

What are the mechanisms of Plasmodium falciparum gametocyte interactions with bone marrow cells during malaria infection?

P. falciparum gametocytes interact with bone marrow cells through multiple stage-specific mechanisms, including receptor-mediated adhesion, extracellular vesicle signaling, and mechanical retention processes.

Abstract

Plasmodium falciparum gametocytes engage bone marrow cells through a range of mechanisms that vary by gametocyte stage and cell type. Messina et al. (2018) report that immature gametocytes adhere to human mesenchymal stem cells via trypsin-sensitive ligands that trigger angiogenic cytokine secretion. Neveu et al. (2020) describe gametocyte development within late erythroblasts, mediated by extracellular vesicles and glycophorin A, which delays erythroid maturation. Suresh et al. (2023) observe that soluble factors from mesenchymal stem cells enhance mosquito infectivity, while Corbett et al. (2021) note that interactions with bone marrow-derived macrophages induce tumor necrosis factor, MIP-2, and nitric oxide.

Additional studies detail alternative sequestration and retention pathways. Rogers et al. (2000) identify cytoadherence of stage III/IV gametocytes to stromal and endothelial cells via receptors such as ICAM-1, CD49c, CD166, and CD164, whereas Silvestrini et al. (2012) report minimal adhesion involving nonclassical mediators. Joice et al. (2014) describe extravascular accumulation with association to erythroblastic islands, and De Niz et al. (2018) along with Peatey et al. (2013) report that homing, transmigration, and physical retention rely on actin-dependent processes and membrane elasticity. None of the studies implicate classical PfEMP1-mediated adhesion.

In sum, the papers collectively indicate that *P. falciparum* gametocyte interactions with bone marrow cells depend on nonclassical adhesion mechanisms, mechanical retention, and host cell modification to secure an extravascular niche conducive to parasite development and transmission.

Paper search

Using your research question "What are the mechanisms of *Plasmodium falciparum* gametocyte interactions with bone marrow cells during malaria infection?", we searched across over 126 million academic papers from the Semantic Scholar corpus. We retrieved the 50 papers most relevant to the query.

Screening

We screened in papers that met these criteria:

- **Parasite Stage:** Does the study investigate *P. falciparum* gametocytes (not solely asexual stages)?
- **Tissue Type:** Does the study examine bone marrow tissue or cells (not exclusively peripheral blood)?
- **Mechanistic Analysis:** Does the study analyze molecular or cellular mechanisms (not purely descriptive or clinical outcomes)?
- **Study Type:** Is the study either primary research with original data OR a systematic review/meta-analysis specifically focused on gametocyte-bone marrow interactions?
- **Experimental Approach:** Does the study include either in vitro or in vivo experimental analysis of gametocyte-cell interactions?
- **Study Design:** Does the study go beyond a pure clinical case report by including mechanistic analysis?

We considered all screening questions together and made a holistic judgement about whether to screen in each paper.

Data extraction

We asked a large language model to extract each data column below from each paper. We gave the model the extraction instructions shown below for each column.

- **Study Design Type:**

Identify the specific type of study design used:

- Specify if it is an in vitro study, in vivo study, animal model, human study, or mixed methodology
- If an animal model, specify the exact animal species used
- Look in the methods section for explicit description of study design
- If multiple design types are used, list all in order of prominence
- If unclear, note "design not clearly specified"

Example answers:

- "In vivo rodent model using Plasmodium berghei"
- "In vitro cell culture study with human primary erythroblasts"
- "Mixed methodology: in vitro and in vivo components"

- **Experimental Model and Techniques:**

Detail the specific experimental techniques and models used:

- List all key methodological approaches (e.g., intravital microscopy, cell-cell adhesion assays, co-culture experiments)
- Specify any specialized techniques used to observe or measure gametocyte interactions
- Include details about imaging or measurement technologies
- Note any genetic modifications or transgenic parasite lines used

Look in methods and techniques sections for comprehensive details.

- **Gametocyte Interaction Mechanisms:**

Extract specific details about:

- Observed interactions between gametocytes and bone marrow cells
- Mechanisms of gametocyte movement, adhesion, or transmission
- Any molecular or cellular processes described
- Specific proteins or cellular structures involved in interactions

Focus on direct descriptions of interaction mechanisms in results and discussion sections. Prioritize direct observations and experimental findings over speculative statements.

- **Cellular Environment and Localization:**

Describe gametocyte localization and cellular environment:

- Specific bone marrow compartments where gametocytes were observed
- Stages of gametocyte development in different cellular contexts

- Interactions with specific cell types (e.g., mesenchymal stem cells, erythroblasts)
- Any changes in gametocyte behavior within different cellular environments

Extract precise location descriptions from methods, results, and discussion sections.

- **Primary Outcomes and Significant Findings:**

Extract the most significant research findings:

- Primary outcomes directly related to gametocyte interactions
- Quantitative measurements of interaction mechanisms
- Novel observations about gametocyte behavior
- Implications for understanding malaria transmission

Focus on results section and author's explicit statements about key findings. Prioritize numerical data and direct experimental observations.

Results

Characteristics of Included Studies

| Study | Study Design | Cell Types Investigated | Gametocyte Stages Studied | Key Mechanisms Examined | Full text retrieved |
|----------------------|---|---|---------------------------------|--|---------------------|
| Messina et al., 2018 | Laboratory-based cell culture with human primary bone marrow mesenchymal stem cells and Plasmodium falciparum gametocytes | Human bone marrow mesenchymal stem cells, Plasmodium falciparum gametocytes | Immature and mature gametocytes | Adhesion to human bone marrow mesenchymal stem cells, cytokine and growth factor secretion, angiogenesis | Yes |
| Neveu et al., 2020 | Mixed methodology: laboratory-based and animal/human components | Human erythroblasts, bone marrow smears | Immature and mature gametocytes | Development within erythroblasts, delay of erythroid differentiation, extracellular vesicle-mediated effects | Yes |

| Study | Study Design | Cell Types Investigated | Gametocyte Stages Studied | Key Mechanisms Examined | Full text retrieved |
|--------------------------|--|--|---|--|---------------------|
| Suresh et al., 2023 | Mixed methodology: laboratory-based cell culture with human primary cells and animal mosquito model | Mesenchymal stem cells, Plasmodium falciparum gametocytes, mosquitoes | Early and mature gametocytes | Mesenchymal stem cell-mediated enhancement of infectivity, soluble factors | No |
| Corbett et al., 2021 | Laboratory-based cell culture with immortalized mouse C57Bl/6 bone marrow-derived macrophages and transgenic Plasmodium falciparum | Mouse bone marrow-derived macrophages, Plasmodium falciparum gametocytes | Early and late gametocytes | Phagocytosis, induction of inflammatory mediators | Yes |
| Rogers et al., 2000 | Laboratory-based study with human bone marrow cells | Human bone marrow stromal and endothelial cells | Stage III, IV, V gametocytes | Cytoadherence, receptor identification (ICAM-1, CD49c, CD166, CD164) | No |
| Silvestrini et al., 2012 | Laboratory-based cell culture with human endothelial cell lines | Endothelial cell lines (including bone marrow-derived) | Immature and mature gametocytes, asexual stages | Adhesion properties, tissue specificity, receptor involvement | Yes |
| Joice et al., 2014 | Mixed methodology: animal/human study and laboratory-based components | Human bone marrow (autopsy), erythroid precursors | All gametocyte stages | Localization, association with erythroblastic islands, phagocytosis | Yes |

| Study | Study Design | Cell Types Investigated | Gametocyte Stages Studied | Key Mechanisms Examined | Full text retrieved |
|---------------------|---|---|---------------------------------|---|---------------------|
| De Niz et al., 2018 | Mixed methodology: animal rodent model (Plasmodium berghei) and human autopsy study | Mouse bone marrow, human autopsy samples | Immature and mature gametocytes | Homing, transmigration, deformability, actin-dependence | Yes |
| Peatey et al., 2013 | Laboratory-based cell culture with erythroid progenitors | Erythroid progenitors, Plasmodium falciparum gametocytes | Developing gametocytes | Enhancement of gametocyto-genesis, membrane elasticity | No |
| Neveu et al., 2018 | Laboratory-based cell culture with human primary erythroblasts and erythroid cell lines | Human erythroblasts, erythroid cell lines, gametocyte-infected erythrocytes | Immature gametocytes | Adhesion assays, STEVOR protein role, membrane topology | Yes |

Study Design:

- Laboratory-based only:6 studies
- Mixed methodology (laboratory-based and animal/human):4 studies
- Animal/human only:We didn't find mention of any studies using only animal or human methods.

Cell Types Investigated:

- Plasmodium falciparum gametocytes:4 studies
- Human bone marrow mesenchymal stem cells or mesenchymal stem cells:2 studies
- Human erythroblasts, erythroid precursors, progenitors, or cell lines:4 studies
- Endothelial cells (including lines):2 studies
- Stromal cells:1 study
- Macrophages (mouse bone marrow-derived):1 study
- Mosquitoes:1 study
- Mouse bone marrow:2 studies
- Human bone marrow (autopsy or smears):3 studies
- Gametocyte-infected erythrocytes:1 study

Gametocyte Stages Studied:

- Immature or early gametocytes:5 studies
- Mature or late gametocytes:5 studies

- All gametocyte stages:1 study
- Specific stages III, IV, and V:1 study
- Developing gametocytes:1 study
- Asexual stages:1 study
- Exclusive focus on only one stage:We didn't find mention of studies focusing exclusively on only one of immature or mature stages; most included both or all.

Key Mechanisms Examined:

- Adhesion or cytoadherence:4 studies
- Cytokine or growth factor secretion:1 study
- Angiogenesis:1 study
- Development or differentiation:2 studies
- Extracellular vesicle-mediated effects:1 study
- Mesenchymal stem cell-mediated enhancement of infectivity or soluble factors:1 study
- Phagocytosis:2 studies
- Induction of inflammatory mediators:1 study
- Receptor identification or involvement:2 studies
- Tissue specificity:1 study
- Localization or association with erythroblastic islands:1 study
- Homing, transmigration, deformability, or actin-dependence:1 study
- Enhancement of gametocytogenesis or membrane elasticity:1 study
- STEVOR protein role or membrane topology:1 study

Thematic Analysis

Physical Interactions and Adhesion Mechanisms

| Study | Interaction Type | Cell Types Involved | Molecular Mediators | Functional Outcome |
|----------------------|--|--|---|---|
| Messina et al., 2018 | Adhesion (trypsin-sensitive, not involving ICAM-1) | Immature gametocytes, human bone marrow mesenchymal stem cells | Trypsin-sensitive ligands (not PfEMP1/ICAM-1) | Induction of angiogenic cytokines, facilitation of gametocyte release |
| Neveu et al., 2020 | Development within host cell | Gametocytes, late erythroblasts | Not PfEMP1; glycophorin A, extracellular vesicles | Delay in erythroid maturation, gametocyte maturation |
| Suresh et al., 2023 | Enhancement of infectivity | Gametocytes, mesenchymal stem cells | Soluble factors (no mention found) | Increased oocyst numbers in mosquitoes |

| Study | Interaction Type | Cell Types Involved | Molecular Mediators | Functional Outcome |
|--------------------------|--------------------------------------|---|--|--|
| Corbett et al., 2021 | Phagocytosis, immune activation | Gametocytes, mouse bone marrow-derived macrophages | Tumor necrosis factor, MIP-2, nitric oxide | Inflammatory mediator production, possible impact on gametocyte survival |
| Rogers et al., 2000 | Cytoadherence (not involving CD36) | Stage III/IV gametocytes, stromal/endothelial cells | ICAM-1, CD49c, CD166, CD164 | Stage-specific adhesion, possible retention in bone marrow |
| Silvestrini et al., 2012 | Minimal adhesion | Immature gametocytes, endothelial cells | Not ICAM-1 or CD36 | Suggests alternative sequestration mechanisms |
| Joice et al., 2014 | Localization, non-adhesive retention | Gametocytes, erythroblastic islands | Pfs16, CD71 | Extravascular accumulation, reduced phagocytosis |
| De Niz et al., 2018 | Homing, transmigration | Gametocytes, bone marrow endothelium | Actin, P-selectin (hypothesized) | Extravascular localization, mobility |
| Peatey et al., 2013 | Physical retention | Gametocytes, erythroid progenitors | Membrane elasticity (no specific protein) | Enhanced gametocytogenesis, retention in marrow |
| Neveu et al., 2018 | Lack of adhesion | Gametocyte-infected erythrocytes, erythroblasts | STEVR (not exposed) | No specific adhesion, alternative mechanisms likely |

Key patterns across these 10 studies:

- Interaction Type:
 - Adhesion, cytoadherence, or retention mechanisms: 6 studies (including minimal, physical, non-adhesive, and lack of adhesion)
 - Development within host cell: 1 study
 - Enhancement of infectivity: 1 study
 - Phagocytosis or immune activation: 1 study
 - Homing or transmigration: 1 study
- Molecular Mediators:
 - Adhesion molecules (ICAM-1, CD49c, CD166, CD164, CD71, P-selectin, glycophorin A): 4 studies

- Non-classical, unknown, or alternative mediators (trypsin-sensitive ligands not PfEMP1/ICAM-1, not ICAM-1/CD36, STEVOR not exposed, membrane elasticity, soluble factors, actin):7 studies
 - Immune mediators (tumor necrosis factor, MIP-2, nitric oxide):1 study
 - Extracellular vesicles:1 study
 - Pfs16:1 study
 - Classical PfEMP1-mediated adhesion:We did not find mention of this in the included studies.
- Functional Outcome:
 - Retention, sequestration, or accumulation in bone marrow or extravascular space:5 studies
 - Effects on gametocyte maturation, development, or enhanced gametocytogenesis:3 studies
 - Facilitation of gametocyte release or increased mobility:2 studies
 - Immune or inflammatory responses:2 studies
 - Increased infectivity to mosquitoes:1 study
 - Induction of angiogenic cytokines:1 study
 - Delay in erythroid maturation:1 study
 - Alternative mechanisms for sequestration or retention:2 studies

Among the included studies, we did not find evidence for classical PfEMP1-mediated adhesion. Several studies suggested alternative or non-classical mechanisms for gametocyte retention and development in the bone marrow.

Cellular Homing and Migration

| Study | Interaction Type | Cell Types Involved | Molecular Mediators | Functional Outcome |
|----------------------|----------------------------|---|----------------------------------|---|
| De Niz et al., 2018 | Homing, transmigration | Gametocytes, bone marrow/spleen endothelium | Actin, P-selectin (hypothesized) | Extravascular localization, high deformability, mobility |
| Joice et al., 2014 | Extravascular accumulation | Gametocytes, bone marrow parenchyma | No mention found | Retention in extravascular space, association with erythroblastic islands |
| Peatey et al., 2013 | Retention via elasticity | Gametocytes, erythroid progenitors | Membrane elasticity | Enhanced retention in marrow |
| Messina et al., 2018 | Angiogenesis induction | Gametocytes, human bone marrow mesenchymal stem cells | Cytokines and growth factors | Potential facilitation of gametocyte release |

Key findings from these 4 studies:

- Interaction types: Each study focused on a distinct interaction: homing/transmigration, extravascular accumulation, retention via elasticity, or angiogenesis induction.
- Cell types: All studies involved gametocytes, with each pairing a different additional cell type.
- Molecular mediators: Actin and P-selectin (hypothesized), membrane elasticity, and cytokines/growth factors were each reported in one study. One study did not mention a specific mediator.
- Functional outcomes: Extravascular localization, high deformability, mobility, retention in extravascular space, association with erythroblastic islands, enhanced retention in marrow, and facilitation of gametocyte release were each reported in only one study.

We did not find mention of any study reporting more than one interaction type or more than one molecular mediator (except for De Niz et al., which listed two mediators). Each study focused on a distinct interaction, cell pairing, mediator, and outcome.

Host Cell Modifications

| Study | Interaction Type | Cell Types Involved | Molecular Mediators | Functional Outcome |
|----------------------|----------------------------------|---|--|--|
| Neveu et al., 2020 | Infection, differentiation delay | Gametocytes, late erythroblasts | Extracellular vesicles, glycophorin A | Delayed erythroid maturation, increased oxidative stress |
| Suresh et al., 2023 | Infectivity enhancement | Gametocytes, mesenchymal stem cells | Soluble factors | Increased mosquito infectivity |
| Messina et al., 2018 | Cytokine induction | Gametocytes, human bone marrow mesenchymal stem cells | Angiogenic cytokines | Enhanced angiogenesis, possible gametocyte release |
| Corbett et al., 2021 | Immune activation | Gametocytes, bone marrow-derived macrophages | Tumor necrosis factor, MIP-2, nitric oxide | Inflammatory mediator production, possible impact on gametocyte survival |

Key findings from these 4 studies:

- Interaction types: Each study examined a different interaction: infection/differentiation delay, infectivity enhancement, cytokine induction, or immune activation.
- Cell types: All studies involved gametocytes, with each pairing a different additional cell type.
- Molecular mediators: Each study identified different mediators: extracellular vesicles, glycophorin A, soluble factors, angiogenic cytokines, tumor necrosis factor, MIP-2, and nitric oxide.
- Functional outcomes: Delayed erythroid maturation/increased oxidative stress, increased mosquito infectivity, enhanced angiogenesis/possible gametocyte release, and inflammatory mediator production/possible impact on gametocyte survival were each unique to a single study.

We did not find mention of any overlap in interaction type, molecular mediator, or functional outcome across these four studies, except that all studies involved gametocytes.

Summary Table: Mechanistic Interactions

| Study | Interaction Type | Cell Types Involved | Molecular Mediators | Functional Outcome |
|--------------------------|--|--|---|---|
| Messina et al., 2018 | Adhesion, cytokine induction | Immature gametocytes, human bone marrow mesenchymal stem cells | Trypsin-sensitive ligands, angiogenic cytokines | Angiogenesis, gametocyte release |
| Neveu et al., 2020 | Intracellular development, differentiation delay | Gametocytes, late erythroblasts | Extracellular vesicles, glycophorin A | Delayed erythroid maturation, gametocyte maturation |
| Suresh et al., 2023 | Infectivity enhancement | Gametocytes, mesenchymal stem cells | Soluble factors | Increased mosquito infectivity |
| Corbett et al., 2021 | Phagocytosis, immune activation | Gametocytes, bone marrow-derived macrophages | Tumor necrosis factor, MIP-2, nitric oxide | Inflammatory mediator production |
| Rogers et al., 2000 | Cytoadherence | Stage III/IV gametocytes, stromal/endothelial cells | ICAM-1, CD49c, CD166, CD164 | Stage-specific adhesion |
| Silvestrini et al., 2012 | Minimal adhesion | Immature gametocytes, endothelial cells | Not ICAM-1 or CD36 | Alternative sequestration mechanisms |
| Joice et al., 2014 | Extravascular localization | Gametocytes, erythroblastic islands | Pfs16, CD71 | Retention, reduced phagocytosis |
| De Niz et al., 2018 | Homing, transmigration | Gametocytes, bone marrow/spleen | Actin, P-selectin (hypothesized) | Extravascular localization, mobility |
| Peatey et al., 2013 | Physical retention | Gametocytes, erythroid progenitors | Membrane elasticity | Enhanced gametocytogenesis, retention |
| Neveu et al., 2018 | Lack of adhesion | Gametocyte-infected erythrocytes, erythroblasts | STEVOR (not exposed) | No specific adhesion |

Interaction Types:

- Adhesion-related interactions (including adhesion, cytoadherence, minimal adhesion, physical retention, and lack of adhesion):5 studies
- Cytokine induction or immune activation:2 studies
- Extravascular localization, homing, or transmigration:3 studies
- Infectivity enhancement, intracellular development, differentiation delay, and phagocytosis:Each reported in 1 study

Cell Types Involved:

- Gametocytes or related forms (including immature gametocytes, stage III/IV gametocytes, and gametocyte-infected erythrocytes):9 studies
- Mesenchymal stromal cells (mesenchymal stem cells or human bone marrow mesenchymal stem cells):2 studies
- Erythroblasts, late erythroblasts, or erythroid progenitors:3 studies
- Endothelial or stromal cells:3 studies
- Bone marrow or spleen:1 study
- Bone marrow-derived macrophages, erythroblastic islands, and gametocyte-infected erythrocytes:Each involved in 1 study

Molecular Mediators:

- Unique molecular mediators in each study, with no mediator reported in more than one study.
- ICAM-1, CD49c, CD166, and CD164:1 study
- Trypsin-sensitive ligands and angiogenic cytokines:1 study
- Extracellular vesicles and glycophorin A:1 study
- Soluble factors, tumor necrosis factor, MIP-2, nitric oxide, Pfs16, CD71, actin, P-selectin, membrane elasticity, and STEVOR (not exposed):Each reported in 1 study
- No molecular mediator was reported in more than one study.

Functional Outcomes:

- Adhesion, retention, or sequestration outcomes (including stage-specific, physical, and alternative mechanisms):6 studies
- Gametocyte release, maturation, or enhanced gametocytogenesis:3 studies
- Extravascular localization or mobility:2 studies
- Angiogenesis, delayed erythroid maturation, increased mosquito infectivity, inflammatory mediator production, reduced phagocytosis, and no specific adhesion:Each reported in 1 study

Synthesis

Key insights from the included studies:

- Diversity of Mechanisms:The included studies indicate that *Plasmodium falciparum* gametocyte interactions with bone marrow cells involve a range of mechanisms, including non-classical adhesion, physical retention, cellular homing, and host cell modification.

- **Stage- and Cell-Type Specificity:** Mechanisms are highly specific to gametocyte stage and the type of bone marrow cell involved. Immature gametocytes are frequently associated with the extravascular bone marrow niche and interact with mesenchymal stem cells, erythroblasts, and macrophages.
- **Absence of Classical Pathways:** Classical cytoadherence pathways, such as those involving PfEMP1, were not identified in the included studies. Instead, alternative receptors (such as ICAM-1, CD49c, CD166, CD164, CD71, P-selectin, and glycophorin A) or physical properties (such as deformability and membrane elasticity) were implicated.
- **Host Cell Responses:** Host cell responses to gametocyte interactions included altered cytokine secretion, delayed erythroid maturation, and enhanced infectivity, which may influence the bone marrow environment and facilitate parasite transmission.
- **Evidence Base Limitations:** The majority of studies used laboratory-based or animal models, with relatively few including animal or human data. The diversity of experimental systems and incomplete mechanistic characterization limit the ability to compare the relative importance of specific pathways across studies.

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