280 ppm N. A summary of the replication scheme after these individuals were removed is included in the *Supplemental Information* (Table S3-4).

A series of linear mixed-effects models were built to investigate the impacts of CO2 concentration, nitrogen fertilization, and inoculation on *G. max* leaf nitrogen content, leaf gas exchange, total leaf area, biomass, biomass allocation, and plant investment in symbiotic nitrogen fixation. All models included CO2 treatment as a categorical fixed effect, inoculation treatment as a categorical fixed effect, and nitrogen fertilization as a continuous fixed effect, with all possible interaction terms between all three fixed effects included. Models accounted for climatic differences between chambers across experiment iterations by including a random intercept term that nested the starting chamber rack within CO2 treatment. Models with this independent variable structure were created for each of the following dependent variables: *N*area, *M*area, *N*mass, *Chl*area, *A*net,420, *A*net,gc, *V*cmax25, *J*max25, *J*max25:*V*cmax25, *R*d25, *PNUE*gc, total leaf area, total biomass, total leaf biomass, stem biomass, root biomass, root nodule biomass, root:shoot ratio, leaf mass fraction, stem mass fraction, root mass fraction, belowground biomass carbon costs to acquire nitrogen, belowground biomass carbon, whole-plant nitrogen biomass, and the root nodule biomass:root biomass ratio.

Shapiro-Wilk tests of normality were used to assess whether linear mixed-effects models satisfied residual normality assumptions. Models for *N*area, *N*mass, *Chl*area, *A*net,420, *A*net,gc, *V*cmax25, *J*max25, *J*max25:*V*cmax25, *R*d25, *PNUE*gc, total leaf area, leaf mass fraction, stem mass fraction, belowground biomass carbon, and whole-plant nitrogen biomass satisfied residual normality assumptions without data transformation. Models for *M*area, root:shoot ratio, belowground biomass carbon costs to acquire nitrogen, and root mass fraction satisfied residual normality assumptions with a natural log data transformation. Models for total biomass, leaf biomass, stem biomass, root biomass, root nodule biomass, and root nodule biomass: root biomass satisfied residual normality assumptions with a square root data transformation.

In all models, the ‘lmer’ function in the ‘lme4’ R package (Bates *et al.*, 2015) was used to fit each model, and the ‘Anova’ function in the ‘car’ R package (Fox and Weisberg, 2019) was used to calculate Type II Wald's χ2 and determine the significance (*α*=0.05) of each fixed effect coefficient. The ‘emmeans’ R package (Lenth, 2019) was used to conduct post-hoc comparisons using Tukey's tests, where degrees of freedom were approximated using the Kenward-Roger approach (Kenward and Roger, 1997). Trendlines and error ribbons representing the 95% confidence intervals were drawn in all figures using ‘emmeans’ outputs across the range in nitrogen fertilization values with a maximum of 36 data points per trendline (Table S4). All analyses and plots were conducted in R version 4.1.0 (R Core Team, 2021). Results for *N*mass and *M*area and organ biomasses are summarized in the *Supplemental Material* (Table S5, S7; Fig. S1).

**Results**

*Leaf nitrogen content*

Elevated CO2 reduced *N*area and *Chl*area by 29% and 30%, respectively (*p*<0.001 in both cases; Table 1; Fig. 1). Increasing nitrogen fertilization increased *N*area (*p*<0.001; Table 1; Fig. 1) more strongly under ambient CO2 than elevated CO2 (CO2-by-nitrogen fertilization interaction: *p*<0.05; Table 1), resulting in a stronger reduction in *N*area under elevated CO2 as nitrogen fertilization increased (Fig. S2). Uninoculated plants experienced a stronger reduction in *N*area under elevated CO2 than inoculated plants (CO2-by-inoculation interaction: *p*<0.05; Table 1). Increasing nitrogen fertilization increased *N*area and *Chl*area (*p*<0.001 in both cases; Table 1; Fig. 1) more strongly in uninoculated plants than inoculated plants (inoculation-by-nitrogen fertilization interaction: *p*<0.001 in both cases; Table 1).

*Gas exchange*

Elevated CO2 decreased *A*net,420 by 17% and increased *A*net,gc by 33% (*p*<0.001 in both cases; Table 2). Increasing nitrogen fertilization increased *A*net,420 and *A*net,gc similarly between CO2 treatments (CO2-by-nitrogen fertilization interaction: *p*>0.05; Table 2; Fig. 2a). Inoculated plants experienced a stronger increase in *A*net,gc under elevated CO2 than uninoculated plants (CO2-by-inoculation interaction: *p*<0.05; Table 2). Increasing nitrogen fertilization increased *A*net,420 and *A*net,gc(*p*<0.001 in both cases; Table 2) more strongly in uninoculated plants than inoculated plants (inoculation-by-nitrogen fertilization interaction: *p*<0.001 in both cases; Fig. 2a-b).

Elevated CO2 decreased *V*cmax25 by 16% and *J*max25 by 10%, increasing *J*max25:*V*cmax25 by 8% (*p*<0.05 in all cases; Table 2). Increasing nitrogen fertilization increased *V*cmax25 and *J*max25, but decreased *J*max25:*V*cmax25, similarly between CO2 (CO2-by-nitrogen fertilization interaction: *p*>0.05 in all cases; Table 2; Fig. 2b-d) and inoculation treatments (CO2-by-inoculation interaction: *p*>0.05 in all cases; Table 2). Increasing nitrogen fertilization increased *V*cmax25 and *J*max25 and decreased *J*max25:*V*cmax25 (*p*<0.001; Table 2), but these patterns were only observed in uninoculated plants (inoculation-by-nitrogen fertilization interaction: *p*<0.05 in all cases).

CO2 treatment did not affect *R*d25 (*p*>0.05; Table S6). Increasing nitrogen fertilization increased *R*d25 (*p*<0.05; Table S6), but this pattern was only observed in uninoculated plants (inoculation-by-nitrogen fertilization interaction: *p*<0.001; Table S6; Fig. S3a). Inoculated plants exhibited marginally greater *R*d25 than uninoculated plants (*p*<0.1; Table S6)

*Photosynthetic nitrogen-use efficiency*

Elevated CO2 increased *PNUE*gc by 97% (*p*<0.001; Table S6; Fig. S3b) due to a 33% increase in *A*net,gc (Fig. 2a) and 29% decrease in *N*area (Fig. 1a). Increasing nitrogen fertilization decreased *PNUE*gc (*p*<0.001; Table S6) more strongly under elevated CO2 (CO2-by-nitrogen fertilization interaction: *p*<0.05; Table S6; Fig. S3b), leading to a weaker increase in *PNUE*gc due to elevated CO2 as nitrogen fertilization increased (Fig. S6). Increasing nitrogen fertilization decreased *PNUE*gc (*p*<0.001; Table S6), but this pattern was only observed in inoculated plants (inoculation-by-nitrogen fertilization interaction: *p*<0.05; Table S6; Fig. S3b).

*Total leaf area and total biomass*

Elevated CO2 increased total leaf area and total biomass by 51% and 102%, respectively (*p*<0.001 in both cases; Table 3). Increasing nitrogen fertilization increased total leaf area and total biomass (*p*<0.001 in both cases; Table 3) more strongly under elevated CO2 than ambient CO2 (CO2-by-nitrogen fertilization interaction: *p*<0.001 in both cases; Table 3), leading to an amplified positive effect of elevated CO2 on total leaf area and total biomass as nitrogen fertilization increased (Fig. 3a-b). Inoculation had no effect on total leaf area or total biomass responses to elevated CO2 (CO2-by-inoculation interaction: *p*>0.05 in both cases; Table 3). Increasing nitrogen fertilization increased total leaf area and total biomass (*p*<0.001 in both cases; Table 3) more strongly in uninoculated plants than inoculated plants (inoculation-by-nitrogen fertilization interaction: *p*<0.001; Table 3; Fig. 3a-b).

*Biomass partitioning*

The root:shoot ratio decreased under elevated CO2 (*p*<0.05; Table 3; Fig. 3c), although this pattern was only observed in inoculated plants (CO2-by-inoculation interaction: *p*<0.05; Table 3, Fig. 3c). Reductions in the root:shoot ratio under elevated CO2 were driven by an increase in the leaf mass fraction under elevated CO2 (*p*<0.001; Table S7) that was only observed in inoculated plants (CO2-by-inoculation interaction: *p*<0.05; Table S7). CO2 treatment did not affect stem mass fraction (*p*>0.05; Table S7), although an interaction between CO2 and inoculation treatment indicated that elevated CO2 increased the root mass fraction in inoculated plants (CO2-by-inoculation interaction: *p*<0.05; Table S7). Increasing nitrogen fertilization decreased the root:shoot ratio (*p*<0.001; Table 3), a pattern that was marginally stronger in uninoculated plants than inoculated plants (CO2-by-inoculation interaction: *p*=0.051; Table 3; Fig. 3c). Increasing nitrogen fertilization increased the leaf mass fraction and decreased the root mass fraction (*p*<0.001 in both cases; Table S7), but these patterns only occurred in uninoculated plants (inoculation-by-nitrogen fertilization interaction: *p*<0.05 in both cases; Table S7). Increasing nitrogen fertilization increased stem mass fraction (*p*<0.001; Table S7), but these patterns only occurred in inoculated plants (inoculation-by-nitrogen fertilization interaction: *p*<0.001; Table S7).

*Belowground biomass carbon cost to acquire nitrogen*

Elevated CO2 increased belowground biomass carbon costs to acquire nitrogen (*p*<0.001; Table 3) more strongly in uninoculated plants than inoculated plants (CO2-by-inoculation interaction: *p*<0.001; Table 3). Increasing nitrogen fertilization decreased carbon costs to acquire nitrogen (*p*<0.001; Table 3) more strongly in uninoculated plants than inoculated plants (inoculation-by-nitrogen fertilization: *p*<0.001; Table 3; Fig. 3d). Interactions between inoculation and nitrogen fertilization treatments were more pronounced when plants were grown under elevated CO2 (CO2-by-inoculation-by-nitrogen fertilization interaction: *p*<0.05; Fig. 3d). This pattern was driven by a strong negative effect of increasing nitrogen fertilization on carbon costs to acquire nitrogen in uninoculated plants grown under elevated CO2 (Tukey: *p*<0.001) coupled with no nitrogen fertilization effect in inoculated plants grown under elevated CO2 (Tukey: *p*<0.001). Under ambient CO2, increasing nitrogen fertilization decreased carbon costs to acquire nitrogen similarly between inoculation treatments (Tukey: *p*>0.05).

Elevated CO2 increased belowground biomass carbon by 93% and increased whole-plant nitrogen biomass by 26% (*p*<0.001 in both cases; Table S8). Increasing nitrogen fertilization increased belowground biomass carbon and whole-plant nitrogen biomass more strongly under elevated CO2 than ambient CO2 (CO2-by-nitrogen fertilization interaction: *p*<0.001; Table S8; Fig. S5). These patterns resulted in an amplified positive effect of elevated CO2 on belowground biomass carbon and whole-plant nitrogen biomass as nitrogen fertilization increased, though this pattern was stronger for whole-plant nitrogen biomass than belowground biomass carbon (Fig. S5). Increasing nitrogen fertilization increased belowground biomass carbon and whole-plant nitrogen biomass (*p*<0.001; Table S8) more strongly in uninoculated plants than inoculated plants (inoculation-by-nitrogen fertilization interaction: *p*<0.001 in both cases; Table S8; Fig. S5).

*Plant investment toward symbiotic nitrogen fixation*

CO2 treatment did not affect root nodule: root biomass (*p*>0.05; Table 3; Fig. 4) despite anecdotally stronger positive effects of elevated CO2 on root biomass (96% increase; *p*<0.001; Table S7) than root nodule biomass (70% increase; *p*<0.001; Table S7). Increasing nitrogen fertilization decreased root nodule: root biomass (*p*<0.001; Table 3) more strongly in inoculated plants than uninoculated plants (inoculation-by-nitrogen fertilization interaction: *p*<0.001; Table 3; Fig. 4).

**Discussion**

*Glycine max* plants were grown under two CO2 concentrations, two inoculation treatments, and nine nitrogen fertilization treatments in a full-factorial growth chamber experiment. We used data collected from this experiment to (1) determine whether plant responses to elevated CO2 aligned more closely with the nitrogen limitation or eco-evolutionary optimality hypothesis and (2) assess how the ability to associate with symbiotic nitrogen-fixing bacteria might influence these responses.

*Leaf photosynthetic responses to elevated CO2 are unrelated to nitrogen availability*

Individuals grown under elevated CO2 experienced a reduction in *A*net,420 (Table 2), leaf nitrogen content (Fig. 1a, S1), *V*cmax25 (Fig. 2b), and *J*max25 (Fig. 2c) compared to plants grown under ambient CO2. These patterns suggest a downregulation of leaf-level investment toward photosynthetic enzymes under elevated CO2. This downregulation was likely driven by increased Rubisco affinity for carboxylation relative to oxygenation, which decreased leaf-level demand to build and maintain photosynthetic enzymes (Bazzaz, 1990; Dong *et al.*, 2022). Despite reduced investment toward photosynthetic enzymes, elevated CO2 increased *A*net,gc (Fig. 2a). This response was associated with a reduction in *N*area and a larger reduction in *V*cmax25 than *J*max25, which increased photosynthetic nitrogen-use efficiency (Fig. S3b) and *J*max25:*V*cmax25 and allowed enhanced *A*net,gc to be achieved by approaching optimal coordination (Chen *et al.*, 1993; Maire *et al.*, 2012; Smith and Keenan, 2020). These patterns are consistent with our expectations and previous studies that have investigated leaf photosynthetic responses to elevated CO2 (Drake *et al.*, 1997; Ainsworth *et al.*, 2002; Ainsworth and Long, 2005; Ainsworth and Rogers, 2007; Crous *et al.*, 2010; Lee *et al.*, 2011; Smith and Dukes, 2013; Poorter *et al.*, 2022; Cui *et al.*, 2023; Stocker *et al.*, 2025).

Positive effects of elevated CO2 on *A*net,gc (Fig. 2a) and *J*max25:*V*cmax25 (Fig. 2d) and negative effects of elevated CO2 on *A*net,420, *V*cmax25, and *J*max25 (Figs. 2a-c) were not modified by nitrogen fertilization, as the slope that explained the effects of increasing nitrogen fertilization on each of these traits was similar between CO2 treatments. Instead, the increase in *J*max25:*V*cmax25 (Fig. 2d) and *PNUE*gc (Fig. S3b) under elevated CO2 provides strong support for the idea that leaves were downregulating *V*cmax25 in response to elevated CO2 such that enhanced *A*net,gccould be achieved by approaching optimal coordination of Rubisco carboxylation and electron transport for RuBP regeneration (Chen *et al.*, 1993; Maire *et al.*, 2012; Smith and Keenan, 2020).

Negative effects of elevated CO2 on mass- and area-based leaf nitrogen content became more pronounced with increasing nitrogen fertilization (Fig. S2a-b). Since nitrogen fertilization did not affect photosynthetic responses to elevated CO2, this decline in leaf nitrogen content may reflect reduced allocation to non-photosynthetic pools, such as structural tissue or chemical pathways that contribute to herbivore defense (Zavala *et al.*, 2013; Onoda *et al.*, 2017; Johnson *et al.*, 2020). While not a primary focus of this study, understanding leaf nitrogen allocation responses to elevated CO2 across nitrogen availability gradients would help clarify the role of leaf nitrogen allocation on leaf-level responses to elevated CO2.

Overall, leaf photosynthetic responses to elevated CO2 showed strong support for the eco-evolutionary optimality hypothesis. Photosynthetic responses to elevated CO2 were independent from nitrogen fertilization, suggesting that these responses were wholly determined through changes in leaf-level demand to build and maintain photosynthetic enzymes. These findings also reinforce previous work showing that leaf photosynthetic responses to elevated CO2 are decoupled from nitrogen availability (Lee *et al.*, 2011; Pastore *et al.*, 2019; Smith and Keenan, 2020; Harrison *et al.*, 2021). Additionally, our results indicate that optimal resource investment to photosynthetic capacity may function as a nitrogen-savings mechanism that allows plants to maximize resource-use efficiency at the leaf-level as a strategy for maximizing resource allocation to whole-plant growth (Smith and Keenan, 2020; Smith *et al.*, 2024).

*Whole-plant responses to elevated CO2 are constrained by nitrogen availability*

Leaf photosynthetic responses to elevated CO2 corresponded with increased total leaf area and total biomass (Fig. 3a-b), supporting previous work (Ainsworth *et al.*, 2002; Ainsworth and Long, 2005; Smith and Dukes, 2013; Poorter *et al.*, 2022; Stocker *et al.*, 2025). Increased total leaf area increased whole-plant capacity for light interception, boosting whole-plant photosynthesis and supporting biomass accumulation when coupled with an increase in leaf-level *A*net,gc. Contrasting expectations and previous work (Nie *et al.*, 2013; Stocker *et al.*, 2025), elevated CO2 decreased the root-to-shoot ratio (Fig. 3c) through an increase in the leaf mass fraction and no change in the stem or root mass fractions (Table S7). Despite this, plants experienced an increase in root biomass (Fig. S6b) and belowground carbon allocation (Fig. S5a) under elevated CO2, suggesting that plants responded to heightened whole-plant demand under elevated CO2 by investing in structures that support nutrient acquisition even if they allocated relatively more biomass aboveground .

Increasing nitrogen fertilization enhanced the positive effects of elevated CO2 on total leaf area and total biomass (Fig. 3a-b). Interestingly, this interaction revealed no effect of CO2 treatment on total leaf area in uninoculated individuals under low nitrogen fertilization, supporting previous work showing that CO2 fertilization effects on traits related to whole-plant growth are often absent under low nutrient availability (Sigurdsson *et al.*, 2013). Similar effects of CO2 treatment on total leaf area under low nitrogen fertilization may have been due to plants being unable to satisfy demand for soil nitrogen similarly between the two CO2 treatments. Stronger positive effects of elevated CO2 on total leaf area and total biomass with increasing nitrogen fertilization were associated with stronger increases in belowground carbon allocation and whole-plant nitrogen uptake (Fig. S5), supporting the nitrogen limitation hypothesis (Luo *et al.*, 2004; Reich *et al.*, 2006; Norby *et al.*, 2010; Feng *et al.*, 2015). These findings indicate that plants grown under elevated CO2 satisfied the greater whole-plant demand to build new tissues by increasing investment to nitrogen acquisition, suggesting that whole-plant responses to elevated CO2 were constrained by nitrogen availability as expected. Despite this, nitrogen availability did not modify whether plants invested in aboveground or belowground tissues, as indicated by similar positive effects of increasing nitrogen fertilization on the root-to-shoot ratio (Fig. 3c) and all organ mass fractions between CO2 treatments (Table S7). These responses indicate that biomass allocation responses to elevated CO2 were more strongly dictated by changes in whole-plant demand to build new tissues than the supply of nutrients.

*Inoculation does not affect leaf or whole-plant responses to elevated* *CO2*

Inoculation increased *N*area (Fig. 1a), *A*net,420, *A*net,gc (Fig. 2a), *V*cmax25 (Fig. 2b), *J*max25 (Fig. 2c), total leaf area (Fig. 3a), and total biomass (Fig. 3b), but decreased *J*max25:*V*cmax25 (Fig. 2d). These results support previous studies suggesting that species forming symbiotic associations with nitrogen-fixing bacteria have greater leaf nitrogen content, photosynthetic capacity, and growth than those that do not (Adams *et al.*, 2016; Bytnerowicz *et al.*, 2023). The positive effects of inoculation on leaf and whole-plant traits were strongest under low nitrogen fertilization and diminished with increasing nitrogen fertilization, likely due to a reduction in plant investment toward symbiotic nitrogen fixation (Fig. 4). These patterns support the idea that forming associations with symbiotic nitrogen-fixing bacteria confers a competitive advantage in nitrogen-limited environments, where access to a less-finite nitrogen pool (i.e., the atmosphere) allows plants to satisfy leaf- and whole-plant demand more efficiently than relying on limited soil nitrogen (Rastetter *et al.*, 2001; Andrews *et al.*, 2011; McCulloch and Porder, 2021).

Inoculation largely had no effect on leaf- or whole-plant responses to elevated CO2, but played a strong role in determining the effect of nitrogen fertilization on measured traits. This null inoculation effect on plant responses to elevated CO2 was consistent across the nitrogen fertilization gradient, which contrasts our hypothesis that inoculation would enhance plant responses to elevated CO2 most strongly under low nitrogen fertilization (Rastetter *et al.*, 2001; Perkowski *et al.*, 2021). Previous research has highlighted that nitrogen-fixing species typically show stronger responses to elevated CO2 than non-fixing species (Ainsworth *et al.*, 2002; Ainsworth and Long, 2005), although some studies question the generality of this pattern (Nowak *et al.*, 2004; Rogers *et al.*, 2009). Our findings assert that the ability to associate with symbiotic nitrogen-fixing bacteria played no role in determining whether plant responses to elevated CO2 aligned with the nitrogen limitation or eco-evolutionary optimality hypotheses, even though inoculated individuals grown under elevated CO2 exhibited greater root nodule biomass (Fig. S6a) and reduced carbon costs to acquire nitrogen (Fig. 3d) compared to those grown under ambient CO2.

As mentioned above, plants grown under elevated CO2 exhibited an increase in root nodule biomass (Fig. S6a). This pattern indicates that plants responded to heightened whole-plant demand for new tissue growth by increasing nitrogen uptake through nitrogen fixation. However, the increase in root nodule biomass was circumvented by a stronger increase in root biomass (Fig. S6b). This pattern indicates an investment shift toward direct uptake with increasing CO2, a response that runs counter to previous work showing that plants increase investment in microbial symbionts when whole-plant demand to build new tissues increases (Taylor and Menge, 2018; Friel and Friesen, 2019; Perkowski *et al.*, 2021). If true, increased relative allocation to root biomass may have been a strategy to prioritize the acquisition of non-nitrogen resources, as nitrogen fixation may increase the extent by which physiology and plant growth becomes limited by other nutrients, such as phosphorus (Finzi and Rodgers, 2009). Previous research has shown that phosphorus plays a key role in shaping plant responses to elevated CO2 and that the benefits of nitrogen fixation under elevated CO2 become more apparent when other nutrients, such as phosphorus, are also available in sufficient supply (van Groenigen *et al.*, 2006; Jiang *et al.*, 2020). Thus, it is possible that the null effects of inoculation on plant responses to elevated CO2 may have been driven by phosphorus colimitation, although future work is needed to test this hypothesis.

*Modeling implications*

Many terrestrial biosphere models predict photosynthetic capacity through parameterized relationships between *N*area and *V*cmax (Smith and Dukes, 2013; Rogers *et al.*, 2017), which assumes that leaf nitrogen-photosynthesis relationships are constant across growing environments. Our results build on previous work suggesting that leaf nitrogen-photosynthesis relationships dynamically change across growing environments (Luo *et al.*, 2021; Waring *et al.*, 2023). Specifically, elevated CO2 reduced leaf nitrogen content (Fig. 1a) more strongly than it increased *A*net,gc (Fig. 2a) and decreased *V*cmax25 (Fig. 2b) and *J*max25 (Fig. 2c), while inoculation increased *V*cmax25 and *J*max25 more strongly than it increased leaf nitrogen content. These patterns indicate that elevated CO2 increased the fractional pool of leaf nitrogen content allocated to Rubisco and bioenergetics, while inoculation decreased the fraction of leaf nitrogen content allocated to Rubisco and bioenergetics (Niinemets and Tenhunen, 1997).

Increasing nitrogen fertilization increased indices of apparent photosynthetic capacity, but this pattern was only observed in uninoculated plants. Increasing nitrogen fertilization also increased *N*area and *Chl*area more strongly in uninoculated plants (Fig. 1). Eco-evolutionary optimality theory predicts that plants should exhibit strong positive effects of increasing nitrogen availability on photosynthetic traits when nitrogen availability is insufficient for satisfying leaf-level demand for building and maintaining photosynthetic enzymes, or when changes in nitrogen availability decrease the relative costs of nitrogen acquisition and use compared to those of water acquisition and use (Wright *et al.*, 2003; Harrison *et al.*, 2021; Stocker *et al.*, 2025). In such cases where nitrogen availability exceeds leaf-level demand for photosynthetic enzymes or costs to acquire nitrogen relative to water increase, the theory predicts that positive effects of increasing nitrogen availability on photosynthesis should diminish, with excess nitrogen not needed to satisfy leaf-level demand for photosynthesis being allocated toward the construction of other plant tissues (e.g., additional leaves). Given this, strong positive effects of increasing nitrogen fertilization on indices of photosynthetic capacity in uninoculated plants were expected, as uninoculated plants were nitrogen-limited under low nitrogen fertilization and could not meet the leaf-level demand for photosynthetic enzymes. We also found some evidence for a diminished positive effect of nitrogen fertilization on photosynthetic traits, with uninoculated plants demonstrating smaller increases in *V*cmax25 between 350 and 630 ppm N (39% increase) than between 0 ppm N and 280 ppm N (79% increase). In contrast, nitrogen fertilization effects on photosynthetic traits were absent in inoculated individuals. This pattern was also expected, as inoculated plants were able to acquire sufficient nitrogen across the nitrogen availability gradient to satisfy leaf-level photosynthetic demand, investing more strongly in microbial symbionts under low nitrogen fertilization and shifting to nitrogen acquisition through direct uptake pathways as nitrogen became more available.

Overall, these results indicate that leaf nitrogen-photosynthesis relationships are context-dependent on nitrogen acquisition strategy, may only be constant in environments where nitrogen availability limits leaf physiology, and will likely shift in response to increasing atmospheric CO2 concentrations. Terrestrial biosphere models that predict photosynthetic capacity through parameterized relationships between *N*area and *V*cmax (Kattge *et al.*, 2009; Walker *et al.*, 2014) may risk overestimating photosynthetic capacity, therefore net primary productivity and the magnitude of the land carbon sink, under future novel growth environments.

Our results demonstrate that optimal resource allocation to photosynthetic capacity defines leaf photosynthetic responses to elevated CO2 and that these responses are not modified by nitrogen availability. Current approaches for simulating photosynthetic responses to CO2 in terrestrial biosphere models with coupled carbon and nitrogen cycles often invoke patterns expected from the nitrogen limitation hypothesis, where nitrogen availability diminishes with time due to increasing CO2 concentrations because whole-plant nitrogen demand continually exceeds supply, depleting the pool of available nitrogen for plants to acquire and allocate to the construction and maintenance of new tissues. This response causes models to simulate a reduction in leaf nitrogen content and therefore photosynthetic capacity, as leaf-level photosynthesis is commonly modeled as a function of positive relationships between nitrogen availability, leaf nitrogen content, and photosynthetic capacity (Smith and Dukes, 2013; Rogers *et al.*, 2017). Findings presented here contradict this framework, suggesting that leaf photosynthetic responses to elevated CO2 result in optimized nitrogen allocation to satisfy reduced leaf nitrogen demand to build and maintain photosynthetic enzymes. Optimality models that use principles from eco-evolutionary optimality theory are capable of capturing photosynthetic responses to CO2 independent of nitrogen availability (Smith and Keenan, 2020; Harrison *et al.*, 2021; Stocker *et al.*, 2025), suggesting that the inclusion of such frameworks may improve the accuracy by which terrestrial biosphere models simulate photosynthetic processes with increasing atmospheric CO2 concentrations.

*Limitations*

Previous work has highlighted that pot experiments restrict belowground rooting volume and may alter plant allocation responses to environmental change (Ainsworth *et al.*, 2002; Poorter *et al.*, 2012). In this study, the ratio of pot volume to total biomass was greater under elevated CO2 and increased with increasing nitrogen fertilization such that several treatment combinations exceeded values recommended to avoid growth limitation imposed by pot volume (<1 g L-1; Table S9; Fig. S7; Poorter et al., 2012). However, there was no evidence to suggest that pot size limited plant growth, as evidenced by the lack of a saturating effect of increasing fertilization on total biomass, belowground carbon biomass, or root biomass under conditions where biomass: pot volume ratios exceeded 1 g L-1 (e.g., individuals of either inoculation status grown under high fertilization and elevated CO2). Field studies that do not restrict belowground rooting volume have observed similar leaf and whole-plant responses to elevated CO2 (Crous *et al.*, 2010; Lee *et al.*, 2011; Pastore *et al.*, 2019; Smith and Keenan, 2020), indicating that the pot volume used in this study (6 L) was sufficient to avoid growth limitation.

Importantly, there are inherent limitations in using a pot experiment to make inferences about how nitrogen availability modifies community- or ecosystem-level responses to elevated CO2. While we caution against using this study to make such extrapolations, a similar experiment conducted under field conditions would help validate the patterns observed here while providing insight into how resource competition within and across species may shape plant responses to nitrogen availability and elevated CO2.

*Conclusions*

Our study provides strong support for the eco-evolutionary optimality hypothesis at the leaf level, where leaf photosynthetic responses to elevated CO2 were independent of nitrogen fertilization and inoculation treatment. Instead, elevated CO2 reduced the maximum rate of Rubisco carboxylation more strongly than it reduced the maximum rate of electron transport for RuBP regeneration, allowing plants to achieve greater net photosynthesis rates under CO2 growth conditions by approaching optimal coordination while reducing leaf nitrogen demand to build and maintain photosynthetic enzymes. In contrast, at the whole-plant level, nitrogen availability played a central role in regulating plant responses to elevated CO2, consistent with the nitrogen limitation hypothesis. Specifically, increases in total leaf area, total biomass, and plant nitrogen under elevated CO2 were all enhanced with increasing nitrogen fertilization.

While inoculation increased root nodulation under elevated CO2, it did not significantly enhance whole-plant responses to elevated CO2, even under low nitrogen conditions where plants were most strongly invested in symbiotic nitrogen-fixing bacteria. This response may have been due to stronger increases in root biomass that caused plants to prioritize direct nitrogen uptake pathways over symbiotic nitrogen fixation as whole-plant demand to build new tissues increased, perhaps as a strategy to reduce colimitation by other nutrients, such as phosphorus.

Overall, plants grown under elevated CO2 responded to increased nitrogen availability by increasing the number of optimally coordinated leaves, while changes in nitrogen availability did not modify the downregulation in apparent photosynthetic capacity under elevated CO2. The differential role of nitrogen availability on leaf and whole-plant responses to elevated CO2 and the dynamic leaf nitrogen-photosynthesis relationships across CO2 and nitrogen fertilization treatments suggests that terrestrial biosphere models may improve simulations of photosynthetic responses to increasing atmospheric CO2 concentrations by adopting frameworks that include optimality principles.

**Supplementary data**

**Text S1** A continuance of the results section that describe the effects of treatment combinations on mass-based leaf nitrogen content and leaf mass per unit leaf area, organ biomass, and the ratio of total biomass to pot volume

**Table S1** Summary table containing volumes of compounds used to create modified Hoagland’s solutions for each soil nitrogen fertilization treatment

**Table S2** Summary of the daily growth chamber growing condition program

**Table S3** Replication scheme for each unique CO2-by-inoculation-by-N fertilization combination

**Table S4** Replication scheme for each unique CO2-by-inoculation combination

**Table S5** Effects of treatment combinations on leaf nitrogen content and leaf mass per area

**Table S6** Effects of treatment combinations on dark respiration and photosynthetic nitrogen-use efficiency

**Table S7** Effects of treatment combinations on biomass partitioning

**Table S8** Effects of treatment combinations on components of the carbon cost to acquire nitrogen

**Table S9** Effects of treatment combinations on the ratio of total biomass to pot volume

**Figure S1** Effects of treatment combinations on mass-based leaf nitrogen content and leaf biomass per unit leaf area

**Figure S2** Effects of CO2 and nitrogen fertilization on area-based leaf nitrogen content, mass-based leaf nitrogen content, and leaf biomass per unit leaf area

**Figure S3** Effects of treatment combinations on dark respiration at 25°C and photosynthetic nitrogen-use efficiency at growth CO2 concentration

**Figure S4** Effects of CO2 and nitrogen fertilization on photosynthetic nitrogen-use efficiency at growth CO2 concentration

**Figure S5** Effects of treatment combinations on belowground biomass carbon and total nitrogen biomass

**Figure S6** Effects of treatment combinations on root nodule biomass and root biomass

**Figure S7** Effects of treatment combinations on the ratio of whole-plant biomass to pot volume

**Acknowledgements**

This study is a contribution to the LEMONTREE (Land Ecosystem Models based On New Theory, obseRvations and ExperimEnts) project, receiving support through Schmidt Sciences, LLC. EAP acknowledges support from a Texas Tech University Doctoral Dissertation Completion Fellowship and a Botanical Society of America Graduate Student Research Award. This work was also supported by US National Science Foundation awards to NGS (DEB-2045968 and DEB-2217353).

**Author contributions**

EAP conceptualized the study objectives and designed the experiment in collaboration with NGS, collected data, conducted data analysis, and wrote the first manuscript draft. EE assisted with data collection and experiment maintenance. NGS conceptualized study objectives and experimental design with EAP and oversaw experiment progress. All authors provided manuscript feedback and approved the manuscript in its current form for submission to *Journal of Experimental Botany*.

**Conflict of Interest**

The authors declare no conflicts of interest.

**Funding**

US National Science Foundation (DEB-2045968 and DEB-2217353), Schmidt Sciences, LLC.

**Data Availability**

All R scripts, data, and metadata are available at <https://doi.org/10.5281/zenodo.12812758> (or on GitHub at: <https://github.com/eaperkowski/NxCO2xI_ms_data>)

**References**

**Adams MA, Turnbull TL, Sprent JI, Buchmann N**. 2016. Legumes are different: Leaf nitrogen, photosynthesis, and water use efficiency. Proceedings of the National Academy of Sciences of the United States of America **113**, 4098–4103.

**Ainsworth EA, Davey PA, Bernacchi CJ, *et al.*** 2002. A meta-analysis of elevated [CO2] effects on soybean (*Glycine max*) physiology, growth and yield. Global Change Biology **8**, 695–709.

**Ainsworth EA, Long SP**. 2005. What have we learned from 15 years of free-air CO2 enrichment (FACE)? A meta-analytic review of the responses of photosynthesis, canopy properties and plant production to rising CO2. New Phytologist **165**, 351–372.

**Ainsworth EA, Rogers A**. 2007. The response of photosynthesis and stomatal conductance to rising [CO2]: mechanisms and environmental interactions. Plant, Cell and Environment **30**, 258–270.

**Allen K, Fisher JB, Phillips RP, Powers JS, Brzostek ER**. 2020. Modeling the carbon cost of plant nitrogen and phosphorus uptake across temperate and tropical forests. Frontiers in Forests and Global Change **3**, 1–12.

**Andrews M, James EK, Sprent JI, Boddey RM, Gross E, dos Reis FB**. 2011. Nitrogen fixation in legumes and actinorhizal plants in natural ecosystems: Values obtained using 15N natural abundance. Plant Ecology and Diversity **4**, 117–130.

**Arora VK, Katavouta A, Williams RG, *et al.*** 2020. Carbon-concentration and carbon-climate feedbacks in CMIP6 models and their comparison to CMIP5 models. Biogeosciences **17**, 4173–4222.

**Barber SA**. 1962. A diffusion and mass-flow concept of soil nutrient availability. Soil Science **93**, 39–49.

**Barnes JD, Balaguer L, Manrique E, Elvira S, Davison AW**. 1992. A reappraisal of the use of DMSO for the extraction and determination of chlorophylls a and b in lichens and higher plants. Environmental and Experimental Botany **32**, 85–100.

**Bates D, Mächler M, Bolker B, Walker S**. 2015. Fitting linear mixed-effects models using lme4. Journal of Statistical Software **67**, 1–48.

**Bazzaz FA**. 1990. The response of natural ecosystems to the rising global CO2 levels. Annual review of ecology and systematics **21**, 167–196.

**Bernacchi CJ, Morgan PB, Ort DR, Long SP**. 2005. The growth of soybean under free air [CO2] enrichment (FACE) stimulates photosynthesis while decreasing in vivo Rubisco capacity. Planta **220**, 434–446.

**Bernacchi CJ, Singsaas EL, Pimentel C, Portis AR, Long SP**. 2001. Improved temperature response functions for models of Rubisco-limited photosynthesis. Plant, Cell and Environment **24**, 253–259.

**Brzostek ER, Fisher JB, Phillips RP**. 2014. Modeling the carbon cost of plant nitrogen acquisition: Mycorrhizal trade-offs and multipath resistance uptake improve predictions of retranslocation. Journal of Geophysical Research: Biogeosciences **119**, 1684–1697.

**Bytnerowicz TA, Funk JL, Menge DNL, Perakis SS, Wolf AA**. 2023. Leaf nitrogen affects photosynthesis and water use efficiency similarly in nitrogen-fixing and non-fixing trees. Journal of Ecology, 1–15.

**Chen J-L, Reynolds JF, Harley PC, Tenhunen JD**. 1993. Coordination theory of leaf nitrogen distribution in a canopy. Oecologia **93**, 63–69.

**Coleman JS, McConnaughay KDM, Bazzaz FA**. 1993. Elevated CO2 and plant nitrogen-use: is reduced tissue nitrogen concentration size-dependent? Oecologia **93**, 195–200.

**Crous KY, Reich PB, Hunter MD, Ellsworth DS**. 2010. Maintenance of leaf N controls the photosynthetic CO2 response of grassland species exposed to 9 years of free-air CO2 enrichment. Global Change Biology **16**, 2076–2088.

**Cui E, Xia J, Luo Y**. 2023. Nitrogen use strategy drives interspecific differences in plant photosynthetic CO2 acclimation. Global Change Biology **29**, 3667–3677.

**Curtis PS**. 1996. A meta-analysis of leaf gas exchange and nitrogen in trees grown under elevated carbon dioxide. Plant, Cell and Environment **19**, 127–137.

**Davies-Barnard T, Meyerholt J, Zaehle S, *et al.*** 2020. Nitrogen cycling in CMIP6 land surface models: progress and limitations. Biogeosciences **17**, 5129–5148.

**Davies-Barnard T, Zaehle S, Friedlingstein P**. 2022. Assessment of the impacts of biological nitrogen fixation structural uncertainty in CMIP6 earth system models. Biogeosciences **19**, 3491–3503.

**Dong N, Wright IJ, Chen JM, Luo X, Wang H, Keenan TF, Smith NG, Prentice IC**. 2022. Rising CO2 and warming reduce global canopy demand for nitrogen. New Phytologist **235**, 1692–1700.

**Drake BG, Gonzàlez-Meler MA, Long SP**. 1997. More efficient plants: a consequence of rising atmospheric CO2? Annual Review of Plant Biology **48**, 609–639.

**Duursma RA**. 2015. Plantecophys - an R package for analysing and modelling leaf gas exchange data. PLOS ONE **10**, e0143346.

**Evans JR**. 1989. Photosynthesis and nitrogen relationships in leaves of C3 plants. Oecologia **78**, 9–19.

**Evans JR, Clarke VC**. 2019. The nitrogen cost of photosynthesis. Journal of Experimental Botany **70**, 7–15.

**Farquhar GD, von Caemmerer S, Berry JA**. 1980. A biochemical model of photosynthetic CO2 assimilation in leaves of C3 species. Planta **149**, 78–90.

**Feng Z, Rütting T, Pleijel H, Wallin G, Reich PB, Kammann CI, Newton PCD, Kobayashi K, Luo Y, Uddling J**. 2015. Constraints to nitrogen acquisition of terrestrial plants under elevated CO2. Global Change Biology **21**, 3152–3168.

**Field CB, Mooney HA**. 1986. Photosynthesis--nitrogen relationship in wild plants. On the Economy of Plant Form and Function: Proceedings of the Sixth Maria Moors Cabot Symposium, Evolutionary Constraints on Primary Productivity, Adaptive Patterns of Energy Capture in Plants, Harvard Forest, August 1983. Cambridge [Cambridgeshire]: Cambridge University Press, c1986.

**Finzi AC, Moore DJP, DeLucia EH, *et al.*** 2006. Progressive nitrogen limitation of ecosystem processes under elevated CO2 in a warm-temperate forest. Ecology **87**, 15–25.

**Finzi AC, Norby RJ, Calfapietra C, *et al.*** 2007. Increases in nitrogen uptake rather than nitrogen-use efficiency support higher rates of temperate forest productivity under elevated CO2. Proceedings of the National Academy of Sciences **104**, 14014–14019.

**Finzi AC, Rodgers VL**. 2009. Bottom-up rather than top-down processes regulate the abundance and activity of nitrogen fixing plants in two Connecticut old-field ecosystems. Biogeochemistry **95**, 309–321.

**Fisher JB, Sitch S, Malhi Y, Fisher RA, Huntingford C, Tan S-Y**. 2010. Carbon cost of plant nitrogen acquisition: A mechanistic, globally applicable model of plant nitrogen uptake, retranslocation, and fixation. Global Biogeochemical Cycles **24**, 1–17.

**Fox J, Weisberg S**. 2019. *An R companion to applied regression*. Thousand Oaks, California: Sage.

**Friel CA, Friesen ML**. 2019. Legumes modulate allocation to rhizobial nitrogen fixation in response to factorial light and nitrogen manipulation. Frontiers in Plant Science **10**, 1316.

**van Groenigen KJ, Six J, Hungate BA, De Graaff MA, Van Breemen N, Van Kessel C**. 2006. Element interactions limit soil carbon storage. Proceedings of the National Academy of Sciences of the United States of America **103**, 6571–6574.

**Gutschick VP**. 1981. Evolved strategies in nitrogen acquisition by plants. The American Naturalist **118**, 607–637.

**Harrison SP, Cramer W, Franklin O, *et al.*** 2021. Eco-evolutionary optimality as a means to improve vegetation and land-surface models. New Phytologist **231**, 2125–2141.

**Hoagland DR, Arnon DI**. 1950. The water-culture method for growing plants without soil. California Agricultural Experiment Station: 347 **347**, 1–32.

**Hungate BA, Dukes JS, Shaw MR, Luo Y, Field CB**. 2003. Nitrogen and climate change. Science **302**, 1512–1513.

**IPCC**. 2021. *Climate Change 2021: The Physical Science Basis. Contribution of Working Group I to the Sixth Assessment Report of the Intergovernmental Panel on Climate Change*. (V Masson-Delmotte, P Zhai, A Pirani, *et al.*, Eds.). Cambridge, UK and New York, USA: Cambridge University Press.

**Iversen CM**. 2010. Digging deeper: Fine-root responses to rising atmospheric CO2 concentration in forested ecosystems. New Phytologist **186**, 346–357.

**Iversen CM, Ledford J, Norby RJ**. 2008. CO2 enrichment increases carbon and nitrogen input from fine roots in a deciduous forest. New Phytologist **179**, 837–847.

**Jiang M, Caldararu S, Zhang H, *et al.*** 2020. Low phosphorus supply constrains plant responses to elevated CO2: A meta‐analysis. Global Change Biology **26**, 5856–5873.

**Johnson SN, Waterman JM, Hall CR**. 2020. Increased insect herbivore performance under elevated CO2 is associated with lower plant defence signalling and minimal declines in nutritional quality. Scientific Reports **10**, 14553.

**Katabuchi M**. 2015. LeafArea: An R package for rapid digital analysis of leaf area. Ecological Research **30**, 1073–1077.

**Kattge J, Knorr W, Raddatz T, Wirth C**. 2009. Quantifying photosynthetic capacity and its relationship to leaf nitrogen content for global-scale terrestrial biosphere models. Global Change Biology **15**, 976–991.

**Kenward MG, Roger JH**. 1997. Small sample inference for fixed effects from restricted maximum likelihood. Biometrics **53**, 983.

**Kou-Giesbrecht S, Arora VK, Seiler C, *et al.*** 2023. Evaluating nitrogen cycling in terrestrial biosphere models: a disconnect between the carbon and nitrogen cycles. Earth System Dynamics **14**, 767–795.

**LeBauer DS, Treseder KK**. 2008. Nitrogen limitation of net primary productivity in terrestrial ecosystems is globally distributed. Ecology **89**, 371–379.

**Lee TD, Barrott SH, Reich PB**. 2011. Photosynthetic responses of 13 grassland species across 11 years of free-air CO2 enrichment is modest, consistent and independent of N supply. Global Change Biology **17**, 2893–2904.

**Lenth R**. 2019. emmeans: estimated marginal means, aka least-squares means. https://cran.r-project.org/package=emmeans.

**Liang J, Qi X, Souza L, Luo Y**. 2016. Processes regulating progressive nitrogen limitation under elevated carbon dioxide: a meta-analysis. Biogeosciences **13**, 2689–2699.

**Lu J, Yang J, Keitel C, Yin L, Wang P, Cheng W, Dijkstra FA**. 2022. Belowground carbon efficiency for nitrogen and phosphorus acquisition varies between *Lolium perenne* and *Trifolium repens* and depends on phosphorus fertilization. Frontiers in Plant Science **13**, 1–9.

**Luo Y, Currie WS, Dukes JS, *et al.*** 2004. Progressive nitrogen limitation of ecosystem responses to rising atmospheric carbon dioxide. BioScience **54**, 731–739.

**Luo Y, Field CB, Mooney HA**. 1994. Predicting responses of photosynthesis and root fraction to elevated [CO2]: interactions among carbon, nitrogen, and growth. Plant, Cell & Environment **17**, 1195–1204.

**Luo X, Keenan TF, Chen JM, *et al.*** 2021. Global variation in the fraction of leaf nitrogen allocated to photosynthesis. Nature Communications **12**, 4866.

**Maire V, Martre P, Kattge J, Gastal F, Esser G, Fontaine S, Soussana J-F**. 2012. The coordination of leaf photosynthesis links C and N fluxes in C3 plant species. PLoS ONE **7**, e38345.

**Makino A, Harada M, Sato T, Nakano H, Mae T**. 1997. Growth and N allocation in rice plants under CO2 enrichment. Plant Physiology **115**, 199–203.

**McCulloch LA, Porder S**. 2021. Light fuels while nitrogen suppresses symbiotic nitrogen fixation hotspots in neotropical canopy gap seedlings. New Phytologist **231**, 1734–1745.

**Medlyn BE, Badeck FW, De Pury DGG, *et al.*** 1999. Effects of elevated [CO2] on photosynthesis in European forest species: A meta-analysis of model parameters. Plant, Cell and Environment **22**, 1475–1495.

**Meyerholt J, Sickel K, Zaehle S**. 2020. Ensemble projections elucidate effects of uncertainty in terrestrial nitrogen limitation on future carbon uptake. Global Change Biology **26**, 3978–3996.

**Moore DJP, Aref S, Ho RM, Pippen JS, Hamilton JG, De Lucia EH**. 2006. Annual basal area increment and growth duration of *Pinus taeda* in response to eight years of free-air carbon dioxide enrichment. Global Change Biology **12**, 1367–1377.

**Nakano H, Makino A, Mae T**. 1997. The effect of elevated partial pressures of CO2 on the relationship between photosynthetic capacity and N content in rice leaves. Plant Physiology **115**, 191–198.

**Nie M, Lu M, Bell J, Raut S, Pendall E**. 2013. Altered root traits due to elevated CO2: A meta-analysis. Global Ecology and Biogeography **22**, 1095–1105.

**Niinemets Ü, Tenhunen JD**. 1997. A model separating leaf structural and physiological effects on carbon gain along light gradients for the shade-tolerant species *Acer saccharum*. Plant, Cell and Environment **20**, 845–866.

**Norby RJ, Warren JM, Iversen CM, Medlyn BE, McMurtrie RE**. 2010. CO2 enhancement of forest productivity constrained by limited nitrogen availability. Proceedings of the National Academy of Sciences **107**, 19368–19373.

**Nowak RS, Ellsworth DS, Smith SD**. 2004. Functional responses of plants to elevated atmospheric CO2 - Do photosynthetic and productivity data from FACE experiments support early predictions? New Phytologist **162**, 253–280.

**Onoda Y, Wright IJ, Evans JR, Hikosaka K, Kitajima K, Niinemets Ü, Poorter H, Tosens T, Westoby M**. 2017. Physiological and structural tradeoffs underlying the leaf economics spectrum. New Phytologist **214**, 1447–1463.

**Pastore MA, Lee TD, Hobbie SE, Reich PB**. 2019. Strong photosynthetic acclimation and enhanced water-use efficiency in grassland functional groups persist over 21 years of CO2 enrichment, independent of nitrogen supply. Global Change Biology **25**, 3031–3044.

**Peng Y, Prentice IC, Bloomfield KJ, Campioli M, Guo Z, Sun Y, Tian D, Wang X, Vicca S, Stocker BD**. 2023. Global terrestrial nitrogen uptake and nitrogen use efficiency. Journal of Ecology, 1–18.

**Perkowski EA, Terrones J, German HL, Smith NG**. 2024. Symbiotic nitrogen fixation reduces belowground biomass carbon costs of nitrogen acquisition under low, but not high, nitrogen availability. AoB PLANTS **16**, 1–22.

**Perkowski EA, Waring EF, Smith NG**. 2021. Root mass carbon costs to acquire nitrogen are determined by nitrogen and light availability in two species with different nitrogen acquisition strategies. Journal of Experimental Botany **72**, 5766–5776.

**Poorter H, Bühler J, Van Dusschoten D, Climent J, Postma JA**. 2012. Pot size matters: A meta-analysis of the effects of rooting volume on plant growth. Functional Plant Biology **39**, 839–850.

**Poorter H, Knopf O, Wright IJ, Temme AA, Hogewoning SW, Graf A, Cernusak LA, Pons TL**. 2022. A meta-analysis of responses of C3 plants to atmospheric CO2: dose–response curves for 85 traits ranging from the molecular to the whole-plant level. New Phytologist **233**, 1560–1596.

**Prentice IC, Dong N, Gleason SM, Maire V, Wright IJ**. 2014. Balancing the costs of carbon gain and water transport: testing a new theoretical framework for plant functional ecology. Ecology Letters **17**, 82–91.

**Prentice IC, Liang X, Medlyn BE, Wang Y-P**. 2015. Reliable, robust and realistic: The three R’s of next-generation land-surface modelling. Atmospheric Chemistry and Physics **15**, 5987–6005.

**R Core Team**. 2021. R: A language and environment for statistical computing. https://www.r-project.org/.

**Rastetter EB, Vitousek PM, Field CB, Shaver GR, Herbert D, Ågren GI**. 2001. Resource optimization and symbiotic nitrogen fixation. Ecosystems **4**, 369–388.

**Reich PB, Hobbie SE, Lee TD, Ellsworth DS, West JB, Tilman D, Knops JMH, Naeem S, Trost J**. 2006. Nitrogen limitation constrains sustainability of ecosystem response to CO2. Nature **440**, 922–925.

**Rogers A, Ainsworth EA, Leakey ADB**. 2009. Will elevated carbon dioxide concentration amplify the benefits of nitrogen fixation in legumes? Plant Physiology **151**, 1009–1016.

**Rogers A, Medlyn BE, Dukes JS, *et al.*** 2017. A roadmap for improving the representation of photosynthesis in Earth system models. New Phytologist **213**, 22–42.

**Saathoff AJ, Welles J**. 2021. Gas exchange measurements in the unsteady state. Plant Cell and Environment **44**, 3509–3523.

**Sage RF**. 1994. Acclimation of photosynthesis to increasing atmospheric CO2: The gas exchange perspective. Photosynthesis Research **39**, 351–368.

**Schneider CA, Rasband WS, Eliceiri KW**. 2012. NIH Image to ImageJ: 25 years of image analysis. Nature Methods **9**, 671–675.

**Sigurdsson BD, Medhurst JL, Wallin G, Eggertsson O, Linder S**. 2013. Growth of mature boreal Norway spruce was not affected by elevated [CO 2] and/or air temperature unless nutrient availability was improved. Tree Physiology **33**, 1192–1205.

**Smith NG, Dukes JS**. 2013. Plant respiration and photosynthesis in global-scale models: incorporating acclimation to temperature and CO2. Global Change Biology **19**, 45–63.

**Smith NG, Keenan TF**. 2020. Mechanisms underlying leaf photosynthetic acclimation to warming and elevated CO2 as inferred from least‐cost optimality theory. Global Change Biology **26**, 5202–5216.

**Smith NG, Keenan TF, Prentice IC, *et al.*** 2019. Global photosynthetic capacity is optimized to the environment. Ecology Letters **22**, 506–517.

**Smith SE, Read DJ**. 2008. *Mycorrhizal Symbiosis*.

**Smith NG, Zhu Q, Keenan TF, Riley WJ**. 2024. Acclimation of photosynthesis to CO2 increases ecosystem carbon storage due to leaf nitrogen savings. Global Change Biology **30**, 1–10.

**Stocker BD, Dong N, Perkowski EA, *et al.*** 2025. Empirical evidence and theoretical understanding of ecosystem carbon and nitrogen cycle interactions. New Phytologist **245**, 49–68.

**Taylor BN, Menge DNL**. 2018. Light regulates tropical symbiotic nitrogen fixation more strongly than soil nitrogen. Nature Plants **4**, 655–661.

**Tejera-Nieves M, Seong DY, Reist L, Walker BJ**. 2024. The Dynamic Assimilation Technique measures photosynthetic CO2 response curves with similar fidelity to steady-state approaches in half the time. Journal of Experimental Botany **75**, 2819–2828.

**Terrer C, Vicca S, Hungate BA, Phillips RP, Prentice IC**. 2016. Mycorrhizal association as a primary control of the CO2 fertilization effect. Science **353**, 72–74.

**Terrer C, Vicca S, Stocker BD, Hungate BA, Phillips RP, Reich PB, Finzi AC, Prentice IC**. 2018. Ecosystem responses to elevated CO2 governed by plant–soil interactions and the cost of nitrogen acquisition. New Phytologist **217**, 507–522.

**Vitousek PM, Howarth RW**. 1991. Nitrogen limitation on land and in the sea: How can it occur? Biogeochemistry **13**, 87–115.

**Walker AP, Beckerman AP, Gu L, Kattge J, Cernusak LA, Domingues TF, Scales JC, Wohlfahrt G, Wullschleger SD, Woodward FI**. 2014. The relationship of leaf photosynthetic traits - Vcmax and Jmax - to leaf nitrogen, leaf phosphorus, and specific leaf area: a meta-analysis and modeling study. Ecology and Evolution **4**, 3218–3235.

**Wang H, Prentice IC, Keenan TF, Davis TW, Wright IJ, Cornwell WK, Evans BJ, Peng C**. 2017. Towards a universal model for carbon dioxide uptake by plants. Nature Plants **3**, 734–741.

**Waring EF, Perkowski EA, Smith NG**. 2023. Soil nitrogen fertilization reduces relative leaf nitrogen allocation to photosynthesis. Journal of Experimental Botany **74**, 5166–5180.

**Wellburn AR**. 1994. The spectral determination of chlorophylls a and b, as well as total carotenoids, using various solvents with spectrophotometers of different resolution. Journal of Plant Physiology **144**, 307–313.

**Wieder WR, Cleveland CC, Smith WK, Todd-Brown K**. 2015. Future productivity and carbon storage limited by terrestrial nutrient availability. Nature Geoscience **8**, 441–444.

**Wright IJ, Reich PB, Westoby M**. 2003. Least-cost input mixtures of water and nitrogen for photosynthesis. The American Naturalist **161**, 98–111.

**Zavala JA, Nabity PD, DeLucia EH**. 2013. An Emerging Understanding of Mechanisms Governing Insect Herbivory Under Elevated CO2. Annual Review of Entomology **58**, 79–97.

**Table 1** Effects of CO2 concentration, inoculation, and nitrogen fertilization on area-based leaf nitrogen content and chlorophyll content\*

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
|  |  | ***N*area** | | ***Chl*area** | |
|  | df | *χ*2 | *p* | *χ*2 | *p* |
| CO2 | 1 | 155.908 | **<0.001** | 62.056 | **<0.001** |
| Inoculation (I) | 1 | 86.029 | **<0.001** | 133.828 | **<0.001** |
| N fertilization (N) | 1 | 316.408 | **<0.001** | 156.659 | **<0.001** |
| CO2 × I | 1 | 4.729 | **0.030** | 1.647 | 0.199 |
| CO2 × N | 1 | 5.723 | **0.017** | 2.780 | 0.095 |
| I × N | 1 | 43.381 | **<0.001** | 73.494 | **<0.001** |
| CO2 × I × N | 1 | 0.489 | 0.484 | 2.123 | 0.145 |

\*Significance determined using Type II Wald χ2 tests (α=0.05). *P*-values less than 0.05 are in bold. Key: df=degrees of freedom, *χ*2=Wald chi-square test statistic, *N*area=leaf nitrogen content per unit leaf area (gN m-2), *Chl*area=chlorophyll content per unit leaf area (mmol m-2)

**Table 2** Effects of CO2 concentration, inoculation, and nitrogen fertilization on leaf gas exchange\*

|  |  |  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
|  |  | ***A*net,420** | | ***A*net,gc** | | ***V*cmax25** | | ***J*max25** | | ***J*max25:*V*cmax25** | |
|  | df | *χ*2 | *p* | *χ*2 | *p* | *χ*2 | *p* | *χ*2 | *p* | *χ*2 | *p* |
| CO2 | 1 | 15.747 | **<0.001** | 52.716 | **<0.001** | 18.039 | **<0.001** | 6.042 | **0.014** | 92.010 | **<0.001** |
| Inoculation (I) | 1 | 77.137 | **<0.001** | 83.008 | **<0.001** | 98.579 | **<0.001** | 85.064 | **<0.001** | 27.768 | **<0.001** |
| N fertilization (N) | 1 | 11.986 | **<0.001** | 14.658 | **<0.001** | 37.053 | **<0.001** | 25.356 | **<0.001** | 28.147 | **<0.001** |
| CO2 × I | 1 | 1.032 | 0.310 | 5.634 | **0.018** | 0.065 | 0.799 | 0.667 | 0.414 | 2.916 | 0.088 |
| CO2 × N | 1 | 1.998 | 0.158 | 0.135 | 0.713 | 1.758 | 0.185 | 0.742 | 0.389 | 3.210 | 0.073 |
| I × N | 1 | 46.800 | **<0.001** | 50.774 | **<0.001** | 60.394 | **<0.001** | 57.41 | **<0.001** | 9.607 | **0.002** |
| CO2 × I × N | 1 | 0.002 | 0.964 | 1.332 | 0.248 | 0.748 | 0.387 | 0.377 | 0.539 | 1.102 | 0.294 |

\*Significance determined using Type II Wald χ2 tests (α=0.05). *P*-values less than 0.05 are in bold. Key: df=degrees of freedom, *χ*2=Wald chi-square test statistic, *A*net,420=net photosynthesis rate at 420 μmol mol-1 CO2 (μmol m-2 s-1), *A*net,gc=net photosynthesis rate at under growth CO2 condition (μmol m-2 s-1), *V*cmax25=apparent maximum rate of Rubisco carboxylation at 25°C (μmol m-2 s-1), *J*max25=apparent maximum rate of electron transport for RuBP regeneration at 25°C (μmol m-2 s-1), *J*max25:*V*cmax25=ratio of *J*max25 to *V*cmax25 (unitless)

**Table 3** Effects of CO2 concentration, inoculation, and nitrogen fertilization on total leaf area, total biomass, carbon costs to acquire nitrogen, and plant investment toward symbiotic nitrogen fixation\*

|  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- |
|  |  | **Total leaf area** | | **Total biomassb** | | **Root:shoot ratioa** | |
|  | df | *χ*2 | *p* | *χ*2 | *p* | *χ*2 | *p* |
| CO2 | 1 | 69.291 | **<0.001** | 131.477 | **<0.001** | 4.892 | **0.027** |
| Inoculation (I) | 1 | 35.715 | **<0.001** | 34.264 | **<0.001** | 9.790 | **0.002** |
| N fertilization (N) | 1 | 274.199 | **<0.001** | 269.046 | **<0.001** | 50.742 | **<0.001** |
| CO2 × I | 1 | 2.064 | 0.151 | 0.518 | 0.472 | 10.467 | **0.001** |
| CO2 × N | 1 | 18.655 | **<0.001** | 16.877 | **<0.001** | 0.012 | 0.914 |
| I × N | 1 | 10.804 | **0.001** | 15.779 | **<0.001** | 3.802 | *0.051* |
| CO2 × I × N | 1 | <0.001 | 0.990 | 0.023 | 0.880 | 0.417 | 0.519 |
|  |  |  |  |  |  |  |  |
|  |  | **Carbon cost to**  **acquire nitrogena** | | **Nodule biomass:**  **root biomass** | |  |  |
|  |  | *χ*2 | *p* | *χ*2 | *p* |  |  |
| CO2 |  | 76.462 | **<0.001** | 0.010 | 0.921 |  |  |
| Inoculation (I) |  | 70.846 | **<0.001** | 902.063 | **<0.001** |  |  |
| N fertilization (N) |  | 74.961 | **<0.001** | 254.741 | **<0.001** |  |  |
| CO2 × I |  | 33.329 | **<0.001** | 21.632 | **<0.001** |  |  |
| CO2 × N |  | 1.889 | 0.169 | 1.590 | 0.207 |  |  |
| I × N |  | 26.719 | **<0.001** | 132.463 | **<0.001** |  |  |
| CO2 × I × N |  | 6.860 | **0.009** | 2.481 | 0.115 |  |  |

\*Significance determined using Type II Wald χ2 tests (*α*=0.05). *P*-values less than 0.05 are in bold and *p*-values where 0.05<*p*<0.1 are in italic font. Key: a=variable was natural log transformed before model fitting, b=variable was square root transformed before model fitting, df=degrees of freedom, *χ*2=Wald chi-square test statistic, total leaf area (cm2), total biomass (g), the ratio of root biomass to shoot biomass (unitless), belowground biomass carbon cost to acquire nitrogen (gC gN-1), the ratio of root nodule biomass to root biomass (unitless)

**Figure Legends**

**Figure 1** Effects of CO2 concentration, inoculation, and nitrogen fertilization on leaf nitrogen per unit leaf area (a) and chlorophyll content per unit leaf area (b). Nitrogen fertilization is on the x-axis in both panels. Red shaded points and trendlines indicate plants grown under elevated CO2, while blue shaded points and trendlines indicate plants grown under ambient CO2. Light blue and light red circular points and trendlines indicate measurements collected from uninoculated plants, while dark blue and dark red triangular points indicate measurements collected from inoculated plants. Solid trendlines indicate regression slopes that are different from zero (*p*<0.05), while dashed trendlines indicate slopes that are not distinguishable from zero (*p*>0.05). Error ribbons of each trendline represent the upper and lower 95% confidence intervals.

**Figure 2** Effects of CO2, inoculation, and nitrogen fertilization on net photosynthesis measured under growth CO2 concentration (a), the apparent maximum rate of Rubisco carboxylation at 25°C (b), the apparent maximum rate of electron transport for RuBP regeneration at 25°C (c), and the ratio of the apparent maximum rate of electron transport for RuBP regeneration to the apparent maximum rate of Rubisco carboxylation (d). Nitrogen fertilization is on the x-axis in all panels. Red shaded points and trendlines indicate plants grown under elevated CO2, while blue shaded points and trendlines indicate plants grown under ambient CO2. Light blue and light red circular points and trendlines indicate measurements collected from uninoculated plants, while dark blue and dark red triangular points indicate measurements collected from inoculated plants. Solid trendlines indicate regression slopes that are different from zero (*p*<0.05), while dashed trendlines indicate slopes that are not distinguishable from zero (*p*>0.05). Error ribbons of each trendline represent the upper and lower 95% confidence intervals.

**Figure 3** Effects of CO2, nitrogen fertilization, and inoculation on total leaf area (a), total biomass (b), the ratio of root biomass to shoot biomass (c), and belowground carbon cost to acquire nitrogen (d). Nitrogen fertilization is on the x-axis in all panels. Red shaded points and trendlines indicate plants grown under elevated CO2, while blue shaded points and trendlines indicate plants grown under ambient CO2. Light blue and light red circular points and trendlines indicate measurements collected from uninoculated plants, while dark blue and dark red triangular points indicate measurements collected from inoculated plants. Solid trendlines indicate regression slopes that are different from zero (*p*<0.05), while dashed trendlines indicate slopes that are not distinguishable from zero (*p*>0.05). Error ribbons of each trendline represent the upper and lower 95% confidence intervals.

**Figure 4** Effects of CO2, nitrogen fertilization, and inoculation on the ratio of root nodule biomass to root biomass. Nitrogen fertilization is on the x-axis. Red shaded points and trendlines indicate plants grown under elevated CO2, while blue shaded points and trendlines indicate plants grown under ambient CO2. Light blue and light red circular points and trendlines indicate measurements collected from uninoculated plants, while dark blue and dark red triangular points indicate measurements collected from inoculated plants. Solid trendlines indicate regression slopes that are different from zero (*p*<0.05), while dashed trendlines indicate slopes that are not distinguishable from zero (*p*>0.05). Error ribbons of each trendline represent the upper and lower 95% confidence intervals.