

#### Review from Lecture 2

- Alpha diversity describes a single community/ sample, and includes metrics of richness, evenness, phylogenetic diversity, and other summative metrics of diversity.
- Because sequencing success can be highly variable, rarefaction is used to ensure an even-depth of sequences across communities that will be compared.
- An OTU table is the input file for community analyses.
  It contains information about the abundance of each
  OTU within every sample. OTU tables can be
  (classic, .txt) or (.biom) format.

#### Tutorial: what did we do?

- Made taxonomic assignments of our sequences: assign\_taxonomy.py
- Made OTU tables (biom + classic): make\_otu\_table.py
- Made a phylogenetic tree of our representative sequences: make\_phylogeny.py
- Rarefied to an equal sequencing depth: alpha\_rarefaction.py
- Calculated & visualized alpha diversity: alpha\_diversity.py, summarize\_taxa\_through\_plots.py

## Questions from this morning?



#### Lecture 3: Beta diversity

- What questions can you ask about your microbial communities?
- Beta-diversity
- Gradients versus categories (clusters)
- Introduction to community resemblance
- Visualizing microbial communities

## Questions about microbial communities

Summary information for each community:
 Alpha diversity

• Differences between communities: Beta diversity

## Comparative diversity

- Space / Time
- Categories (e.g., treatment v control)
- Gradients/empirical measurements (e.g., pH, blood sugar levels, temperature)

 Look forward to the R lecture on category/ gradient analyses!

# Beta diversity requires a measure of pair-wise community resemblance

- Resemblance = distance, similarity, dissimilarity
- Important decisions in choosing a resemblance metric:
  - Weighted v. Unweighted
  - Phylogenetic v. Taxonomic
- All pairs of resemblances are included in a sample by sample resemblance (distance/similarity) matrix
  - Simplifies the data and the analysis
- Choice of resemblance metric will influence the outcome of community analysis

#### Making a Resemblance Matrix

## 1. OTU table (usually relativized)

	Caterpillar 1	Caterpillar 2	Caterpillar 3
OTU 1	0	0.966	0.179
оти з	0.047	0.002	0.039
оти з	0.953	0.032	0.782

2. Chose appropriate resemblance (e.g., Bray Curtis, Unifrac)



3. Create a square (observation x observation) resemblance matrix from pair-wise comparisons.

	Caterpillar 1	Caterpillar 2	Caterpillar 3
Caterpillar 1	0		
Caterpillar 2	0.966	0	
Caterpillar 3	0.179	0.787	9

### **Examples of Resemblance metrics**

Phylogighted constricts







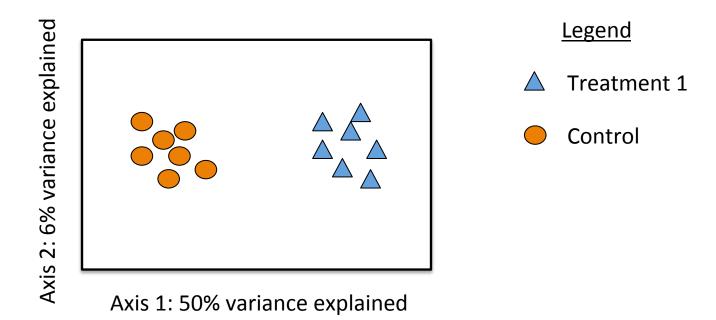


Metric name	Sørenson	Bray- Curtis	Weighted Unifrac	Unweighted Unifrac
Accounts for				
Composition	X	X	X	X
OTU abundances?		X	X	
Phylogenetic diversity?			X	<b>X</b>

#### What is the purpose of the analysis?

- 1. Exploration: hypothesis generating, perfect for observational studies, includes visualizations like ordinations and clustering
- 2. Hypothesis testing: address a specific question (e.g., are there differences among treatment groups?), and usually permutation-based p-value

#### Visualizing communities: ordination



2 or 3 dimensional representation of the data Each symbol is one community (compared by the chosen resemblance metric) The distance between symbols represents the extent of differences between communities First axis often explains most variance in the data, should be labeled.

### Types of ordinations

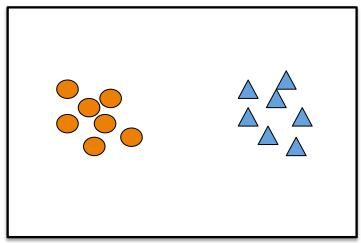
- Non-metric multidimensional scaling (NMDS)
- Principle coordinates analysis (PCoA)
- Correspondence analysis (CA)

Avoid: Principle components analysis (PCA),
 Redundancy analysis (RDA) in some situations,
 and constrained analyses unless you really
 know what you are doing

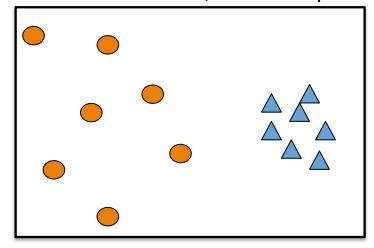
#### How do we look at ordinations?

Think about: CENTROID (mean) or DISPERSION (spread, variability)

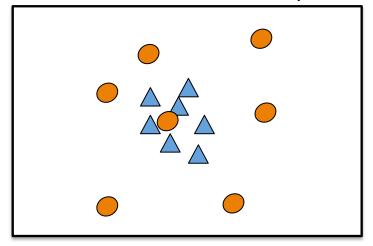
A. Different centroid, same spread



B. Different centroid, different spread



C. Same centroid, different spread



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#### Visualizing communities: clustering

A different way of visualizing the same data

