**Global response patterns of plant functional traits to nitrogen and phosphorus addition are governed by additive interactions, climatic demand, and species acquisition strategy**

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**Abstract**

The availability of nutrients such as nitrogen and phosphorus plays a central role in shaping plant ecophysiological responses to global change. While nitrogen availability has been asserted as a key driver of plant responses to global change, the role of phosphorus – both individually and in combination with nitrogen – remains less understood due to a lack of data syntheses that precludes the development of a mechanistic framework. To address this knowledge gap, we compiled data from full-factorial nitrogen and phosphorus addition experiments across the globe and conducted a meta-analysis. We quantified the individual and interactive effects of nitrogen and phosphorus addition on leaf nutrient content, leaf photosynthetic traits, and biomass partitioning and assessed how climate and species acquisition strategy mediate these responses. Nutrient addition played a minimal role in shaping leaf-level photosynthetic traits, despite nitrogen and phosphorus addition each increasing foliar nutrient concentrations, increasing aboveground biomass, and decreasing the root:shoot ratio. Most trait responses to combined nitrogen and phosphorus addition were driven by additive interactions except for aboveground biomass, which demonstrated a stronger response to combined nitrogen and phosphorus addition than the sum of the individual nitrogen and phosphorus effects. Some trait responses were mediated by climate, with foliar nitrogen responses to nitrogen addition and foliar phosphorus responses to phosphorus addition each being strongest in regions with greater demand for soil resources. Additionally, some trait responses to nutrient addition were mediated by acquisition strategy, with nitrogen-fixing species or species with mining acquisition strategies often demonstrating dampened responses to nutrient addition than non-fixing species or species with scavenging acquisition strategies. These findings clarify the role of phosphorus in shaping plant ecophysiological trait variation, highlight possible mechanisms governing trait responses across climatic gradients and acquisition strategies, and provide a foundation for improving the representation of C-N-P interactions in terrestrial biosphere models.

**Introduction**

Terrestrial ecosystems are regulated by complex carbon, nitrogen, and phosphorus cycles. Recent terrestrial biosphere model development has focused on incorporating fully coupled nitrogen and phosphorus cycles, following from evidence that nutrient availability constrains primary productivity under present and future climates (Hungate *et al.*, 2003; LeBauer & Treseder, 2008; Wieder *et al.*, 2015; Fay *et al.*, 2015). Models with fully coupled nutrient cycles generally reduce the magnitude of the land carbon sink (Goll *et al.*, 2012; Wieder *et al.*, 2015; Braghiere *et al.*, 2022), and ensemble analyses often show stronger agreement with observations compared to carbon-only models (Kou-Giesbrecht *et al.*, 2023; Stocker *et al.*, 2025). However, land carbon sink trends still diverge across models, with some studies showing a stronger divergence among nutrient-coupled models than carbon-only models (Davies-Barnard *et al.*, 2020; Kou-Giesbrecht *et al.*, 2023; Stocker *et al.*, 2025). This model disparity is likely due to an incomplete understanding of how nutrient availability influences plant traits that drive ecosystem carbon and nutrient fluxes, highlighting the need for global data syntheses to resolve broad-scale patterns of nutrient effects on plant traits.

Nitrogen availability has long been recognized as an important driver of plant ecophysiological trait variation. Soil nitrogen availability is often positively correlated with leaf nitrogen content, while leaf nitrogen content is often positively correlated with photosynthetic capacity (Field & Mooney, 1986; Evans, 1989; Walker *et al.*, 2014; Firn *et al.*, 2019; Liang *et al.*, 2020). At the whole-plant level, increased nitrogen availability is commonly associated with greater biomass production, typically through stronger increases in aboveground than belowground biomass (Liang *et al.*, 2020; Stocker *et al.*, 2025). Leaf nitrogen-photosynthesis relationships arise because a large fraction of leaf nitrogen is allocated toward the maintenance and construction of photosynthetic enzymes (Evans & Seemann, 1989; Waring *et al.*, 2023), and is part of the reason why leaf nitrogen is often used as a key predictor of photosynthesis in terrestrial biosphere models (Rogers *et al.*, 2017). However, direct correlations between soil nitrogen availability and photosynthetic capacity are often weaker than correlations between soil nitrogen availability and leaf nitrogen content (Liang *et al.*, 2020; Waring *et al.*, 2023; Stocker *et al.*, 2025), likely because investment in photosynthetic capacity is more strongly regulated by climatic factors that influence leaf-level demand for soil resources than soil nitrogen supply (Smith *et al.*, 2019; Dong *et al.*, 2020; Peng *et al.*, 2021; Westerband *et al.*, 2023). Leaf and whole-plant responses to nitrogen availability are shaped by climate and species identity, with stronger plant responses expected under conditions that increase the leaf-level demand for soil resources (Cheaib *et al.*, 2025b; Perkowski *et al.*, 2025) and in species with nutrient acquisition strategies that rely on soil inorganic nitrogen (Cheaib *et al.*, 2025a; Perkowski *et al.*, 2025).

Phosphorus availability is also an important determinant of plant ecophysiological trait variation, though has historically garnered less attention than nitrogen. Soil phosphorus availability is often positively correlated with leaf phosphorus content, and leaf phosphorus content is often positively correlated with photosynthetic capacity (Walker *et al.*, 2014; Norby *et al.*, 2017; Firn *et al.*, 2019; Ellsworth *et al.*, 2022), leading to an increase in biomass production that stems from stronger increases in aboveground biomass than belowground biomass (Cleland *et al.*, 2019; Keller *et al.*, 2023). Leaf phosphorus-photosynthesis relationships arise because phosphorus is required to build and maintain energy transfer compounds (e.g., ATP, NADPH), substrate (e.g., Ribulose-1,5-bisphosphate), and triose phosphate compounds that play a central role in driving the Calvin-Benson cycle. However, direct relationships between soil phosphorus availability and photosynthesis are inconsistently observed (Mo *et al.*, 2019; Yu *et al.*, 2022; Hersch-Green *et al.*, 2024), a discrepancy that may reflect shifts in the fraction of leaf phosphorus allocated to metabolic versus non-metabolic pools across phosphorus gradients (Wang *et al.*, 2019; Yu *et al.*, 2022) and complicates model predictions when photosynthesis is predicted using leaf nutrients (Rogers *et al.*, 2017). As with nitrogen, climate and species identity likely play an important role in determining plant trait responses to phosphorus availability, with stronger trait responses expected in regions where demand for phosphorus is high (e.g., low temperature, high aridity, high light) or in species with acquisition strategies that increase phosphorus limitation (e.g., species that associate with N2-fixing bacteria). However, the effects of climate and species identity on leaf and whole-plant responses to phosphorus availability have not been well-documented. Broad-scale syntheses that assess phosphorus effects on leaf and whole-plant traits, both individually and interactively with nitrogen, would be useful for identifying mechanisms that drive phosphorus effects on leaf and whole-plant trait variation, and for advancing frameworks that predict carbon and nutrient fluxes in phosphorus-coupled terrestrial biosphere models.

Nitrogen and phosphorus often colimit primary productivity (Du *et al.*, 2020; Braghiere *et al.*, 2022), likely due to their complementary role in driving photosynthetic processes and biomass partitioning. As mentioned, nitrogen supports the construction and maintenance of photosynthetic enzymes, while phosphorus supplies the energy transfer compounds and substrate needed to drive the Calvin-Benson cycle. Nitrogen and phosphorus addition also each contribute to biomass production through enhanced aboveground biomass relative to belowground biomass. This complementarity suggests that combined enrichment of nitrogen and phosphorus could jointly enhance leaf nutrient content, photosynthesis, and biomass production (Niinemets & Kull, 2005; Walker *et al.*, 2014; Fay *et al.*, 2015). Indeed, some studies suggest that plants respond to combined nitrogen and phosphorus addition by increasing aboveground biomass more strongly than expected from individual effects, highlighting the likelihood that plant responses to nitrogen and phosphorus availability may be synergistic (Harpole *et al.*, 2011; Fay *et al.*, 2015; Fang *et al.*, 2024). Synergistic interactions may be particularly apparent in ecosystems where nitrogen and phosphorus strongly co-limit growth, such as tropical ecosystems (Vitousek *et al.*, 2010; Du *et al.*, 2020). Yet, plant responses to multiple environmental change factors tend to be additive (Yue *et al.*, 2017; Ding *et al.*, 2025), and existing syntheses have only quantified synergistic nitrogen and phosphorus interactions for a subset of traits (Elser *et al.*, 2007; Harpole *et al.*, 2011; Fang *et al.*, 2024; Ding *et al.*, 2025). Beyond analyses from coordinated nutrient manipulation experiments (i.e., the Nutrient Network; Firn et al., 2019; Cheaib et al., 2025b), no data syntheses to our knowledge have explicitly quantified the interactive effects of nitrogen and phosphorus on leaf nutrient content and photosynthesis, representing a key knowledge gap in understanding how nutrient interactions regulate primary productivity and the land carbon sink. Thus, broad-scale analyses that compile data from multiple experiments are needed to clarify these dynamics and to develop mechanistic frameworks for predicting how nitrogen-phosphorus interactions influence leaf and whole-plant traits.

Here, we conducted a global meta-analysis using experiments that manipulated nitrogen and phosphorus in full-factorial designs to address three primary objectives. First, we quantified the effects of nitrogen, phosphorus, and nitrogen+phosphorus addition on leaf nutrient content, net photosynthesis, photosynthetic capacity, leaf nitrogen and phosphorus use efficiency, biomass production, and biomass partitioning. Second, we quantified the interaction effect size of each trait to understand whether the effects of nitrogen+phosphorus addition were the product of additive, synergistic, or antagonistic interactions. Finally, we evaluated how climate and species identity traits (e.g., photosynthetic pathway, ability to associate with nitrogen-fixing bacteria, mycorrhizal nutrient acquisition strategy) moderate trait responses to nutrient addition. We used this meta-analytic approach and study objectives to test the following hypotheses: (1) nitrogen and phosphorus addition will increase leaf nutrient content and biomass production, with minimal impacts on investment in leaf-level photosynthesis; (2) the effects of combined nitrogen and phosphorus addition will be no different than the sum of the individual effects (i.e. these responses will be driven by additive interactions); (3) leaf and whole-plant responses to nutrient addition will be strongest in regions that amplify demand for soil resources; and (4) leaf and whole-plant responses to nutrient addition will be moderated by nutrient acquisition strategy and photosynthetic pathway.

**Materials and Methods**

*Data compilation*

Initial data for the meta-analysis were collected using citations listed in the Manipulation Experiments Synthesis Initiative (MESI) database (Van Sundert *et al.*, 2023). Manipulation experiments that added nitrogen and phosphorus in a full-factorial design were only selected in this database to ensure that any comparisons made between nitrogen, phosphorus, and nitrogen+phosphorus addition responses were from the same subset of experiments. All data for manuscripts included in the MESI database that fit these criteria were downloaded or extracted using a plot digitizer to ensure that all relevant traits were included in the meta-analysis and undergo a round of quality control to avoid any data entry issues that may arise when using large ecological datasets (Augustine *et al.*, 2024). To supplement studies included in the MESI database, studies that reported data from Nutrient Network experiments were also included in the meta-analysis, including only measurements collected from control, nitrogen, phosphorus, and nitrogen+phosphorus addition plots. Each site in each paper that reported data from Nutrient Network experiments was treated as an independent experiment, following that the Nutrient Network is a globally distributed experiment where independent sites share the same nutrient addition and experimental design scheme (Borer *et al.*, 2014). Specifically, we added leaf nutrient data from Firn et al. (2019), biomass partitioning data from Cleland et al. (2019), and photosynthetic data from Hersch-Green et al. (2024).

To supplement MESI and Nutrient Network datasets, we added additional manipulation experiments using journal articles published on or before March 2025. We selected manipulation experiments where nitrogen and phosphorus were added in a full-factorial design. From this, we selected experiments that measured traits related to leaf photosynthesis (e.g., net photosynthesis maximum rates of Rubisco carboxylation and electron transport for RuBP regeneration), leaf nutrient content (e.g., mass- or area-based leaf nitrogen content, mass- or area-based leaf phosphorus content), biomass (e.g., above-ground or belowground biomass), or biomass partitioning (e.g., root mass fraction, root:shoot ratio). Finally, we selected experiments that included explicit explanations of treatment replication schemes to accurately calculate summary statistics. We first searched for studies that followed these guidelines using citations included in the MESI and Nutrient Network papers. To supplement these studies, we also created a search query in Web of Science using similar search terms as in (Liang *et al.*, 2020). Specifically, our query mined for the following topics: (nitrogen AND phosphorus) AND (fertiliz\* OR addition) AND (effect\* OR respon\* OR affect\* OR impact\* OR increas\* OR decreas\* OR alter\* OR deposition OR enrich\*) AND (leaf nitrogen\* OR leaf phosphorus\* OR \*use efficiency OR biomass OR mass fraction OR root:shoot OR LMA OR SLA OR photosynthesis OR Vcmax OR Jmax) NOT (animal\* OR medic\* OR chemist\*).

*Data extraction*

One mean value ± standard deviation per trait per species per nutrient fertilization treatment per experimental site was considered one observation for experiments that reported results at the species level. Observations for different species from the same study were considered independent, allowing us to determine the effects of species identity traits (e.g., mycorrhizal type, photosynthetic pathway, growth form) in modifying plant responses to nutrient treatments. For experiments that reported results at the treatment level, one mean value ± standard deviation per trait per nutrient fertilization treatment per experimental site was considered one observation.

Observations were integrated into a compiled dataset through multiple pathways. First, summary statistics (mean, standard deviation, replication scheme) were calculated directly from published datasets from studies that adopted open data practices. Where possible, summary statistics were extracted from tables included in the main text or supplemental information if studies did not explicitly publish their data. If studies did not include their data or provide summary statistics in tables, we digitized plots using information about treatment and sample replication information. Plots were digitized in R (version 4.4.2) using the ‘metadigitise’ package (Pick *et al.*, 2019). Studies that did not include clear descriptions about the replication scheme were not included in the dataset. Overall, this data extraction approach rendered 4843 observations (1614 observations each for nitrogen, phosphorus, and nitrogen+phosphorus treatments) from 87 studies. Of these studies, 80 were field experiments, 6 were greenhouse experiments, and 1 was a growth chamber experiment. Of the field experiments, 168 independent sites were represented, spanning a broad global gradient with diverse climate types (Table S1). The dataset also includes data comprising 175 species from 55 families, representing diverse growth forms, growth durations, nutrient acquisition strategy, and photosynthetic pathway.

*Moderator variables*

Mean monthly temperature, precipitation, and solar radiation information spanning 1970-2000 were extracted using site latitude and longitude data for all field experiments using the Climatic Research Unit Time Series v4.09 (CRU TS v4.09) gridded data product (Harris *et al.*, 2020) downscaled to a 30 arc-second spatial resolution with WorldClim 2.1 (Fick & Hijmans, 2017). Site aridity was extracted using a complementary gridded data product (Global-AI\_PET\_v3) using the same period (1970-2000) and spatial resolution (30 arc-seconds) (Zomer *et al.*, 2022). This aridity product uses WorldClim 2.1 to calculate a monthly aridity index as a function of mean monthly precipitation per unit mean monthly potential evapotranspiration, estimating potential evapotranspiration using the Penman-Monteith approach. Given that increasing aridity index values indicate increasing wetness, we use the term “moisture index” to avoid confusion.

Mean monthly temperature (°C), mean monthly precipitation (mm), mean daily solar radiation (kJ m-2 day-1), and mean monthly moisture index (unitless) values were extracted from the grid cell containing each site using the “extract” function in the “raster” R package (Hijmans, 2010). Solar radiation values (from kJ m-2 day-1) were converted to photosynthetically active radiation (μmol m-2 s-1) assuming a conversion factor of 2.1 μmol m-2 s-1per unit W m-2. Site mean growing season temperature (*T*g; °C), growing season moisture index (*MI*g; unitless), and growing season PAR (*PAR*g; μmol m-2 s-1) were estimated using the months where mean temperature was above 0°C. All growing season climate data are reported in Table S1.

Species identity traits were included for all measurements that were collected at the species level. Specifically, we included information about species family, growth form (tree/shrub, graminoid, forb), growth duration (annual, perennial), photosynthetic pathway (C3, C4), ability to associate with N2 fixing bacteria (N2-fixer or non-fixer), and mycorrhizal type (AM, EcM, dual AM-EcM, facultative AM, ErM, and non-mycorrhizal). Mycorrhizal type was assigned from the FungalRoot database using the genus of each species (Soudzilovskaia *et al.*, 2020) and used to determine the mycorrhizal nutrient acquisition strategy following (Cheaib *et al.*, 2025a). Specifically, EcM, ErM, and dual AM-EcM species were assigned a scavenging mycorrhizal nutrient acquisition strategy while AM, facultative AM, and non-mycorrhizal species were assigned a mining mycorrhizal nutrient acquisition strategy. Ability to associate with N2-fixing bacteria was determined based on whether species were in the *Fabaceae* family.

*Determination and analysis of individual and interaction effect sizes*

We used the natural logarithm of the response ratio (ln RR) to determine the individual effects of nitrogen, phosphorus, and nitrogen+phosphorus addition on leaf and whole-plant traits. For each observation *i* (i.e., trait per species per site per experiment), we calculated the natural logarithm of the response ratio (ln RR) as:

(1)

Where is the mean value of a treatment (i.e., nitrogen, phosphorus, or nitrogen+phosphorus addition) and is the mean value of the control treatment for each observation. We determined the weighted log-response ratio () across observations as:

(2)

Where ln RRi is the log-response ratio of observation *i* given in (1), *w*i is the weight of each log-response ratio, and *k* is the total number of observations. *w*i was calculated as the inverse of the variance (*v*i) of observation *i* (that is, *w*i = 1 / *v*i). *v*i was calculated as:

(3)

Where *s*t and *s*c are the standard deviations of the treatment and control groups, respectively, and *n*t and *n*c are the sample sizes of the treatment and control groups.

Next, we used Hedge’s *d* to determine the interactive effect of nitrogen and phosphorus addition on leaf and whole-plant traits (Gurevitch *et al.*, 2000; Yue *et al.*, 2017; Ding *et al.*, 2025). For each observation *i*, the interactive effect size of nitrogen and phosphorus addition (dNPi) was calculated as:

(4)

Where ,,, and refer to the mean of the nitrogen, phosphorus, nitrogen+ phosphorus, and control treatments, respectively, for each observation *i*. *s*int\_i refers to the pooled standard deviation across treatments, calculated as:

(5)

Where *N*ci, *N*ni, *N*pi, and *N*npi refer to the sample sizes of control, nitrogen, phosphorus, and nitrogen+phosphorus treatments, respectively. *S*ci, *S*ni, *S*pi, and *S*npi refer to the sample sizes of control, nitrogen, phosphorus, and nitrogen+phosphorus treatments, respectively, for each observation. In (6), *J*int\_i refers to a correction term for small sample size bias, calculated as:

(6)

We determined the weighted interaction effect size () of each trait across experiments as:

(7)

Where *d*NPi is the interaction effect size of observation *i* given in (6), *w*dnpi is the associated weight of each interaction effect size, and *k* represents the total number of observations. *w*dnpi was calculated as the inverse of the variance (*vd*NPi) of observation *i* (that is, *w*dnpi = 1 / *vd*NPi). *vd*NPi was calculated as:

(8)

*Data analysis*

We constructed a series of mixed-effects meta-regression models to understand the individual and interactive effects of nitrogen and phosphorus addition on leaf and whole-plant traits. Three separate models were created for each trait to assess the individual effects of nitrogen, phosphorus, and nitrogen+phosphorus addition using log-response ratios and their associated variances. We created a fourth model for each trait to assess the interactive effect of nitrogen and phosphorus addition using *d*NPi values and their associated variances and weights. We also assessed the role of climate or species identity moderated the response of each trait to nitrogen, phosphorus, or nitrogen+phosphorus addition by including *T*g, *MI*g, *PAR*g, photosynthetic pathway (C3, C4), N2-fixation ability (N2-fixer, non-fixer), and mycorrhizal nutrient acquisition strategy (mining, scavenging) as moderator variables. We also assessed the role of climate and species identity on the interaction effect size of each trait. In all cases, we built mixed-effects meta-regression models using the ‘rma.mv’ function in the ‘metafor’ R package (Viechtbauer, 2010), manually specifying the weights of each observation as explained above, fitting each model using restricted maximum likelihood estimation, and including experiment as a random intercept term. We used the ‘orchaRd’ R package to assess and visualize moderator effects (Nakagawa *et al.*, 2023). We could not assess the role of climate in modifying leaf photosynthetic traits, as there was not enough data collected from field experiments to test patterns with confidence.

Interactions between nitrogen and phosphorus addition on leaf and whole-plant traits were classified into three categories: additive, synergistic, and antagonistic interactions. Null interaction effects (i.e. the 95% confidence intervals overlapped with zero) were classified as additive interactions, where the combined effect of nitrogen and phosphorus addition had similar effects as the sum of the individual effects of nitrogen and phosphorus addition. An interaction was classified as synergistic (i.e. the combined effect of nitrogen and phosphorus addition was stronger than predicted through individual effects) if positive individual effects of nitrogen and phosphorus addition corresponded with a significant positive interaction effect, if negative individual effects of nitrogen and phosphorus addition correspond with a significant negative interaction effect, or if mixed sign individual effects (e.g., one positive and one negative effect) corresponded with a significant negative interaction effect. An interaction was classified as antagonistic (that is, the combined effect of nitrogen and phosphorus addition was weaker than predicted through individual effects) if positive individual effects of nitrogen and phosphorus addition corresponded with a significant negative interaction effect, if negative individual effects of nitrogen and phosphorus addition corresponded with a significant positive interaction effect, or if mixed sign individual effects corresponded with a significant positive interaction effect.

All plots and statistics were completed using R version 4.4.2 (R Core Team, 2021). Plots visualize individual and interaction effects for each response variable as the percentage change between control and treatment groups, calculated as .

**Results**

*Nitrogen addition*

Nitrogen addition increased *N*mass and *N*area by 13.2% and 13.4%, respectively, but decreased *M*area by 3.6% and *P*mass by 7.2% and had no effect on *P*area (Table S2; Fig. 2a). Photosynthetic traits were not modified by nitrogen addition (Table S3; Fig 2b). However, nitrogen addition increased aboveground biomass by 38.5% without affecting belowground biomass, leading to a 14.6% decrease in the root mass fraction and 28.9% decrease root:shoot ratio (Table S4; Fig. 2c).

Individual effects of nitrogen addition were modified by climate. The positive effects of nitrogen addition on *N*mass and *N*area diminished with increasing growing season temperature, though this response was stronger for *N*mass than *N*area (Fig. 4a; Table S6). Greater temperatures were also associated with a reduction in the response ratio that explained the effect of nitrogen addition on total biomass despite no general nitrogen addition effect on total biomass (Table S7). Increasing growing season moisture index also diminished the positive effects of nitrogen addition on *N*mass and *N*area, with temperature playing a stronger role in modifying *N*mass responses than *N*area (Fig 4b; Table S6). Finally, increased growing season light availability weakened the negative effect of nitrogen addition on the root mass fraction (Table S7). Greater light availability was also associated with an increase in the response ratio that explained the effect of nitrogen addition on *P*area, despite no general nitrogen addition effect on *P*area (Table S6).

Species identity traits modified some responses to nitrogen addition (Tables S8-S10). For example, positive effects of nitrogen addition on *N*mass were only observed in species that could not associate with N2-fixing bacteria and were generally stronger in C4 species compared to C3 species, though responses did not differ between mining and scavenging mycorrhizal acquisition strategies (Fig 4c-e; Tables S8-S10).

*Phosphorus addition*

Phosphorus addition yielded a 56.7% and 71.2% increase in *P*mass and *P*area, respectively, with no effect detected for *M*area, *N*mass, or *N*area (Table S2; Fig 2d). Phosphorus addition increased *J*max by 19.4%, but there was no effect detected for any other photosynthetic trait (Table S2; Fig 2d). Phosphorus addition increased aboveground biomass by 21.1% without affecting belowground biomass, yielding a 16.8% increase in total biomass and 20.3% reduction in the root:shoot ratio (Table S4; Fig. 2f).

Climate shaped several trait responses to phosphorus addition. Increasing growing season temperature diminished the positive effects of phosphorus addition on *P*mass and *P*area, though this response was stronger for *P*mass than *P*area (Fig. 4f; Table S6). Greater growing season temperatures also slightly diminished the positive effect of phosphorus addition on total biomass (Table S6). Increasing growing season moisture index slightly weakened the positive effects of phosphorus addition on *P*mass (Fig. 4g; Table S6), though this did not scale to modify the phosphorus addition effect on *P*area (Table S6). Interestingly, increasing growing season moisture index strengthened the negative effect of phosphorus addition on the root mass fraction and increased the response ratio that explained the effect of phosphorus addition on *M*area despite no general phosphorus addition effect on *M*area (Table S6-7).

Species identity also modified some plant responses to phosphorus addition. For example, positive effects of phosphorus addition on *P*mass and *J*max were stronger in species that could associate with N2-fixing bacteria than those that could not, in species with mining rather than scavenging mycorrhizal strategies, and in C3 compared to C4 species (Fig. 4h-j; Tables S8-S10).

*Nitrogen + phosphorus addition*

Nitrogen+phosphorus addition increased *N*mass, *N*area, *P*mass, and *P*area by 12.5%, 16.6%, 44.1%, and 47.6%, respectively, and decreased *M*area by 5% (Fig. 2g). Nitrogen+phosphorus addition increased *A*sat by 27.9% through a stronger increase in *J*max (29.9% increase) than *V*cmax (marginal 17.9% increase; Table SX; Fig. 2h). Nitrogen addition increased total biomass by 46% through an 87.2% increase in aboveground biomass and no change in belowground biomass, leading to a 13.7% decrease in the root mass fraction and 33% decrease in the root:shoot ratio (Table SX; Fig 2i).

As with nitrogen and phosphorus addition, climate shaped some plant responses to nitrogen+phosphorus addition. Increasing growing season temperature diminished the positive effects of nitrogen+phosphorus addition on *N*mass and *P*mass, with weak diminishing positive effects observed for total biomass (Table S6-S7). Growing season moisture index had no effect on trait responses to nitrogen+phosphorus addition except for *M*area, where increasing moisture index values diminished the negative effect of nitrogen+phosphorus on *M*area (Table S6). Increasing growing season light availability enhanced the positive effect of nitrogen+phosphorus addition on *P*mass and *P*area and increased the response ratio that explained the effect of nitrogen+phosphorus addition on belowground biomass despite no detectable general effect of nitrogen+phosphorus addition (Table S6-S7).

Species identity modifies some plant response to nitrogen+phosphorus addition. For example, positive effects of nitrogen+phosphorus addition on *N*mass were only observed in plants that could not associate with N2-fixing bacteria and only in C3 species, but were similar between mycorrhizal nutrient acquisition strategies (Tables S8-S10). In contrast, *P*mass responses to nitrogen+phosphorus addition were stronger in species that could associate with N2-fixing bacteria than species that could not and in species with scavenging mycorrhizal acquisition strategies compared to mining acquisition strategies, but were not modified by photosynthetic pathway (Tables S8-S10).

*Interaction effects*

Interaction effect sizes indicated that all trait responses to nitrogen+phosphorus addition except for aboveground biomass were the product of additive interactions (Table S5; Fig. 3). Aboveground biomass demonstrated a synergistic interaction to nitrogen and phosphorus addition, where there was a stronger effect of nitrogen+phosphorus addition on aboveground (87.2% increase; Table S4) than predicted through the sum of individual nitrogen (38.5% increase; Table S4) and phosphorus (21.1% increase; Table S4) addition effects.

Climate did not modify traits for which there was an additive interaction. However, the interaction effect size for aboveground biomass decreased with increasing growing season temperature and increasing moisture index (Table SX; Fig. 4a-b).

**Figure 2**

A collage of graphs

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**Figure 2**. Individual effects of nitrogen addition (top row of panels), phosphorus addition (middle row of panels), and nitrogen+phosphorus addition (bottom row of panels) on traits related to leaf nutrient content (left column of panels), leaf photosynthesis (middle column of panels), and biomass partitioning (right column of panels). Large points indicate the meta-analytic mean (represented as % change, calculated as), error bars indicate 95% confidence intervals, and sample sizes are reported in parentheses. Significant treatment effects are noted with asterisks next to sample sizes (based on *Z*-score; \*: *p*<0.05, \*\*: *p*<0.01; \*\*\*: *p*<0.001). Key: *M*area=leaf biomass per unit leaf area, *N*mass=leaf nitrogen content per unit leaf biomass, *N*area=leaf nitrogen content per unit leaf area, *P*mass=leaf phosphorus content per unit leaf biomass, *P*area=leaf phosphorus content per unit leaf area, *A*sat=light-saturated net photosynthesis rate, *V*cmax=maximum rate of Rubisco carboxylation, *J*max=maximum rate of electron transport for RuBP regeneration, PNUE=photosynthetic nitrogen-use efficiency, PPUE=photosynthetic phosphorus-use efficiency, AGB=aboveground biomass, BGB=belowground biomass, RMF=root mass fraction, root:shoot=ratio of root biomass to shoot biomass.

**Figure 3**

A graph of a number of individuals

AI-generated content may be incorrect.

**Figure 3**. Interaction effects of nitrogen and phosphorus addition on traits related to leaf nutrient content (a), leaf photosynthesis (b), and biomass partitioning (c). Large points indicate the meta-analytic mean (represented as % change, calculated as ), error bars indicate 95% confidence intervals, and sample sizes are reported in parentheses. Black colored points indicate additive interactions while red colored points indicate synergistic interactions. Significant interaction effects are also noted with asterisks next to sample sizes (based on *Z*-score; \*: *p*<0.05). All acronyms are as explained in Figure 2.

*Moderator effects on plant responses to nutrient addition*

Climate and species identity modified plant responses to nitrogen+phosphorus addition. Increasing growing season temperature decreased the effect of nitrogen+phosphorus addition on *N*mass, but there was no effect of moisture index (Table S6). Additionally, *N*mass responses to nitrogen+phosphorus addition were only observed in species that could not form associations with nitrogen-fixing bacteria (Table S8), did not differ between mycorrhizal acquisition strategies (Table S9), and were stronger in C4 species compared to C3 species (Table S10). Increasing growing season temperature did not modify the effect of nitrogen+phosphorus addition on *P*mass, though increasing moisture index decreased the effect of nitrogen+phosphorus on *P*mass (Table S6). *P*mass responses to nitrogen+phosphorus addition were stronger in species that could form associations with nitrogen-fixing bacteria compared to species that could not form these associations (Table S8), in scavenging strategies compared to mining strategies (Table S9), and in C3 species compared to C4 species (Table S10).

All other nitrogen, phosphorus, nitrogen+phosphorus addition, and interaction responses to moderator variables are explained in the *Supplement* (Text S1; Table S6-14).

**Figure 4**

**A collage of graphs

AI-generated content may be incorrect.**

**Figure 4** Effects of growing season climate and species identity on mass-based leaf nitrogen responses to nitrogen addition (top row of panels) and leaf phosphorus responses to phosphorus addition (bottom row of panels). From left to right, the top row of panels describes the growing season temperature (a), growing season moisture index (b), ability to associate with nitrogen fixing bacteria (c), mycorrhizal nutrient acquisition strategy (d), and photosynthetic pathway effect on the leaf nitrogen content response to nitrogen addition. The bottom row of panels describes the growing season temperature (f), growing season moisture index (g), ability to associate with nitrogen fixing bacteria (h), mycorrhizal nutrient acquisition strategy (i), and photosynthetic pathway (j) effect on the leaf phosphorus content response to phosphorus addition. For climate moderator plots, grey points indicate individual data points. Solid trendlines are drawn if climate significantly modified leaf nutrient responses to nutrient addition (*p*<0.05) and dashed trendlines are drawn if climate marginally modified leaf nutrient responses to nutrient addition (*p*<0.1). For species identity traits, large points indicate the meta-analytic mean (represented as % change, calculated as ) and error bars indicate 95% confidence intervals, with significant pairwise differences noted with asterisks above brackets (based on *Z*-score; \*: *p*<0.05, \*\*: *p*<0.01; \*\*\*: *p*<0.001).

**Discussion**

We compiled a global dataset of full-factorial nitrogen and phosphorus addition experiments to evaluate how nitrogen and phosphorus enrichment alters leaf and whole-plant functional traits. Consistent with our first hypothesis, nitrogen addition, phosphorus addition, and nitrogen+ phosphorus addition primarily influenced leaf nutrient content and biomass production, with limited effects on leaf-level photosynthetic traits. Most trait responses to combined nitrogen+ phosphorus addition were driven by additive interactions, supporting our second hypothesis. However, aboveground biomass demonstrated a synergistic interaction, with stronger increases under nitrogen+phosphorus addition than predicted through the sum of the individual nitrogen and phosphorus effects. Trait responses to nutrient addition were generally strongest in regions with greater demand for soil resources, in line with our third hypothesis. Finally, trait responses were moderated by species identity traits such as photosynthetic pathway, ability to associate with N2-fixing bacteria, and mycorrhizal acquisition strategy, supporting our fourth hypothesis. Overall, these findings are consistent with previous work showing limited impacts of nutrient addition on leaf-level photosynthesis (Liang *et al.*, 2020; Waring *et al.*, 2023; Stocker *et al.*, 2025), stronger plant responses in regions with high climatic demand for soil resources (Cheaib *et al.*, 2025b) and responses to nutrient addition that are context dependent on photosynthetic pathway (Liang *et al.*, 2020) and acquisition strategy (Perkowski *et al.*, 2021, 2025; Cheaib *et al.*, 2025a). Collectively, these findings provide the groundwork for mechanistic frameworks that could be used to incorporate more realistic C-N-P cycle interactions in terrestrial biosphere models.

*Nitrogen and phosphorus addition play a stronger role in modifying leaf nutrient content and whole-plant traits than photosynthetic traits*

Leaf nutrient responses to nitrogen and phosphorus additions were generally stronger in magnitude than photosynthetic responses. Specifically, nitrogen addition played no role in modifying photosynthetic traits, while phosphorus addition weakly increased *J*max and had no effect on any other photosynthetic trait. Despite this, we observed strong positive effects of nitrogen addition on leaf nitrogen content and phosphorus addition on leaf phosphorus content. This decoupling between leaf nutrients and photosynthesis implies that nutrient addition may have decreased the relative fraction of leaf nurtrients allocated to photosynthesis and therefore increased the fraction of nutrients allocated to non-photosynthetic pools (e.g., structure). These changes in the fractional pool of leaf nutrients allocated to photosynthesis has been observed in response to nitrogen addition (Waring *et al.*, 2023; Perkowski *et al.*, 2025), but not phosphorus, in part because determining fractional pools of leaf phosphorus beyond metabolic processes remains a challenge. Regardless, these patterns contrast the current formulation of photosynthesis in many nutrient-coupled models, where photosynthetic capacity is often predicted as a function of leaf nitrogen and phosphorus content across environmental gradients (Achat *et al.*, 2016; Rogers *et al.*, 2017; Stocker *et al.*, 2025). Our results suggest that static leaf nutrient-photosynthesis relationships may lead to an overestimation of the terrestrial carbon sink, as these static relationships will likely overestimate leaf-level photosynthesis and whole plant productivity in environments with elevated nutrient inputs.

Additionally, whole-plant responses to nutrient additions were generally stronger than photosynthetic responses to nutrient additions. These responses seem paradoxical on the surface, as increased biomass accumulation cannot occur without an increase in carbon assimilation. However, these patterns could be driven by optimized resource allocation to photosynthetic enzymes that maximizes resource allocation to whole-plant growth, following patterns expected from eco-evolutionary optimality theory (Stocker *et al.*, 2025; Perkowski *et al.*, 2025). Eco-evolutionary optimality theory predicts that investment in photosynthetic enzymes is determined by photosynthetic demand for soil resources associated with climate (Smith *et al.*, 2019, 2024; Harrison *et al.*, 2021; Stocker *et al.*, 2025; Perkowski *et al.*, 2025), where changes in soil resource supply should only increase photosynthesis when demand remains unsatisfied (Stocker *et al.*, 2025). The theory posits that plants should allocate excess nutrients not needed to satisfy leaf-level demand toward the construction of additional leaves, leading to a stronger increase in total leaf area and whole-plant productivity than leaf-level photosynthesis. These responses indicate that limited photosynthetic responses to nutrient addition may be attributed to leaf-level nutrient demand for photosynthetic enzymes being satisfied, leading to stronger investment in nutrients toward the construction of additional optimally coordinated leaves.vTo date, support for this theory has primarily been observed across nitrogen availability gradients and nitrogen manipulation experiments (Dong *et al.*, 2017, 2020, 2022; Liang *et al.*, 2020; Paillassa *et al.*, 2020; Stocker *et al.*, 2025; Perkowski *et al.*, 2025), though some work indicates that these patterns may also apply across phosphorus availability gradients (Westerband *et al.*, 2023). Our findings suggest that these same optimization patterns may also apply phosphorus manipulation experiments, a key result given recent efforts to incorporate eco-evolutionary optimality (Harrison *et al.*, 2021; Smith *et al.*, 2024; Ren *et al.*, 2025) and the phosphorus cycle (Allen *et al.*, 2020; Braghiere *et al.*, 2022) into terrestrial biosphere models.

*Plant responses to* *nitrogen+phosphorus addition are largely governed by additive interactions*

A central focus of this meta-analysis was to assess whether plant responses to nitrogen+ phosphorus addition were driven by additive, synergistic, or antagonistic interactions. Existing syntheses suggest that ecosystem and plant responses to multiple global change factors are often driven by additive interactions (Yue *et al.*, 2017; Ding *et al.*, 2025). Except for a recent meta-analysis showing that aboveground biomass responds synergistically to nitrogen+phosphorus addition (Fang *et al.*, 2024), global syntheses have rarely addressed the interactive effects of nitrogen and phosphorus addition on leaf nutrient, photosynthetic, or whole-plant traits. Our analysis shows that leaf and whole-plant responses to nitrogen+phosphorus addition were indeed largely driven by additive interactions. This was the case for all traits except for aboveground biomass, which demonstrated a synergistic interaction. These findings indicate that most plant trait responses to nitrogen+phosphorus addition are no different than the sum of the individual effects of nitrogen and phosphorus addition, with stronger effects of nitrogen+phosphorus addition on aboveground biomass than the sum of its individual effects.

Our results revealed a synergistic interaction that defined the aboveground biomass response to nitrogen+phosphorus addition, supporting findings from previous work (Fay *et al.*, 2015; Fang *et al.*, 2024)

*Plant responses to nutrient additions are mediated through climatic demand and species identity*

Following expectations, trait responses to nutrient addition were strongest under climatic conditions where photosynthetic demand was highest. Specifically, the effects of nitrogen addition on *N*mass and *N*area were strongest in colder, drier climates, as indicated by a negative effect of increasing temperature and moisture index on the responses of *N*mass and *N*area to nitrogen addition. Similar patterns were observed with phosphorus, where the effects of phosphorus addition on *P*mass and *P*area were strongest in colder and drier climates due to negative effects of increasing temperature and moisture index on the responses of *P*mass and *P*area to phosphorus addition.

Additionally, leaf nutrient responses to n

*Future research needs and directions*

Unfortunately, we could not investigate the role of climate in modulating photosynthetic responses to nitrogen and phosphorus addition. This constraint was due to the limited number of full-factorial nitrogen and phosphorus experiments that are conducted in the field, representing a clear future area of needed research and key knowledge gap that remains in our understanding of how photosynthetic processes respond to nutrient additions. Previous work has shown that climatic factors which influence demand play a predictable and key role in determining leaf nitrogen allocation responses to nitrogen and phosphorus addition (Cheaib et al., 2025). Our work supports these findings by showing that leaf nutrient allocation responses to nitrogen and phosphorus addition are dependent on climate and associated demands for soil resources. However, similar field analyses that scale these patterns to photosynthetic traits remain lacking. Quantifying these responses is particularly important, as recent work has highlighted that the fraction of leaf nutrients (nitrogen in this case) allocated to photosynthetic tissues decrease in response to increasing nitrogen availability and are dependent on climate-related demand for soil resources (Waring et al., 2023; Cheaib et al., 2025; Perkowski et al., 2025). Without field experiments that quantify leaf photosynthetic responses to nutrient treatments, we unable to comment on whether these responses scale with leaf nutrient allocation responses in ways that are predicted through eco-evolutionary optimality.

**References**

**Achat DL, Augusto L, Gallet-Budynek A, Loustau D**. **2016**. Future challenges in coupled C–N–P cycle models for terrestrial ecosystems under global change: a review. *Biogeochemistry* **131**: 173–202.

**Allen K, Fisher JB, Phillips RP, Powers JS, Brzostek ER**. **2020**. Modeling the carbon cost of plant nitrogen and phosphorus uptake across temperate and tropical forests. *Frontiers in Forests and Global Change* **3**: 1–12.

**Augustine SP, Bailey-Marren I, Charton KT, Kiel NG, Peyton MS**. **2024**. Improper data practices erode the quality of global ecological databases and impede the progress of ecological research. *Global Change Biology* **30**: 1–11.

**Borer ET, Harpole WS, Adler PB, Lind EM, Orrock JL, Seabloom EW, Smith MD**. **2014**. Finding generality in ecology: A model for globally distributed experiments. *Methods in Ecology and Evolution* **5**: 65–73.

**Braghiere RK, Fisher JB, Allen K, Brzostek ER, Shi M, Yang X, Ricciuto DM, Fisher RA, Zhu Q, Phillips RP**. **2022**. Modeling global carbon costs of plant nitrogen and phosphorus acquisition. *Journal of Advances in Modeling Earth Systems* **14**: 1–23.

**Cheaib A, Chieppa J, Perkowski EA, Smith NG**. **2025a**. Soil resource acquisition strategy modulates global plant nutrient and water economics. *New Phytologist* **246**: 1536–1553.

**Cheaib A, Waring EF, McNellis R, Perkowski EA, Martina JP, Seabloom EW, Borer ET, Wilfahrt PA, Dong N, Prentice IC, *et al.*** **2025b**. Soil Nitrogen Supply Exerts Largest Influence on Leaf Nitrogen in Environments with the Greatest Leaf Nitrogen Demand. *Ecology Letters* **28**: 1–13.

**Cleland EE, Lind EM, DeCrappeo NM, DeLorenze E, Wilkins RA, Adler PB, Bakker JD, Brown CS, Davies KF, Esch E, *et al.*** **2019**. Belowground Biomass Response to Nutrient Enrichment Depends on Light Limitation Across Globally Distributed Grasslands. *Ecosystems* **22**: 1466–1477.

**Davies-Barnard T, Meyerholt J, Zaehle S, Friedlingstein P, Brovkin V, Fan Y, Fisher RA, Jones CD, Lee H, Peano D, *et al.*** **2020**. Nitrogen cycling in CMIP6 land surface models: progress and limitations. *Biogeosciences* **17**: 5129–5148.

**Ding B, Xu D, Wang S, Liu W, Zhang Q**. **2025**. Additive Effects of Multiple Global Change Drivers on Terrestrial Nitrogen Cycling Worldwide. *Global Change Biology* **31**: 1–16.

**Dong N, Prentice IC, Evans BJ, Caddy-Retalic S, Lowe AJ, Wright IJ**. **2017**. Leaf nitrogen from first principles: field evidence for adaptive variation with climate. *Biogeosciences* **14**: 481–495.

**Dong N, Prentice IC, Wright IJ, Evans BJ, Togashi HF, Caddy-Retalic S, McInerney FA, Sparrow B, Leitch E, Lowe AJ**. **2020**. Components of leaf‐trait variation along environmental gradients. *New Phytologist* **228**: 82–94.

**Dong N, Prentice IC, Wright IJ, Wang H, Atkin OK, Bloomfield KJ, Domingues TF, Gleason SM, Maire V, Onoda Y, *et al.*** **2022**. Leaf nitrogen from the perspective of optimal plant function. *Journal of Ecology* **110**: 2585–2602.

**Du E, Terrer C, Pellegrini AFA, Ahlström A, van Lissa CJ, Zhao X, Xia N, Wu X, Jackson RB**. **2020**. Global patterns of terrestrial nitrogen and phosphorus limitation. *Nature Geoscience* **13**: 221–226.

**Ellsworth DS, Crous KY, De Kauwe MG, Verryckt LT, Goll D, Zaehle S, Bloomfield KJ, Ciais P, Cernusak LA, Domingues TF, *et al.*** **2022**. Convergence in phosphorus constraints to photosynthesis in forests around the world. *Nature Communications* **13**: 5005.

**Elser JJ, Bracken MES, Cleland EE, Gruner DS, Harpole WS, Hillebrand H, Ngai JT, Seabloom EW, Shurin JB, Smith JE**. **2007**. Global analysis of nitrogen and phosphorus limitation of primary producers in freshwater, marine and terrestrial ecosystems. *Ecology Letters* **10**: 1135–1142.

**Evans JR**. **1989**. Photosynthesis and nitrogen relationships in leaves of C3 plants. *Oecologia* **78**: 9–19.

**Evans JR, Seemann JR**. **1989**. The allocation of protein nitrogen in the photosynthetic apparatus: costs, consequences, and control. *Photosynthesis* **8**: 183–205.

**Fang J, Chen B, Wang F, Li W, Zhang H, Fang J, Liu S, Zheng Z, Guo M, Niu S**. **2024**. Nitrogen, phosphorus, and potassium co‐limitation in terrestrial ecosystems: A global meta‐analysis. *Plants, People, Planet* **6**: 1329–1340.

**Fay PA, Prober SM, Harpole WS, Knops JMH, Bakker JD, Borer ET, Lind EM, MacDougall AM, Seabloom EW, Wragg PD, *et al.*** **2015**. Grassland productivity limited by multiple nutrients. *Nature Plants* **1**: 15080.

**Fick SE, Hijmans RJ**. **2017**. WorldClim 2: new 1‐km spatial resolution climate surfaces for global land areas. *International Journal of Climatology* **37**: 4302–4315.

**Field CB, Mooney HA**. **1986**. Photosynthesis--nitrogen relationship in wild plants. In: On the Economy of Plant Form and Function: Proceedings of the Sixth Maria Moors Cabot Symposium, Evolutionary Constraints on Primary Productivity, Adaptive Patterns of Energy Capture in Plants, Harvard Forest, August 1983. Cambridge [Cambridgeshire]: Cambridge University Press, c1986.

**Firn J, McGree JM, Harvey E, Flores-Moreno H, Schütz M, Buckley YM, Borer ET, Seabloom EW, La Pierre KJ, MacDougall AM, *et al.*** **2019**. Leaf nutrients, not specific leaf area, are consistent indicators of elevated nutrient inputs. *Nature Ecology & Evolution* **3**: 400–406.

**Goll DS, Brovkin V, Parida BR, Reick CH, Kattge J, Reich PB, van Bodegom PM, Niinemets Ü**. **2012**. Nutrient limitation reduces land carbon uptake in simulations with a model of combined carbon, nitrogen and phosphorus cycling. *Biogeosciences* **9**: 3547–3569.

**Gurevitch J, Morrison JA, Hedges L V**. **2000**. *The Interaction between Competition and Predation: A Meta-analysis of Field Experiments*.

**Harpole WS, Ngai JT, Cleland EE, Seabloom EW, Borer ET, Bracken MES, Elser JJ, Gruner DS, Hillebrand H, Shurin JB, *et al.*** **2011**. Nutrient co‐limitation of primary producer communities. *Ecology Letters* **14**: 852–862.

**Harris I, Osborn TJ, Jones P, Lister D**. **2020**. Version 4 of the CRU TS monthly high-resolution gridded multivariate climate dataset. *Scientific Data* **7**.

**Harrison SP, Cramer W, Franklin O, Prentice IC, Wang H, Brännström Å, de Boer H, Dieckmann U, Joshi J, Keenan TF, *et al.*** **2021**. Eco-evolutionary optimality as a means to improve vegetation and land-surface models. *New Phytologist* **231**: 2125–2141.

**Hersch-Green EI, Fay PA, Hass HB, Smith NG**. **2024**. Mechanistic insights into plant community responses to environmental variables: genome size, cellular nutrient investments, and metabolic tradeoffs. *New Phytologist*.

**Hijmans RJ**. **2010**. raster: Geographic Data Analysis and Modeling. *CRAN: Contributed Packages*.

**Hungate BA, Dukes JS, Shaw MR, Luo Y, Field CB**. **2003**. Nitrogen and climate change. *Science* **302**: 1512–1513.

**Keller AB, Walter CA, Blumenthal DM, Borer ET, Collins SL, DeLancey LC, Fay PA, Hofmockel KS, Knops JMH, Leakey ADB, *et al.*** **2023**. Stronger fertilization effects on aboveground versus belowground plant properties across nine U.S. grasslands. *Ecology* **104**.

**Kou-Giesbrecht S, Arora VK, Seiler C, Arneth A, Falk S, Jain AK, Joos F, Kennedy D, Knauer J, Sitch S, *et al.*** **2023**. Evaluating nitrogen cycling in terrestrial biosphere models: a disconnect between the carbon and nitrogen cycles. *Earth System Dynamics* **14**: 767–795.

**LeBauer DS, Treseder KK**. **2008**. Nitrogen limitation of net primary productivity in terrestrial ecosystems is globally distributed. *Ecology* **89**: 371–379.

**Liang X, Zhang T, Lu X, Ellsworth DS, BassiriRad H, You C, Wang D, He P, Deng Q, Liu H, *et al.*** **2020**. Global response patterns of plant photosynthesis to nitrogen addition: A meta‐analysis. *Global Change Biology* **26**: 3585–3600.

**Mo Q, Li Z, Sayer EJ, Lambers H, Li Y, Zou B, Tang J, Heskel M, Ding Y, Wang F**. **2019**. Foliar phosphorus fractions reveal how tropical plants maintain photosynthetic rates despite low soil phosphorus availability. *Functional Ecology* **33**: 503–513.

**Nakagawa S, Lagisz M, O’Dea RE, Pottier P, Rutkowska J, Senior AM, Yang Y, Noble DWA**. **2023**. orchaRd 2.0: An R package for visualising meta‐analyses with orchard plots. *Methods in Ecology and Evolution* **14**: 2003–2010.

**Niinemets Ü, Kull K**. **2005**. Colimitation of plant primary productivity by nitrogen and phosphorus in a species-rich wooded meadow on calcareous soils. *Acta Oecologica* **28**: 345–356.

**Paillassa J, Wright IJ, Prentice IC, Pepin S, Smith NG, Ethier G, Westerband AC, Lamarque LJ, Wang H, Cornwell WK, *et al.*** **2020**. When and where soil is important to modify the carbon and water economy of leaves. *New Phytologist* **228**: 121–135.

**Peng Y, Bloomfield KJ, Cernusak LA, Domingues TF, Prentice IC**. **2021**. Global climate and nutrient controls of photosynthetic capacity. *Communications Biology* **4**: 462.

**Perkowski EA, Ezekannagha E, Smith NG**. **2025**. Nitrogen demand, availability, and acquisition strategy control plant responses to elevated CO2 (A Rogers, Ed.). *Journal of Experimental Botany* **76**: 2908–2923.

**Perkowski EA, Waring EF, Smith NG**. **2021**. Root mass carbon costs to acquire nitrogen are determined by nitrogen and light availability in two species with different nitrogen acquisition strategies. *Journal of Experimental Botany* **72**: 5766–5776.

**Pick JL, Nakagawa S, Noble DWA**. **2019**. Reproducible, flexible and high-throughput data extraction from primary literature: The metaDigitise r package. *Methods in Ecology and Evolution* **10**: 426–431.

**R Core Team**. **2021**. R: A language and environment for statistical computing.

**Reich PB, Oleksyn J, Wright IJ**. **2009**. Leaf phosphorus influences the photosynthesis–nitrogen relation: a cross-biome analysis of 314 species. *Oecologia* **160**: 207–212.

**Ren Y, Wang H, Harrison SP, Prentice IC, Mengoli G, Zhao L, Reich PB, Yang K**. **2025**. Incorporating the Acclimation of Photosynthesis and Leaf Respiration in the Noah‐MP Land Surface Model: Model Development and Evaluation. *Journal of Advances in Modeling Earth Systems* **17**.

**Rogers A, Medlyn BE, Dukes JS, Bonan GB, Caemmerer S, Dietze MC, Kattge J, Leakey ADB, Mercado LM, Niinemets Ü, *et al.*** **2017**. A roadmap for improving the representation of photosynthesis in Earth system models. *New Phytologist* **213**: 22–42.

**Smith NG, Keenan TF, Prentice IC, Wang H, Wright IJ, Niinemets Ü, Crous KY, Domingues TF, Guerrieri R, Ishida FY, *et al.*** **2019**. Global photosynthetic capacity is optimized to the environment. *Ecology Letters* **22**: 506–517.

**Smith NG, Zhu Q, Keenan TF, Riley WJ**. **2024**. Acclimation of photosynthesis to CO2 increases ecosystem carbon storage due to leaf nitrogen savings. *Global Change Biology* **30**: 1–10.

**Soudzilovskaia NA, Vaessen S, Barceló M, He J, Rahimlou S, Abarenkov K, Brundrett MC, Gomes SIF, Merckx VSFT, Tedersoo L**. **2020**. FungalRoot: global online database of plant mycorrhizal associations. *New Phytologist* **227**: 955–966.

**Stocker BD, Dong N, Perkowski EA, Schneider PD, Xu H, de Boer HJ, Rebel KT, Smith NG, Van Sundert K, Wang H, *et al.*** **2025**. Empirical evidence and theoretical understanding of ecosystem carbon and nitrogen cycle interactions. *New Phytologist* **245**: 49–68.

**Van Sundert K, Leuzinger S, Bader MKF, Chang SX, De Kauwe MG, Dukes JS, Langley JA, Ma Z, Mariën B, Reynaert S, *et al.*** **2023**. When things get MESI: The Manipulation Experiments Synthesis Initiative—A coordinated effort to synthesize terrestrial global change experiments. *Global Change Biology* **29**: 1922–1938.

**Viechtbauer W**. **2010**. Conducting Meta-Analyses in R with the metafor Package. *Journal of Statistical Software* **36**: 1–48.

**Vitousek PM, Porder S, Houlton BZ, Chadwick OA**. **2010**. Terrestrial phosphorus limitation: mechanisms, implications, and nitrogen–phosphorus interactions. *Ecological Applications* **20**: 5–15.

**Walker AP, Beckerman AP, Gu L, Kattge J, Cernusak LA, Domingues TF, Scales JC, Wohlfahrt G, Wullschleger SD, Woodward FI**. **2014**. The relationship of leaf photosynthetic traits - Vcmax and Jmax - to leaf nitrogen, leaf phosphorus, and specific leaf area: a meta-analysis and modeling study. *Ecology and Evolution* **4**: 3218–3235.

**Wang FC, Fang XM, Wang GG, Mao R, Lin XF, Wang H, Chen FS**. **2019**. Effects of nutrient addition on foliar phosphorus fractions and their resorption in different-aged leaves of Chinese fir in subtropical China. *Plant and Soil* **443**: 41–54.

**Waring EF, Perkowski EA, Smith NG**. **2023**. Soil nitrogen fertilization reduces relative leaf nitrogen allocation to photosynthesis. *Journal of Experimental Botany* **74**: 5166–5180.

**Westerband AC, Wright IJ, Maire V, Paillassa J, Prentice IC, Atkin OK, Bloomfield KJ, Cernusak LA, Dong N, Gleason SM, *et al.*** **2023**. Coordination of photosynthetic traits across soil and climate gradients. *Global Change Biology* **29**: 856–873.

**Wieder WR, Cleveland CC, Smith WK, Todd-Brown K**. **2015**. Future productivity and carbon storage limited by terrestrial nutrient availability. *Nature Geoscience* **8**: 441–444.

**Yu Q, Ni X, Cheng X, Ma S, Tian D, Zhu B, Zhu J, Ji C, Tang Z, Fang J**. **2022**. Foliar phosphorus allocation and photosynthesis reveal plants’ adaptative strategies to phosphorus limitation in tropical forests at different successional stages. *Science of the Total Environment* **846**: 157456.

**Yue K, Fornara DA, Yang W, Peng Y, Peng C, Liu Z, Wu F**. **2017**. Influence of multiple global change drivers on terrestrial carbon storage: additive effects are common. *Ecology Letters* **20**: 663–672.

**Zomer RJ, Xu J, Trabucco A**. **2022**. Version 3 of the Global Aridity Index and Potential Evapotranspiration Database. *Scientific Data* **9**: 409.