**Title**: Symbiotic nitrogen fixation reduces carbon costs of nitrogen acquisition under low, but not high, nitrogen availability

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**Abstract**

Many plant species form symbiotic associations with nitrogen fixing bacteria. Through this symbiosis, plants give photosynthate to the bacteria in exchange for nitrogen fixed from the atmosphere. This symbiosis forms an important link between carbon and nitrogen cycles in many ecosystems. However, the economics of this relationship under different background soil nitrogen availabilities are not well understood. Here, we used a manipulation experiment to examine how the costs of nitrogen acquisition vary under a factorial combination of soil nitrogen availability and nitrogen fixing bacteria inoculation in *Glycine max* L. (Merr.). After a 7-week growth period, we measured root, stem, leaf, and nodule biomass as well as carbon and nitrogen amounts of each organ. We used this information to assess structural carbon costs to acquire nitrogen, plant investments to nitrogen fixation, leaf nitrogen allocation, and whole plant growth. We found that structural carbon costs to acquire nitrogen decreased with inoculation in the low soil nitrogen availability treatment, but were unaffected by inoculation in the high soil nitrogen fertilization treatment. The treatment differences were the result of greater plant nitrogen, rather than any change in belowground carbon allocation. These results suggest that symbioses with nitrogen fixing bacteria reduce carbon costs of nitrogen acquisition, but only when soil nitrogen is low. This helps to explain the prevalence of plants capable of forming these associations in less fertile areas and demonstrates patterns that can help guide models linking carbon and nitrogen cycles in terrestrial ecosystems.

**Keywords**

nitrogen fixation; whole plant growth; greenhouse; crops; nutrient acquisition strategy

**Introduction**

Terrestrial ecosystem processes are regulated, in part, by interactions between carbon and nitrogen cycles. As a result, terrestrial biosphere models are beginning to include coupled carbon and nitrogen cycles to more realistically simulate past, present, and future atmosphere-biosphere fluxes (Oreskes *et al.*, 1994; Hungate *et al.*, 2003; Prentice *et al.*, 2015). Carbon and nutrient flux simulations tend to converge across terrestrial biosphere model products using past and present climate scenarios; however, often diverge under future environmental change scenarios (Friedlingstein *et al.*, 2014; Davies-Barnard *et al.*, 2020). This divergence could be due to an incomplete understanding of how changing environments modify processes that link ecosystem carbon and nitrogen cycles (Wieder *et al.*, 2015; Fay *et al.*, 2015; Meyerholt *et al.*, 2016).

Plant nitrogen acquisition is one process in terrestrial ecosystems that links carbon and nitrogen cycles. Plants acquire nutrients by allocating photosynthetically derived carbon belowground in exchange for nitrogen through different nitrogen acquisition strategies. These nitrogen acquisition strategies can include direct uptake pathways such as mass flow or diffusion (Barber, 1962), symbioses with mycorrhizal fungi or symbiotic nitrogen-fixing bacteria (Vance & Heichel, 1991; Marschner & Dell, 1994; Smith & Read, 2008; Udvardi & Poole, 2013), or root exudates that supply carbon to free-living soil microbial communities (Phillips *et al.*, 2011; Wen *et al.*, 2022).

In principle, plants cannot acquire nitrogen without first allocating carbon belowground, which implies an inherent carbon cost to the plant for acquiring nitrogen. These nitrogen return on carbon invested belowground may vary in species with different nitrogen acquisition strategies. For instance, carbon investment in roots for direct nitrogen uptake does not require costs beyond root development, as is the cases for acquisition strategies that involve other organisms. However, the nitrogen return from a given belowground carbon investment may be greater if carbon is given to decomposers who produce inorganic nitrogen than can be taken up by roots (CITE), fungal symbionts that mine the soil for nitrogen (CITE), or bacteria symbionts that can provide nitrogen fixed from the atmosphere (CITE). The variability in costs to acquire nitrogen may help to explain the prevalence of different nitrogen acquisition strategies in different environments, but these have not been well quantified outside of a few studies (Terrer *et al.*, 2018; Lu *et al.*, 2022)

Nitrogen acquisition costs for a given nitrogen acquisition strategy is likely dependent on external environmental factors such as atmospheric CO2, light availability, and soil nutrient availability (Brzostek *et al.*, 2014; Terrer *et al.*, 2018; Allen *et al.*, 2020; Perkowski *et al.*, 2021; Lu *et al.*, 2022). For instance, the amount of photosynthate allocated belowground in exchange for nitrogen may increase with increased light and CO2, as these factors reduce the cost to produce photosynthate (Perkowski et al., 2021, Terrer et al., 2018, OTHERS??). However, soil nitrogen availability is likely to reduce costs for nitrogen acquisition due a reduction in soil resourcing mining (by roots or symbionts) needed to meet plant nitrogen demand. However, this may not play out in plant species with strong and specialized symbiotic relationships with nitrogen-acquiring partners, such as plants that associate with nitrogen fixing bacteria.

In a recent study, Perkowski *et al.* (2021) show that increasing soil nitrogen fertilization decreased carbon costs to acquire nitrogen in *Gossypium hirsutum* and *Glycine max*. *Gossypium hirsutum* can acquire nutrients via direct uptake pathways or through symbioses with arbuscular mycorrhizal fungi, while *G. max* can acquire nutrients via direct uptake pathways or through symbioses with nitrogen-fixing bacteria. In the experiment, the authors noted that carbon costs to acquire nitrogen in *G. max* were generally less responsive to increasing soil nitrogen fertilization than *G. hirsutum*, a pattern that coincided with a reduction in *G. max* root nodulation with increasing fertilization. The authors speculated that this response may have been driven by resource optimization, where *G. max* shifted their dominant mode of nitrogen acquisition from nitrogen fixation to direct uptake with increasing fertilization once costs to acquire nitrogen via direct uptake became less than the costs to acquire nitrogen via nitrogen fixation (Rastetter *et al.*, 2001). However, the authors were not able to make robust conclusions about whether the carbon cost to acquire nitrogen responses to soil nitrogen fertilization differed between *G. hirsutum* and *G. max* due to differences in species nutrient acquisition strategy because the two species are not phylogenetically related and adopt different growth forms and growth durations.

To better understand how nitrogen fixation and soil nitrogen fertilization interact to influence carbon costs to acquire nitrogen, we grew *Glycine max* L. (Merr.) under two soil nitrogen fertilization treatments and two inoculation treatments in a full factorial greenhouse experiment. We used this experiment to test the following hypotheses:

1. Soil nitrogen fertilization will decrease carbon costs of nitrogen acquisition in both uninoculated and inoculated individuals. This will manifest as an increase the amount of nitrogen acquired per belowground carbon investment, or a stronger increase in plant nitrogen uptake than belowground carbon allocation.
2. Inoculation with nitrogen-fixing bacteria will decrease carbon costs to acquire nitrogen under low soil nitrogen availability, as carbon costs to acquire nitrogen through symbiotic nitrogen fixation will be less than the carbon cost to acquire nitrogen via direct uptake. However, there will be no effect of inoculation under high soil nitrogen availability due to all plants shifting toward a similar, direct uptake-dominated mode of nitrogen acquisition.
3. There will be a decrease in nodulation with increasing soil nitrogen availability due to a reduction in carbon costs to obtain nitrogen from direct uptake with increasing soil nitrogen fertilization.

**Methods**

*Experimental Design*

*Glycine max* seeds were planted in 64, 6-liter pots (NS-600, Nursery Supplies, Orange, CA, USA) containing unfertilized potting mix (Sungro Sunshine Mix #2, Agawam, MA, USA). Pots and potting mix were steam sterilized at 95C for three hours to eliminate any bacterial or fungal growth. Thirty-two randomly selected pots were planted with seeds inoculated with *Bradyrhizobium japonicum* (Verdesian N-Dure™ Soybean, Cary, NC, USA) following a brief surface sterilization in 20,000 ppm sodium hypochlorite for 5 minutes followed by three washes in ultrapure water (Scouten & Beuchat, 2002; Montville & Schaffner, 2004). The remaining 32 pots were planted with seeds that did not receive any inoculation treatment. Uninoculated seeds were also surface sterilized in 20,000 ppm sodium hypochlorite for 5 minutes followed by three ultrapure water washes to ensure that the only difference between seed treatments was the inoculation treatment.

Upon planting, all pots were immediately placed in one of four random blocks in a greenhouse and received one of two nitrogen fertilization treatments as 150 mL of a modified Hoagland’s solution (Hoagland & Arnon, 1950) equivalent to either 70 or 630 ppm N twice per week for seven weeks. Nitrogen fertilization doses were received as topical agents to the soil surface and were modified to keep concentrations of other macronutrients and micronutrients equivalent (Table S1). Throughout the experiment, plants were routinely well-watered to minimize any chance of water stress. There was no evidence of pot size induced growth limitation at the time of biomass harvest, indicated by total biomass: pot volume ratios less than 1 g L-1 within each treatment combination (Table S2; Fig. S1; Poorter *et al.*, 2012)

*Plant trait measurements*

All experimental individuals were harvested and biomass was separated into major organ types (leaves, stems, roots, and root nodules when present) approximately seven weeks after experiment initiation. Leaf areas of all harvested leaves were measured using an LI-3100C (Li-COR Biosciences, Lincoln, Nebraska, USA). Total leaf area (cm2) was calculated as the sum of all leaf areas. All harvested material was dried in an oven set to 65°C for at least 48 hours, weighed, and ground to homogeneity. Total dry biomass (g) was calculated as the sum of dry leaf, stem, root, and root nodule biomass. Carbon and nitrogen content of each respective organ was quantified through elemental combustion (Costech-4010, Costech, Inc., Valencia, CA, USA) using subsamples of ground and homogenized organ tissue.

Following the approach explained in Perkowski *et al.* (2021), we calculated structural carbon costs to acquire nitrogen as the ratio of total belowground carbon biomass to whole plant nitrogen biomass (g C g-1 N). Belowground carbon biomass (g C) was calculated as the sum of total root carbon biomass and total root nodule carbon biomass. Total root carbon biomass was calculated by multiplying the carbon content of roots by total root biomass, while total root nodule carbon biomass was calculated by multiplying the carbon content of root nodules by total root nodule biomass. Whole-plant nitrogen biomass (g N) was calculated by multiplying the nitrogen content of leaves, stems, roots, and root nodules by biomass of each respective organ type, then calculating the sum of nitrogen biomass of each organ type. This calculation only quantifies plant structural carbon costs to acquire nitrogen and does not include additional carbon costs of nitrogen acquisition associated with root respiration, root exudation, or root turnover. An explicit explanation of the limitations for interpreting this calculation can be found in Perkowski *et al.* (2021) and Terrer *et al.* (2018).

*Statistical analyses*

We built a series of linear mixed-effects models to investigate the impacts of soil nitrogen fertilization and inoculation on *G. max* traits. All models included soil nitrogen fertilization, inoculation, and interactions between soil nitrogen fertilization and inoculation as categorical fixed effects. Block number was included as a random intercept term to account for any environmental heterogeneity within the greenhouse room. Models with this independent variable structure were constructed to quantify relationships between soil nitrogen fertilization and inoculation on structural carbon costs to acquire nitrogen, belowground carbon biomass, whole plant nitrogen biomass, total biomass, total leaf area, root nodule biomass: root biomass, root nodule biomass, and root biomass.

We used Shapiro-Wilk tests of normality to determine whether linear mixed-effects models satisfied residual normality assumptions. Whole-plant nitrogen biomass and total leaf area were the only models that satisfied residual normality assumptions. We attempted to satisfy residuals. We attempted to satisfy residual normality assumptions by fitting the other models using dependent variables that were natural log transformed. If residual normality assumptions were still not met after a natural-log transformation (Shapiro-Wilk: p<0.05), then models were fit using dependent variables that were square root transformed. All residual normality assumptions were met with either a natural log or square root data transformation (Shapiro-Wilk: p>0.05 in all cases). Specifically, models for structural carbon costs to acquire nitrogen, belowground carbon biomass, total biomass, root biomass, and biomass: pot volume satisfied normality assumptions when fit using natural log transformed data, while the models for nodule biomass: root biomass and root nodule biomass were satisfied when fit using square-root transformations.

In all statistical models, we used the 'lmer' function in the 'lme4' R package (Bates *et al.*, 2015) to fit each model and the 'Anova' function in the 'car' R package (Fox & Weisberg, 2019) to calculate Type II Wald's χ2 and determine the significance (α=0.05) of each fixed effect coefficient. We then used the 'emmeans' R package (Lenth, 2019) to conduct post-hoc comparisons using Tukey's tests, where degrees of freedom were approximated using the Kenward-Roger approach (Kenward & Roger, 1997) All analyses and plots were conducted in R version 4.2.0 (R Core Team, 2021).

**Results**

*Structural carbon costs to acquire nitrogen*

An interaction between nitrogen fertilization and inoculation (*p*<0.05;Table 1) indicated that negative effects of inoculation (*p*<0.001; Table 1) on structural carbon costs to acquire nitrogen were only apparent under low soil nitrogen fertilization (Tukey: *p*<0.001), as there was no inoculation effect on structural carbon costs to acquire nitrogen under high soil nitrogen fertilization (Tukey: *p*>0.05; Fig. 1A). Structural carbon costs to acquire nitrogen also generally decreased with increasing soil nitrogen fertilization (*p*<0.001; Table 1; Fig. 1A)*.* Structural carbon costs to acquire nitrogen responses to inoculation and fertilization treatments were more strongly driven by treatment impacts on whole-plant nitrogen biomass than belowground carbon biomass.

Inoculation decreased belowground carbon biomass (*p*<0.05; Table 1), a response that was not modified by soil nitrogen fertilization (inoculation-by-fertilization interaction: *p*>0.05; Table 1; Fig. 1B). There was no effect of soil nitrogen fertilization on belowground carbon biomass (*p*>0.05; Table 1).

An interaction between soil nitrogen fertilization and inoculation (*p*<0.001; Table 1) indicated that positive effects of inoculation on whole-plant nitrogen biomass (*p*<0.001; Table 1) were only apparent under low soil nitrogen fertilization (Tukey: *p*<0.001), as there was no effect of inoculation on whole-plant nitrogen biomass under high soil nitrogen fertilization (Tukey: *p*>0.05; Fig. 1C). Whole-plant nitrogen biomass generally increased with increasing soil nitrogen fertilization (*p*<0.001; Table 1).

**Table 1** Analysis of variance results exploring effect of soil nitrogen fertilization, inoculation with *B. japonicum*, and interactions between soil nitrogen fertilization and inoculation on structural carbon costs to acquire nitrogen, whole plant growth, and root nodulation\*

|  |  |  |  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- | --- |
|  | | **Carbon cost to acquire nitrogen** | | **Belowground carbon biomass** | | **Whole-plant nitrogen biomass** | | **Total**  **leaf area** | | **Whole plant biomass** | |
|  | df | χ2 | *p* | χ2 | *p* | χ2 | *p* | χ2 | *p* | χ2 | *p* |
| N fertilization (N) | 1 | 23.340 | **<0.001** | 0.076 | 0.782 | 358.695 | **<0.001** | 292.458 | **<0.001** | 52.427 | **<0.001** |
| Inoculation (I) | 1 | 16.749 | **<0.001** | 4.166 | **0.041** | 24.113 | **<0.001** | 35.095 | **<0.001** | 2.042 | 0.153 |
| N\*I | 1 | 4.833 | **0.028** | 0.265 | 0.607 | 13.515 | **<0.001** | 17.898 | **<0.001** | 1.230 | 0.267 |
|  |  |  |  |  |  |  |  |  |  |  |  |
|  |  | **Nodule biomass: root biomass** | | **Nodule**  **biomass** | | **Root**  **biomass** | |  | |  | |
|  | df | χ2 | *p* | χ2 | *p* | χ2 | *p* |  |  |  |  |
| N fertilization (N) | 1 | 1.291 | 0.256 | 1.364 | 0.243 | 0.011 | 0.918 |  |  |  |  |
| Inoculation (I) | 1 | 27.375 | **<0.001** | 30.788 | **<0.001** | 3.268 | *0.071* |  |  |  |  |
| N\*I | 1 | 0.493 | 0.483 | 1.005 | 0.316 | 0.254 | 0.614 |  |  |  |  |

\*Significance determined using Type II Wald χ2 tests (α=0.05). *P*-values less than 0.05 are in bold and *P*-values between 0.05 and 0.1 are italicized.

**Figure 1**

**A graph of different types of fertilizers

Description automatically generated**

**Figure 1** Effects of soil nitrogen fertilization and inoculation on *G. max* structural carbon costs to acquire nitrogen (“*N*cost”; panel A), belowground carbon biomass (“*C*bg”; panel B), and whole-plant nitrogen biomass (“*N*wp”; panel C). Soil nitrogen fertilization is represented on the x-axis, while inoculation treatment is represented by colored boxplots. Yellow shaded boxplots indicate individuals that were not inoculated with *B. japonicum*, while red shaded boxplots indicate individuals that were inoculated with *B. japonicum*. Boxes are the upper (75% percentile) and lower (25% percentile) quartile. The whiskers are the minimum and maximum value, calculated as 1.5 times the upper and lower quartile value. Colored dots are individual data points, jittered for visibility. The lettering above each box indicates the results from post-hoc Tukey’s tests with different lettering indicating statistically different groups (Tukey: *p*<0.05).

*Whole plant growth and plant investments to nitrogen fixation*

An interaction between nitrogen fertilization and inoculation (*p*<0.001; Table 1) indicated that positive effects of inoculation on total leaf area (*p*<0.001; Table 1) were only apparent under low nitrogen fertilization (Tukey: *p*<0.001), as there was no inoculation effect on total leaf area under high nitrogen fertilization (Tukey: *p*>0.05; Fig. 2A). Increasing soil nitrogen fertilization increased total leaf area (*p*<0.001; Table 1; Fig. 2A).

Increasing nitrogen fertilization increased total biomass (*p*<0.001; Table 1; Fig. 2B), a pattern that was not modified by inoculation (inoculation-by-fertilization interaction: *p*>0.05; Table 1). Inoculation had no effect on total biomass (*p*>0.05; Table 1; Fig. 2B).

**Figure 2**

**A graph of different stages of fertilization

Description automatically generated**

**Figure 2** Effects of soil nitrogen fertilization and inoculation on *G. max* total leaf area (panel A), total biomass (panel B). Soil nitrogen fertilization is represented on the x-axis, while inoculation treatment is represented by colored boxplots. Yellow shaded boxplots indicate individuals that were not inoculated with *B. japonicum*, while red shaded boxplots indicate individuals that were inoculated with *B. japonicum*. Boxes are the upper (75% percentile) and lower (25% percentile) quartile. The whiskers are the minimum and maximum value, calculated as 1.5 times the upper and lower quartile value. Colored dots are individual data points, jittered for visibility. The lettering above each box indicates the results from post-hoc Tukey’s tests with different lettering indicating statistically different groups (Tukey: *p*<0.05).

*Plant investment in symbiotic nitrogen fixation*

Inoculation increased root nodule biomass: root biomass (*p*<0.001; Table 1; Fig 3A), a pattern that was not modified by fertilization (inoculation-by-fertilization interaction: *p*>0.05; Table 1). There was no effect of soil nitrogen fertilization on root nodule biomass: root biomass (*p*>0.05; Table 1; Fig 3A).

Inoculation increased root nodule biomass (*p*<0.001; Table 1; Fig 3B), a pattern that was not modified by fertilization (inoculation-by-fertilization interaction: *p*>0.05; Table 1). There was also no effect of soil nitrogen fertilization on root nodule biomass (*p*>0.05; Table 1; Fig. 3B).

Inoculation had a marginal negative effect on root biomass (*p*<0.1; Table 1; Fig. 3C), a pattern that was not modified by fertilization (inoculation-by-fertilization interaction: *p*>0.05; Table 1). Fertilization had no effect on root biomass (*p*>0.05; Table 1; Fig. 3C).

**Figure 3**

**A diagram of different types of plants

Description automatically generated with medium confidence**

**Figure 3** Effects of soil nitrogen fertilization and inoculation on *G. max* nodule biomass: root biomass (panel A), nodule biomass (panel B), and root biomass (panel C). Soil nitrogen fertilization is represented on the x-axis, while inoculation treatment is represented by colored boxplots. Yellow shaded boxplots indicate individuals that were not inoculated with *B. japonicum*, while red shaded boxplots indicate individuals that were inoculated with *B. japonicum*. Boxes are the upper (75% percentile) and lower (25% percentile) quartile. The whiskers are the minimum and maximum value, calculated as 1.5 times the upper and lower quartile value. Colored dots are individual data points, jittered for visibility. The lettering above each box indicates the results from post-hoc Tukey’s tests with different lettering indicating statistically different groups (Tukey: *p*<0.05).

**Discussion**

Here, we quantified the interactive effect of soil nitrogen fertilization and inoculation by symbiotic nitrogen-fixing bacteria on *G. max* structural carbon costs to acquire nitrogen using a fully factorial manipulation experiment. We found that inoculation reduced carbon costs to acquire nitrogen under low, but not high, levels of soil nitrogen fertilization. This pattern was observed despite no significant differences in belowground root investment across the treatments. Instead, this pattern was driven by greater whole-plant nitrogen uptake in inoculated plants than uninoculated plants when soil nitrogen fertilization was low. This suggests that symbioses with nitrogen-fixing bacteria reduce nitrogen acquisition costs under low soil nitrogen availability, allowing plants to increase nitrogen uptake efficiency compared to individuals restricted to direct uptake methods. That said, structural carbon costs to acquire nitrogen were the lowest under high soil nitrogen availability as a result of high amounts of plant nitrogen uptake. Interestingly, investment in root nodules did not vary with soil nitrogen fertilization, though variability in root nodule investment was greater under low soil nitrogen availability. We expand on and contextualize these key results below.

*The impact of inoculation on plant carbon costs to acquire nitrogen depend on soil nitrogen availability*

Our results provide direct evidence that symbioses with nitrogen-fixing bacteria reduce carbon costs to acquire nitrogen when soil nitrogen availability is low. This corroborates results from past theory (Vitousek *et al.*, 2002) and cross-species experimental studies (Perkowski *et al.*, 2021). Here, we used individuals of the same species to confirm that symbioses with nitrogen-fixing bacteria are the primary driver of this response.

Despite a large inoculation effect on carbon costs to acquire nitrogen at low soil nitrogen availability, there was no impact (positive or negative) of inoculation at high levels of soil nitrogen availability. Similar results were shown in a previous cross-species study that found that plants with and without symbioses with nitrogen fixing bacteria had more similar carbon costs to acquire nitrogen when soil nitrogen availability was high, compared to that when it was low (Perkowski *et al.*, 2021). The difference may help to explain the greater prevalence of plants capable of symbiotic nitrogen fixation where soil nitrogen availability is low (e.g., Monks *et al.*, 2012), as expected from theory (Vitousek & Field, 1999; Vitousek *et al.*, 2002; Menge *et al.*, 2008).

Our results indicate that symbiotic nitrogen fixation may provide a competitive advantage in nitrogen-poor soils by reducing plant carbon costs for acquiring nitrogen. However, the longer-term outcomes of this advantage are difficult to predict because nitrogen fixation brings in nitrogen to the ecosystem that may alleviate nitrogen limitation in non-fixing species following recycling. Additionally, other bottom-up (e.g., soil resources) and top-down (e.g., herbivory) may limit the competitive ability of species that associate with symbiotic nitrogen-fixing bacteria in terrestrial ecosystems (Eisele *et al.*, 1989; Ritchie *et al.*, 1998; Vitousek & Field, 1999; Rastetter *et al.*, 2001; Vitousek *et al.*, 2002, 2013). Longer term field and mesocosm experiments (e.g., Finzi and Rodgers, 2009), coupled with targeted model experiments (e.g., Brzostek et al., 2014) could help to clarify the role of different drivers.

*Soil nitrogen availability and inoculation modify whole plant nitrogen, but not belowground structural carbon*

Reductions in carbon costs to acquire nitrogen with both increasing soil nitrogen fertilization and inoculation under low soil nitrogen were the result of increased plant nitrogen uptake, but not due to a change in belowground investment in structural carbon. In fact, belowground structural carbon was not impacted by any of our treatments.

The increase in nitrogen uptake in our study was predominantly used to support aboveground tissue, which showed large stimulation under increasing soil nitrogen availability and with inoculation when soil nitrogen was low. Theory suggests that increasing nitrogen availability (from soil or symbionts) should increase relative plant investment in aboveground tissues (Ågren & Franklin, 2003), as we observed here. And, indeed, meta-analyses find consistent positive stimulation of aboveground biomass with increasing soil nitrogen availability, but inconsistent impacts on belowground biomass (Li *et al.*, 2020).

Our findings provide a possible benchmark for models that use carbon costs of nitrogen acquisition to simulate terrestrial carbon-nitrogen dynamics (e.g., Brzostek et al., 2014). From our results, changes in these costs should be the result of differences in the amount of nitrogen acquired by plants rather than changes in belowground carbon costs. However, it must be noted that we were not able to capture additional carbon costs that resulted from differences in root exudation or respiration under our different treatments. It is unclear whether these are proportional to structural belowground carbon costs and future studies should be performed to validate this assumption.

*Soil nitrogen fertilization does not significantly reduce plant investment in nitrogen fixing bacteria symbiosis*

We found that inoculated plants had similar levels of nodulation under both of our soil nitrogen availability treatments. This indicates that, in this study, the level of nitrogen availability did not impact the strength of the symbiosis between *G. max* and *B. japonicum*. This result was counter to expectation that greater soil nitrogen availability would reduce plant reliance on nitrogen fixing symbionts (Vitousek *et al.*, 2002; Perkowski *et al.*, 2021). The results may imply stronger bacterial control over the symbiosis than previously thought. In fact, if there were greater amounts of unquantified plant carbon going to bacterial respiration, we may have actually observed higher carbon costs for nitrogen acquisition in inoculated plants under high soil nitrogen. Carbon and nitrogen tracing experiments would be useful for further examining this result.

*Study limitations*

This study has a few limitations that deserve recognition and limit the generality of our observed responses. First, effects of soil nitrogen fertilization on root nodulation may be nonlinear, and a two-point fertilization experiment such as the one done here is not equipped to address possible nonlinearities that might explain the interaction between soil nitrogen fertilization and root nodulation. Future work should consider conducting similar experiments using a larger suite of nitrogen fertilization treatments than presented here. Additionally, this study used a single plant species and an inoculant comprising a single bacterial species. While this did allow us to isolate mechanisms that drove *G. max* responses to nitrogen fertilization and inoculation independent of phylogeny or genetic diversity (a factor that limited inferences in Perkowski *et al.*, 2021), future work should consider conducting similar experiments using a suite of leguminous species, as well as a suite of different *Rhizobium* or other *Actinobacteria* mixtures. Doing so would better allow us to generalize patterns observed here and would better replicate soil microbial communities observed in nature.

*Conclusions*

Here, we used a single pair symbiosis to quantify the impact of symbiotic nitrogen fixation on the structural carbon costs to acquire nitrogen under varying soil nitrogen environments. We find that symbiotic nitrogen fixing bacteria reduced structural carbon costs to acquire nitrogen when soil nitrogen availability was low, but had no impact when soil nitrogen availability was high. Carbon cost to acquire nitrogen differences between treatment combinations were entirely due to changes in plant nitrogen uptake rather than belowground structural carbon investments, suggesting that symbiotic nitrogen fixation allowed plants to maximize nitrogen uptake efficiency under low soil nitrogen environments. These results indicate that symbiotic nitrogen fixation may provide a competitive advantage to plants growing in nitrogen-poor soils. The findings can be used to help improve simulations of carbon-nitrogen economics in terrestrial biosphere models.

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**Author contributions**

EAP conducted data analysis, wrote the first draft of the manuscript with NGS, and made revisions based on collaborator and reviewer feedback with NGS. JT designed the experiment with NGS and EAP, carried out the experiment, and contributed to manuscript revisions. HG assisted with post-experiment harvest and contributed to manuscript revisions. NGS oversaw experiment progress, assisted with the post-experiment harvest, and contributed to manuscript revisions.

**Data Availability Statement**

All statistical analyses and plots were created in R version 4.2.0. All R code and data for this manuscript are available in a GitHub repository at <insert URL here> (<insert DOI from Zenodo here>).

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