



RESEARCH ARTICLE

Plant phylogenetic history explains in-stream decomposition at a global scale

Carri J. LeRoy¹  | Andrew L. Hipp^{2,3} | Kate Lueders² | Jennifer J. Follstad Shah⁴ | John S. Kominoski⁵ | Marcelo Ardón^{6*} | Walter K. Dodds⁷ | Mark O. Gessner^{8,9} | Natalie A. Griffiths¹⁰ | Antoine Lecerf¹¹  | David W. P. Manning¹² | Robert L. Sinsabaugh¹³ | Jack R. Webster¹⁴

¹Environmental Studies Program, The Evergreen State College, Olympia, WA, USA; ²The Morton Arboretum, Lisle, IL, USA; ³The Field Museum, Chicago, IL, USA; ⁴Environmental & Sustainability Studies and Department of Geography, University of Utah, Salt Lake City, UT, USA; ⁵Department of Biological Sciences, Florida International University, Miami, FL, USA; ⁶Department of Forestry and Environmental Resources, North Carolina State University, Raleigh, NC, USA; ⁷Division of Biology, Kansas State University, Manhattan, KS, USA; ⁸Department of Experimental Limnology, Leibniz-Institute of Freshwater Ecology and Inland Fisheries, Stechlin, Germany; ⁹Department of Ecology, Berlin Institute of Technology (TU Berlin), Berlin, Germany; ¹⁰Environmental Science Division, Climate Change Science Institute, Oak Ridge National Laboratory, Oak Ridge, TN, USA; ¹¹EcoLab (Laboratoire d'Écologie Fonctionnelle), Université de Toulouse, UPS, INP, NCRS, Toulouse, France; ¹²Department of Biology, University of Nebraska at Omaha, Omaha, NE, USA; ¹³Department of Biology, University of New Mexico, Albuquerque, NM, USA and ¹⁴Department of Biological Sciences, Virginia Polytechnic Institute and State University, Blacksburg, Virginia, USA

Correspondence

Carri J. LeRoy
Email: LeRoyC@evergreen.edu

Funding information

Oak Ridge National Laboratory, Grant/
Award Number: DE-AC05-00OR22725

Handling Editor: David Wardle

Abstract

1. Evolutionary history and adaptation to climate shape plant traits. Some include leaf traits that influence litter quality. Thus, evolutionary history should affect litter decomposition, a crucial ecosystem process. In addition, litter decomposition is directly influenced by climate. We consequently expect plant phylogeny, adaptation and climate to jointly influence litter decomposition. These effects and their interactions have yet to be untangled at a global scale.
2. Here we present an analysis of variation in litter decomposition rates in rivers and streams across 285 published studies for 239 species (from ferns to angiosperms) distributed at 494 locations world-wide. We estimated the relative contributions of climatic conditions and phylogenetic heritage on litter decomposition rates, partitioning phylogenetic from climatic effects at the site and species levels using phylogenetic eigenvector analysis and phylogenetic linear mixed models. In addition, we modelled transitions in decomposition rates under a suite of multiple adaptive-regime Ornstein–Uhlenbeck models to test the hypothesis that natural selection has shaped clade-level litter decomposition rates.
3. Leaf litter decomposition rate exhibited a significant phylogenetic signal. Modelling decomposition rate as a function of location, climatic niche and phylogeny

Carri J. LeRoy and Andrew L. Hipp contributed equally to this work.

*Authors in alphabetical order beyond this point.

Copyright Statement: This manuscript has been co-authored by UT-Battelle, LLC under Contract No. DE-AC05-00OR22725 with the U.S. Department of Energy. The United States Government retains and the publisher, by accepting the article for publication, acknowledges that the United States Government retains a non-exclusive, paid-up, irrevocable, worldwide license to publish or reproduce the published form of this manuscript, or allow others to do so, for United States Government purposes. The Department of Energy will provide public access to these results of federally sponsored research in accordance with the DOE Public Access Plan (<http://energy.gov/downloads/doe-public-access-plan>).

consistently recovered phylogeny alone as one of the top models in species-level analyses. Since many previous studies have focused on the same species across many locations, we also conducted analyses at the species \times site level. Both phylogenetic and climatic factors were favoured in models of this analysis, but the single most important predictor for angiosperms and for all taxa analysed together was phylogeny alone.

4. *Synthesis.* For plant species distributed globally at nearly 500 locations we found that plant phylogenetic history is a critically important predictor of litter decomposition rate in rivers and streams, explaining more of the variance in decomposition than site or climatic regime. Thus, our study demonstrates the influence of evolutionary history on suites of plant traits that shape a key ecosystem process.

KEYWORDS

climate, decomposition rate, ecosystem process, evolutionary ecology, global ecology, leaf litter, phylogenetic comparative methods

1 | INTRODUCTION

The past 20 years have seen a dramatic increase in our understanding of the influence of biodiversity on ecosystem functioning (Hooper et al., 2005; Cardinale et al., 2012; Gessner et al., 2010; Weisser et al., 2017). Expanding beyond the early emphasis of species richness effects on plant productivity, phylogenetic and genetic diversity have also been shown to influence ecosystem processes (Hughes, Inouye, Johnson, Underwood, & Vellend, 2008; Latzel et al., 2013; LeRoy, Whitham, Wooley, & Marks, 2007; Schweitzer et al., 2004; Sundqvist, Giesler, & Wardle, 2011). The exploration of the role of genetic patterns across species has generally taken one of two routes: (1) a diversity approach that uses phylogenetic diversity as a predictor of biodiversity–ecosystem function relationships (Flynn, Mirotchnick, Jain, Palmer, & Naeem, 2011; Srivastava, Cadotte, MacDonald, Marushia, & Mirotchnick, 2012), or (2) a phylogenetic approach that investigates how the evolution of traits and species relationships may influence ecosystem functioning (Cadotte et al., 2017; Donovan, Mason, Bowsher, Goolsby, & Ishibashi, 2014; Edwards, Still, & Donoghue, 2007). Studies taking the first approach have demonstrated that plant phylogenetic diversity influences plant productivity (Cadotte, Cardinale, & Oakley, 2008; Gravel et al., 2012), nutrient cycling (Cornelissen & Cornwell, 2014) and numerous species traits that subsequently affect ecosystem functioning (e.g. Cavender-Bares, Kozak, Fine, & Kembel, 2009; Díaz et al., 2013; Matthews et al., 2011; Senior et al., 2016). Despite a rapidly growing body of literature addressing these relationships, there have only been a few studies examining the influence of plant phylogenetic diversity on litter decomposition (Boyer et al., 2015; Cornwell et al., 2008; Liu et al., 2014; Makkonen et al., 2012; Pan et al., 2014). Even fewer studies have taken the second approach.

Only two previous studies in terrestrial ecosystems have explicitly investigated how plant phylogeny (and thus evolutionary history)

shapes litter decomposition (Liu et al., 2014; Pan et al., 2014). Liu et al. (2014) applied a Brownian motion model to analyse data collected at a single site and found that mass loss was slower in basal angiosperms than in eudicot trees. Pan et al. (2014) compared three phylogenetic models for decomposition data also collected at a single site and found evidence that a constrained single-optimum Ornstein–Uhlenbeck (OU) model best fit their data, suggesting that decomposition rate is under stabilizing selection. No phylogenetic approaches have attempted to address the question of how important evolutionary history is in explaining litter decomposition rates across multiple sites. Understanding the relative contributions of contemporary environment and evolutionary heritage to decomposition rates globally has the potential to improve modelling carbon cycling on geological time-scales, by allowing us to estimate the decomposition rates characteristic of lineages that have variously dominated different habitats over deep time.

Plant litter decomposition is a crucial ecosystem process that drives carbon cycling in many terrestrial and aquatic environments. Shaded headwater streams make up the vast majority of river lengths globally (73.2%, Leopold, Wolman, & Miller, 1964) and strongly rely on plant litter inputs as a resource (Wallace, Eggert, Meyer, & Webster, 1997), relative to all lotic habitats. Plant litter decomposition at a global scale has been explained by both extrinsic (climate, latitude, altitude) and intrinsic (litter quality) factors (Aerts, 1997), but most previous studies have focused on terrestrial decomposition (Cornwell et al., 2008; Pietsch et al., 2014; Weedon et al., 2009). Results of these previous meta-analyses have generally shown that intrinsic species-level traits explain as much variation as extrinsic factors, if not more. Based on a review of 16 studies, Cornwell et al. (2008) found that species differences resulted in larger variation in decomposition rates than climatic differences. Specifically, litter traits like nitrogen and phosphorus concentrations led to faster decomposition and traits like lignin content, leaf

mass area and the concentrations of water- and acid-soluble polysaccharides were associated with slower decomposition (Cornwell et al., 2008). Weedon et al. (2009) examined wood decomposition globally and found a phylogenetic distinction between angiosperms and gymnosperms with lower lignin, and higher N and P in the former, and a higher lignin-to-N ratio in the latter. Pietsch et al. (2014) explored the relationships between litter and wood traits and decomposition published in the two previous studies (Cornwell et al., 2008; Weedon et al., 2009) and found that wood and leaf decomposition rates were decoupled.

Despite decades of investigations into leaf litter decomposition in freshwaters, resulting in a wealth of information (Kaushik & Hynes, 1971; Petersen & Cummins, 1974; Tank, Rosi-Marshall, Griffiths, Entekin, & Stephen, 2010; Webster & Benfield, 1986), previous global decomposition syntheses (Cornwell et al., 2008; Pietsch et al., 2014; Weedon et al., 2009) have until recently excluded aquatic studies. Recent global analyses of aquatic litter decomposition (Boyer et al., 2011, 2015; Follstad Shah et al., 2017; Handa et al., 2014; Tiegs et al., 2019) provide a strong framework for new comparative studies. The oldest of these (Boyer et al., 2011) compared breakdown rates at 26 globally distributed sites and found increased decomposition rates with increasing temperature and, when adjusted for temperature effects, with increasing latitude as a result of higher detritivore abundance. A follow-up study (Boyer et al., 2015) found an influence of phylogenetic diversity on breakdown rates of litter mixtures across 24 sites and 70 species. Follstad Shah et al. (2017) found that aquatic decomposition rates for hundreds of species across the globe tend to increase with latitude when adjusted for temperature effects using a large meta-analysis. Finally, using cotton strips as a standardized substrate at 514 stream and adjacent riparian sites world-wide, Tiegs et al. (2019) found wide variation in in-stream breakdown rates and decreased decomposition rates with increasing latitude.

However, none of these large-scale studies of aquatic leaf litter decomposition explored how the evolutionary history of individual species has shaped leaf litter decomposition. Phylogenetic diversity of plant communities has been addressed, but phylogenetic history of the constituent species integrates leaf trait evolution—indeed, the evolution of all plant traits—and as such may provide improved predictions of decomposition rates globally in linked terrestrial-aquatic ecosystems, where connections are mitigated by traits we may not predict *a priori*. Phylogenetic diversity of communities alone tells only part of the story of how evolutionary history shapes ecosystem function.

Here, we explore how both plant phylogenetic history and climatic variation among sites influence rates of leaf litter decomposition in streams distributed across the globe. Since climatic variables (mean, maximum and minimum temperature; precipitation; seasonality; isothermality) can influence both plant biogeography and the decomposition environment directly, we detangle these two influences (Dodds, Gido, Whiles, Daniels, & Grudinski, 2015), testing the predictions that: (a) across broad spatial scales, decomposition rates will be strongly influenced by site \times species

interactions, with both factors influenced by climate, and (b) phylogenetic history will affect decomposition rates through its influences on species traits but also on the site \times species interaction. As these predictions are not mutually exclusive, we tested them in a modelling framework that addresses the joint influences of phylogeny and climate at both levels of analysis, providing novel insights into how plant evolutionary history shapes stream ecosystem processes globally.

2 | MATERIALS AND METHODS

2.1 | Plant litter decomposition database compilation

Data compiled for this study were extracted from values reported in the scientific literature as detailed in Follstad Shah et al. (2017). Briefly, we used search terms “(leaf OR litter) AND (breakdown OR decomposition OR processing) AND (stream OR river)” to identify studies. This provided us with an initial list of articles published on May 13, 2011, which was updated by articles cited in review papers on leaf litter decomposition in streams by Follstad Shah et al., (2017). Our final list included 636 papers published between 1966 and 2011, of which 285 met our criteria that: (a) rates of leaf litter decomposition were measured in natural streams and rivers with perennial flow; and (b) stream temperature and decomposition rate constants were reported or could be calculated. We discarded studies including factors beyond local species differences (e.g. nutrient addition, predator exclusion, exotic species, long-distance reciprocal transplantations). However, we included in our dataset a variety of leaf litter decomposition methods used across studies, particularly litter bags of various size and mesh size, deployment periods and processing methods, to ensure a large sample size. Studies may have also included measurements of water chemistry, velocity, shredder abundance and microbial measures, but the inconsistency of these measurements made it impossible to comprehensively address these other factors.

For each article, we recorded the scientific name of the plant yielding the leaf litter and the corresponding decomposition rate constants as reported or calculated (see Follstad Shah et al., 2017 and methods therein), where missing, as $m_t = m_0 e^{-k_D t}$, where m_t is leaf litter mass remaining at time t and m_0 is the initial leaf litter mass. Data compilation resulted in 3,189 records of leaf litter decomposition from 494 sites (Figure 1) for 239 plant taxa from 124 genera, 70 families and 34 orders. Decomposition rates ranged from 0.0002 to 0.7890 per day and resulted in a global, average in-stream decomposition rate of 0.0240 per day (Follstad Shah et al., 2017) with a 95% confidence interval of 0.0227–0.0253, and a median rate of 0.0127 per day.

Latitude and longitude data were extracted from each paper and mapped to identify potential errors which were subsequently rectified. Many of the studies lacked detailed data on environmental conditions (stream discharge, water chemistry, etc.), and a previous study exploring patterns in this dataset examined stream

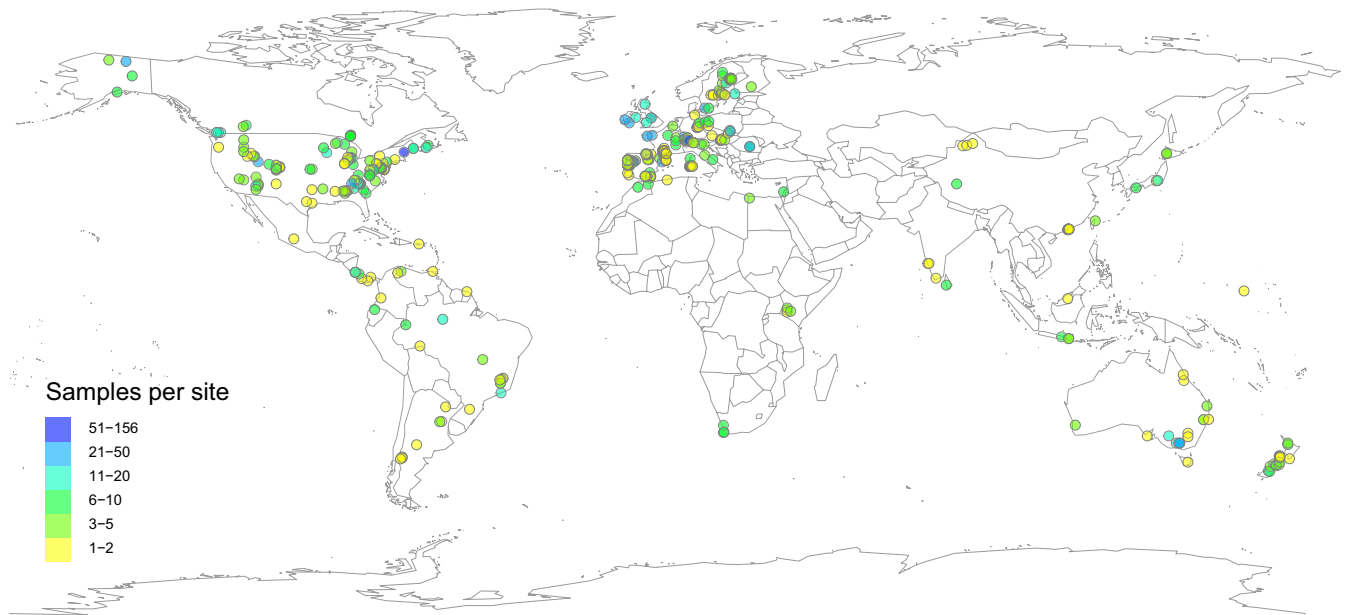


FIGURE 1 Map of sampling locations and sample sizes, includes all 494 sites included in our study. The numbers of decomposition rate values (samples) for each site ranges from 1 to 156 [Colour figure can be viewed at [wileyonlinelibrary.com](https://onlinelibrary.wiley.com)]

temperature (Follstad Shah et al., 2017), so we focused on broad climatic variables in this analysis. Climatic data were inferred for each site from the 10 min (~340 km²) resolution WorldClim v 2.0 dataset (Fick & Hijmans, 2017). All 19 of the WorldClim bioclimatic ("bioclim") variables (<https://www.worldclim.org/bioclim>) were downloaded and used individually or summarized as ordination axes (see methods for *Comparing environmental and phylogenetic drivers* below). R scripts for executing analyses and downloading data have been archived at (<https://github.com/andrew-hipp/decomposition-phylogeny-2019>).

2.2 | Plant phylogenetic data organization

We used Phylomatic (Beaulieu, Ree, Cavender-Bares, Weiblen, & Donoghue, 2012; Webb & Donoghue, 2005) to assemble a base phylogenetic tree for all species for which we had litter decomposition rate data (Appendix S1), using Phylomatic base tree R20120829 as the starting megatree, based on Angiosperm Phylogeny Group III (Haston, Richardson, Stevens, Chase, & Harris, 2009; Qian & Jin, 2016). We normalized scientific names using the Taxonomic Name Resolution Service (Boyle et al., 2013) and corrected as needed with names used in published phylogenies (<https://github.com/andrew-hipp/decomposition-phylogeny-2019>). Mesquite (Maddison & Maddison, 2018) was used to manually resolve branches within families, as the supertree is not fully resolved for all families, based on those same phylogenetic studies (Appendix S1). Node ages of the resulting tree were then calibrated using more recent angiosperm phylogenies (Bell, Soltis, & Soltis, 2010) and the simple branch length adjuster tool (BLADJ) to even out node spacing between calibration points. Initial analyses conducted on the tree with unmodified ages did not suggest different interpretations than analyses conducted on the Phylomatic tree with updated ages, suggesting that our results

are robust to a range of branch length assumptions. Throughout the paper, we report analyses conducted on the tree with revised clade ages and results for the entire dataset as well as a dataset including just the angiosperms. The tree was visualized and exported for publication in R using the GGPLOT2 and GGRTREE packages (Yu et al., 2016) and custom scripts (<https://github.com/andrew-hipp/decomposition-phylogeny-2019>). Site-level phylogenetic diversity was calculated for all sites where more than one taxon occurred using mean pairwise distance (MPD) and mean nearest taxon distance (MNTD) in picante v. 1.7 for R (Kembel et al., 2010).

For analyses at the site level using generalized linear mixed models, we created a composite tree with one tip per species per site, creating a polytomy for each species with a depth of 1.5 Ma to ensure that all species polytomies were younger than the youngest node in the phylogeny. Thus, each species × site combination had one tip in the tree, allowing us to partition the site and phylogenetic contributions to variance in litter decomposition rates.

2.3 | Phylogenetic signal and phylogenetic transitions on decomposition rate

We estimated phylogenetic signal, defined as the "tendency for related species to resemble each other more than they resemble species drawn at random from the tree", (Blomberg & Garland, 2002) in litter decomposition rates using **Blomberg's K** (Blomberg, Garland, & Ives, 2003). Blomberg's K typically scales from 0 for a character with no phylogenetic autocorrelation to 1 for a character that has phylogenetic autocorrelation (i.e. similarity in trait value between close relatives) equivalent to expectations for a trait evolving according to a Brownian motion (random walk) process on the tree being evaluated (Figure 2). K may range to greater than 1 for traits that exhibit

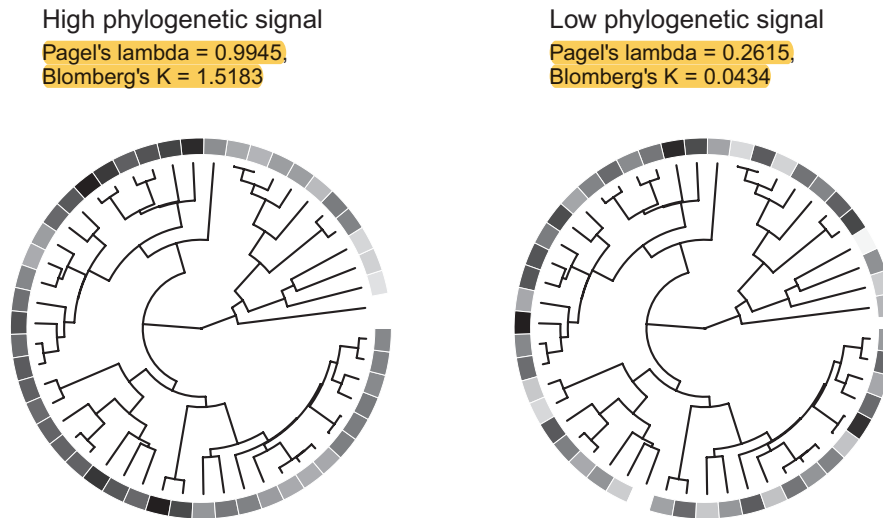


FIGURE 2 Conceptual diagram showing phylogenetic signal strength compared to ecosystem function. Two phylogenetic trees are shown with hypothetical decomposition rates plotted along the tips of the tree, ranging from slow (white squares) to fast (black squares) decomposition rates (k /day). If decomposition rates did not vary across the phylogenetic tree, all tips would be the same colour (null model—not shown). If decomposition rates were distributed randomly across the phylogenetic tree, there would be a low phylogenetic signal as in the panel on the right. The panel on the left shows an example with a high phylogenetic signal for decomposition. Representative Pagel's lambda and Blomberg's K values for each scenario are included

greater phylogenetic autocorrelation than expected under the assumption of Brownian motion. Stated another way, $K > 1$ indicates that close relatives are more similar than expected if the trait being studied evolves according to a random walk. Significance was assessed by permuting tip states 4,999 times, calculating K for each permutation, then calculating p as $2 \times$ the rank position of K_{observed} ; thus yielding a two-tailed p -value, reflecting the fact that both clustering (relatives more similar than expected) and overdispersion (close relatives less similar than expected) are possible evolutionary outcomes. These analyses were conducted with the package `PICANTE` 1.7 (Kembel et al., 2010) in R 3.4.4 (R Core Team, 2018). For comparison with previous studies, we also calculated Pagel's lambda (Münkemüller et al., 2012; Pagel, 1999), a scalar of the off-diagonal cells of the covariance matrix estimated from the tree, which scales from 0 to 1 (or a bit higher, depending on the tree structure). At $\lambda = 0$, the covariance is equivalent to a star-shaped phylogeny, where all species are equally related, reflecting the case where phylogeny has no effect on litter decomposition rates. At $\lambda = 1$, the covariance between any pair of tips is predicted directly by the relative distance from the root of the tree to the most recent common ancestor of those tips; this models a Brownian motion process in which the branch lengths of the tree are known without error. The λ model was fitted using the `fitContinuous` function in the package `GEIGER` 2.0.6 (Harmon, Weir, Brock, Glor, & Challenger, 2008) in R 3.4.4.

We identified phylogenetic shifts in litter decomposition rate by evaluating the relative support for alternative OU models, which model transitions in trait values as responses to shifting selective regimes (Butler & King, 2004; Hansen, 1997; Martins & Hansen, 1997). Analysis was performed using the `l1ou + IC` method (Khabbazian, Kriebel, Rohe, & Ané, 2016), a stepwise model-selection and evaluation approach implemented using the least absolute shrinkage

and selection operator (lasso) (Tibshirani, 1996, 2011) method and a modified information criterion that accounts for clade sizes entailed by shifts in litter decomposition rate, modelled as shifting optima in trait space (where the trait is litter decomposition rate). We compared results with an Expectation Maximization (EM) search algorithm (Bastide, Ané, Robin, & Mariadassou, 2018), which generally searches parameter space more rapidly. Analyses were conducted in the `l1ou` (Khabbazian et al., 2016) and `PHYLOGENETICEM` (Bastide, Mariadassou, & Robin, 2017) packages of R 3.4.4.

We reconstructed phylogenetic transitions in the rate of evolution of litter decomposition rate (i.e. the rate of evolution of $\log(k)$) using reversible-jump Markov-chain Monte Carlo (rjMCMC; Green, 1995) as implemented in the `rjmcmbm` function of `GEIGER` 2.0.6 (Eastman, Alfaro, Joyce, Hipp, & Harmon, 2011; Harmon et al., 2008). In this method, every branch on the phylogeny was assigned to a class of rates, corresponding to presumed shifts in the rate of evolution on the phylogenetic tree. The relative rates of all branches within a taxonomic class were allowed to vary. The assignments of branches to classes were also allowed to vary by allowing transitions in rate to appear or disappear from the tree.

2.4 | Comparing environmental and phylogenetic drivers

We used two methods to compare phylogenetic and climatic models to predict global patterns of litter decomposition rates. First, we used an approach based on phylogenetic eigenvector regression (PVR; Diniz-Filho, Sant'Ana, & Bini, 1998) to partition variance in litter decomposition rate into (a) an environmental component based on the four strongest bioclim variables (Bioclim1: annual mean temperature; Bioclim4: temperature seasonality; Bioclim12: annual

TABLE 1 Average in-stream decomposition rate constants (k /day) for global plant families. N indicates number of species sampled per clade.

Clade	Order	Family	N	mean k /day	SD	SEM
Fern/fern allies	eupolypod II	Blechnaceae	1	0.0066	–	–
Fern/fern allies	Marattiales	Marattiaceae	1	0.013	–	–
Gymnosperm	Cupressales	Cupressaceae	1	0.008	–	–
Gymnosperm	Cupressales	Podocarpaceae	1	0.016	–	–
Gymnosperm	Pinales	Pinaceae	8	0.0058	0.0054	0.0019
Monocot	Alismatales	Potamogetonaceae	2	0.0091	0.0077	0.0054
Monocot	Arecales	Arecaceae	1	0.008	–	–
Monocot	Commelinales	Pontederiaceae	1	0.0122	–	–
Monocot	Pandanales	Pandanaceae	1	0.0144	–	–
Monocot	Poales	Cyperaceae	3	0.0127	0.011	0.0063
Monocot	Poales	Poaceae	12	0.0108	0.0086	0.0025
Monocot	Poales	Typhaceae	1	0.0252	–	–
Magnoliid	Laurales	Atherospermataceae	1	0.0072	–	–
Magnoliid	Laurales	Lauraceae	4	0.006	0.0043	0.0022
Magnoliid	Magnoliales	Magnoliaceae	1	0.0155	–	–
Eudicot	Apiales	Araliaceae	2	0.0291	0.01	0.0071
Eudicot	Apiales	Pittosporaceae	2	0.0296	0.0195	0.0138
Eudicot	Aquifoliales	Aquifoliaceae	2	0.0303	0.0293	0.0207
Eudicot	Asterales	Asteraceae	1	0.0216	–	–
Eudicot	Brassicales	Brassicaceae	1	0.0845	–	–
Eudicot	Caryophyllales	Polygonaceae	2	0.0078	0.0032	0.0023
Eudicot	Caryophyllales	Tamaricaceae	2	0.0064	0.0022	0.0016
Eudicot	Cornales	Cornaceae	3	0.04	0.0274	0.0158
Eudicot	Cornales	Nyssaceae	1	0.0313	–	–
Eudicot	Dipsacales	Caprifoliaceae	1	0.1018	–	–
Eudicot	Ericales	Ericaceae	3	0.004	0.0032	0.0019
Eudicot	Ericales	Primulaceae	1	0.007	–	–
Eudicot	Fabales	Fabaceae	6	0.0311	0.035	0.0143
Eudicot	Fagales	Betulaceae	17	0.0229	0.0233	0.0057
Eudicot	Fagales	Casuarinaceae	1	0.0209	–	–
Eudicot	Fagales	Fagaceae	18	0.0114	0.0055	0.0013
Eudicot	Fagales	Juglandaceae	3	0.0095	0.0039	0.0022
Eudicot	Fagales	Myricaceae	1	0.039	–	–
Eudicot	Fagales	Nothofagaceae	4	0.0106	0.0071	0.0035
Eudicot	Gentianales	Apocynaceae	2	0.0237	0.0181	0.0128
Eudicot	Gentianales	Rubiaceae	2	0.0476	6.00E-04	4.00E-04
Eudicot	Icacinales	Icacinaceae	1	0.006	–	–
Eudicot	Lamiales	Oleaceae	4	0.0182	0.0085	0.0042
Eudicot	Lamiales	Scrophulariaceae	1	0.1522	–	–
Eudicot	Lamiales	Verbenaceae	1	0.0182	–	–
Eudicot	Malpighiales	Euphorbiaceae	5	0.0411	0.023	0.0103
Eudicot	Malpighiales	Salicaceae	28	0.0158	0.0133	0.0025
Eudicot	Malpighiales	Violaceae	1	0.0477	–	–
Eudicot	Malvales	Malvaceae	5	0.0858	0.1407	0.0629

(Continues)

TABLE 1 (Continued)

Clade	Order	Family	N	mean k/day	SD	SEM
Eudicot	Myrtales	Combretaceae	2	0.025	0.0198	0.014
Eudicot	Myrtales	Lythraceae	1	0.0333	–	–
Eudicot	Myrtales	Melastomataceae	2	0.0064	0.0027	0.0019
Eudicot	Myrtales	Myrtaceae	20	0.0115	0.0091	0.002
Eudicot	Myrtales	Onagraceae	1	0.0165	–	–
Eudicot	Oxalidales	Cunoniaceae	2	0.0363	0.0471	0.0333
Eudicot	Oxalidales	Elaeocarpaceae	2	0.0286	0.0369	0.0261
Eudicot	Proteales	Platanaceae	6	0.0057	0.0019	8.00E-04
Eudicot	Proteales	Proteaceae	2	0.009	0.0022	0.0016
Eudicot	Ranunculales	Eupteleaceae	1	0.0133	–	–
Eudicot	Rosales	Elaeagnaceae	1	0.024	–	–
Eudicot	Rosales	Moraceae	6	0.0388	0.0308	0.0126
Eudicot	Rosales	Rhamnaceae	1	0.0286	–	–
Eudicot	Rosales	Rosaceae	4	0.0394	0.0443	0.0222
Eudicot	Rosales	Ulmaceae	7	0.0705	0.0727	0.0275
Eudicot	Rosales	Urticaceae	1	0.02	–	–
Eudicot	Sapindales	Anacardiaceae	2	0.0202	0.0078	0.0056
Eudicot	Sapindales	Burseraceae	2	0.0036	4.00E-04	2.00E-04
Eudicot	Sapindales	Hippocastanaceae	1	0.0066	–	–
Eudicot	Sapindales	Meliaceae	1	0.023	–	–
Eudicot	Sapindales	Rutaceae	1	0.019	–	–
Eudicot	Sapindales	Sapindaceae	8	0.0232	0.0146	0.0052
Eudicot	Sapindales	Simaroubaceae	1	0.0945	–	–
Eudicot	Saxifragales	Haloragaceae	1	0.025	–	–
Eudicot	Saxifragales	Hamamelidaceae	2	0.0172	0.0029	0.002
Eudicot	Vitales	Vitaceae	1	0.0142	–	–

Note: Standard deviation (SD) and standard error (SEM) are provided for families with $N > 1$.

precipitation; and Bioclim14: precipitation of the driest month); (b) four non-metric multidimensional scaling (NMDS) ordination axes created using a Euclidean distance measure from all 19 bioclim variables (<https://www.worldclim.org/bioclim>) (and for comparison, four principle component analysis (PCA) axes created using Euclidean distance measures with all 19 variables); (c) a phylogenetic component based on the first four eigenvectors from principal coordinate analysis (PCoA) of a pairwise phylogenetic distance matrix; and (d) site identity for analyses conducted at the site level. Site identity was added as a random effect to account for any aspects of site that are not directly measured; in this way, site identity in our model is analogous to plot or block in a designed experiment. A stepwise model selection process is generally employed to select phylogenetic eigenvectors for analysis (Diniz-Filho et al., 2012; Gonçalves-Souza, Diniz-Filho, & Romero, 2014). However, the lack of an explicit evolutionary model underlying phylogenetic eigenvector analysis (Freckleton, Cooper, & Jetz, 2011) combined with the relative arbitrariness of eigenvector selection—selecting all 238 eigenvectors would overfit the data, while selecting anything less than all fails to capture all phylogenetic data—makes PVR an approximate approach.

Consequently, we used only four phylogenetic eigenvectors, which cumulatively explain 55% of the total variance in the phylogeny when the relative eigenvalues are used as estimates of the variance explained by each PCoA axis. Axes 2 and 3 of the angiosperm ordination recovered differences among broadly overlapping clades; consequently, we used PCoA axes 1, 2, 4 and 5 for model evaluation in the angiosperms by species models, but this had a negligible effect on model ranking and support. These four eigenvectors were included in models with either four bioclim variables or four bioclim NMDS axes for a comparison of the relative predictive value of both climate and phylogeny. We selected the same number of axes, four, from both the phylogeny PCoA and the climatic NMDS to avoid biasing our linear regressions towards detecting either climate or phylogeny as a better predictor of decomposition rate. In both cases, the axes explain a fair amount of variation in the data. For the phylogenetic PCoA, the first four axes explain 33.0, 8.6, 8.0 and 5.4% of variation respectively. For NMDS, variance partitioning is not the target of each axis, but ordination stress—a measure of the mismatch between the K -dimensional ordination and the distance matrix used to generate it. Ordination stress drops from 0.214 at

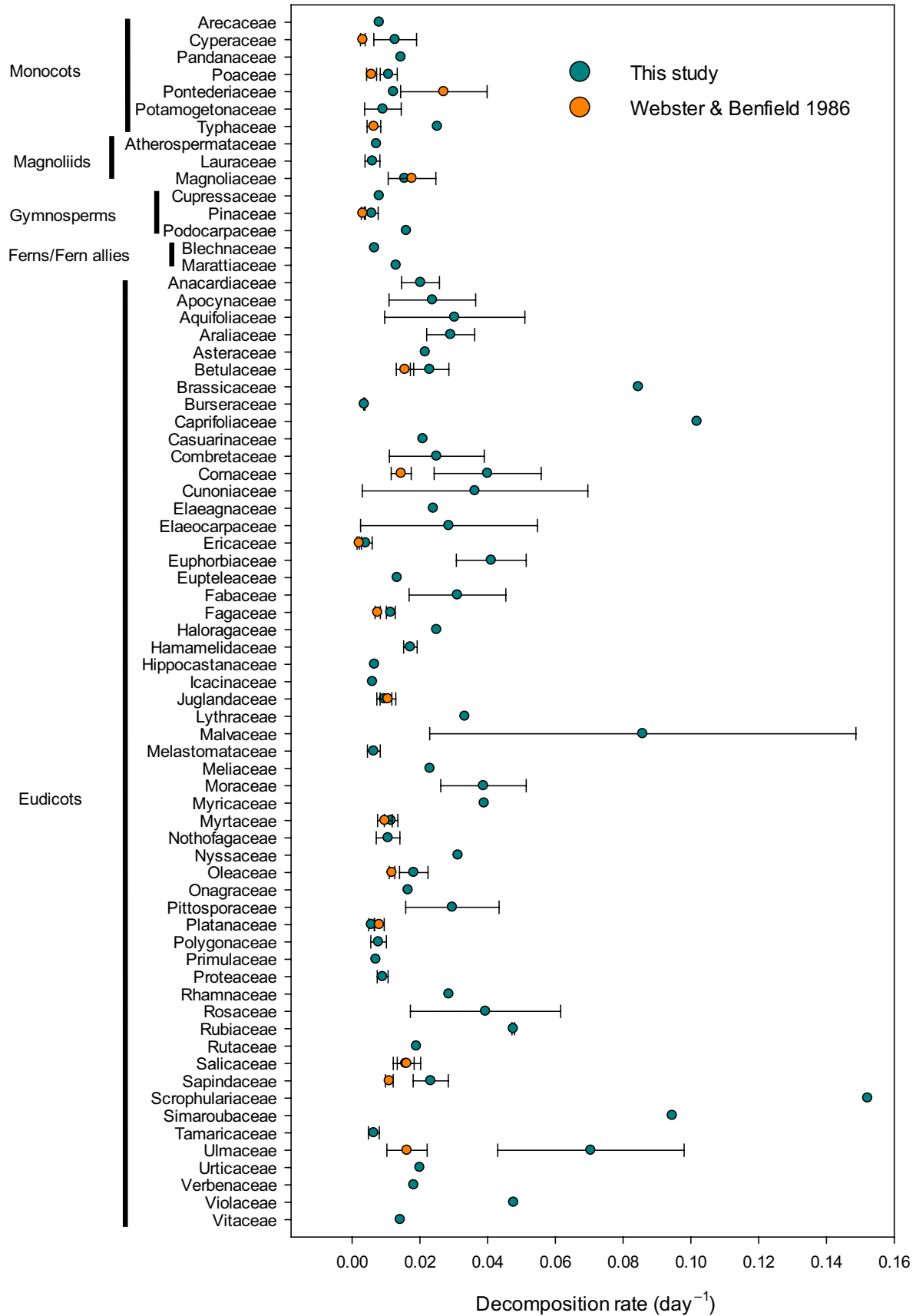


FIGURE 3 Comparison of decomposition rates by plant family between Webster and Benfield (1986) and this study. Symbols represent mean decomposition rates (k/day) for both Webster and Benfield (1986; orange circles) and this study (green circles). Error bars represent standard error of the mean calculated by sample for Webster and Benfield (1986) and by species for this study [Colour figure can be viewed at wileyonlinelibrary.com]

$K = 1$ to 0.035 at $K = 4$; all dimensions above $K = 4$ reduce stress by less than 0.02 per increase in dimension. To assess the effect of ordination method on model support and parameter estimates, we replicated the analyses with the first four axes of a PCA using the 19 bioclim climatic predictors and found that there is no difference in model support and negligible difference in the estimated regression parameters (Supplemental Table S1). Code to do all analyses is

in the repository for this paper (<https://github.com/andrew-hipp/decomposition-phylogeny-2019>). Linear regression models were compared based on Akaike's Information Criterion (AIC; Akaike, 1974) and coefficients of determination (R^2), and model confidence intervals were constructed using cumulative AIC weights (Burnham & Anderson, 2002; Burnham, Anderson, & Huyvaert, 2011). All analyses were conducted using the ape (Paradis, Claude, & Strimmer,

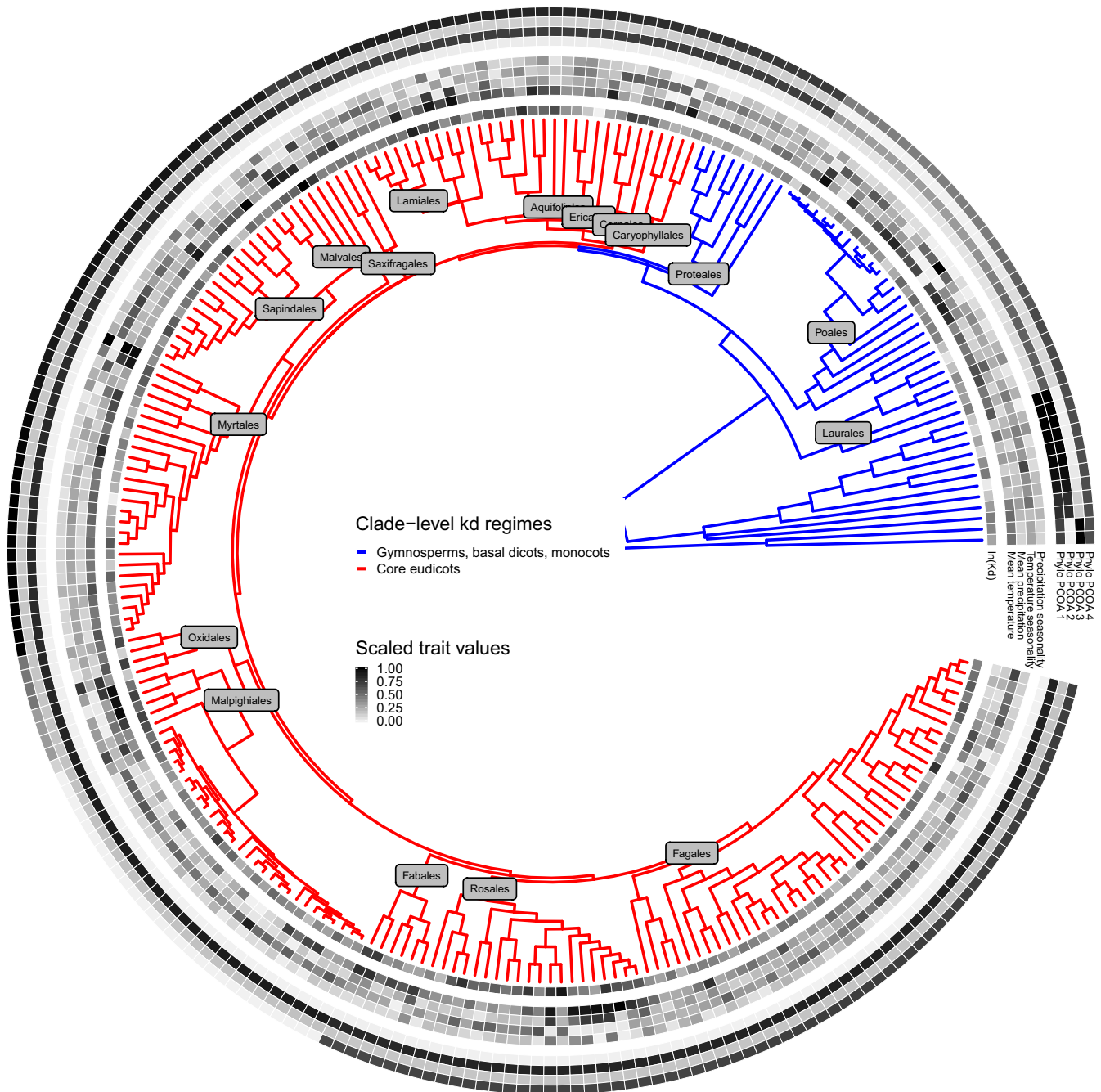


FIGURE 4 Phylogeny of species included in our analyses, with natural-log transformed global in-stream leaf litter decomposition rates ($k/\text{day} = kd$) and bioclimatic and phylogenetic predictors included in models. Tip states are scaled to range from 0.0 to 1.0 . Transitions among clade-level leaf litter decomposition rates were modelled using a multiple-regime Ornstein–Uhlenbeck model with a phylogenetic lasso approach and expectation maximization (EM) algorithm for model fitting. The two approaches converged on all transitions represented here except the decrease in $\ln(kd)$ observed in *Rhododendron*, which was recovered using the phylogenetic lasso but not the EM algorithm (see Figure S2). Variables plotted along the tips include: decomposition rate, $\ln(kd)$; mean annual temperature (BIO1), mean precipitation (BIO12), temperature seasonality (BIO4), precipitation seasonality (BIO14) and four phylogenetic PCoA axis scores [Colour figure can be viewed at wileyonlinelibrary.com]

2004) and vegan (Oksanen, Blanchet, Kindt, Legendre, & O'Hara, 2016) packages of R and custom scripts available at <https://github.com/andrew-hipp/decomposition-phylogeny-2019>.

Our second approach used phylogenetic generalized linear mixed models (PGLMM; Hadfield & Nakagawa, 2010) in a Bayesian framework to quantify the effects of phylogeny, climate, and site on litter decomposition rate more precisely. This approach uses the phylogenetic covariance matrix as a predictor of covariance in trait value among tips, and it scales the importance of the phylogeny by rescaling the covariance matrix. For a species-level analysis, this is analogous to fitting a phylogenetic generalized least squares model while simultaneously estimating a scalar of the off-diagonal elements of the covariance matrix (i.e. Pagel's λ) (Housworth, Martins, & Lynch, 2004; Pagel, 1997, 1999; Revell, 2010). In species-level analyses, we treat phylogeny as a random effect and bioclim variables as fixed effects. For a site-level analysis, PGLMM offers the flexibility of treating both phylogeny and site as random effects and bioclim variables as fixed effects. We compared PGLMM models based on Deviance Information Criterion (DIC; van der Linde, 2005; Spiegelhalter, Best, Carlin, & Linde, 2002) and residual model variance, and model intervals were constructed using DIC weights. Analyses were conducted in R using the MCMCGLMM package (Hadfield, 2010) and custom scripts available at <https://github.com/andrew-hipp/decomposition-phylogeny-2019>.

3 | RESULTS

Litter decomposition rates (k/day) in stream environments showed a wide range of variation across plant families (min-max: 0.0036–0.1522) and plant orders globally (Table 1). Our results (averaged by plant family) are compared to an early synthesis of decomposition rates (Webster & Benfield, 1986; Figure 3) to provide a more complete picture of global decomposition across plant families.

3.1 | Phylogenetic signal and phylogenetic transitions in litter decomposition rate

The litter decomposition rate data exhibited a significant phylogenetic signal in both the all-taxa tree (Figure 4; $K = 0.177$, $p = .01$; $\lambda = 0.977$) and the angiosperm tree ($K = 0.376$, $p = .004$; $\lambda = 0.961$; Table 2). Because plants are distributed non-randomly across the globe, climatic variables averaged for each species all exhibited some phylogenetic signal. Mean annual temperature (BIO1) and temperature seasonality (BIO4) each exhibited significant phylogenetic signal (on the angiosperm tree, $K = 0.230$ – 0.266 , $p \leq .002$, $\lambda = 0.59$ – 0.70 ; on the all-taxa tree, $K = 0.097$ – 0.117 , $p \leq .006$, $\lambda = 0.74$ – 0.83), and precipitation of the driest month (BIO14) exhibited significant phylogenetic signal on the angiosperm tree ($K = 0.227$, $p = .038$, $\lambda = 0.23$), but a non-significant phylogenetic signal on the all-taxa tree ($K = 0.1$, $p = .054$, $\lambda = 0.0$), while mean annual precipitation (BIO12) exhibited weak and, in our dataset, non-significant phylogenetic

signal ($K = 0.094$ – 0.208 , $p = .078$ – 0.112 , $\lambda = 0.51$ – 0.64 ; Table 2). Ordination of these data in four dimensions (Figure S1) was well-supported (stress = 0.035), and axes 1 and 3 exhibited weak but significant phylogenetic signal on the angiosperm tree ($K = 0.200$ – 0.255 , $p < .042$, $\lambda = 0.35$ – 0.70) and the all-taxa tree ($K = 0.093$ – 0.111 , $p < .032$, $\lambda = 0.52$ – 0.83).

Analyses of alternative multiple-regime OU models identified a significant increase in litter decomposition rate at the base of the eudicots (Figure S2). Both inferred a relatively high rate of adaptation, corresponding to 5.0%–5.5% of the total tree length (for the phylogenetic lasso, $\alpha = 14.36$ [$t_{1/2} = 0.0483$] on a tree arbitrarily rescaled to height 1.0; for the E-M algorithm, $\alpha = 0.0305$ [$t_{1/2} = 22.73$] on a tree spanning 432 million years, $t_{1/2} = 5.26\%$ of total tree depth). While interpreting the rate of adaptation (α) is not perfectly straightforward (Bastide et al., 2018), this rate of adaptation suggests that the transition to a higher litter decomposition rate at the base of the eudicots occurred too rapidly to be explained by a purely random walk model, that is, by the variance we expected as lineages diverge from one another (Felsenstein, 1985). This in turn suggests that natural selection on leaf traits shaped this transition, driving accelerated rates of litter decomposition in the angiosperms. By contrast, analysis of evolutionary rates using rjMCMC suggests that the evolution of litter decomposition rates is relatively homogeneous: no lineages were recovered as evolving at a significantly higher or lower rate than other clades (Figure S3).

TABLE 2 Phylogenetic signal (Blomberg's K and Pagel's λ) for key decomposition variables (decomposition rate $k/\text{day} = \text{kd}$; natural log of decomposition rate $k/\text{day} = \ln \text{kd}$), and bioclimatic variables: Bio1 (mean annual air temperature), Bio12 (mean annual precipitation), Bio4 (temperature seasonality), Bio14 (precipitation of the driest month), NMDS axes 1–4 (from ordination of all 19 bioclim variables)

	Angiosperm tree	All taxa tree
$k (\text{day}^{-1})$	$K = 0.376, p = 0.004, \lambda = 0.961$	$K = 0.177, p = 0.010, \lambda = 0.977$
$\ln k (\text{day}^{-1})$	$K = 0.226, p = 0.010, \lambda = 0.536$	$K = 0.122, p = 0.004, \lambda = 0.716$
Bio1	$K = 0.230, p = 0.002, \lambda = 0.587$	$K = 0.097, p = 0.006, \lambda = 0.743$
Bio12	$K = 0.208, p = 0.078, \lambda = 0.505$	$K = 0.094, p = 0.122, \lambda = 0.638$
Bio4	$K = 0.266, p < 0.001, \lambda = 0.703$	$K = 0.117, p < 0.001, \lambda = 0.833$
Bio14	$K = 0.227, p = 0.038, \lambda = 0.225$	$K = 0.100, p = 0.054, \lambda = 0.000$
NMDS1	$K = 0.255, p = 0.002, \lambda = 0.697$	$K = 0.111, p < 0.001, \lambda = 0.826$
NMDS2	$K = 0.186, p = 0.204, \lambda = 0.106$	$K = 0.080, p = 0.262, \lambda = 0.000$
NMDS3	$K = 0.200, p = 0.042, \lambda = 0.343$	$K = 0.093, p = 0.032, \lambda = 0.510$
NMDS4	$K = 0.192, p = 0.058, \lambda = 0.000$	$K = 0.090, p = 0.072, \lambda = 0.000$

3.2 | Comparing environmental and phylogenetic drivers

Top-ranked linear models for both the species-level and site-level analyses included phylogenetic PCoA axes, irrespective of whether the all-taxa phylogeny or angiosperms-only phylogeny was analysed (Table 3). Climatic predictors, whether included directly in the model as bioclim variables, or indirectly as the first four NMDS axes, appeared in the 95% model confidence set (based on cumulative AIC weight) only in combination with phylogeny, whereas phylogeny alone had the lowest AIC of any of the individual models in the species-level analysis of the angiosperm dataset. The first axis of the phylogenetic PCoA had a stronger effect than climatic predictors in multiple regression as estimated using regression coefficients on rescaled data for the all-taxa analyses and the angiosperm dataset analysed by species. For the angiosperm dataset analysed by site, climatic predictors had a stronger effect (Table 3). Among all models in the 95% confidence set, mean temperature (BIO1) had the strongest climatic effect on litter decomposition rate as estimated by normalized partial regression coefficients, followed by either mean annual precipitation (BIO12) or temperature seasonality (BIO4) (Table 3).

Including phylogeny as a random effect in analyses by species reduced the residual variance by approximately 40% in the linear mixed models (Table 4). Including just site as a random effect reduced residual variance from 1.0 to 0.58–0.59 in analyses conducted by site; including just phylogeny (also as a random effect) reduced residual variance from 1.0 to 0.62 or 0.65 (Table 4). The single most important predictor in both sets of taxa and both analysis levels was phylogeny, which was for all sets of analyses the lowest-DIC single-predictor model (Table 4). Including climate as a suite of fixed effects did not improve model fit over that of the model including only phylogeny as a random effect in analyses conducted by species. In analyses conducted by site, the model including site and phylogeny as random effects and climate as a suite of fixed effects was by far the best fit model, with phylogeny explaining ca. $6 \times$ the variance explained by site in the all taxa dataset, and ca. $2 \times$ the variance explained by site in the angiosperm dataset (Table 4, site var. and phylo var. columns). In both datasets, mean annual temperature (BIO1) had approximately twice the effect size of mean annual rainfall (BIO12), but neither effect was significant (Table 4). Using ordination to determine the influence of climatic factors on species-specific decomposition rates (239 species plotted in climatic space, Figure 5), demonstrated that climate is a relatively poor predictor of decomposition rates at a global scale.

Average phylogenetic MNTD for the sites where more than one taxon occurred ($N = 179$) was 210.4 Ma, and average phylogenetic MPD was 232.5 Ma. Eight sites showed significant MPD and 10 showed significant MNTD (two-tailed test, $p < .01$, Figure S4). Forty-one sites sampled had at least one eudicot and one species from a different major clade.

4 | DISCUSSION

The analyses presented here demonstrate that phylogenetic history—the evolutionary history that led to the plants we observe across the globe today—is a more powerful predictor of global variation in rates of in-stream leaf litter decomposition than site or the range of climate variables considered in this study. While this is perhaps not surprising at the species level, where we might expect intrinsic properties of species to swamp climatic conditions at the sites where individual plants are collected, it is remarkable that even data analysed at the site level suggests that phylogeny explains 2.2–5.8 times the variation explained by site (Table 4). This is only possible because sites are phylogenetically diverse: of the 179 sites in our study where two or more taxa were measured, the average phylogenetic MNTD and average phylogenetic MPD corresponded to divergence times of 105 and 116 Ma respectively, demonstrating that sites captured large swaths of phylogenetic history. In fact, 41 sites sampled had at least one eudicot and one species from a different major clade. As the major evolutionary transition in decomposition rates appears to have been on the branch leading to the eudicots, this co-occurrence of eudicots with non-eudicot lineages is key to the fact that site conditions as well as climatic conditions predicted less variance than even the relatively small proportion of phylogenetic variance captured by four phylogenetic eigenvectors (Table 3). Evolutionary history explains variation in traits that are highly relevant to explaining global litter decomposition rates.

Phylogenetic effects observed in this study may be related to variation in whole suites of plant traits that influence microbial decomposers and invertebrate litter consumers and that act across ecosystems, from the terrestrial environments where the plants evolved to the aquatic ecosystems where their leaves decompose. Leaf litter decomposability is influenced by a plant's position on the leaf economics spectrum (Díaz et al., 2016; Wright et al., 2004). In terrestrial systems, plant traits have been found to explain the decomposition rate of leaves (Cornwell et al., 2008) and wood (Hu et al., 2018) more than climatic factors. We were unable to include specific leaf traits as explanatory variables in our study due to inconsistent reporting of leaf traits in the published studies. However, plant traits are phylogenetically heritable (Cavender-Bares et al., 2018; Flores et al., 2014; Hao, Kuang, & Kang, 2015; Pearse & Hipp, 2012; Schmerler et al., 2012; Zanne et al., 2014), and plant phylogeny is thus expected to integrate across the entire suite of litter quality traits. Hence, our study implies that the traits that predict leaf decomposability track phylogenetic history more closely than climatic transitions among species. Our results demonstrate that phylogeny explains an important fraction of variance in plant traits similar to other studies where they were either correlated with a subset of climatic predictors (e.g. Willis, Ruhfel, Primack, Miller-Rushing, & Davis, 2008) or incompletely explained by the full set of climatic predictors expected to explain them (Li, Ives, & Waller, 2017; Pearse & Hipp, 2009).

The phylogenetic effect we find in this study is among the strongest reported to date for decomposition rates. Phylogenetic

TABLE 3 Phylogenetic eigenvector regression model results including both phylogenetic and bioclimatic predictors of decomposition rates

Taxon set	Analysis level	Model	R ²	AIC	delta AIC	Sum AICw	bio1	bio12	bio14
allTaxa	bySp	NMDS & phyloPCOA	0.114	668.205	0	0.394	–	–	–
allTaxa	bySp	bioclim & phyloPCOA	0.114	668.302	0.097	0.77	0.129 (<i>p</i> = .224)	0.164 (<i>p</i> = .205)	–0.008 (<i>p</i> = .939)
allTaxa	bySp	phyloPCOA	0.08	669.303	1.098	0.998	–	–	–
allTaxa	bySp	NMDS	0.04	679.394	11.189	0.999	–	–	–
allTaxa	bySp	bioclim	0.035	680.678	12.473	1	0.13 (<i>p</i> = .222)	0.131 (<i>p</i> = .325)	0.032 (<i>p</i> = .775)
allTaxa	bySpxSite	bioclim & phyloPCOA	0.096	2,523.736	0	1	0.131 (<i>p</i> = .012)	0.047 (<i>p</i> = .41)	0.002 (<i>p</i> = .958)
allTaxa	bySpxSite	bioclim	0.048	2,562.93	39.194	1	0.12 (<i>p</i> = .019)	0.05 (<i>p</i> = .387)	–0.015 (<i>p</i> = .757)
allTaxa	bySpxSite	phyloPCOA	0.047	2,563.876	40.14	1	–	–	–
allTaxa	bySpxSite	NMDS & phyloPCOA	0.052	2,566.611	42.875	1	–	–	–
allTaxa	bySpxSite	NMDS	0.005	2,603.493	79.757	1	–	–	–
angio-sperms	bySp	NMDS & phyloPCOA	0.082	643.89	0	0.356	–	–	–
angio-sperms	bySp	phyloPCOA	0.047	644.313	0.423	0.643	–	–	–
angio-sperms	bySp	bioclim & phyloPCOA	0.08	644.384	0.494	0.921	0.13 (<i>p</i> = .245)	0.122 (<i>p</i> = .387)	0.023 (<i>p</i> = .842)
angio-sperms	bySp	NMDS	0.031	648.027	4.137	0.966	–	–	–
angio-sperms	bySp	bioclim	0.029	648.583	4.693	1	0.074 (<i>p</i> = .505)	0.132 (<i>p</i> = .341)	0.036 (<i>p</i> = .757)
angio-sperms	bySpxSite	bioclim & phyloPCOA	0.076	2,484.951	0	1	0.156 (<i>p</i> = .004)	0.032 (<i>p</i> = .576)	0.024 (<i>p</i> = .618)
angio-sperms	bySpxSite	bioclim	0.047	2,504.595	19.644	1	0.089 (<i>p</i> = .089)	0.04 (<i>p</i> = .499)	0.002 (<i>p</i> = .959)
angio-sperms	bySpxSite	phyloPCOA	0.02	2,529.733	44.782	1	–	–	–
angio-sperms	bySpxSite	NMDS & phyloPCOA	0.027	2,531.992	47.041	1	–	–	–
angio-sperms	bySpxSite	NMDS	0.005	2,543.133	58.182	1	–	–	–

Note: All predictors used in the models are indicated by the presence of a regression coefficient; parameters not included in each model are indicated by blanks in the table. Models were run using two sets of data: (a) all taxa in the dataset, and (b) just the angiosperms in the dataset. Analyses were conducted by species and by species × site. Results include coefficients of determination (R²), Akaike's Information Criterion (AIC), differences between the top-ranked model and others (delta AIC), and the cumulative AIC weight (Sum AICw), with lowest AIC scores reported at the top. Regression coefficients are all estimated on data rescaled to unit variance and mean of 0 and are highlighted in bold when *p*-values are less than 0.05. "NMDS" in the model name indicates that nonmetric multidimensional scaling axes for the Bioclim data were used in lieu of individual Bioclim variables.

heritability of leaf decomposition in streams analysed in this study (Pagel's λ = 0.977 for 239 species globally) is higher than λ values found for the decomposability of angiosperm species studied in terrestrial environments in the UK (45 species; λ = 0.32) and central

Argentina (24 species; λ = 0.70) (Díaz et al., 2013), the eastern US (λ = 0.39 for 78 leaf species, Jo, Fridley, & Frank, 2016), and Ecuador (17 species; λ = 0.80, for undamaged leaves; Cárdenas, Hättenschwiler, Valencia, Argot, & Dangles, 2015). These smaller studies had lower

bioclim4	NMDS1	NMDS2	NMDS3	NMDS4	phylo Axis1	phylo Axis2	phylo Axis3	phylo Axis4	phylo Axis5
–	0.137 (<i>p</i> = .033)	0.006 (<i>p</i> = .929)	0.075 (<i>p</i> = .238)	–0.105 (<i>p</i> = .093)	–0.213 (<i>p</i> = .001)	0.13 (<i>p</i> = .042)	0.113 (<i>p</i> = .074)	0.022 (<i>p</i> = .727)	–
0.122 (<i>p</i> = .198)	–	–	–	–	–0.22 (<i>p</i> = .001)	0.13 (<i>p</i> = .043)	0.118 (<i>p</i> = .062)	0.019 (<i>p</i> = .771)	–
–	–	–	–	–	–0.236 (<i>p</i> < .001)	0.112 (<i>p</i> = .075)	0.101 (<i>p</i> = .109)	0.038 (<i>p</i> = .544)	–
–	0.143 (<i>p</i> = .026)	–0.005 (<i>p</i> = .937)	0.051 (<i>p</i> = .428)	–0.131 (<i>p</i> = .041)	–	–	–	–	–
0.11 (<i>p</i> = .256)	–	–	–	–	–	–	–	–	–
–0.077 (<i>p</i> = .111)	–	–	–	–	–0.197 (<i>p</i> < .001)	0.032 (<i>p</i> = .39)	0.046 (<i>p</i> = .206)	–0.039 (<i>p</i> = .23)	–
–0.084 (<i>p</i> = .085)	–	–	–	–	–	–	–	–	–
–	–	–	–	–	–0.202 (<i>p</i> < .001)	0.008 (<i>p</i> = .839)	0.045 (<i>p</i> = .225)	0.001 (<i>p</i> = .978)	–
–	0.074 (<i>p</i> = .031)	–0.029 (<i>p</i> = .395)	0.011 (<i>p</i> = .733)	–0.014 (<i>p</i> = .664)	–0.203 (<i>p</i> < .001)	0.012 (<i>p</i> = .748)	0.048 (<i>p</i> = .203)	–0.015 (<i>p</i> = .66)	–
–	0.068 (<i>p</i> = .044)	–0.019 (<i>p</i> = .578)	–0.001 (<i>p</i> = .984)	–0.016 (<i>p</i> = .635)	–	–	–	–	–
–	0.149 (<i>p</i> = .027)	0.003 (<i>p</i> = .963)	0.055 (<i>p</i> = .421)	–0.103 (<i>p</i> = .116)	–0.249 (<i>p</i> = .088)	–0.03 (<i>p</i> = .837)	–	0.023 (<i>p</i> = .729)	–0.04 (<i>p</i> = .555)
–	–	–	–	–	–0.249 (<i>p</i> = .09)	–0.047 (<i>p</i> = .747)	–	0.044 (<i>p</i> = .506)	–0.035 (<i>p</i> = .591)
0.087 (<i>p</i> = .396)	–	–	–	–	–0.252 (<i>p</i> = .085)	–0.03 (<i>p</i> = .837)	–	0.018 (<i>p</i> = .786)	–0.033 (<i>p</i> = .623)
–	0.13 (<i>p</i> = .051)	–0.031 (<i>p</i> = .639)	0.04 (<i>p</i> = .545)	–0.108 (<i>p</i> = .102)	–	–	–	–	–
0.072 (<i>p</i> = .476)	–	–	–	–	–	–	–	–	–
–0.076 (<i>p</i> = .129)	–	–	–	–	0.068 (<i>p</i> = .045)	–0.061 (<i>p</i> = .071)	0.123 (<i>p</i> < .001)	0.085 (<i>p</i> = .009)	–
–0.117 (<i>p</i> = .018)	–	–	–	–	–	–	–	–	–
–	–	–	–	–	0.046 (<i>p</i> = .175)	–0.011 (<i>p</i> = .745)	0.087 (<i>p</i> = .011)	0.095 (<i>p</i> = .004)	–
–	0.083 (<i>p</i> = .022)	–0.022 (<i>p</i> = .523)	0.002 (<i>p</i> = .945)	–0.031 (<i>p</i> = .382)	0.054 (<i>p</i> = .132)	–0.032 (<i>p</i> = .359)	0.103 (<i>p</i> = .004)	0.087 (<i>p</i> = .01)	–
–	0.064 (<i>p</i> = .061)	–0.05 (<i>p</i> = .145)	0.0 (<i>p</i> = .994)	0.001 (<i>p</i> = .975)	–	–	–	–	–

power to detect divergence from Brownian motion processes of trait evolution (Boettiger, Coop, & Ralph, 2012) and are biased towards reporting high λ values by the long average length of their terminal branches. Therefore, our results are particularly remarkable in the

light of the fact that the high species density of our phylogenetic tree and resultant relative shortening of terminal branches increases statistical power to detect divergence from Brownian motion (compare, for example, Figure 4 of the present study with Figure 4 of

TABLE 4 Phylogenetic generalized linear mixed model results including phylogeny as a random effect along with bioclimatic predictors of decomposition rates

Taxon set	Analysis level	Model	DIC	delta DIC	Sum DICw	Residual var	Site var	Phylo var	bio1	bio12	bio4	bio14
allTaxa	bySp	phyloPCOA	587.107	0	0.977	0.524	-	1.42	-	-	-	-
allTaxa	bySp	NMDS & phyloPCOA	594.644	7.537	1	0.542	-	1.31	0.002 (<i>p</i> = .238)	0.000 (<i>p</i> = .972)	0.000 (<i>p</i> = .471)	0.001 (<i>p</i> = .591)
allTaxa	bySp	NMDS	659.939	72.832	1	0.907	-	-	0.002 (<i>p</i> = .228)	0.000 (<i>p</i> = .323)	0.000 (<i>p</i> = .258)	0.001 (<i>p</i> = .774)
allTaxa	bySp	no Predictors	660.457	73.35	1	0.922	-	-	-	-	-	-
allTaxa	bySpxSite	phyloPCOA & NMDS & site	1612	0	1	0.203	0.394	2.263	0.122 (<i>p</i> = .045)	-0.053 (<i>p</i> = .436)	-0.125 (<i>p</i> = .050)	-0.014 (<i>p</i> = .822)
allTaxa	bySpxSite	phyloPCOA & site	1627.434	15.434	1	0.206	0.414	2.182	-	-	-	-
allTaxa	bySpxSite	phyloPCOA & NMDS	2,193.091	581.091	1	0.563	-	2.37	0.182 (<i>p</i> = .000)	-0.016 (<i>p</i> = .786)	-0.121 (<i>p</i> = .026)	0.008 (<i>p</i> = .868)
allTaxa	bySpxSite	phyloPCOA	2,274.177	662.177	1	0.627	-	1.92	-	-	-	-
allTaxa	bySpxSite	NMDS & site	2,341.481	729.481	1	0.587	0.354	-	0.121 (<i>p</i> = .036)	-0.009 (<i>p</i> = .905)	-0.109 (<i>p</i> = .078)	-0.012 (<i>p</i> = .832)
allTaxa	bySpxSite	site	2,354.479	742.479	1	0.591	0.38	-	-	-	-	-
allTaxa	bySpxSite	NMDS	2,562.968	950.968	1	0.956	-	-	0.120 (<i>p</i> = .018)	0.050 (<i>p</i> = .409)	-0.083 (<i>p</i> = .084)	-0.013 (<i>p</i> = .792)
allTaxa	bySpxSite	no Predictors	2,599.644	987.644	1	1.002	-	-	-	-	-	-
Angiosperms	bySp	phyloPCOA	574.868	0	0.939	0.559	-	0.681	-	-	-	-
Angiosperms	bySp	NMDS & phyloPCOA	580.337	5.469	1	0.569	-	0.68	0.102 (<i>p</i> = .324)	-0.022 (<i>p</i> = .854)	0.038 (<i>p</i> = .683)	0.068 (<i>p</i> = .516)
Angiosperms	bySp	no Predictors	640.431	65.563	1	0.978	-	-	-	-	-	-
Angiosperms	bySp	NMDS	641.865	66.997	1	0.966	-	-	0.073 (<i>p</i> = .488)	0.129 (<i>p</i> = .346)	0.071 (<i>p</i> = .462)	0.035 (<i>p</i> = .780)
Angiosperms	bySpxSite	phyloPCOA & NMDS & site	1604.314	0	0.999	0.209	0.41	0.98	0.111 (<i>p</i> = .075)	-0.050 (<i>p</i> = .478)	-0.139 (<i>p</i> = .041)	-0.019 (<i>p</i> = .739)
Angiosperms	bySpxSite	phyloPCOA & site	1619.028	14.714	1	0.213	0.431	0.934	-	-	-	-
Angiosperms	bySpxSite	phyloPCOA & NMDS	2,172.502	568.188	1	0.583	-	1.04	0.181 (<i>p</i> = .000)	-0.019 (<i>p</i> = .750)	-0.141 (<i>p</i> = .012)	0.004 (<i>p</i> = .962)
Angiosperms	bySpxSite	phyloPCOA	2,255.31	650.996	1	0.653	-	0.816	-	-	-	-
Angiosperms	bySpxSite	NMDS & site	2,279.589	675.275	1	0.578	0.361	-	0.089 (<i>p</i> = .147)	-0.014 (<i>p</i> = .839)	-0.141 (<i>p</i> = .026)	-0.002 (<i>p</i> = .968)
Angiosperms	bySpxSite	site	2,290.683	686.369	1	0.581	0.389	-	-	-	-	-
Angiosperms	bySpxSite	NMDS	2,504.622	900.308	1	0.958	-	-	0.089 (<i>p</i> = .085)	0.040 (<i>p</i> = .471)	-0.117 (<i>p</i> = .022)	0.002 (<i>p</i> = .960)
Angiosperms	bySpxSite	no Predictors	2,540.015	935.701	1	1.002	-	-	-	-	-	-

Note: All predictors used in the models are indicated by presence of regression coefficient; parameters not included in each model are indicated by blanks in the table. Models were run using two sets of data: (a) all taxa in the dataset, and (b) just the angiosperms in the dataset. Analyses were conducted by species and by site × species. Results include Deviance Information Criterion (DIC), differences between the top-ranked model and others (delta DIC), and the cumulative DIC weight (Sum DICw), *p*-values < 0.05 are highlighted in bold.

Díaz et al., 2013). In fact, the phylogenetic signal we observed for in-stream decomposition is greater than the phylogenetic signals for all leaf traits ($\lambda = 0.15$ – 0.70 for chlorophyll, P, Ca, tannins, phenols, cellulose, N and lignin, in increasing order of λ) in a tropical forest for a recent analysis of 184 species (McManus et al., 2016) and a number of leaf traits ($\lambda = 0.27$ – 0.76 for leaf area, chlorophyll and leaf thickness) across 229 species in tropical and subtropical China (Yang et al., 2014).

Our large-scale synthesis demonstrates that phylogenetic variation shapes a fundamental ecosystem process across terrestrial–aquatic ecosystem boundaries, despite the fact that the species being studied evolved in one ecosystem while the process of interest—in-stream decomposition rate—is measured in another. This phylogenetic effect is detectable even at a significant distance from the source of the leaf litter and despite a broad set of environmental factors that also influence decomposition rates—water velocity, water temperature, microbial colonization and macroinvertebrate shredders. The patterns we observe here are thus not a consequence of phylogenetic niche conservatism; rather, the entire suite of phylogenetically heritable plant traits (e.g. leaf chemistry, toughness) that shape in-stream litter decomposition rates.

Our study further illustrates that the tree of life is not simply a product of adaptation to changing environments, but that evolutionary history of species and, presumably, their traits drive ecosystem processes (Cornelissen & Cornwell, 2014). The analyses presented here highlight the central role of phylogenetic history's influence on litter decomposition at a global scale and this work represents an important merger of ecosystem science with evolutionary history

(Mouquet et al., 2012; Narwani, Matthews, Fox, & Venail, 2015). The result that phylogenetic signals can be observed across terrestrial–aquatic boundaries is novel and strengthens evidence for the organizing power of genes on communities and ecosystems (Jackrel & Wootton, 2014; LeRoy & Fischer, 2019; LeRoy, Whitham, Keim, & Marks, 2006; LeRoy et al., 2007; LeRoy, Wooley, & Lindroth, 2012; Whitham et al., 2006).

This study also provides further context for our continued exploration of how climate change and global warming are predicted to increase rates of aquatic decomposition through increased stream temperatures (Boyer et al., 2011; Follstad Shah et al., 2017; Tiegs et al., 2019). There are both direct and indirect influences of temperature on this key ecosystem process. Temperature can directly increase rates of decomposition through accelerated microbial growth and litter consumption; however, elevated CO_2 concentrations driving temperature increases could also lower leaf litter quality via higher C:nutrient ratios and thus suppress decomposition rates (Boyer et al., 2017; Tuchman, Wetzel, Rier, Wahtera, & Teeri, 2002). Temperature can also indirectly affect decomposition through plant species' plastic responses to climate change in terms of expressed phenotypes, adaptation in situ and shifts in species ranges (Christmas, Breed, & Lowe, 2016; Hooper et al., 2012; Kominoski et al., 2013). How these indirect changes may influence stream ecosystems is not well understood. Finally, there may be feedbacks between climate change and in-stream decomposition through carbon evasion from streams. Riverine ecosystems are important contributors to the breakdown of terrestrially derived carbon (Cole et al., 2007), and as a result, streams and rivers have been identified as major sources of carbon dioxide in the atmosphere (Raymond et al., 2013).

Streams are influenced by the biomes they are embedded in (Dodds et al., 2015; Tiegs et al., 2019), but biomes and the species assemblages within them will both shift under predicted climate change models. The selective pressures on plants that might influence litter decomposition rates (e.g. predation, defences against microbial attack, desiccation resistance) are decoupled from the physical, chemical and biological processes occurring in streams. In spite of these complexities, our study highlights the importance of riparian plant community composition on stream litter decomposition. Thus, the interactive changes in phenotypic expression, phenology and species range shifts could directly and indirectly influence global litter decomposition rates. Taken as a whole, this work highlights the significant effect of phylogenetic history on key ecosystem processes.

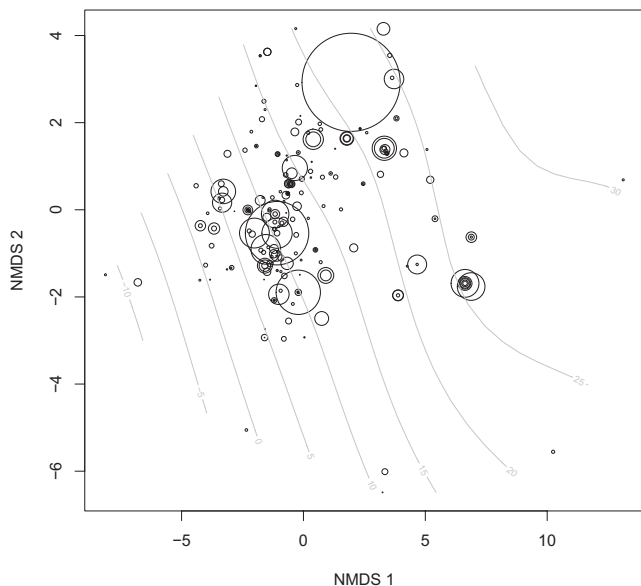


FIGURE 5 Ordination of 239 species in climatic space. Point size corresponds to natural log decomposition rate (k/day). Isoclines represent mean annual air temperature in degrees centigrade ($\text{BIO1}/10$). The first two axes of climatic niche space (using all 19 bioclim climatic variables from WorldClim) demonstrate that climate is a relatively poor predictor of decomposition rates, at least in our study

ACKNOWLEDGEMENTS

We thank the LTER Network for funding an initial collaboration in 2011. NAG was supported by the Department of Energy's Office of Science, Biological and Environmental Research. Oak Ridge National Laboratory is managed by UT-Battelle, LLC, for the US DOE under contract DE-AC05-00OR22725. We thank Chuck

Hawkins, Sherri Johnson, Chris Swan, Amy Rosemond and Lydia Zeglin for insightful conversations about this research and for participating in the initial data compilation. We thank Bruce McCune for advice on statistical analyses. We thank Editor Dr. David Wardle and two anonymous reviewers for their comprehensive reviews of this work. All authors have confirmed that they have no relationships, financial or otherwise, that might be perceived as influencing their objectivity or that could be considered a potential source of conflict of interest.

AUTHORS' CONTRIBUTIONS

C.J.L. and A.L.H. contributed equally to writing this paper and are joint first authors. C.J.L. conceived the study idea and coordinated collaborators. A.L.H. and K.L. compiled the phylogenetic data, and A.L.H. performed the phylogenetic comparative analyses and statistical analyses. J.J.F.S. led the initial data compilation on litter decomposition in streams, which also involved J.S.K., M.A., W.K.D., M.O.G., N.A.G., A.L., C.J.L., D.W.P.M., and J.R.W. C.J.L., A.L.H. and K.L. wrote the initial draft manuscript and all authors (C.J.L., A.L.H., K.L., J.J.F.S., J.S.K., M.A., W.K.D., M.O.G., N.A.G., A.L., D.W.P.M., R.L.S. and J.R.W.) contributed critically to subsequent drafts and gave final approval for publication. C.J.L. coordinated inputs and edits by all co-authors and reviewers.

DATA AVAILABILITY STATEMENT

All raw and processed data, R-scripts, and figures are available at: <https://github.com/andrew-hipp/decomposition-phylogeny-2019>.

ORCID

Carri J. LeRoy  <https://orcid.org/0000-0002-1185-4437>

Antoine Lecerf  <https://orcid.org/0000-0002-7802-9773>

REFERENCES

- Aerts, R. (1997). Climate, leaf litter chemistry and leaf litter decomposition in terrestrial ecosystems: A triangular relationship. *Oikos*, 79, 439–449. <https://doi.org/10.2307/3546886>
- Akaike, H. (1974). A new look at the statistical model identification. *IEEE Transactions on Automatic Control*, 19(6), 716–723. <https://doi.org/10.1109/TAC.1974.1100705>
- Bastide, P., Ané, C., Robin, S., & Mariadassou, M. (2018). Inference of adaptive shifts for multivariate correlated traits. *Systematic Biology*, 67(4), 662–680. <https://doi.org/10.1093/sysbio/syy005>
- Bastide, P., Mariadassou, M., & Robin, S. (2017). Detection of adaptive shifts on phylogenies by using shifted stochastic processes on a tree. *Journal of the Royal Statistical Society, B*, 79(4), 1067–1093. <https://doi.org/10.1111/rssb.12206>
- Beaulieu, J. M., Ree, R. H., Cavender-Bares, J., Weiblen, G. D., & Donoghue, M. J. (2012). Synthesizing phylogenetic knowledge for ecological research. *Ecology*, 93(sp8), S4–S13. <https://doi.org/10.1890/11-0638.1>
- Bell, C. D., Soltis, D. E., & Soltis, P. S. (2010). The age and diversification of the angiosperms re-visited. *American Journal of Botany*, 97(8), 1296–1303. <https://doi.org/10.3732/ajb.0900346>
- Blomberg, S. P., & Garland, T. Jr (2002). Tempo and mode in evolution: Phylogenetic inertia, adaptation and comparative methods. *Journal of Evolutionary Biology*, 15(6), 899–910. <https://doi.org/10.1046/j.1420-9101.2002.00472.x>
- Blomberg, S. P., Garland, T. Jr, & Ives, A. A. (2003). Testing for phylogenetic signal in comparative data: Behavioral traits are more labile. *Evolution*, 57(4), 717–745. <https://doi.org/10.1111/j.0014-3820.2003.tb00285.x>
- Boettiger, C., Coop, G., & Ralph, P. (2012). Is your phylogeny informative? Measuring the power of comparative methods. *Evolution*, 66(7), 2240–2251. <https://doi.org/10.1111/j.1558-5646.2011.01574.x>
- Boyero, L., Graça, M. A. S., Tonin, A. M., Pérez, J., J. Swafford, A., Ferreira, V., ... Pearson, R. G. (2017). Riparian plant litter quality increases with latitude. *Scientific Reports*, 7, 10562. <https://doi.org/10.1038/s41598-017-10640-3>
- Boyero, L., Pearson, R. G., Gessner, M. O., Barmuta, L. A., Ferreira, V., Graça, M. A. S., ... West, D. C. (2011). A global experiment suggests climate warming will not accelerate litter decomposition in streams but might reduce carbon sequestration. *Ecology Letters*, 14(3), 289–294. <https://doi.org/10.1111/j.1461-0248.2010.01578.x>
- Boyero, L., Pearson, R. G., Hui, C., Gessner, M. O., Pérez, J., Alexandrou, M. A., ... Yule, C. M. (2015). Biotic and abiotic variables influencing plant litter breakdown in streams: a global study. *Proceedings of the Royal Society B: Biological Sciences*, 283, 20152664. <https://doi.org/10.1098/rspb.2015.2664>
- Boyle, B., Hopkins, N., Lu, Z., Raygoza Garay, J. A., Mozzherin, D., Rees, T., ... Enquist, B. J. (2013). The taxonomic name resolution service: An online tool for automated standardization of plant names. *BMC Bioinformatics*, 14(1), 16. <https://doi.org/10.1186/1471-2105-14-16>
- Burnham, K. P., & Anderson, D. R. (2002). *Model selection and multimodel inference: A practical information-theoretic approach* (2nd ed.). New York, NY: Springer.
- Burnham, K., Anderson, D., & Huyvaert, K. (2011). AIC model selection and multimodel inference in behavioral ecology: Some background, observations, and comparisons. *Behavioral Ecology and Sociobiology*, 65(1), 23–35. <https://doi.org/10.1007/s00265-010-1029-6>
- Butler, M., & King, A. A. (2004). Phylogenetic comparative analysis: A modeling approach for adaptive evolution. *American Naturalist*, 164, 683–695. <https://doi.org/10.1086/426002>
- Cadotte, M. W., Cardinale, B. J., & Oakley, T. H. (2008). Evolutionary history and the effect of biodiversity on plant productivity. *Proceedings of the National Academy of Sciences USA*, 105(44), 17012–17017. <https://doi.org/10.1073/pnas.0805962105>
- Cadotte, M. W., Livingstone, S. W., Yasui, S.-L.-E., Dinnage, R., Li, J.-T., Marushia, R., ... Shu, W. (2017). Explaining ecosystem multifunction with evolutionary models. *Ecology*, 98(12), 3175–3187. <https://doi.org/10.1002/ecy.2045>
- Cárdenas, R. E., Hättenschwiler, S., Valencia, R., Argot, A., & Dangles, O. (2015). Plant herbivory responses through changes in leaf quality have no effect on subsequent leaf-litter decomposition in a neotropical rain forest tree community. *New Phytologist*, 207(3), 817–829. <https://doi.org/10.1111/nph.13368>
- Cardinale, B. J., Duffy, E., Gonzalez, A., Hooper, D. U., Perrings, C., Venail, P., ... Naeem, S. (2012). Biodiversity loss and its impact on humanity. *Nature*, 486(7401), 59–67. <https://doi.org/10.1038/nature11148>
- Cavender-Bares, J., Kothari, S., Meireles, J. E., Kaproth, M. A., Manos, P. S., & Hipp, A. L. (2018). The role of diversification in community assembly of the oaks (*Quercus* L.) across the continental U.S. *American Journal of Botany*, 105(3), 565–586. <https://doi.org/10.1002/ajb2.1049>
- Cavender-Bares, J., Kozak, K. H., Fine, P. V. A., & Kembel, S. W. (2009). The merging of community ecology and phylogenetic biology. *Ecology Letters*, 12(7), 693–715. <https://doi.org/10.1111/j.1461-0248.2009.01314.x>

- Christmas, M. J., Breed, M. F., & Lowe, A. J. (2016). Constraints to and conservation implications for climate change adaptation in plants. *Conservation Genetics*, 17(2), 305–320. <https://doi.org/10.1007/s10592-015-0782-5>
- Cole, J. J., Prairie, Y. T., Caraco, N. F., McDowell, W. H., Tranvik, L. J., Striegl, R. G., ... Melack, J. (2007). Plumbing the global carbon cycle: Integrating inland waters into the terrestrial carbon budget. *Ecosystems*, 10, 171–184. <https://doi.org/10.1007/s10021-006-9013-8>
- Cornelissen, J. H. C., & Cornwell, W. K. (2014). The tree of life in ecosystems: Evolution of plant effects on carbon and nutrient cycling. *Journal of Ecology*, 102(2), 269–274. <https://doi.org/10.1111/1365-2745.12217>
- Cornwell, W. K., Cornelissen, J. H. C., Amatangelo, K., Dorrepaal, E., Eviner, V. T., Godoy, O., ... Westoby, M. (2008). Plant species traits are the predominant control on litter decomposition rates within biomes worldwide. *Ecology Letters*, 11(10), 1065–1071. <https://doi.org/10.1111/j.1461-0248.2008.01219.x>
- Díaz, S., Kattge, J., Cornelissen, J. H. C., Wright, I. J., Lavorel, S., Dray, S., ... Gorné, L. D. (2016). The global spectrum of plant form and function. *Nature*, 529, 167–171. <https://doi.org/10.1038/nature16489>
- Díaz, S., Purvis, A., Cornelissen, J. H. C., Mace, G. M., Donoghue, M. J., Ewers, R. M., ... Pearse, W. D. (2013). Functional traits, the phylogeny of function, and ecosystem service vulnerability. *Ecology and Evolution*, 3(9), 2958–2975. <https://doi.org/10.1002/ece3.601>
- Diniz-Filho, J. A. F., Bini, L. M., Rangel, T. F., Morales-Castilla, I., Olalla-Tárraga, M. Á., Rodríguez, M. Á., & Hawkins, B. A. (2012). On the selection of phylogenetic eigenvectors for ecological analyses. *Ecography*, 35, 239–249. <https://doi.org/10.1111/j.1600-0587.2011.06949.x>
- Diniz-Filho, J. A. F., de Sant'Ana, C. E. R., & Bini, L. M. (1998). An eigenvector method for estimating phylogenetic inertia. *Evolution*, 52(5), 1247–1262. <https://doi.org/10.1111/j.1558-5646.1998.tb02006.x>
- Dodds, W. K., Gido, K., Whiles, M. R., Daniels, M. D., & Grudinski, B. P. (2015). The stream biome gradient concept: Factors controlling lotic systems across broad biogeographic scales. *Freshwater Science*, 34, 1–19. <https://doi.org/10.1086/679756>
- Donovan, L. A., Mason, C. M., Bowshe, A. W., Goolsby, E. W., & Ishibashi, C. D. A. (2014). Ecological and evolutionary lability of plant traits affecting carbon and nutrient cycling. *Journal of Ecology*, 102(2), 302–314. <https://doi.org/10.1111/1365-2745.12193>
- Eastman, J. M., Alfaro, M. E., Joyce, P., Hipp, A. L., & Harmon, L. J. (2011). A novel comparative method for modeling shifts in the rate of character evolution on trees. *Evolution*, 65(12), 3578–3589. <https://doi.org/10.1111/j.1558-5646.2011.01401.x>
- Edwards, E. J., Still, C. J., & Donoghue, M. J. (2007). The relevance of phylogeny to studies of global change. *Trends in Ecology and Evolution*, 22(5), 243–249. <https://doi.org/10.1016/j.tree.2007.02.002>
- Felsenstein, J. (1985). Phylogenies and the comparative method. *American Naturalist*, 125, 1–15. <https://doi.org/10.1086/284325>
- Fick, S. E., & Hijmans, R. J. (2017). WorldClim 2: New 1-km spatial resolution climate surfaces for global land areas. *International Journal of Climatology*, 37(12), 4302–4315. <https://doi.org/10.1002/joc.5086>
- Flores, O., Garnier, E., Wright, I. J., Reich, P. B., Pierce, S., Díaz, S., ... Weiher, E. (2014). An evolutionary perspective on leaf economics: Phylogenetics of leaf mass per area in vascular plants. *Ecology and Evolution*, 4(14), 2799–2811. <https://doi.org/10.1002/ece3.1087>
- Flynn, D. F. B., Mirotchnick, N., Jain, M., Palmer, M. I., & Naeem, S. (2011). Functional and phylogenetic diversity as predictors of biodiversity–ecosystem-function relationships. *Ecology*, 92, 1573–1581. <https://doi.org/10.1890/10-1245.1>
- Follstad Shah, J. J., Kominoski, J. S., Ardón, M., Dodds, W. K., Gessner, M. O., Griffiths, N. A., ... Zeglin, L. H. (2017). Global synthesis of the temperature sensitivity of leaf litter breakdown in streams and rivers. *Global Change Biology*, 23, 3064–3075. <https://doi.org/10.1111/gcb.13609>
- Freckleton, R. P., Cooper, N., & Jetz, W. (2011). Comparative methods as a statistical fix: The dangers of ignoring an evolutionary model. *The American Naturalist*, 178, E10–E17. <https://doi.org/10.1086/660272>
- Gessner, M. O., Swan, C. M., Dang, C. K., McKie, B. G., Bardgett, R. D., Wall, D. H., & Hättenschwiler, S. (2010). Diversity meets decomposition. *Trends in Ecology and Evolution*, 25, 372–380. <https://doi.org/10.1016/j.tree.2010.01.010>
- Gonçalves-Souza, T., Diniz-Filho, J. A. F., & Romero, G. Q. (2014). Disentangling the phylogenetic and ecological components of spider phenotypic variation. *PLoS ONE*, 9(2), e89314. <https://doi.org/10.1371/journal.pone.0089314>
- Gravel, D., Bell, T., Barbera, C., Combe, M., Pommier, T., & Mouquet, N. (2012). Phylogenetic constraints on ecosystem functioning. *Nature Communications*, 3, 1117. <https://doi.org/10.1038/ncomms2123>
- Green, P. J. (1995). Reversible jump Markov chain Monte Carlo computation and Bayesian model determination. *Biometrika*, 82, 711–732. <https://doi.org/10.1093/biomet/82.4.711>
- Hadfield, J. D. (2010). MCMC methods for multi-response generalized linear mixed models: The MCMCglmm R package. *Journal of Statistical Software*, 33, 1–22. <https://doi.org/10.18637/jss.v033.i02>
- Hadfield, J. D., & Nakagawa, S. (2010). General quantitative genetic methods for comparative biology: Phylogenies, taxonomies and multi-trait models for continuous and categorical characters. *Journal of Evolutionary Biology*, 23, 494–508. <https://doi.org/10.1111/j.1420-9101.2009.01915.x>
- Handa, I. T., Aerts, R., Berendse, F., Berg, M. P., Bruder, A., Butenschoten, O., ... Hättenschwiler, S. (2014). Consequences of biodiversity loss for litter decomposition across biomes. *Nature*, 509, 218–221. <https://doi.org/10.1038/nature13247>
- Hansen, T. F. (1997). Stabilizing selection and the comparative analysis of adaptation. *Evolution*, 51(5), 1341–1351. <https://doi.org/10.1111/j.1558-5646.1997.tb01457.x>
- Hao, Z., Kuang, Y., & Kang, M. (2015). Untangling the influence of phylogeny, soil and climate on leaf element concentrations in a biodiversity hotspot. *Functional Ecology*, 29, 165–176. <https://doi.org/10.1111/1365-2435.12344>
- Harmon, L. J., Weir, J. T., Brock, C. D., Glor, R. E., & Challenger, W. (2008). GEIGER: Investigating evolutionary radiations. *Bioinformatics*, 24, 129–131. <https://doi.org/10.1093/bioinformatics/btm538>
- Haston, E., Richardson, J. E., Stevens, P. F., Chase, M. W., & Harris, D. J. (2009). The Linear Angiosperm Phylogeny Group (LAPG) III: A linear sequence of the families in APG III. *Botanical Journal of the Linnean Society*, 161(2), 128–131. <https://doi.org/10.1111/j.1095-8339.2009.01000.x>
- Hooper, D. U., Adair, E. C., Cardinale, B. J., Byrnes, J. E. K., Hungate, B. A., Matulich, K. L., ... O'Connor, M. I. (2012). A global synthesis reveals biodiversity loss as a major driver of ecosystem change. *Nature*, 486, 105–109. <https://doi.org/10.1038/nature11118>
- Hooper, D. U., Chapin, F. S., Ewel, J. J., Hector, A., Inchausti, P., Lavorel, S., ... Wardle, D. A. (2005). Effects of biodiversity on ecosystem functioning: A consensus of current knowledge. *Ecological Monographs*, 75, 3–35. <https://doi.org/10.1890/04-0922>
- Housworth, E. A., Martins, E. P., & Lynch, M. (2004). The phylogenetic mixed model. *The American Naturalist*, 163, 84–96. <https://doi.org/10.1086/380570>
- Hu, Z., Michaletz, S. T., Johnson, D. J., McDowell, N. G., Huang, Z., Zhou, X., & Xu, C. (2018). Traits drive global wood decomposition rates more than climate. *Global Change Biology*, 24, 5259–5269. <https://doi.org/10.1111/gcb.14357>
- Hughes, A. R., Inouye, B. D., Johnson, M. T. J., Underwood, N., & Vellend, M. (2008). Ecological consequences of genetic diversity. *Ecology Letters*, 11, 609–623. <https://doi.org/10.1111/j.1461-0248.2008.01179.x>
- Jackrel, S. L., & Wootton, J. T. (2014). Local adaptation of stream communities to intraspecific variation in a terrestrial ecosystem subsidy. *Ecology*, 95, 37–43. <https://doi.org/10.1890/13-0804.1>

- Jo, I., Fridley, J. D., & Frank, D. A. (2016). More of the same? In situ leaf and root decomposition rates do not vary between 80 native and nonnative deciduous forest species. *New Phytologist*, 209, 115–122. <https://doi.org/10.1111/nph.13619>
- Kaushik, N. K., & Hynes, H. B. N. (1971). The fate of the dead leaves that fall into streams. *Archiv für Hydrobiologie*, 68, 465–515.
- Kembel, S. W., Cowan, P. D., Helmus, M. R., Cornwell, W. K., Morlon, H., Ackerly, D. D., ... Webb, C. O. (2010). Picante: R tools for integrating phylogenies and ecology. *Bioinformatics*, 26, 1463–1464. <https://doi.org/10.1093/bioinformatics/btq166>
- Khabbaziyan, M., Kriebel, R., Rohe, K., & Ané, C. (2016). Fast and accurate detection of evolutionary shifts in Ornstein-Uhlenbeck models. *Methods in Ecology and Evolution*, 7, 811–824. <https://doi.org/10.1111/2041-210X.12534>
- Kominoski, J. S., Shah, J. J. F., Canhoto, C., Fischer, D. G., Gilling, D. P., González, E., ... Tiegs, S. D. (2013). Forecasting functional implications of global changes in riparian plant communities. *Frontiers in Ecology and the Environment*, 11, 423–432. <https://doi.org/10.1890/120056>
- Latzel, V., Allan, E., Bortolini Silveira, A., Colot, V., Fischer, M., & Bossdorf, O. (2013). Epigenetic diversity increases the productivity and stability of plant populations. *Nature Communications*, 4, 2875. <https://doi.org/10.1038/ncomms3875>
- Leopold, L. B., Wolman, M. G., & Miller, J. P. (1964). *Fluvial processes in geomorphology* (p. 522). San Francisco, CA: Freeman.
- LeRoy, C. J., & Fischer, D. G. (2019). Do genetically-specific tree canopy environments feed back to affect genetically specific leaf decomposition rates? *Plant and Soil*, 437, 1–10. <https://doi.org/10.1007/s11104-019-03952-y>
- LeRoy, C. J., Whitham, T. G., Keim, P., & Marks, J. C. (2006). Plant genes link forests and streams. *Ecology*, 87, 255–261. <https://doi.org/10.1890/05-0159>
- LeRoy, C. J., Whitham, T. G., Wooley, S. C., & Marks, J. C. (2007). Within-species variation in foliar chemistry influences leaf-litter decomposition in a Utah river. *Journal of the North American Benthological Society*, 26, 426–438. <https://doi.org/10.1899/06-113.1>
- LeRoy, C. J., Wooley, S. C., & Lindroth, R. L. (2012). Genotype and soil nutrient environment influence aspen litter chemistry and in-stream decomposition. *Freshwater Science*, 31, 1244–1253. <https://doi.org/10.1899/12-029.1>
- Li, D., Ives, A. R., & Waller, D. M. (2017). Can functional traits account for phylogenetic signal in community composition? *New Phytologist*, 214, 607–618. <https://doi.org/10.1111/nph.14397>
- Liu, G., Cornwell, W. K., Pan, X., Cao, K., Ye, X., Huang, Z., ... Cornelissen, J. H. C. (2014). Understanding the ecosystem implications of the angiosperm rise to dominance: Leaf litter decomposability among magnoliids and other basal angiosperms. *Journal of Ecology*, 102, 337–344. <https://doi.org/10.1111/1365-2745.12192>
- Maddison, W. P., & Maddison, D. R. (2018). Mesquite: A modular system for evolutionary analysis. Version 3.40. Retrieved from <http://mesquitemproject.org>
- Makkonen, M., Berg, M. P., Handa, I. T., Hättenschwiler, S., van Ruijven, J., van Bodegom, P. M., & Aerts, R. (2012). Highly consistent effects of plant litter identity and functional traits on decomposition across a latitudinal gradient. *Ecology Letters*, 15, 1033–1041. <https://doi.org/10.1111/j.1461-0248.2012.01826.x>
- Martins, E. P., & Hansen, T. F. (1997). Phylogenies and the comparative method: A general approach to incorporating phylogenetic information into analysis of interspecific data. *American Naturalist*, 149, 646–667.
- Matthews, B., Narwani, A., Hausch, S., Nonaka, E., Peter, H., Yamamichi, M., ... Turner, C. B. (2011). Toward an integration of evolutionary biology and ecosystem science. *Ecology Letters*, 14, 690–701. <https://doi.org/10.1111/j.1461-0248.2011.01627.x>
- McManus, K. M., Asner, G. P., Martin, R. E., Dexter, K. G., Kress, W. J., & Field, C. B. (2016). Phylogenetic structure of foliar spectral traits in tropical forest canopies. *Remote Sensing*, 8, 196. <https://doi.org/10.3390/rs8030196>
- Mouquet, N., Devictor, V., Meynard, C. N., Munoz, F., Bersier, L.-F., Chave, J., ... Thuiller, W. (2012). Ecophylogenetics: Advances and perspectives. *Biological Reviews*, 87, 769–785. <https://doi.org/10.1111/j.1469-185X.2012.00224.x>
- Münkemüller, T., Lavergne, S., Bzeznik, B., Dray, S., Jombart, T., Schiffrers, K., & Thuiller, W. (2012). How to measure and test phylogenetic signal. *Methods in Ecology and Evolution*, 3, 43–756. <https://doi.org/10.1111/j.2041-210X.2012.00196.x>
- Narwani, A., Matthews, B., Fox, J., & Venail, P. (2015). Using phylogenetics in community assembly and ecosystem function research. *Functional Ecology*, 29, 589–591. <https://doi.org/10.1111/1365-2435.12431>
- Oksanen, J., Blanchet, F., Kindt, R., Legendre, P., & O'Hara, R. (2016). Vegan: Community ecology package. R package version 2.3-3. Retrieved from <https://CRAN.R-project.org/package=vegan>
- Pagel, M. (1997). Inferring evolutionary processes from phylogenies. *Zoologica Scripta*, 26, 331–348. <https://doi.org/10.1111/j.1463-6409.1997.tb00423.x>
- Pagel, M. (1999). Inferring the historical patterns of biological evolution. *Nature*, 401, 877–884. <https://doi.org/10.1038/44766>
- Pan, X., Cornelissen, J. H. C., Zhao, W.-W., Liu, G.-F., Hu, Y.-K., Prinzing, A., ... Cornwell, W. K. (2014). Experimental evidence that the Ornstein-Uhlenbeck model best describes the evolution of leaf litter decomposability. *Ecology and Evolution*, 4, 3339–3349. <https://doi.org/10.1002/ece3.1115>
- Paradis, E., Claude, J., & Strimmer, K. (2004). APE: Analyses of phylogenetics and evolution in R language. *Bioinformatics*, 20, 289–290. <https://doi.org/10.1093/bioinformatics/btg412>
- Pearse, I. S., & Hipp, A. L. (2009). Phylogenetic and trait similarity to a native species predict herbivory on non-native oaks. *Proceedings of the National Academy of Science of the USA*, 106, 18097–18102. <https://doi.org/10.1073/pnas.0904867106>
- Pearse, I. S., & Hipp, A. L. (2012). Global patterns of leaf defenses in oak species. *Evolution*, 66, 2272–2286. <https://doi.org/10.1111/j.1558-5646.2012.01591.x>
- Petersen, R. C., & Cummins, K. W. (1974). Leaf processing in a woodland stream. *Freshwater Biology*, 4, 343–368. <https://doi.org/10.1111/j.1365-2427.1974.tb00103.x>
- Pietsch, K. A., Ogle, K., Cornelissen, J. H. C., Cornwell, W. K., Bönisch, G., Craine, J. M., ... Wirth, C. (2014). Global relationship of wood and leaf litter decomposability: The role of functional traits within and across plant organs. *Global Ecology and Biogeography*, 23, 1046–1057. <https://doi.org/10.1111/geb.12172>
- Qian, H., & Jin, Y. (2016). An updated megaphylogeny of plants, a tool for generating plant phylogenies and an analysis of phylogenetic community structure. *Journal of Plant Ecology*, 9(2), 233–239. <https://doi.org/10.1093/jpe/rtv047>
- R Core Team. (2018). R: A language and environment for statistical computing version 3.4.4. Vienna, Austria: R Foundation for Statistical Computing. Retrieved from <http://www.R-project.org/>
- Raymond, P. A., Hartmann, J., Lauerwald, R., Sobek, S., McDonald, C., Hoover, M., ... Guth, P. (2013). Global carbon dioxide emissions from inland waters. *Nature*, 503, 355–359. <https://doi.org/10.1038/nature12760>
- Revell, L. J. (2010). Phylogenetic signal and linear regression on species data. *Methods in Ecology and Evolution*, 1, 319–329. <https://doi.org/10.1111/j.2041-210X.2010.00044.x>
- Schmerler, S. B., Clement, W. L., Beaulieu, J. M., Chatelet, D. S., Sack, L., Donoghue, M. J., & Edwards, E. J. (2012). Evolution of leaf form correlates with tropical-temperate transitions in Viburnum (Adoxaceae). *Proceedings of the Royal Society B*, 279, 3905–3913. <https://doi.org/10.1098/rspb.2012.1110>
- Schweitzer, J. A., Bailey, J. K., Rehill, B. J., Martinsen, G. D., Hart, S. C., Lindroth, R. L., ... Whitham, T. G. (2004). Genetically based trait in

- dominant tree affects ecosystem processes. *Ecology Letters*, 7, 127–134. <https://doi.org/10.1111/j.1461-0248.2003.00562.x>
- Senior, J. K., Potts, B. M., Davies, N. W., Wooliver, R. C., Schweitzer, J. A., Bailey, J. K., & O'Reilly-Wapstra, J. M. (2016). Phylogeny explains variation in the root chemistry of Eucalyptus species. *Journal of Chemical Ecology*, 42, 1086–1097. <https://doi.org/10.1007/s10886-016-0750-7>
- Spiegelhalter, D. J., Best, N. G., Carlin, B. P., & van der Linde, A. (2002). Bayesian measures of model complexity and fit. *Journal of the Royal Statistical Society B*, 64, 583–639. <https://doi.org/10.1111/1467-9868.00353>
- Srivastava, D. S., Cadotte, M. W., MacDonald, A. A. M., Marushia, R. G., & Mirotchnick, N. (2012). Phylogenetic diversity and the functioning of ecosystems. *Ecology Letters*, 15, 637–648. <https://doi.org/10.1111/j.1461-0248.2012.01795.x>
- Sundqvist, M. K., Giesler, R., & Wardle, D. A. (2011). Within- and across-species responses of plant traits and litter decomposition to elevation across contrasting vegetation types in subarctic tundra. *PLoS ONE*, 6, e27056. <https://doi.org/10.1371/journal.pone.0027056>
- Tank, J. L., Rosi-Marshall, E. J., Griffiths, N. A., Entek, S. A., & Stephen, M. L. (2010). A review of allochthonous organic matter dynamics and metabolism in streams. *Journal of the North American Benthological Society*, 29, 118–146. <https://doi.org/10.1899/08-170.1>
- Tibshirani, R. (1996). Regression shrinkage and selection via the lasso. *Journal of the Royal Statistical Society B*, 58, 267–288. <https://doi.org/10.1111/j.2517-6161.1996.tb02080.x>
- Tibshirani, R. (2011). Regression shrinkage and selection via the lasso: A retrospective. *Journal of the Royal Statistical Society B*, 73, 273–282. <https://doi.org/10.1111/j.1467-9868.2011.00771.x>
- Tiegs, S. D., Costello, D. M., Isken, M. W., Woodward, G., McIntyre, P. B., Gessner, M. O., ... Zwart, J. A. (2019). Global patterns and drivers of ecosystem functioning in rivers and riparian zones. *Science Advances*, 5(1), eaav0486. <https://doi.org/10.1126/sciadv.aav0486>
- Tuchman, N. C., Wetzel, R. G., Rier, S. T., Wahtera, K. A., & Teeri, J. A. (2002). Elevated atmospheric CO₂ lowers leaf litter nutritional quality for stream ecosystem food webs. *Global Change Biology*, 8, 163–170. <https://doi.org/10.1046/j.1365-2486.2002.00460.x>
- van der Linde, A. (2005). DIC in variable selection. *Statistica Neerlandica*, 59, 45–56. <https://doi.org/10.1111/j.1467-9574.2005.00278.x>
- Wallace, J. B., Eggert, S. L., Meyer, J. L., & Webster, J. R. (1997). Multiple trophic levels of a forest stream linked to terrestrial litter inputs. *Science*, 277(5322), 102–104. <https://doi.org/10.1126/science.277.5322.102>
- Webb, C., & Donoghue, M. (2005). Phylomatic: Tree assembly for applied phylogenetics. *Molecular Ecology Notes*, 5, 181–183. <https://doi.org/10.1111/j.1471-8286.2004.00829.x>
- Webster, J. R., & Benfield, E. F. (1986). Vascular plant breakdown in freshwater ecosystems. *Annual Review of Ecology and Systematics*, 17, 567–594. <https://doi.org/10.1146/annurev.es.17.110186.003031>
- Weedon, J. T., Cornwell, W. K., Cornelissen, J. H. C., Zanne, A. E., Wirth, C., & Coomes, D. A. (2009). Global meta-analysis of wood decomposition rates: A role for trait variation among tree species? *Ecology Letters*, 12, 45–56. <https://doi.org/10.1111/j.1461-0248.2008.01259.x>
- Weisser, W. W., Roscher, C., Meyer, S. T., Ebeling, A., Luo, G., Allan, E., ... Eisenhauer, N. (2017). Biodiversity effects on ecosystem functioning in a 15-year grassland experiment: Patterns, mechanisms, and open questions. *Basic and Applied Ecology*, 23, 1–73. <https://doi.org/10.1016/j.baae.2017.06.002>
- Whitham, T. G., Bailey, J. K., Schweitzer, J. A., Shuster, S. M., Bangert, R. K., LeRoy, C. J., ... Wooley, S. C. (2006). A framework for community and ecosystem genetics: From genes to ecosystems. *Nature Reviews Genetics*, 7, 510–523. <https://doi.org/10.1038/nrg1877>
- Willis, C. G., Ruhfel, B., Primack, R. B., Miller-Rushing, A. J., & Davis, C. C. (2008). Phylogenetic patterns of species loss in Thoreau's woods are driven by climate change. *Proceedings of the National Academy of Sciences USA*, 105, 17029–17033. <https://doi.org/10.1073/pnas.0806446105>
- Wright, I. J., Reich, P. B., Westoby, M., Ackerly, D. D., Baruch, Z., Bongers, F., ... Villar, R. (2004). The worldwide leaf economics spectrum. *Nature*, 428, 821–827. <https://doi.org/10.1038/nature02403>
- Yang, J., Ci, X., Lu, M., Zhang, G., Cao, M., Li, J., & Lin, L. (2014). Functional traits of tree species with phylogenetic signal co-vary with environmental niches in two large forest dynamics plots. *Journal of Plant Ecology*, 7, 115–125. <https://doi.org/10.1093/jpe/rtt070>
- Yu, G., Smith, D. K., Zhu, H., Guan, Y., Lam, T.-T.-Y., & McInerney, G. (2016). ggtree: An R package for visualization and annotation of phylogenetic trees with their covariates and other associated data. *Methods in Ecology and Evolution*, 8, 28–36. <https://doi.org/10.1111/2041-210X.12628>
- Zanne, A. E., Tank, D. C., Cornwell, W. K., Eastman, J. M., Smith, S. A., FitzJohn, R. G., ... Beaulieu, J. M. (2014). Three keys to the radiation of angiosperms into freezing environments. *Nature*, 506, 89–92. <https://doi.org/10.1038/nature12872>

SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section at the end of the article.

How to cite this article: LeRoy CJ, Hipp AL, Lueders K, et al. Plant phylogenetic history explains in-stream decomposition at a global scale. *J Ecol.* 2020;108:17–35. <https://doi.org/10.1111/1365-2745.13262>