

Meta-analysis investigating P and N*P addition impacts on plant functional traits

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Script that explores the effect of P addition on leaf and whole-plant functional traits using P fertilization and N*P fertilization experiments. The meta-analysis includes data from the MESI database as of January 07, 2025 and additional experiments compiled by Evan Perkowski.

Script summarizes the number of observations per trait and then conducts a meta-analysis to summarize plant responses to P addition, then conducts a second meta-analysis that summarizes responses when P is added in concert with N.

NOTE: NEED TO CHECK THAT ADDED SAMPLE SIZE OF ADDED EXPERIMENTS IS APPROPRIATE FOR MESI (SHOULD BE LISTED AS 'PLOT' REPS, NOT INDIVIDUAL REP NUMBER)

```
# Libraries
library(dplyr)
library(tidyr)
library(stringr)
library(ggplot2)
library(readr)
library(metafor)
library(MAd)
library(patchwork)

# MESI data
df_mesi <- read_csv("../data/mesi_main.csv")

# Manual data compilation
df_manual <- read_csv("../data/CNP_compiled_data.csv") %>%
  mutate(sampling_year = as.character(sampling_year),
         treatment = ifelse(treatment == FALSE, "f", treatment),
         sampling_date = as.character(sampling_date))

# Merge MESI database with manual data compilation
df_total <- df_mesi %>%
  full_join(df_manual)
```

Explore data availability in combined dataset for N-fertilization experiments

```
explore_nfert_exps <- df_total %>%

# field experiments only
```

```

filter(experiment_type == "field") %>%

# fertilisation experiments only
filter(treatment == "f") %>%

# P-fertilisation only (without N or K addition)
filter(npk == "_100")

head(explore_nfert_exps)

## # A tibble: 6 x 60
##   db      id  duplicate_id citation    response site study exp    lat    lon
##   <chr>   <chr> <chr>          <chr>      <chr>   <chr> <chr> <chr> <dbl> <dbl>
## 1 hebei  h962  h962      stape_et_a~ agb      -11.~ -11.~ -11.~ -12.0 -38.1
## 2 hebei  h963  h963      stape_et_a~ agb      -11.~ -11.~ -11.~ -12.0 -38.1
## 3 hebei  h964  h964      stape_et_a~ agb      -11.~ -11.~ -11.~ -12.0 -38.1
## 4 hebei  h965  h965      stape_et_a~ agb      -11.~ -11.~ -11.~ -12.0 -38.1
## 5 alberta a72   a72      collins_et~ agb      34.2~ 34.2~ 34.2~ 34.2 -106.
## 6 alberta a71   a71      collins_et~ agb      34.2~ 34.2~ 34.2~ 34.2 -106.
## # i 50 more variables: elevation <dbl>, mat <dbl>, map <dbl>,
## # ecosystem_type <chr>, vegetation_type <chr>, experiment_type <chr>,
## # community_type <chr>, dominant_species <chr>, growth_form <chr>, age <dbl>,
## # disturbance_type <chr>, treatment <chr>, npk <chr>, w_t1 <chr>, c_c <dbl>,
## # c_t <dbl>, d_t <dbl>, d_t2 <dbl>, n_c <dbl>, n_t <dbl>, p_c <dbl>,
## # p_t <dbl>, k_c <dbl>, k_t <dbl>, i_c <dbl>, i_t <dbl>, i_t2 <dbl>,
## # s_c <dbl>, s_t <dbl>, w_t2 <dbl>, w_t3 <dbl>, start_year <dbl>, ...

## How many experiments?
length(unique(explore_nfert_exps$exp))

## [1] 417

## What traits are available?
unique(explore_nfert_exps$response)

## [1] "agb"
## [2] "fine_root_biomass"
## [3] "total_biomass"
## [4] "bgb"
## [5] "r_soil"
## [6] "npp"
## [7] "agb_group"
## [8] "anpp_group"
## [9] "soil_total_c_org_layer"
## [10] "soil_total_c_min_layer"
## [11] "soil_total_c"
## [12] "mbc"
## [13] "anpp"
## [14] "mbn"
## [15] "litter_decomposition"
## [16] "bgb_n"
## [17] "leaf_c"
## [18] "leaf_cn"
## [19] "leaf_n_mass"
## [20] "soil_nh4_min_layer"

```

```

## [21] "soil_no3_min_layer"
## [22] "bgb_n_stock"
## [23] "soil_total_n_min_layer"
## [24] "soil_n2o_flux"
## [25] "agb_n_stock"
## [26] "som_n"
## [27] "soil_nh4"
## [28] "soil_no3"
## [29] "root_production"
## [30] "root_shoot_ratio"
## [31] "mb"
## [32] "anet"
## [33] "agb_n"
## [34] "asat"
## [35] "leaf_n_area"
## [36] "soil_potential_net_n_mineralization"
## [37] "soil_nh4-n"
## [38] "soil_no3-n"
## [39] "soil_total_n"
## [40] "soil_total_cn"
## [41] "soil_n_immobilization"
## [42] "mbcn"
## [43] "lwp"
## [44] "soil_total_p"
## [45] "r_soilh"
## [46] "fine_root_turnover"
## [47] "leaf_biomass_eco"
## [48] "stem_biomass"
## [49] "litter_n"
## [50] "litter_p"
## [51] "litter_k"
## [52] "fine_root_production"
## [53] "r_eco"
## [54] "soc"
## [55] "soil_p"
## [56] "leaf_p_mass"
## [57] "leaf_k_mass"
## [58] "leaf_np"
## [59] "bgb_coarse"
## [60] "r_root"
## [61] "som"
## [62] "vcmax"
## [63] "anpp_leaf"
## [64] "fine_root_respiration"
## [65] "jmax"
## [66] "lai"
## [67] "leaf_litterfall"
## [68] "gpp"
## [69] "nep"
## [70] "swc"
## [71] "nee"
## [72] "amax"
## [73] "litter_biomass"
## [74] "bgb_group"

```

```

## [75] "total_biomass_group"
## [76] "litterfall"
## [77] "leaf_nue"
## [78] "soil_total_c_profile"
## [79] "r_leaf"
## [80] "lai_max"
## [81] "wue_leaf"
## [82] "gs"
## [83] "wood_n"
## [84] "soil_net_n_mineralization"
## [85] "wue"
## [86] "wue_eco"
## [87] "grain_n"
## [88] "root_n"
## [89] "root_n_uptake"
## [90] "grain_c"
## [91] "bgb_c"
## [92] "stem_n"
## [93] "soil_total_cn_org_layer"
## [94] "soil_total_cn_min_layer"
## [95] "anpp_grain"
## [96] "bnpp"
## [97] "soil_in"
## [98] "leaf_cp"
## [99] "leaf_litter_p"
## [100] "leaf_litter_np"
## [101] "agb_p_stock"
## [102] "agb_ndvi"
## [103] "agb_height"
## [104] "agb_p"
## [105] "agb_cn"
## [106] "bgb_cn"
## [107] "leaf_biomass_plant"
## [108] "root_length"
## [109] "soil_gross_nitrification"
## [110] "soil_net_nitrification"
## [111] "soil_denitrification"
## [112] "anpp_woody"
## [113] "leaf_biomass_leaf"
## [114] "coarse_root_n"
## [115] "fine_root_n"
## [116] "leaf_litter_n"
## [117] "bai"
## [118] "leaf_area_leaf"
## [119] "soil_phkcl"
## [120] "soil_pbray"
## [121] "agb_np"
## [122] "soil_n_leaching"
## [123] "soil_net_ammonification"
## [124] "agb_c"
## [125] "root_p_uptake"
## [126] "rmf"
## [127] "rootshoot"
## [128] "lma"

```

```
## [129] "leaf_p_area"
## [130] "spad"
## [131] "tpu"
## [132] "rd"
## [133] "leaf_pue"
## [134] "leaf_structure_p"
## [135] "leaf_metabolic_p"
## [136] "leaf_nucleic_p"
## [137] "leaf_residual_p"
## [138] "leaf_thickness"
## [139] "sla"
## [140] "ldmc"
## [141] "leaf_pi"
## [142] "leaf_sugar_p"
## [143] "E"
## [144] "leaf_wue"
```

Select variables

```
use_response_n <- c("total_biomass",
                    "agb",
                    "bgb",
                    "leaf_n_mass",
                    "leaf_n_area",
                    "leaf_p_mass",
                    "leaf_p_area",
                    "leaf_np",
                    "gs",
                    "lma",
                    "sla",
                    "spad",
                    "amax",
                    "vcmax",
                    "jmax",
                    "leaf_nue",
                    "leaf_pue",
                    "rd",
                    "tpu",
                    "asat",
                    "cica",
                    "ci",
                    "leaf_structre_p",
                    "leaf_metabolic_p",
                    "leaf_nucleic_p",
                    "leaf_residual_p",
                    "leaf_pi",
                    "leaf_sugar_p"
)

nfert_responses <- explore_nfert_exps %>%
  filter(response %in% use_response_n) %>%
  mutate(myvar = response) %>%
  mutate(myvar = ifelse(myvar %in% c("cica", "ci"),
```

```

      "cica", myvar)) %>%
mutate(myvar = ifelse(myvar %in% c("leaf_", "amax"),
      "anet", myvar))

use_vars_n <- unique(nfert_responses$myvar)

```

Analysis

Calculate "ROM" - the log transformed ratio of means (Hedges et al., 1999; Lajeunesse, 2011) for each observation pair (ambient and elevated).

```

nfert_responses2 <- nfert_responses %>%

  ## keep only essential variables and drop rows containing missing values for
  ## essential variables
  select(id, duplicate_id, exp, myvar, treatment, sampling_year,
    x_t, x_c, sd_t, sd_c, rep_t, rep_c) %>%

  ## Get logarithm of response ratio and its variance
  metafor::escalc(
    measure = "ROM",
    m1i = x_t, sd1i = sd_t, n1i = rep_t,
    m2i = x_c, sd2i = sd_c, n2i = rep_c,
    data = .,
    append = TRUE,
    var.names = c("logr", "logr_var")
  ) %>%

  ## to keep the output readable from the console output
  as_tibble() %>%

  ## get standard error
  mutate( logr_se = sqrt(logr_var) / sqrt(rep_t) )

head(nfert_responses2)

## # A tibble: 6 x 15
##   id    duplicate_id exp      myvar treatment sampling_year   x_t   x_c   sd_t
##   <chr> <chr>      <chr>    <chr> <chr>      <chr>      <dbl> <dbl> <dbl>
## 1 h962  h962      -11.97_~ agb    f          <NA>      3.89e3 3.38e3 4.62e2
## 2 h963  h963      -11.97_~ agb    f          <NA>      7.86e3 7.26e3 7.04e2
## 3 h964  h964      -11.97_~ agb    f          <NA>      1.00e4 9.48e3 7.46e2
## 4 h965  h965      -11.97_~ agb    f          <NA>      1.10e4 1.06e4 1.48e3
## 5 a72   a72       34.2_10~ agb    f          2        7.72e1 4.89e1 6.07e0
## 6 a71   a71       34.2_10~ agb    f          3        5.6 e1 3.75e1 4.36e0
## # i 6 more variables: sd_c <dbl>, rep_t <dbl>, rep_c <dbl>, logr <dbl>,
## #   logr_var <dbl>, logr_se <dbl>

# Aggregate all measurements (multiple years, sampling dates and plots) by experiment (and response var
nfert_responses3 <- nfert_responses2 %>%

  # suggested addition by Kevin, email 02.10.2023 10:03
  dplyr::distinct(duplicate_id, x_t, x_c, .keep_all = TRUE) |>

```

```

filter(!is.na(logr_var) & !is.na(logr)) %>%

# re-create ID (common ID per experiment and response variable)
select(-id) %>%
mutate( id = paste(exp, myvar, sep = "_XXX_")) %>%

MAAd::agg(
  id = id,
  es = logr,
  var = logr_var,
  cor = 1.0,
  method = "BHHR",
  data = .
) %>%

## to keep the output readable from the console output
as_tibble() %>%

# separate ID again for ease of data use
mutate( id = str_split(id, "_XXX_") ) %>%
mutate( exp = purrr::map_chr(id, 1),
        myvar = purrr::map_chr(id, 2) ) %>%

## rename again
select(exp, myvar, logr = es, logr_var = var) %>%

## add number of observations (sum of plots and repeated samplings)
left_join(
  nfert_responses2 %>%
    group_by(exp, myvar, treatment) %>%
    summarise(n_c = sum(rep_c), n_t = sum(rep_t)),
  by = c("exp", "myvar")
) %>%

## get standard error. Verify if number available observations are identical
## for ambient and elevated. Use N from control here (n_c).
mutate( logr_se = sqrt(logr_var) / sqrt(n_c) ,

        # merge SLA and LMA measurements by taking inverse of logr (keep SE)
        logr = ifelse(myvar == "sla", -logr, logr),
        myvar = ifelse(myvar == "sla", "lma", myvar))

```

`summarise()` has grouped output by 'exp', 'myvar'. You can override using the
`.groups` argument.

```
head(nfert_responses3)
```

```
## # A tibble: 6 x 8
```

	exp	myvar	logr	logr_var	treatment	n_c	n_t	logr_se
	<chr>	<chr>	<dbl>	<dbl>	<chr>	<dbl>	<dbl>	<dbl>
## 1	-11.97_-38.12_f	agb	0.0789	0.00489	f	16	16	0.0175
## 2	34.2_106.43_f	agb	0.429	0.00271	f	10	10	0.0165
## 3	37.25_-121.75_forb_fn	agb	0	0.00292	f	3	3	0.0312
## 4	37.25_-121.75_grass_fn	agb	-0.121	0.00305	f	3	3	0.0319

```
## 5 38.53_-76.33_f      agb      1.01   0.00544 f      35      35 0.0125
## 6 38.53_-76.33_f      total_b~ 0.465   0.000154 f      15      15 0.00320
```

Meta-analysis

Aggregate log-ratios across multiple experiments, taking into account their respective variance and using the experiment identity as a grouping factor for random intercepts.

```
source("../helper_fxns/analyse_meta.R")

out_n <- purrr::map(as.list(use_vars_n),
  ~analyse_meta(nfert_responses3 %>%
    rename(var = myvar), nam_target = .))

names(out_n) <- use_vars_n
df_box_n <- purrr::map_dfr(out_n, "df_box") |>
  left_join(
    nfert_responses3 |>
      group_by(myvar) |>
      summarise(logr_min = min(logr), logr_max = max(logr)) |>
      rename(var = myvar),
    by = "var"
  )
saveRDS(df_box_n, file = paste0(here::here(), "df_box_nfert.rds"))
```

Final data size

Number of data points (plot-level measurements) per variable:

```
nfert_responses3 %>%
  group_by(myvar) %>%
  summarise(n_plots = sum(n_c, na.rm = TRUE), n_exp = n()) %>%
  rename("Variable"="myvar", "N observations"="n_plots", "N experiments"="n_exp") %>%
  knitr::kable()
```

Variable	N observations	N experiments
agb	3764	154
anet	12	1
asat	284	11
bgb	1262	104
gs	12	1
jmax	162	6
leaf_metabolic_p	25	1
leaf_n_area	442	33
leaf_n_mass	808	50
leaf_np	511	40
leaf_nucleic_p	35	2
leaf_nue	52	4
leaf_p_area	316	28
leaf_p_mass	620	51
leaf_pi	10	1
leaf_pue	25	1
leaf_residual_p	35	2
leaf_sugar_p	10	1
lma	382	29

Variable	N observations	N experiments
rd	85	2
spad	107	3
total_biomass	870	28
tpu	33	2
vcmax	158	6

Some quick plots:

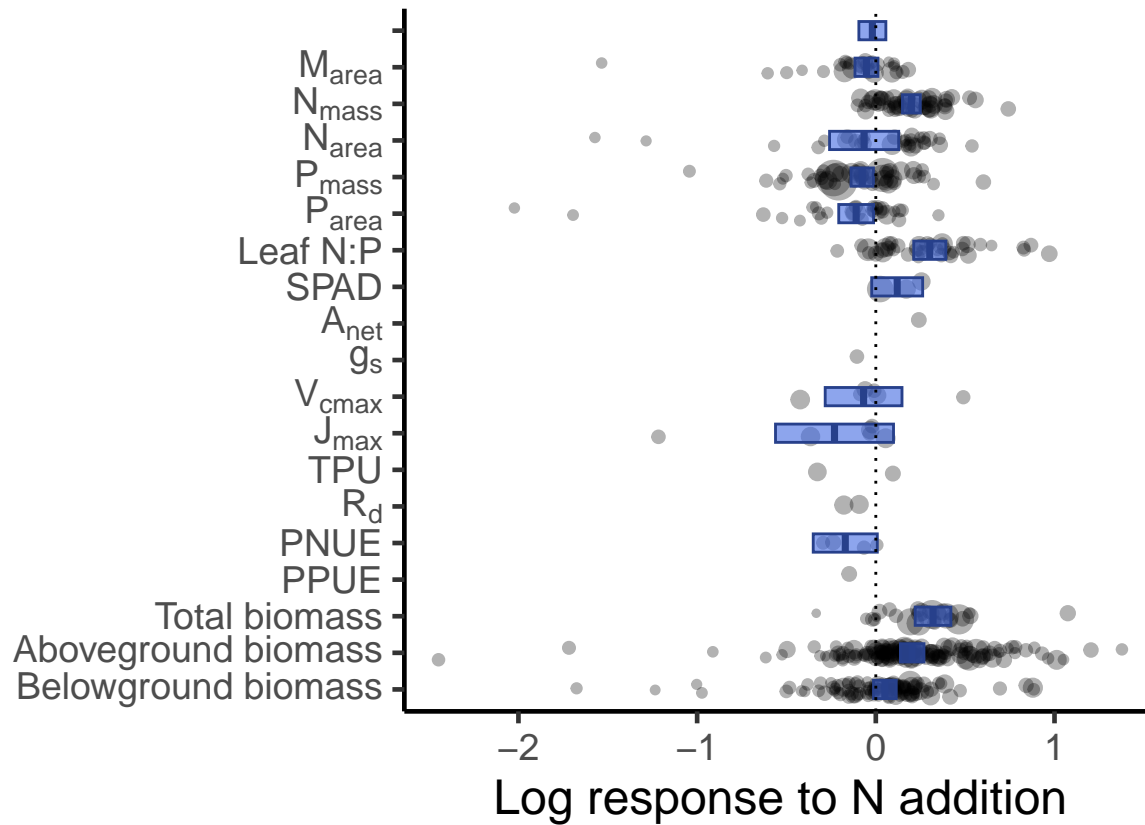
```
nfert_responses3 <- nfert_responses3 %>%
  mutate(myvar = factor(myvar, levels = c("bgb", "agb", "total_biomass", "leaf_pue",
    "leaf_nue", "rd", "tpu", "jmax", "vcmax",
    "cica", "gs", "anet", "spad", "leaf_np",
    "leaf_p_area", "leaf_p_mass", "leaf_n_area",
    "leaf_n_mass", "lma")))

meta_plot_n <- ggplot(data = subset(nfert_responses3, myvar != "cica"),
  aes(x = myvar, y = logr)) +
  geom_jitter(color = rgb(0,0,0,0.3),
    aes( size = 1/logr_se ),
    position = position_jitter(w = 0.2, h = 0),
    show.legend = FALSE) +
  geom_crossbar( data = df_box_n %>% drop_na(var),
    aes(x = var, y = middle, ymin = ymin, ymax = ymax),
    fill = "royalblue",
    color = "royalblue4",
    alpha = 0.6,
    width = 0.5 ) +
  geom_hline( yintercept = 0.0, linewidth = 0.5, linetype = "dotted" ) +
  scale_x_discrete(labels = c("Belowground biomass",
    "Aboveground biomass",
    "Total biomass",
    "PPUE",
    "PNUE",
    expression("R"["d"]),
    "TPU",
    expression("J"["max"]),
    expression("V"["cmax"]),
    expression("g"["s"]),
    expression("A"["net"]),
    "SPAD",
    "Leaf N:P",
    expression("P"["area"]),
    expression("P"["mass"]),
    expression("N"["area"]),
    expression("N"["mass"]),
    expression("M"["area"]))) +

  #scale_x_discrete("", labels = mylabl) +
  labs(x = "",
    y = "Log response to N addition") +
  coord_flip() +
```

```
theme_classic(base_size = 18)
meta_plot_n
```

```
## Warning: Removed 2 rows containing missing values (`geom_point()`).
```



Explore data availability in combined dataset for P-fertilization experiments

```
explore_pfert_exps <- df_total %>%
  # field experiments only
  # filter(experiment_type == "field") %>%

  # fertilisation experiments only
  filter(treatment == "f") %>%

  # P-fertilisation only (without N or K addition)
  filter(npk == "_010")

head(explore_pfert_exps)
```

```
## # A tibble: 6 x 60
##   db      id duplicate_id citation      response site study exp      lat lon
##   <chr>   <chr> <chr>         <chr>      <chr>    <chr> <chr> <chr> <dbl> <dbl>
## 1 sichuan s885 s885      lawrence_2~ total_b~ 0.12~ 0.12~ 0.12~ 0.12 110.
## 2 sichuan s888 s888      lawrence_2~ total_b~ 0.12~ 0.12~ 0.12~ 0.12 110.
```

```
## 3 sichuan s879 s879      lawrence_2~ total_b~ 0.12~ 0.12~ 0.12~ 0.12 110.
## 4 sichuan s882 s882      lawrence_2~ total_b~ 0.12~ 0.12~ 0.12~ 0.12 110.
## 5 sichuan s385 s385      augustine_~ anpp      0.28~ 0.28~ 0.28~ 0.28 37.9
## 6 sichuan s172 s172      ohalloran_~ agb       -15.~ -15.~ -15.~ -15.4 23.2
## # i 50 more variables: elevation <dbl>, mat <dbl>, map <dbl>,
## # ecosystem_type <chr>, vegetation_type <chr>, experiment_type <chr>,
## # community_type <chr>, dominant_species <chr>, growth_form <chr>, age <dbl>,
## # disturbance_type <chr>, treatment <chr>, npk <chr>, w_t1 <chr>, c_c <dbl>,
## # c_t <dbl>, d_t <dbl>, d_t2 <dbl>, n_c <dbl>, n_t <dbl>, p_c <dbl>,
## # p_t <dbl>, k_c <dbl>, k_t <dbl>, i_c <dbl>, i_t <dbl>, i_t2 <dbl>,
## # s_c <dbl>, s_t <dbl>, w_t2 <dbl>, w_t3 <dbl>, start_year <dbl>, ...
```

How many experiments?

```
length(unique(explore_pfert_exps$exp))
```

```
## [1] 255
```

What traits are available?

```
unique(explore_pfert_exps$response)
```

```
## [1] "total_biomass_group"      "anpp"                      "agb"
## [4] "total_biomass"           "mbc"                       "soil_total_c"
## [7] "bgb"                     "leaf_c"                    "agb_group"
## [10] "anpp_group"              "agb_n"                     "agb_p"
## [13] "soil_total_n"            "agb_p_stock"               "gpp"
## [16] "fine_root_biomass"       "leaf_n_mass"               "leaf_p_mass"
## [19] "lai"                     "mbn"                       "r_soil"
## [22] "gs"                      "lwp"                       "fine_root_production"
## [25] "soil_no3-n_supply_rate"  "soil_nh4-n_supply_rate"    "soil_p_supply_rate"
## [28] "leaf_biomass_eco"        "root_shoot_ratio"          "bpe"
## [31] "total_biomass_p_stock"   "total_biomass_n_stock"     "total_biomass_np"
## [34] "agb_pod_n"               "wood_n"                    "agb_pod_p"
## [37] "wood_p"                  "agb_grain_n"               "agb_grain_p"
## [40] "anpp_grain"              "soil_p"                    "agb_ndvi"
## [43] "agb_height"              "bgb_n"                     "bgb_p"
## [46] "root_n_uptake"           "nee"                       "soil_np"
## [49] "leaf_np"                 "litter_decomposition"      "r_root"
## [52] "nep"                     "lma"                       "leaf_n_area"
## [55] "leaf_p_area"             "rgr"                       "amax"
## [58] "vcmax"                   "jmax"                      "leaf_nue"
## [61] "leaf_pue"                "leaf_pi"                   "leaf_ester_p"
## [64] "leaf_nucleic_p"          "leaf_lipid_p"              "leaf_residual_p"
## [67] "asat"                    "tpu"                       "rd"
## [70] "cica"                    "rmf"                       "rootshoot"
## [73] "sla"                     "stom_lim"                  "gsw"
## [76] "spad"                    "anet"                      "leaf_structure_p"
## [79] "leaf_metabolic_p"        "leaf_thickness"            "ldmc"
## [82] "fine_root_p_mass"        "coarse_root_p_mass"        "stem_p_mass"
## [85] "total_p_uptake_eff"      "E"                         "iwue"
## [88] "leaf_sugar_p"            "leaf_wue"
```

Select variables

```
use_response_p <- c("total_biomass",
                    "agb",
```

```

        "bgb",
        "rmf",
        "rootshoot",
        "leaf_n_mass",
        "leaf_n_area",
        "leaf_p_mass",
        "leaf_p_area",
        "leaf_np",
        "gs",
        "lma",
        "sla",
        "spad",
        "amax",
        "vcmax",
        "jmax",
        "leaf_nue",
        "leaf_pue",
        "rd",
        "tpu",
        "asat",
        "cica",
        "ci",
        "leaf_strucure_p",
        "leaf_metabolic_p",
        "leaf_nucleic_p",
        "leaf_residual_p",
        "leaf_pi",
        "leaf_sugar_p"
    )

pfert_responses <- explore_pfert_exps %>%
  filter(response %in% use_response_p) %>%
  mutate(myvar = response) %>%
  mutate(myvar = ifelse(myvar %in% c("cica", "ci"),
                        "cica", myvar)) %>%
  mutate(myvar = ifelse(myvar %in% c("asat", "amax"),
                        "anet", myvar))

use_vars_p <- unique(pfert_responses$myvar)

```

Analysis

Calculate "ROM" - the log transformed ratio of means (Hedges et al., 1999; Lajeunesse, 2011) for each observation pair (ambient and elevated).

```

pfert_responses2 <- pfert_responses %>%

  ## keep only essential variables and drop rows containing missing values for
  ## essential variables
  select(id, duplicate_id, exp, myvar, treatment, sampling_year,
         x_t, x_c, sd_t, sd_c, rep_t, rep_c) %>%

  ## Get logarithm of response ratio and its variance
  metafor::escalc(

```

```

measure = "ROM",
m1i = x_t, sd1i = sd_t, n1i = rep_t,
m2i = x_c, sd2i = sd_c, n2i = rep_c,
data = .,
append = TRUE,
var.names = c("logr", "logr_var")
) %>%

## to keep the output readable from the console output
as_tibble() %>%

## get standard error
mutate( logr_se = sqrt(logr_var) / sqrt(rep_t) )

head(pfert_responses2)

## # A tibble: 6 x 15
##   id      duplicate_id exp      myvar treatment sampling_year   x_t   x_c sd_t
##   <chr> <chr>          <chr>    <chr> <chr>         <chr>   <dbl> <dbl> <dbl>
## 1 s172  s172          -15.44_2~ agb    f             2        6.29  9.43  5.76
## 2 s3497 s3497          17.25_-8~ agb    f             3       488.  106.  43.7
## 3 s3500 s3500          17.25_-8~ agb    f             3       418.  200.  83.7
## 4 s3503 s3503          17.25_-8~ agb    f             3       247.   80.1  43.7
## 5 s1524 s1524          -18.66_2~ tota~ f             2       76.4  46.7  18.8
## 6 s168  s168          -18.66_2~ agb    f             1       80.0  51.1  19.6
## # i 6 more variables: sd_c <dbl>, rep_t <dbl>, rep_c <dbl>, logr <dbl>,
## #   logr_var <dbl>, logr_se <dbl>

# Aggregate all measurements (multiple years, sampling dates and plots) by experiment (and response var
pfert_responses3 <- pfert_responses2 %>%

# suggested addition by Kevin, email 02.10.2023 10:03
dplyr::distinct(duplicate_id, x_t, x_c, .keep_all = TRUE) |>

filter(!is.na(logr_var) & !is.na(logr)) %>%

# re-create ID (common ID per experiment and response variable)
select(-id) %>%
mutate( id = paste(exp, myvar, sep = "_XXX_") ) %>%

MAAd::agg(
  id = id,
  es = logr,
  var = logr_var,
  cor = 1.0,
  method = "BHHR",
  data = .
) %>%

## to keep the output readable from the console output
as_tibble() %>%

# separate ID again for ease of data use
mutate( id = str_split(id, "_XXX_") ) %>%

```

```

mutate( exp = purrr::map_chr(id, 1),
        myvar = purrr::map_chr(id, 2) ) %>%

## rename again
select(exp, myvar, logr = es, logr_var = var) %>%

## add number of observations (sum of plots and repeated samplings)
left_join(
  pfert_responses2 %>%
    group_by(exp, myvar, treatment) %>%
    summarise(n_c = sum(rep_c), n_t = sum(rep_t)),
  by = c("exp", "myvar")
) %>%

## get standard error. Verify if number available observations are identical
## for ambient and elevated. Use N from control here (n_c).
mutate( logr_se = sqrt(logr_var) / sqrt(n_c) ,

        # merge SLA and LMA measurements by taking inverse of logr (keep SE)
        logr = ifelse(myvar == "sla", -logr, logr),
        myvar = ifelse(myvar == "sla", "lma", myvar))

```

`summarise()` has grouped output by 'exp', 'myvar'. You can override using the
`.groups` argument.

```
head(pfert_responses3)
```

```
## # A tibble: 6 x 8
```

##	exp	myvar	logr	logr_var	treatment	n_c	n_t	logr_se
##	<chr>	<chr>	<dbl>	<dbl>	<chr>	<dbl>	<dbl>	<dbl>
## 1	-15.44_23.25_f	agb	-0.405	0.287	f	4	4	0.268
## 2	17.25_-88.77_f	agb	1.13	0.0572	f	9	9	0.0797
## 3	-18.66_25.5_f	total_biomass	0.491	0.0285	f	3	3	0.0974
## 4	-18.66_25.5_f	agb	0.158	0.418	f	8	8	0.229
## 5	19.6_-155.33_fp	bgb	0.142	0.0124	f	3	3	0.0644
## 6	-2.98_-47.52_f	agb	0.269	0.000881	f	4	4	0.0148

Meta-analysis

Aggregate log-ratios across multiple experiments, taking into account their respective variance and using the experiment identity as a grouping factor for random intercepts.

```

source("../helper_fxns/analyse_meta.R")

out_p <- purrr::map(as.list(use_vars_p),
  ~analyse_meta(pfert_responses3 %>%
    rename(var = myvar), nam_target = .))

names(out_p) <- use_vars_p
df_box_p <- purrr::map_dfr(out_p, "df_box") |>
  left_join(
    pfert_responses3 |>
      group_by(myvar) |>
      summarise(logr_min = min(logr), logr_max = max(logr)) |>
      rename(var = myvar),

```

```

    by = "var"
  )
saveRDS(df_box_p, file = paste0(here::here(), "df_box_pfert.rds"))

```

Final data size

Number of data points (plot-level measurements) per variable:

```

pfert_responses3 %>%
  group_by(myvar) %>%
  summarise(n_plots = sum(n_c, na.rm = TRUE), n_exp = n()) %>%
  rename("Variable"="myvar", "N observations"="n_plots", "N experiments"="n_exp") %>%
  knitr::kable()

```

Variable	N observations	N experiments
agb	918	126
anet	608	12
bgb	250	56
cica	116	3
gs	69	5
jmax	473	8
leaf_metabolic_p	25	1
leaf_n_area	727	33
leaf_n_mass	885	44
leaf_np	807	44
leaf_nucleic_p	87	3
leaf_nue	382	5
leaf_p_area	701	33
leaf_p_mass	821	47
leaf_pi	62	2
leaf_pue	381	5
leaf_residual_p	87	3
leaf_sugar_p	5	1
lma	761	33
rd	309	4
rmf	91	29
rootshoot	91	29
spad	107	3
total_biomass	101	14
tpu	160	5
vcmax	473	8

Number of data points (plot-level measurements) per experiment:

```

pfert_responses3 %>%
  group_by(exp) %>%
  summarise(n_plots = sum(n_c), n_exp = n()) %>%
  rename_("Experiment"="exp", "N observations"="n_plots", "N experiments"="n_exp") %>%
  knitr::kable()

```

Experiment	N observations	N experiments
-15.44_23.25_f	4	1
-18.66_25.5_f	11	2

Experiment	N observations	N experiments
-2.98_-47.52_f	4	1
-21.65_21.81_f	3	1
-22.283_117.666_f	6	1
-22.41_21.71_f	4	1
-22.78_31.25_f	16	2
-23.75_31.43_f	16	2
-24.17_21.89_f	7	2
-24.4_31.75_f	16	2
-25.12_31.23_f	16	2
-25.29_31.91_f	16	2
-3.5_36_f	6	1
-3.95_-79.03_f	12	2
-33.35_150.3_f	9	3
-35.73_-58.05_f	12	1
17.25_-88.77_f	9	1
19.6_-155.33_fp	3	1
22.13_-159.63_f	11	1
26.52_109.78_fp	3	1
31.37_90.02_f	24	1
31.3_-81.28_f	4	1
31.55_-81.78_f	4	1
32.54_-116.7_fp	12	1
33.7_120.3_f	15	1
34.92_102.88_f2p	5	1
34.92_102.88_f3p	5	1
34.92_102.88_fp	5	1
37.25_-121.75_forb_fp	3	1
37.25_-121.75_grass_fp	3	1
37.48_101.2_fp	18	3
37.55_-122.3_f	3	1
37.6_101.32_fp	20	3
37.87_-122.52_f	18	2
39.25_-121.28_fp	20	1
39.75_-74.75_fp	4	1
41.35_36.25_fp	4	1
41.35_36.25_fp2	4	1
41.62_-71.32_fp	8	2
42.28_-85.58_f	6	2
42.58_122.21_fp	12	2
44.8_116.03_fp	6	2
47.57_7.6_f	6	1
51.85_5.62_fp	3	1
52.07_5.58_fp	3	1
52.37_5.1_fp	6	1
52.5_5.7_fp	6	1
53.83_-8.83_fp	10	1
54.63_8.83_fp	5	1
64.83_-147.72_fp	3	1
69.43_-133.02_fp	10	1
69.43_-133.02_fp2	10	1
9.6_-79.5_f	3	1
alpflifx_fp	60	1

Experiment	N observations	N experiments
bennekom_drained_fp	5	1
bennekom_undrained_fp	5	1
bloomfield2014_fp	1462	14
bordeaux_fp	40	3
bown2007_fnp	80	8
buitengoor_1992_fp	5	1
buitengoor_1993_fp	5	1
carswell2005_fnp	80	4
cleland2019_bldr.us_fnp	8	4
cleland2019_bnch.us_fnp	12	4
cleland2019_bogong.au_fnp	12	4
cleland2019_burrawan.au_fnp	12	4
cleland2019_cbgb.us_fnp	24	4
cleland2019_cdc.us_fnp	12	4
cleland2019_cdpt.us_fnp	12	4
cleland2019_cowi.ca_fnp	12	4
cleland2019_elliot.us_fnp	12	4
cleland2019_frue.ch_fnp	12	4
cleland2019_gilb.za_fnp	12	4
cleland2019_hall.us_fnp	12	4
cleland2019_hart.us_fnp	12	4
cleland2019_konz.us_fnp	12	4
cleland2019_lancaster.uk_fnp	8	4
cleland2019_look.us_fnp	12	4
cleland2019_mtca.au_fnp	12	4
cleland2019_sage.us_fnp	12	4
cleland2019_saline.us_fnp	12	4
cleland2019_sgs.us_fnp	12	4
cleland2019_shps.us_fnp	12	4
cleland2019_sier.us_fnp	12	4
cleland2019_smith.us_fnp	12	4
cleland2019_spin.us_fnp	12	4
cleland2019_summ.za_fnp	12	4
cleland2019_trel.us_fnp	12	4
cleland2019_ukul.za_fnp	24	4
cleland2019_unc.us_fnp	12	4
cleland2019_valm.ch_fnp	12	4
crous2017_fnp	2351	12
daqinggou_fp	12	2
drentsche_aa_drained_fp	5	1
drentsche_aa_wet_fp	5	1
ewenke_f_p	6	1
fan2024_fp	773	14
firn2019_bldr.us_fnp	21	6
firn2019_bnch.us_fnp	51	6
firn2019_bogong.au_fnp	72	6
firn2019_burrawan.au_fnp	73	6
firn2019_cbgb.us_fnp	54	6
firn2019_comp.pt_fnp	78	6
firn2019_cowi.ca_fnp	45	6
firn2019_elliot.us_fnp	54	6
firn2019_frue.ch_fnp	46	6

Experiment	N observations	N experiments
firn2019_gilb.za_fnp	48	6
firn2019_hopl.us_fnp	51	6
firn2019_kiny.au_fnp	45	6
firn2019_konz.us_fnp	29	6
firn2019_lancaster.uk_fnp	51	6
firn2019_look.us_fnp	54	6
firn2019_mcla.us_fnp	51	6
firn2019_mtca.au_fnp	53	6
firn2019_sage.us_fnp	31	6
firn2019_sgs.us_fnp	39	6
firn2019_shps.us_fnp	24	6
firn2019_sier.us_fnp	48	6
firn2019_smith.us_fnp	54	6
firn2019_summ.za_fnp	72	6
firn2019_unc.us_fnp	54	6
firn2019_valm.ch_fnp	120	6
flottbek_fp	30	1
gusewell_s1_fp	4	1
gusewell_s2_fp	4	1
gusewell_t1_fp	4	1
gusewell_t2_fp	4	1
gusewell_v1_fp	4	1
gusewell_v2_fp	4	1
gusewell_v3_fp	4	1
gusewell_w1_p	4	1
gusewell_w2_p	4	1
gusewell_w3_p	4	1
gusewell_w4_p	4	1
herschgreen2024_cbgb.us_fnp	88	4
herschgreen2024_cdc.us_fnp	70	4
herschgreen2024_churn.us_fnp	50	4
herschgreen2024_kbs.us_fnp	50	4
herschgreen2024_konz.us_fnp	69	4
herschgreen2024_spin.us_fnp	77	4
hol_kortenhoef_fp	5	1
ingers_ng_2006np_fp	6	1
indoneisa_f	54	3
kansasf_fp	12	1
kansask_fp	12	1
katelijne_2016_fp	5	1
katelijne_2017_f2	10	1
katelijne_2017_f3	10	1
kisa_grahamiana_f	20	1
luneburg_field_2006_fp	10	1
luneburg_field_2008_fp	10	1
luneburg_gh_drought_molinia_fp	NA	2
luneburg_gh_fert_molinia_fp	10	1
michigan_underc_bog_fp	19	1
michigan_underc_intermfen_fp	17	1
michigan_underc_richfen_fp	20	1
mo2019_fnp	275	11
mo2021_fnp	10	2

Experiment	N observations	N experiments
molenpolder_fp	5	1
nashfield_pooled_fp	8	2
niwot_ridge2_dm_fp	5	1
niwot_ridge2_wm_fp	5	1
sanjiang_mire_pfert_fp	9	1
sanjiang_mire_pfert_fp2	9	1
sanjiang_mire_pfert_fp3	9	1
sanpedro_fp	6	3
schiermonnikoog_old_fp	12	1
schiermonnikoog_old_fp2	12	1
schiermonnikoog_young_fp	12	1
schiermonnikoog_young_fp2	12	1
shaaxi_330_fp	3	1
shaaxi_6_fp	3	1
tambopata_fp	6	3
teberda_fp	8	1
tono_fp	6	3
toolik_nonacidic_fp	6	2
verryckt2022_nouragues_fnp	472	11
verryckt2022_paracou_fnp	420	11
wang2019_fnp	25	5
warren2002_fp	30	2
warren2011_fp	120	8
wayqecha_fp	6	3
westbroek_polder_fp	5	1
yu2022_fnp	120	5
yucatan_marsh_highsalinity_fp	30	1
yucatan_marsh_lowsalinity_fp	30	1
yucatan_marsh_mediumsalityity_fp	30	1
zavistic2018_Bad_Brukenau_fp	26	4
zavistic2018_Unterluus_fp	26	4
zwarte_beek_drained_fp	5	1
zwarte_beek_wet_fp	5	1

Some quick plots:

```
pfert_responses3 <- pfert_responses3 %>%
  mutate(myvar = factor(myvar, levels = c("bgb", "agb", "total_biomass", "rmf", "rootshoot",
    "leaf_pue", "leaf_nue", "rd", "tpu", "jmax", "vcmax",
    "cica", "gs", "anet", "spad", "leaf_np",
    "leaf_p_area", "leaf_p_mass", "leaf_n_area",
    "leaf_n_mass", "lma", "leaf_strucure_p", "leaf_pi",
    "leaf_metabolic_p", "leaf_sugar_p", "leaf_nucleic_p",
    "leaf_residual_p"))))

meta_plot_p <- ggplot(data = subset(pfert_responses3, myvar != "cica"),
  aes(x = myvar, y = logr)) +
  geom_jitter(color = rgb(0,0,0,0.3),
    aes( size = 1/logr_se ),
    position = position_jitter(w = 0.2, h = 0),
    show.legend = FALSE) +
```

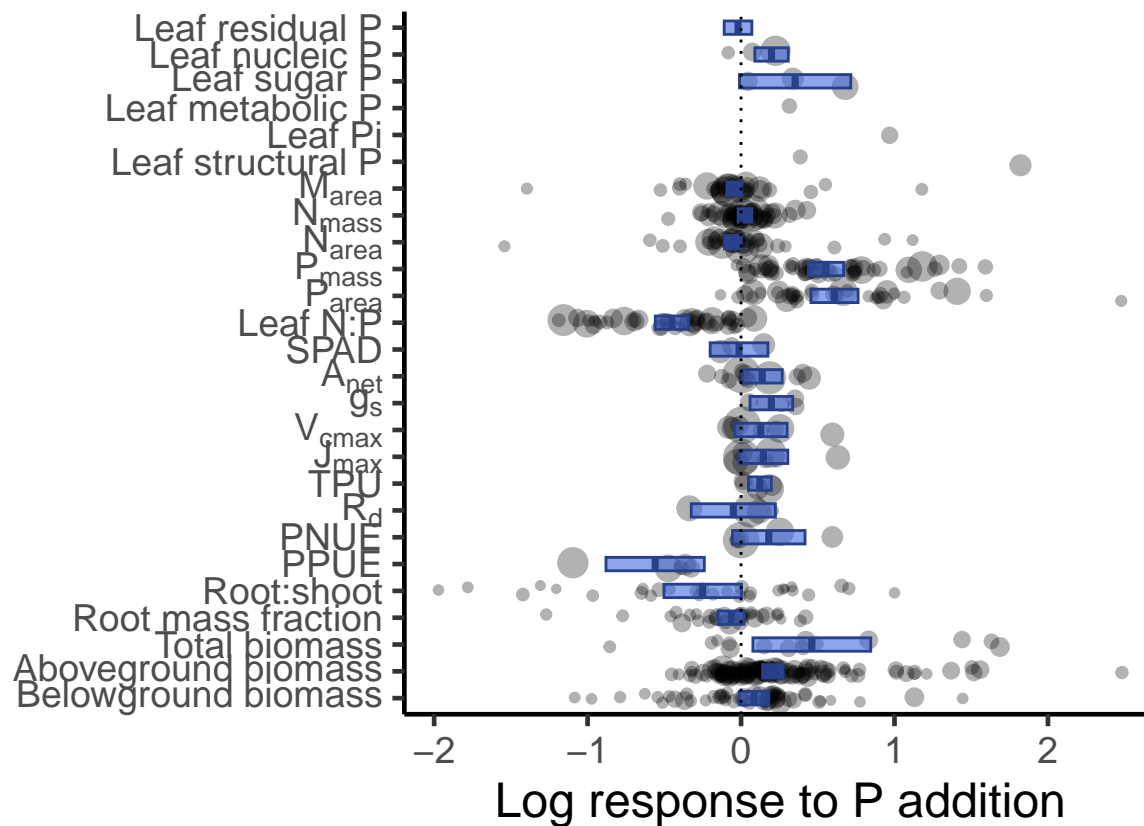
```

geom_crossbar( data = df_box_p %>% drop_na(var),
  aes(x = var, y = middle, ymin = ymin, ymax = ymax),
  fill = "royalblue",
  color = "royalblue4",
  alpha = 0.6,
  width = 0.5 ) +
geom_hline( yintercept = 0.0, linewidth = 0.5, linetype = "dotted" ) +
scale_x_discrete(labels = c("Belowground biomass",
  "Aboveground biomass",
  "Total biomass",
  "Root mass fraction",
  "Root:shoot",
  "PPUE",
  "PNUE",
  expression("R"["d"]),
  "TPU",
  expression("J"["max"]),
  expression("V"["cmax"]),
  expression("g"["s"]),
  expression("A"["net"]),
  "SPAD",
  "Leaf N:P",
  expression("P"["area"]),
  expression("P"["mass"]),
  expression("N"["area"]),
  expression("N"["mass"]),
  expression("M"["area"]),
  "Leaf structural P",
  "Leaf Pi",
  "Leaf metabolic P",
  "Leaf sugar P",
  "Leaf nucleic P",
  "Leaf residual P")) +

#scale_x_discrete("", labels = mylabl) +
labs(x = "",
  y = "Log response to P addition") +
coord_flip() +
theme_classic(base_size = 18)
meta_plot_p

```

```
## Warning: Removed 2 rows containing missing values (`geom_point()`).
```



Explore data availability in combined dataset for N*P-fertilization experiments

```
explore_npfert_exps <- df_total %>%

# fertilisation experiments only
filter(treatment == "f") %>%

# P-fertilization in concert with N-fertilization (without K addition)
filter(npk == "_110")

head(explore_npfert_exps)
```

```
## # A tibble: 6 x 60
##   db      id  duplicate_id citation    response site  study exp    lat  lon
##   <chr>  <chr> <chr>         <chr>      <chr>   <chr> <chr> <chr>  <dbl> <dbl>
## 1 sichuan s886 s886      lawrence_2~ total_b~ 0.12~ 0.12~ 0.12~ 0.12 110.
## 2 sichuan s889 s889      lawrence_2~ total_b~ 0.12~ 0.12~ 0.12~ 0.12 110.
## 3 sichuan s880 s880      lawrence_2~ total_b~ 0.12~ 0.12~ 0.12~ 0.12 110.
## 4 sichuan s883 s883      lawrence_2~ total_b~ 0.12~ 0.12~ 0.12~ 0.12 110.
## 5 sichuan s386 s386      augustine_~ anpp    0.28~ 0.28~ 0.28~ 0.28 37.9
## 6 sichuan s171 s171      ohalloran_~ agb     -15.~ -15.~ -15.~ -15.4 23.2
## # i 50 more variables: elevation <dbl>, mat <dbl>, map <dbl>,
## # ecosystem_type <chr>, vegetation_type <chr>, experiment_type <chr>,
## # community_type <chr>, dominant_species <chr>, growth_form <chr>, age <dbl>,
## # disturbance_type <chr>, treatment <chr>, npk <chr>, w_t1 <chr>, c_c <dbl>,
```

```
## #   c_t <dbl>, d_t <dbl>, d_t2 <dbl>, n_c <dbl>, n_t <dbl>, p_c <dbl>,
## #   p_t <dbl>, k_c <dbl>, k_t <dbl>, i_c <dbl>, i_t <dbl>, i_t2 <dbl>,
## #   s_c <dbl>, s_t <dbl>, w_t2 <dbl>, w_t3 <dbl>, start_year <dbl>, ...
```

```
## How many experiments?
```

```
length(unique(explore_npfert_exps$exp))
```

```
## [1] 286
```

```
## What traits are available?
```

```
unique(explore_npfert_exps$response)
```

```
## [1] "total_biomass_group"      "anpp"
## [3] "agb"                     "total_biomass"
## [5] "agb_c"                   "bgb_c"
## [7] "soil_total_c"            "mbc"
## [9] "bgb"                     "leaf_c"
## [11] "leaf_biomass_eco"        "agb_group"
## [13] "stem_biomass"            "agb_n"
## [15] "agb_p"                   "soil_total_n"
## [17] "agb_p_stock"             "gpp"
## [19] "fine_root_biomass"       "soil_no3"
## [21] "r_soil"                  "total_biomass_production"
## [23] "soil_total_cn"           "root_production"
## [25] "soc"                     "lwp"
## [27] "gs"                      "soil_no3_leaching"
## [29] "soil_nh4"                "soil_drp"
## [31] "soil_p"                  "fine_root_production"
## [33] "soil_no3-n_supply_rate"  "soil_nh4-n_supply_rate"
## [35] "soil_p_supply_rate"      "root_shoot_ratio"
## [37] "bpe"                     "total_biomass_np"
## [39] "agb_cn"                  "agb_cp"
## [41] "agb_ndvi"                "agb_height"
## [43] "bgb_n"                   "bgb_p"
## [45] "root_n_uptake"           "nee"
## [47] "swc"                     "soil_phkcl"
## [49] "soil_pbray"              "som_cn"
## [51] "soil_np"                 "leaf_n_mass"
## [53] "leaf_p_mass"             "leaf_np"
## [55] "litter_decomposition"    "r_root"
## [57] "nep"                     "agb_k_stock"
## [59] "rmf"                     "rootshoot"
## [61] "lma"                     "leaf_n_area"
## [63] "leaf_p_area"             "sla"
## [65] "stom_lim"                "vcmax"
## [67] "jmax"                    "tpu"
## [69] "leaf_nue"                "leaf_pue"
## [71] "asat"                    "rd"
## [73] "gsw"                     "spad"
## [75] "anet"                    "leaf_structure_p"
## [77] "leaf_metabolic_p"        "leaf_nucleic_p"
## [79] "leaf_residual_p"         "leaf_thickness"
## [81] "ldmc"                    "leaf_pi"
## [83] "leaf_sugar_p"            "E"
## [85] "leaf_wue"
```

Select variables

```
use_response_np <- c("total_biomass",
                    "agb",
                    "bgb",
                    "leaf_n_mass",
                    "leaf_n_area",
                    "leaf_p_mass",
                    "leaf_p_area",
                    "leaf_np",
                    "gs",
                    "lma",
                    "sla",
                    "spad",
                    "amax",
                    "vcmax",
                    "jmax",
                    "leaf_nue",
                    "leaf_pue",
                    "rd",
                    "tpu",
                    "asat",
                    "cica",
                    "ci"
)

npfert_responses <- explore_npfert_exps %>%
  filter(response %in% use_response_np) %>%
  mutate(myvar = response) %>%
  mutate(myvar = ifelse(myvar %in% c("cica", "ci"),
                        "cica", myvar)) %>%
  mutate(myvar = ifelse(myvar %in% c("asat", "amax"),
                        "anet", myvar))

use_vars_np <- unique(npfert_responses$myvar)
```

Analysis

Calculate "ROM" - the log transformed ratio of means (Hedges et al., 1999; Lajeunesse, 2011) for each observation pair (ambient and elevated).

```
npfert_responses2 <- npfert_responses %>%

  ## keep only essential variables and drop rows containing missing values for
  ## essential variables
  select(id, duplicate_id, exp, myvar, treatment, sampling_year,
         x_t, x_c, sd_t, sd_c, rep_t, rep_c) %>%

  ## Get logarithm of response ratio and its variance
  metafor::escalc(
    measure = "ROM",
    m1i = x_t, sd1i = sd_t, n1i = rep_t,
    m2i = x_c, sd2i = sd_c, n2i = rep_c,
    data = .,
```

```

    append = TRUE,
    var.names = c("logr", "logr_var")
  ) %>%

  ## to keep the output readable from the console output
  as_tibble() %>%

  ## get standard error
  mutate( logr_se = sqrt(logr_var) / sqrt(rep_t) )

head(npfert_responses2)

## # A tibble: 6 x 15
##   id      duplicate_id exp      myvar treatment sampling_year   x_t   x_c sd_t
##   <chr> <chr>          <chr>    <chr> <chr>      <chr>      <dbl> <dbl> <dbl>
## 1 s171  s171          -15.44_2~ agb    f          2          8.38   9.43  5.76
## 2 s3499 s3499          17.25_-8~ agb    f          3         506.   106.  40.0
## 3 s3502 s3502          17.25_-8~ agb    f          3         364.   200.  76.4
## 4 s3505 s3505          17.25_-8~ agb    f          3         262.    80.1  76.4
## 5 s1525 s1525          -18.66_2~ tota~ f          2          59.5   46.7  27.1
## 6 s169  s169          -18.66_2~ agb    f          1         163.    51.1  64.3
## # i 6 more variables: sd_c <dbl>, rep_t <dbl>, rep_c <dbl>, logr <dbl>,
## #   logr_var <dbl>, logr_se <dbl>

# Aggregate all measurements (multiple years, sampling dates and plots) by experiment (and response var
npfert_responses3 <- npfert_responses2 %>%

  # suggested addition by Kevin, email 02.10.2023 10:03
  dplyr::distinct(duplicate_id, x_t, x_c, .keep_all = TRUE) |>

  filter(!is.na(logr_var) & !is.na(logr)) %>%

  # re-create ID (common ID per experiment and response variable)
  select(-id) %>%
  mutate( id = paste(exp, myvar, sep = "_XXX_")) %>%

  MAd::agg(
    id = id,
    es = logr,
    var = logr_var,
    cor = 1.0,
    method = "BHHR",
    data = .
  ) %>%

  ## to keep the output readable from the console output
  as_tibble() %>%

  # separate ID again for ease of data use
  mutate( id = str_split(id, "_XXX_") ) %>%
  mutate( exp = purrr::map_chr(id, 1),
         myvar = purrr::map_chr(id, 2) ) %>%

  ## rename again

```



```

select(exp, myvar, logr = es, logr_var = var) %>%

## add number of observations (sum of plots and repeated samplings)
left_join(
  npfert_responses2 %>%
    group_by(exp, myvar, treatment) %>%
    summarise(n_c = sum(rep_c), n_t = sum(rep_t)),
  by = c("exp", "myvar")
) %>%

## get standard error. Verify if number available observations are identical
## for ambient and elevated. Use N from control here (n_c).
mutate( logr_se = sqrt(logr_var) / sqrt(n_c) ,

# merge SLA and LMA measurements by taking inverse of logr (keep SE)
  logr = ifelse(myvar == "sla", -logr, logr),
  myvar = ifelse(myvar == "sla", "lma", myvar))

## `summarise()` has grouped output by 'exp', 'myvar'. You can override using the
## `.groups` argument.
head(npfert_responses3)

## # A tibble: 6 x 8
##   exp          myvar      logr logr_var treatment   n_c   n_t logr_se
##   <chr>        <chr>    <dbl>   <dbl>   <chr>    <dbl> <dbl>   <dbl>
## 1 -15.44_23.25_f agb      -0.118 0.195     f         4     4   0.221
## 2 17.25_-88.77_f agb       1.11 0.0634    f         9     9   0.0839
## 3 -18.66_25.5_f  total_biomass 0.242 0.0776    f         3     3   0.161
## 4 -18.66_25.5_f agb       1.01 0.287     f         8     8   0.190
## 5 19.6_-155.33_fnp bgb      0.636 0.0174    f         3     3   0.0762
## 6 -2.98_-47.52_f agb      0.245 0.000560 f         4     4   0.0118

```

Meta-analysis

Aggregate log-ratios across multiple experiments, taking into account their respective variance and using the experiment identity as a grouping factor for random intercepts.

```

out_np <- purrr::map(as.list(use_vars_np),
  ~analyse_meta(npfert_responses3 %>%
    rename(var = myvar), nam_target = .))
names(out_np) <- use_vars_np
df_box_np <- purrr::map_dfr(out_np, "df_box") |>
  left_join(
    npfert_responses3 |>
      group_by(myvar) |>
      summarise(logr_min = min(logr), logr_max = max(logr)) |>
      rename(var = myvar),
    by = "var"
  )
saveRDS(df_box_np, file = paste0(here::here(), "df_box_npfert.rds"))

```

Final data size

Number of data points (plot-level measurements) per variable:

```
npfert_responses3 %>%
  group_by(myvar) %>%
  summarise(n_plots = sum(n_c, na.rm = TRUE), n_exp = n()) %>%
  rename("Variable"="myvar", "N observations"="n_plots", "N experiments"="n_exp") %>%
  knitr::kable()
```

Variable	N observations	N experiments
agb	1439	140
anet	323	7
bgb	385	68
gs	35	2
jmax	304	5
leaf_n_area	552	30
leaf_n_mass	710	41
leaf_np	633	41
leaf_nue	230	3
leaf_p_area	527	30
leaf_p_mass	642	42
leaf_pue	230	3
lma	599	31
rd	275	3
spad	107	3
total_biomass	90	14
tpu	43	3
vcmax	304	5

Number of data points (plot-level measurements) per experiment:

```
npfert_responses3 %>%
  group_by(exp) %>%
  summarise(n_plots = sum(n_c), n_exp = n()) %>%
  rename_("Experiment"="exp", "N observations"="n_plots", "N experiments"="n_exp") %>%
  knitr::kable()
```

Experiment	N observations	N experiments
-15.44_23.25_f	4	1
-18.66_25.5_f	11	2
-2.98_-47.52_f	4	1
-21.65_21.81_f	3	1
-22.283_117.666_f	6	1
-22.41_21.71_f	4	1
-22.78_31.25_f	16	2
-23.75_31.43_f	16	2
-24.17_21.89_f	7	2
-24.4_31.75_f	16	2
-25.12_31.23_f	16	2
-25.29_31.91_f	16	2
-3.5_36_f	6	1
-3.95_-79.03_f	12	2
17.25_-88.77_f	9	1

Experiment	N observations	N experiments
19.6_-155.33_fnp	3	1
22.13_-159.63_f	11	1
26.52_109.78_fn2p	3	1
31.37_90.02_f	48	1
31.3_-81.28_f	4	1
31.42_-88.45_f	8	2
31.55_-81.78_f	4	1
32.54_-116.7_fnp	12	1
33.7_120.3_f	15	1
34.92_102.88_f2np	5	1
34.92_102.88_f3np	5	1
34.92_102.88_fnp	5	1
35.97_101.88_f	40	2
37.25_-121.75_forb_fnp	3	1
37.25_-121.75_grass_fnp	3	1
37.48_101.2_fnp	18	3
37.55_-122.3_f	3	1
37.6_101.32_fnp	20	3
37.87_-122.52_f	14	2
39.25_-121.28_fnp	20	1
41.35_36.25_fn2p	4	1
41.35_36.25_fn2p2	4	1
41.35_36.25_fn3p	4	1
41.35_36.25_fn3p2	4	1
41.35_36.25_fnp	4	1
41.35_36.25_fnp2	4	1
41.62_-71.32_fnp	8	2
42.58_122.21_fnp	12	2
44.8_116.03_fnp	24	2
51.85_5.62_fnp	3	1
52.07_5.58_fnp	3	1
52.37_5.1_fnp	6	1
52.5_5.7_fnp	6	1
53.83_-8.83_fnp	10	1
54.63_8.83_fnp	5	1
64.83_-147.72_fnp	3	1
68.2_-149.6_f	16	2
68.38_-104.54_f	4	1
69.43_-133.02_fn2p2	10	1
69.43_-133.02_fnp	10	1
9.6_-79.5_f	3	1
alpflix_fnp	60	1
bennekom_drained_fnp	5	1
bennekom_undrained_fnp	5	1
bown2007_fnp	80	8
buitengoor_1992_fnp	5	1
carswell2005_fnp	80	4
cleland2019_blldr.us_fnp	4	2
cleland2019_bnch.us_fnp	6	2
cleland2019_bogong.au_fnp	6	2
cleland2019_burrawan.au_fnp	6	2
cleland2019_cbgb.us_fnp	12	2

Experiment	N observations	N experiments
cleland2019_cdc.us_fnp	6	2
cleland2019_cdpt.us_fnp	6	2
cleland2019_cowi.ca_fnp	6	2
cleland2019_elliot.us_fnp	6	2
cleland2019_frue.ch_fnp	6	2
cleland2019_gilb.za_fnp	6	2
cleland2019_hall.us_fnp	6	2
cleland2019_hart.us_fnp	6	2
cleland2019_konz.us_fnp	6	2
cleland2019_lancaster.uk_fnp	4	2
cleland2019_look.us_fnp	6	2
cleland2019_mtca.au_fnp	6	2
cleland2019_sage.us_fnp	6	2
cleland2019_saline.us_fnp	6	2
cleland2019_sgs.us_fnp	6	2
cleland2019_shps.us_fnp	6	2
cleland2019_sier.us_fnp	6	2
cleland2019_smith.us_fnp	6	2
cleland2019_spin.us_fnp	6	2
cleland2019_summ.za_fnp	6	2
cleland2019_trel.us_fnp	6	2
cleland2019_ukul.za_fnp	12	2
cleland2019_unc.us_fnp	6	2
cleland2019_valm.ch_fnp	6	2
crous2017_fnp	2351	12
cuiliugou_f	80	1
cuiliugou_f2	72	1
cuiliugou_f3	72	1
damxung_f	30	1
damxung_f2	30	1
daqinggou_fnp	12	2
drentsche_aa_drained_fnp	5	1
drentsche_aa_wet_fnp	5	1
duolun15_fn2p	4	1
duolun15_fnp	4	1
duolun1_f	4	1
escambia_county_f	8	2
ewenke_f_np	6	1
firn2019_bldr.us_fnp	21	6
firn2019_bnch.us_fnp	51	6
firn2019_bogong.au_fnp	72	6
firn2019_burrawan.au_fnp	73	6
firn2019_cbgb.us_fnp	54	6
firn2019_comp.pt_fnp	78	6
firn2019_cowi.ca_fnp	45	6
firn2019_elliot.us_fnp	54	6
firn2019_frue.ch_fnp	46	6
firn2019_gilb.za_fnp	48	6
firn2019_hopl.us_fnp	51	6
firn2019_kiny.au_fnp	45	6
firn2019_konz.us_fnp	29	6
firn2019_lancaster.uk_fnp	51	6

Experiment	N observations	N experiments
firn2019_look.us_fnp	54	6
firn2019_mcla.us_fnp	51	6
firn2019_mtca.au_fnp	53	6
firn2019_sage.us_fnp	31	6
firn2019_sgs.us_fnp	39	6
firn2019_shps.us_fnp	24	6
firn2019_sier.us_fnp	48	6
firn2019_smith.us_fnp	54	6
firn2019_summ.za_fnp	72	6
firn2019_unc.us_fnp	54	6
firn2019_valm.ch_fnp	120	6
flottbek_fnfp	25	1
gusewell_s1_fnp	4	1
gusewell_s2_fnp	4	1
gusewell_t1_fnp	4	1
gusewell_t2_fnp	4	1
gusewell_v1_fnp	4	1
gusewell_v2_fnp	4	1
gusewell_v3_fnp	4	1
gusewell_w1_fnp	4	1
gusewell_w2_fnp	4	1
gusewell_w3_fnp	4	1
gusewell_w4_fnp	4	1
haibei_fn1p	33	1
haibei_fn1pp	33	1
haibei_fn2p	33	1
haibei_fn2pp	33	1
haibei_fn3p	33	1
haibei_fn3pp	33	1
herschgreen2024_cbgb.us_fnp	88	4
herschgreen2024_cdcr.us_fnp	70	4
herschgreen2024_churn.us_fnp	50	4
herschgreen2024_kbs.us_fnp	50	4
herschgreen2024_konz.us_fnp	69	4
herschgreen2024_spin.us_fnp	77	4
imgers_ng_2006np_fn2	6	1
imgers_ng_2006np_fn3	6	1
imgers_ng_2006np_fn4	6	1
imgers_ng_2006np_fn5	6	1
imgers_ng_2006np_fn6	6	1
imgers_ng_2006np_fp2	6	1
imgers_ng_2006np_fp3	6	1
imgers_ng_2006np_fp4	6	1
imgers_ng_2006np_fp5	6	1
imgers_ng_2006np_fp6	6	1
jingtai_f	32	1
jingtai_f2	32	1
jingtai_f3	32	1
kansasf_fnp	12	1
kansask_fnp	12	1
katelijne_2016_fnp	5	1
katelijne_2017_f	5	1

Experiment	N observations	N experiments
lunenburg_field_2006_fnp	10	1
lunenburg_field_2008_fnp	10	1
lunenburg_gh_drought_molinia_fnp	NA	2
lunenburg_gh_fert_molinia_fnp	10	1
michigan_underc_bog_fnp	19	1
michigan_underc_intermfen_fnp	17	1
michigan_underc_richfen_fnp	20	1
mo2019_fnp	200	8
mo2021_fnp	10	2
nashfield_pooled_fnp	8	2
niwot_ridge2_dm_fnp	10	1
niwot_ridge2_wm_fnp	10	1
sanpedro_fnp	6	3
schiermonnikoog_old_fn2p	12	1
schiermonnikoog_old_fn2p2	12	1
schiermonnikoog_old_fnp	12	1
schiermonnikoog_old_fnp2	12	1
schiermonnikoog_young_fn2p	12	1
schiermonnikoog_young_fn2p2	12	1
schiermonnikoog_young_fnp	12	1
schiermonnikoog_young_fnp2	12	1
shaaxi_330_fnp	3	1
shaaxi_6_fnp	3	1
tambopata_fnp	6	3
teberda_fnp	8	1
tono_fnp	6	3
toolik_acidic_1981_f	4	1
toolik_acidic_f	4	1
toolik_nonacidic_fnp	18	3
toolik_shrub_f	6	3
verryckt2022_nouragues_fnp	472	11
verryckt2022_paracou_fnp	420	11
wang2019_fnp	10	1
wayqecha_fnp	6	3
yu2022_fnp	120	5
yucatan_marsh_highsalinity_fnp	30	1
yucatan_marsh_lowsalinity_fnp	30	1
yucatan_marsh_mediumsalinity_fnp	30	1
zwarte_beek_drained_fnp	5	1
zwarte_beek_wet_fnp	5	1

Some quick plots:

```
npfert_responses3 <- npfert_responses3 %>%
  mutate(myvar = factor(myvar, levels = c("bgb", "agb", "total_biomass", "leaf_pue",
    "leaf_nue", "rd", "tpu", "jmax", "vcmax",
    "cica", "gs", "anet", "spad", "leaf_np",
    "leaf_p_area", "leaf_p_mass", "leaf_n_area",
    "leaf_n_mass", "lma")))

meta_plot_np <- ggplot(data = subset(npfert_responses3, myvar != "cica"),
```

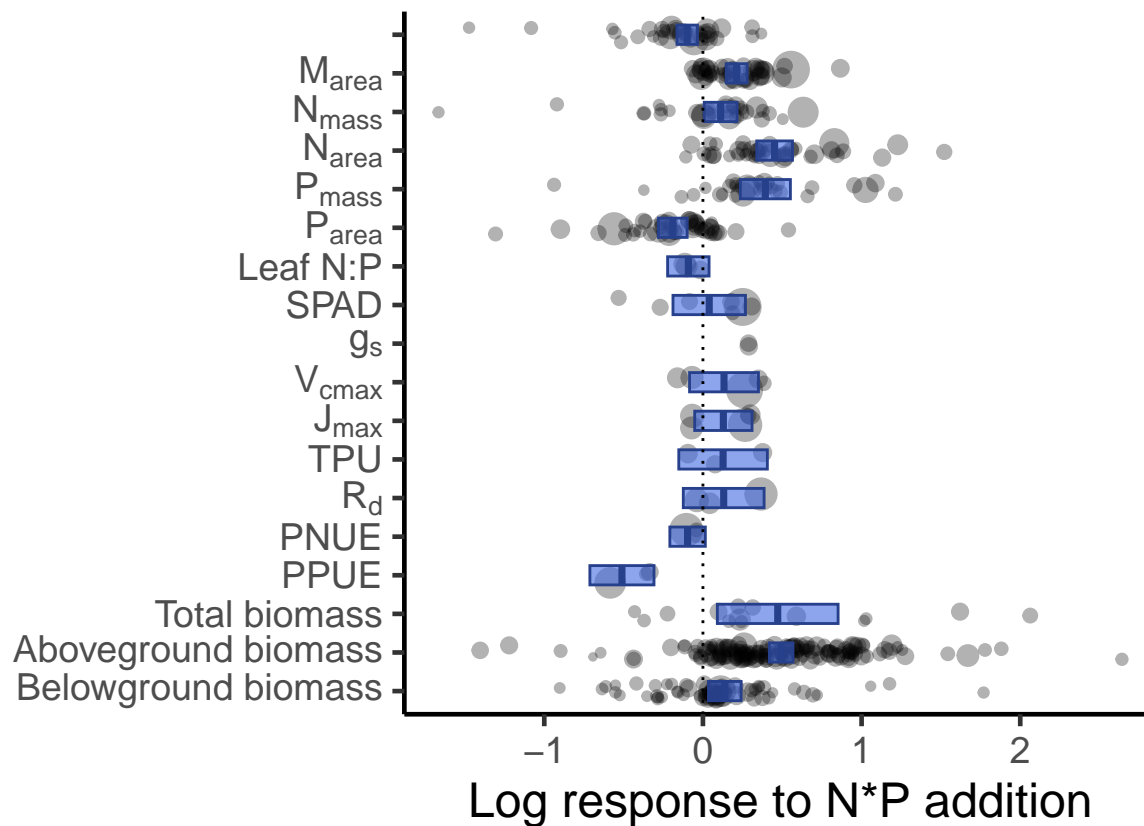
```

      aes(x = myvar, y = logr)) +
geom_jitter(color = rgb(0,0,0,0.3),
  aes( size = 1/logr_se ),
  position = position_jitter(w = 0.2, h = 0),
  show.legend = FALSE) +
geom_crossbar( data = df_box_np %>% drop_na(var),
  aes(x = var, y = middle, ymin = ymin, ymax = ymax),
  fill = "royalblue",
  color = "royalblue4",
  alpha = 0.6,
  width = 0.5 ) +
geom_hline( yintercept = 0.0, linewidth = 0.5, linetype = "dotted" ) +
scale_x_discrete(labels = c("Belowground biomass",
  "Aboveground biomass",
  "Total biomass",
  "PPUE",
  "PNUE",
  expression("R"["d"]),
  "TPU",
  expression("J"["max"]),
  expression("V"["cmax"]),
  expression("g"["s"]),
  "SPAD",
  "Leaf N:P",
  expression("P"["area"]),
  expression("P"["mass"]),
  expression("N"["area"]),
  expression("N"["mass"]),
  expression("M"["area"]))) +

#scale_x_discrete("", labels = mylabl) +
labs(x = "",
  y = "Log response to N:P addition") +
coord_flip() +
theme_classic(base_size = 18)
meta_plot_np

```

```
## Warning: Removed 2 rows containing missing values (`geom_point()`).
```



```
# Add exp type to all data frames to merge together
df_box_np$manip_type <- "np"
df_box_p$manip_type <- "p"
df_box_n$manip_type <- "n"

npfert_responses3$manip_type <- "np"
pfert_responses3$manip_type <- "p"
nfert_responses3$manip_type <- "n"

# Merge P and NP meta results
df_box_all <- df_box_n %>%
  full_join(df_box_p) %>%
  full_join(df_box_np) %>%
  mutate(manip_type = factor(manip_type, levels = c("np", "p", "n")))

## Joining with `by = join_by(var, middle, ymin, ymax, middle_scaled, ymin_scaled,
## ymax_scaled, logr_min, logr_max, manip_type)`
## Joining with `by = join_by(var, middle, ymin, ymax, middle_scaled, ymin_scaled,
## ymax_scaled, logr_min, logr_max, manip_type)`

fert_exp_responses_all <- nfert_responses3 %>%
  full_join(pfert_responses3) %>%
  full_join(npfert_responses3) %>%
  mutate(manip_type = factor(manip_type, levels = c("np", "p", "n")))

## Joining with `by = join_by(exp, myvar, logr, logr_var, treatment, n_c, n_t,
## logr_se, manip_type)`
## Joining with `by = join_by(exp, myvar, logr, logr_var, treatment, n_c, n_t,
```

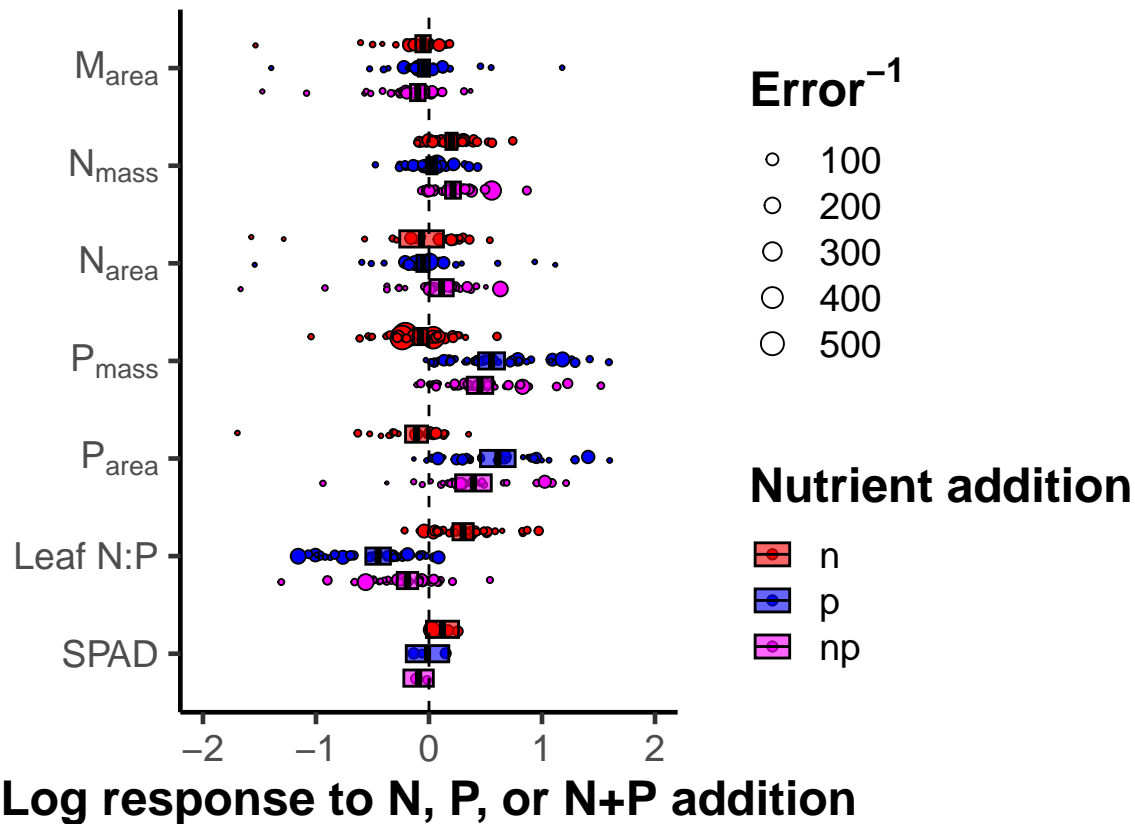


```
## logr_se, manip_type)`
head(fert_exp_responses_all)

## # A tibble: 6 x 9
##   exp      myvar    logr logr_var treatment    n_c    n_t logr_se manip_type
##   <chr>      <fct>    <dbl>    <dbl> <chr>      <dbl> <dbl>    <dbl> <fct>
## 1 -11.97_-38.12~ agb    0.0789 0.00489 f         16    16 0.0175 n
## 2 34.2_106.43_f agb    0.429 0.00271 f         10    10 0.0165 n
## 3 37.25_-121.75~ agb    0      0.00292 f          3     3 0.0312 n
## 4 37.25_-121.75~ agb   -0.121 0.00305 f          3     3 0.0319 n
## 5 38.53_-76.33_f agb    1.01  0.00544 f         35    35 0.0125 n
## 6 38.53_-76.33_f tota~ 0.465 0.000154 f         15    15 0.00320 n

# Plot nutrients. Separating by trait type to avoid plot overwhelm
meta_plot_all_leaf_nutrients <- ggplot(
  data = subset(fert_exp_responses_all,
    myvar %in% c("lma", "leaf_n_mass", "leaf_n_area", "leaf_p_mass",
      "leaf_p_area", "leaf_np", "spad")),
  aes(x = myvar, y = logr, fill = manip_type)) +
  geom_jitter(position = position_jitterdodge(jitter.width = 0.1,
    dodge.width = 0.75),
    shape = 21, aes(size = 1/logr_se)) +
  geom_crossbar(data = df_box_all %>% drop_na(var) %>%
    filter(var %in% c("lma", "leaf_n_mass", "leaf_n_area",
      "leaf_p_mass", "leaf_p_area", "leaf_np",
      "spad")),
    aes(x = var, y = middle, ymin = ymin, ymax = ymax),
    alpha = 0.6, width = 0.5,
    position = position_dodge(width = 0.75)) +
  geom_hline(yintercept = 0, linewidth = 0.5, linetype = "dashed") +
  scale_x_discrete(labels = c("SPAD",
    "Leaf N:P",
    expression("P"["area"]),
    expression("P"["mass"]),
    expression("N"["area"]),
    expression("N"["mass"]),
    expression("M"["area"]))) +
  scale_y_continuous(limits = c(-2, 2), breaks = seq(-2, 2, 1)) +
  scale_fill_manual(limits = c("n", "p", "np"),
    values = c("red", "blue", "magenta")) +
  scale_size(range = c(0.25, 4)) +
  labs(x = "",
    y = "Log response to N, P, or N+P addition",
    fill = "Nutrient addition",
    size = expression(bold("Error"^-1))) +
  coord_flip() +
  theme_classic(base_size = 18) +
  theme(legend.position = "right",
    legend.title = element_text(face = "bold"),
    axis.title.x = element_text(face = "bold"))
meta_plot_all_leaf_nutrients

## Warning: Removed 4 rows containing missing values (`geom_point()`).
```



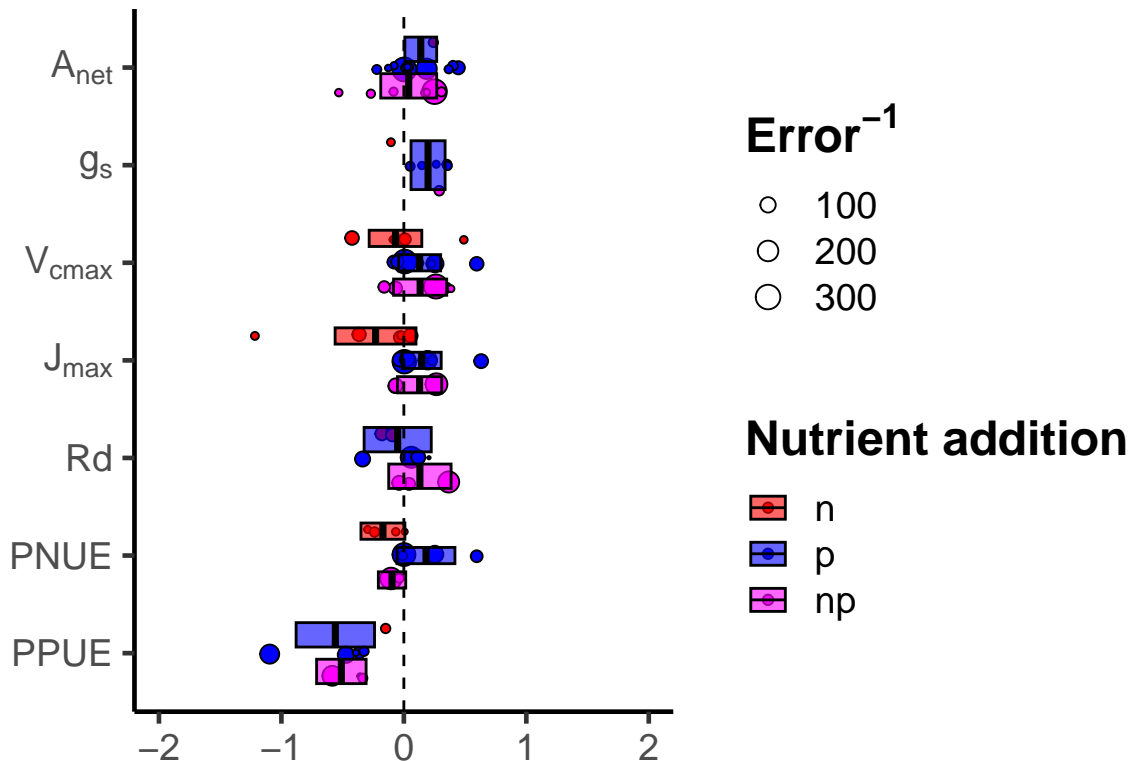
Plot photosynthetic traits. Separating by trait type to avoid plot overwhelm

```
meta_plot_all_photo <- ggplot(
  data = subset(fert_exp_responses_all,
    myvar %in% c("anet", "gs", "vcmax", "jmax",
      "rd", "leaf_nue", "leaf_pue")),
  aes(x = myvar, y = logr, fill = manip_type)) +
  geom_jitter(position = position_jitterdodge(jitter.width = 0.1,
    dodge.width = 0.75),
    shape = 21, aes(size = 1/logr_se)) +
  geom_crossbar(data = df_box_all %>% drop_na(var) %>%
    filter(var %in% c("anet", "gs", "vcmax", "jmax",
      "rd", "leaf_nue", "leaf_pue")),
    aes(x = var, y = middle, ymin = ymin, ymax = ymax),
    alpha = 0.6, width = 0.5,
    position = position_dodge(width = 0.75)) +
  geom_hline(yintercept = 0, linewidth = 0.5, linetype = "dashed") +
  scale_x_discrete(labels = c("PPUE",
    "PNUE",
    expression("Rd"),
    expression("J"["max"]),
    expression("V"["cmax"]),
    expression("g"["s"]),
    expression("A"["net"]))) +
  scale_y_continuous(limits = c(-2, 2), breaks = seq(-2, 2, 1)) +
  scale_fill_manual(limits = c("n", "p", "np"),
    values = c("red", "blue", "magenta")) +
  scale_size(range = c(0.25, 4)) +
```

```

labs(x = "",
     y = "Log response to N, P, or N+P addition",
     fill = "Nutrient addition",
     size = expression(bold("Error"-1))) +
coord_flip() +
theme_classic(base_size = 18) +
theme(legend.position = "right",
      legend.title = element_text(face = "bold"),
      axis.title.x = element_text(face = "bold"))
meta_plot_all_photo

```



Log response to N, P, or N+P addition

```

# Plot biomass traits. Separating by trait type to avoid plot overwhelm
meta_plot_all_biomass <- ggplot(
  data = subset(fert_exp_responses_all,
               myvar %in% c("bgb", "agb", "total_biomass")),
  aes(x = myvar, y = logr, fill = manip_type)) +
  geom_jitter(position = position_jitterdodge(jitter.width = 0.1,
                                             dodge.width = 0.75),
             shape = 21, aes(size = 1/logr_se)) +
  geom_crossbar(data = df_box_all %>% drop_na(var) %>%
               filter(var %in% c("bgb", "agb", "total_biomass")),
               aes(x = var, y = middle, ymin = ymin, ymax = ymax),
               alpha = 0.6, width = 0.5,
               position = position_dodge(width = 0.75)) +
  geom_hline(yintercept = 0, linewidth = 0.5, linetype = "dashed") +
  scale_x_discrete(labels = c("Belowground biomass",
                             "Aboveground biomass"),

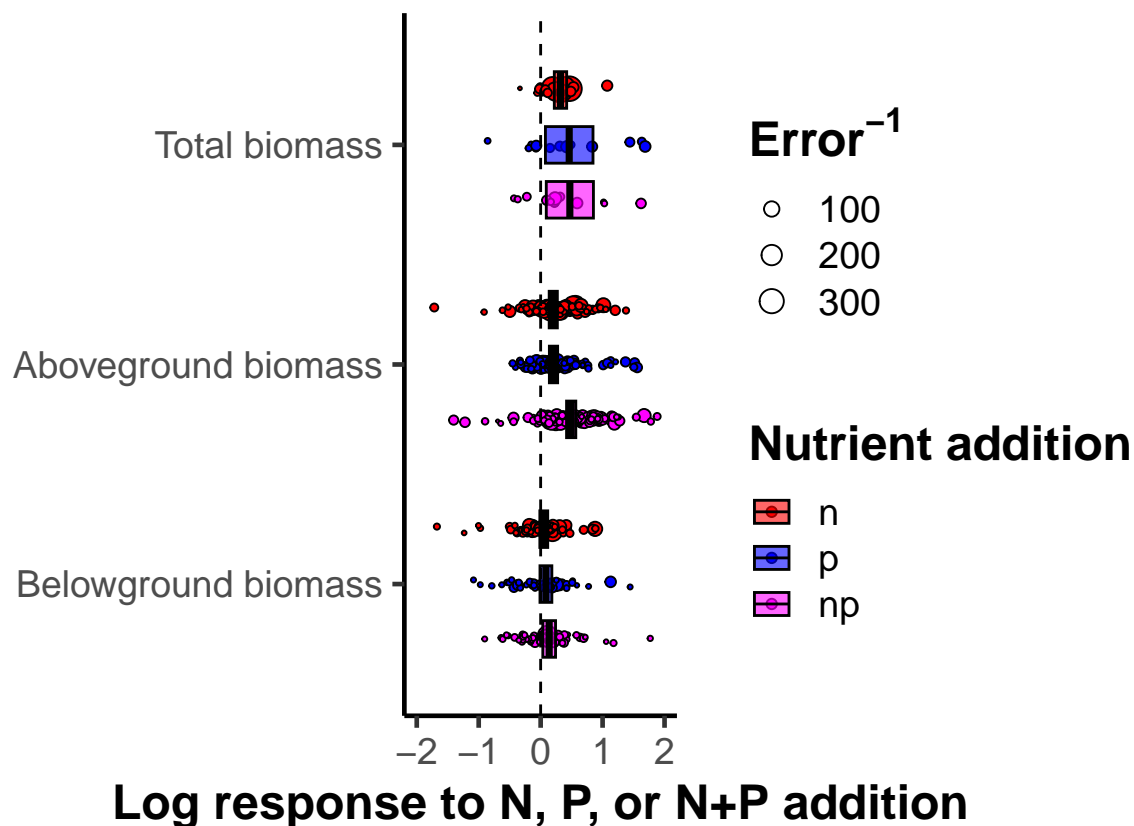
```

```

    "Total biomass")) +
scale_y_continuous(limits = c(-2, 2), breaks = seq(-2, 2, 1)) +
scale_fill_manual(limits = c("n", "p", "np"),
  values = c("red", "blue", "magenta")) +
scale_size(range = c(0.25, 4)) +
labs(x = "",
  y = "Log response to N, P, or N+P addition",
  fill = "Nutrient addition",
  size = expression(bold("Error"^-1))) +
coord_flip() +
theme_classic(base_size = 18) +
theme(legend.position = "right",
  legend.title = element_text(face = "bold"),
  axis.title.x = element_text(face = "bold"))
meta_plot_all_biomass

```

Warning: Removed 8 rows containing missing values (``geom_point()``).



Write plots

```

# N meta-analysis
# png("../plots/meta_results_n.png", width = 9, height = 6,
#   units = "in", res = 600)
# meta_plot_n
# dev.off()
#
# # P meta-analysis

```

```
png("../plots/meta_results_p.png", width = 9, height = 6,
     units = "in", res = 600)
meta_plot_p
```

```
## Warning: Removed 2 rows containing missing values (`geom_point()`).
```

```
dev.off()
```

```
## pdf
```

```
## 2
```

```
#
# # NP meta-analysis
# png("../plots/meta_results_np.png", width = 9, height = 6,
#      units = "in", res = 600)
# meta_plot_np
# dev.off()
#
# # Combine N, P, NP meta-analysis into single figure. First, leaf nutrients
# png("../plots/CNPmeta_plot_nutrients_combined.png", width = 9, height = 6,
#      units = "in", res = 600)
# meta_plot_all_leaf_nutrients
# dev.off()
#
# # Second, photosynthetic traits
# png("../plots/CNPmeta_plot_photo_combined.png", width = 9, height = 6,
#      units = "in", res = 600)
# meta_plot_all_photo
# dev.off()
#
# # Third, biomass
# png("../plots/CNPmeta_plot_biomass_combined.png", width = 9, height = 6,
#      units = "in", res = 600)
# meta_plot_all_biomass
# dev.off()
#
# # Finally, lets arrange all of the combined plots into a 3-panel figure
# png("../plots/CNPmeta_plot_all_combined.png", height = 16, width = 9,
#      units = "in", res = 600)
# ggarrange(meta_plot_all_leaf_nutrients, meta_plot_all_photo,
#            meta_plot_all_biomass, nrow = 3, ncol = 1, common.legend = TRUE,
#            legend = "right", labels = c("(a)", "(b)", "(c)", align = "hv"))
# dev.off()
```