

# Introduction to R Statistics

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An introduction to using the R statistics package and the RStudio interface.

## Learning objectives

1. Read data from files and output results to files
2. Extract relevant portions of datasets
3. Run standard statistical tests in R, including Student's  $t$ , linear regression and analysis of variance

## Setup

First we need to setup our development environment. We need to create two folders: 'data' will store the data we will be analyzing, and 'output' will store the results of our analyses.

```
dir.create(path = "data")
dir.create(path = "output")
```

## Analysis of Variance (ANOVA)

For our first set of analyses, we'll use a dataset that comes pre-loaded in R. The `iris` data is from early statistical work of R.A. Fisher doi: 10.1111/2Fj.1469-1809.1936.tb02137.x, who used three species of *Iris* flowers to develop linear discriminant analysis.

Start by looking at the data with the `head` command:

```
head(x = iris)
```

##	Sepal.Length	Sepal.Width	Petal.Length	Petal.Width	Species
## 1	5.1	3.5	1.4	0.2	setosa
## 2	4.9	3.0	1.4	0.2	setosa
## 3	4.7	3.2	1.3	0.2	setosa
## 4	4.6	3.1	1.5	0.2	setosa
## 5	5.0	3.6	1.4	0.2	setosa
## 6	5.4	3.9	1.7	0.4	setosa

`iris` is a `data.frame`, which is probably the most commonly used data type in R. It is basically a table where each column is a variable and each row has one set of values for each of those variables. In the `iris` data, there are five columns: `Sepal.Length`, `Sepal.Width`, `Petal.Length`, `Petal.Width`, and `Species`. Each row corresponds to the measurements for an individual flower. Note that all the values in a column of a `data.frame` must be of the same type - if you try to mix numbers and words in the same column, R will "coerce" the data to a single type, which may cause problems for downstream analyses.

An investigation of our call to the `head` command illustrates two fundamental concepts in R: variables and functions.

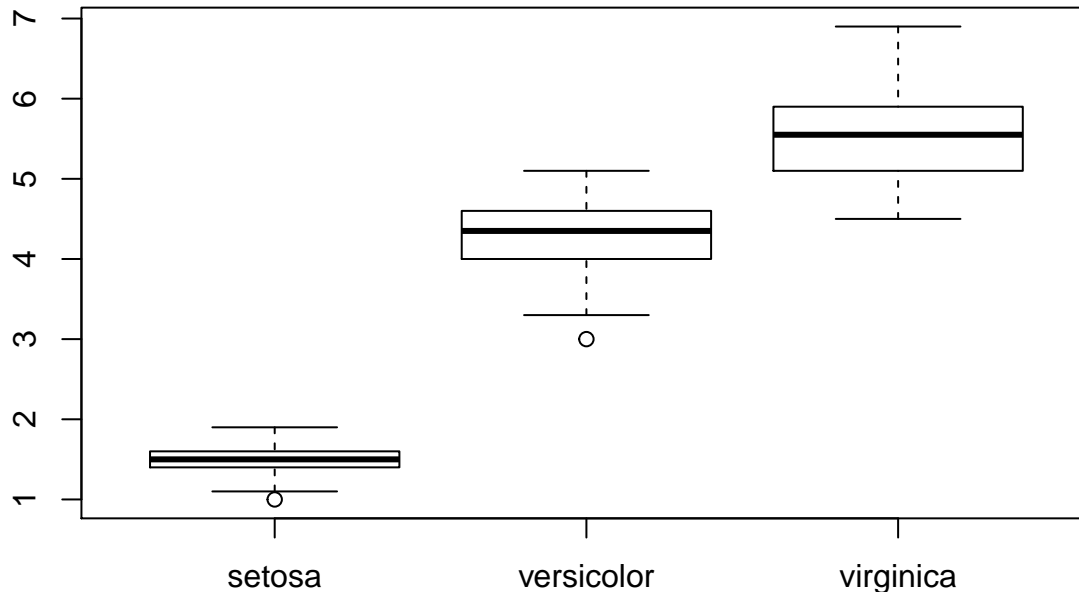
```
head(x = iris)
```

- `iris` is a variable. That is, it is a name we use to refer to some information in our computer's memory. In this case, the information is a table of flower measurements.

- `head` is the name of the function that prints out the first six rows of a `data.frame`. Most functions require some form of input; in this example, we provided one piece of input to `head`: the name of the variable for which we want the first six lines.

Another great idea when investigating data is to plot it out to see if there are any odd values. Here we use `boxplot` to show the data for each species.

```
boxplot(formula = Petal.Length ~ Species, data = iris)
```



`boxplot` uses the syntax `y ~ group`, where the reference to the left of the tilde (`~`) is the value to plot on the y-axis (here we are plotting the values of `Petal.length`) and the reference to the right indicates how to group the data (here we group by the value in the `Species` column of `iris`). Find out more about the plot by typing `?boxplot` into the console.

To keep track of what we do, we will switch from running commands directly in the console to writing R scripts that we can execute. These scripts are just text files with R commands. Create a new script file called `iris-anova.R`. Now add some key information to the top of the script, using the comment character, `#`, so R will know to ignore these lines.

```
# ANOVA on iris data set
# Jeff Oliver
# jcoliver@email.arizona.edu
# 2016-09-09
```

Commenting your code is critical in understanding why and how you did analyses when you return to the code two years from now.

The question we will first address is: *are there differences in petal length among the three species?*

We start by building an analysis of variance model with the `aov` function:

```
aov(formula = Petal.Length ~ Species, data = iris)
```

```
## Call:
##   aov(formula = Petal.Length ~ Species, data = iris)
##
## Terms:
##              Species Residuals
## Sum of Squares  437.1028    27.2226
## Deg. of Freedom      2         147
```

```
##
## Residual standard error: 0.4303345
## Estimated effects may be unbalanced
```

In this case, we pass *two* arguments to the `aov` function:

1. For the `formula` parameter, we pass `Petal.Length ~ Species`. This format is used throughout R for describing relationships we are testing. The format is `y ~ x`, where the response variables (e.g. `y`) are to the left of the tilde (`~`) and the predictor variables (e.g. `x`) are to the right of the tilde. In this example, we are asking if petal length is significantly different among the three species.
2. We also need to tell R where to find the `Petal.Length` and `Species` data, so we pass the variable name of the `iris` `data.frame` to the `data` parameter.

But we want to store the model, not just print it to the screen, so we use the assignment operator `<-` to store the product of the `aov` function in a variable of our choice

```
petal.length.aov <- aov(formula = Petal.Length ~ Species, data = iris)
```

Notice how when we execute this command, nothing printed in the console. This is because we instead sent the output of the `aov` call to a variable. If you just type the variable name,

```
petal.length.aov
```

you will see the familiar output from the `aov` function:

```
## Call:
##   aov(formula = Petal.Length ~ Species, data = iris)
##
## Terms:
##              Species Residuals
## Sum of Squares  437.1028    27.2226
## Deg. of Freedom      2        147
##
## Residual standard error: 0.4303345
## Estimated effects may be unbalanced
```

To see the results of the ANOVA, we call the `summary` function:

```
summary(object = petal.length.aov)

##              Df Sum Sq Mean Sq F value Pr(>F)
## Species        2  437.1   218.55    1180 <2e-16 ***
## Residuals     147   27.2     0.19
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

The species *do* have significantly different petal lengths ( $P < 0.001$ ).

The last thing we want to do with this code is save our results to a file. To do so, we put the call to `summary` between a pair of calls to `sink`:

```
sink(file = "output/petal-length-anova.txt")
summary(object = petal.length.aov)
sink()
```

Notice now that because we have directed output to the file “petal-length-anova.txt”, the output of `summary` will *not* be output to the console. Open the file to make sure the output was saved correctly.

Our script should look like this:

```

# ANOVA on iris data set
# Jeff Oliver
# jcoliver@email.arizona.edu
# 2016-09-09

# Run ANOVA on petal length
petal.length.aov <- aov(formula = Petal.Length ~ Species, data = iris)

# Save results to file
sink(file = "output/petal-length-anova.txt")
summary(object = petal.length.aov)
sink()

```

### Challenge 1

Use ANOVA to test for differences in sepal width among the three species. What is the value of the  $F$ -statistic?

---

### Student's $t$

So the species are different, but how? To investigate this, we need to perform pairwise comparisons between the species. We use a  $t$ -test to ask whether or not the values for two species were likely drawn from two separate populations. Just looking at the data for two species of irises, it looks like the petal lengths are different, but are the *significantly* different?

<i>I. setosa</i>	<i>I. versicolor</i>
1.4	4.7
1.4	4.5
1.3	4.9
1.5	4.0
1.4	4.6
...	...

Start by making a new R script file called ‘iris-t-test.R’ and add the header information.

```

# T-test on iris petal lengths
# Jeff Oliver
# jcoliver@email.arizona.edu
# 2016-09-09

# Compare setosa and versicolor

```

We’ll start by comparing the data of *Iris setosa* and *Iris versicolor*, so we need to create two new data objects, one corresponding to the *I. setosa* data and one for the *I. versicolor* data.

```

setosa <- iris[iris$Species == "setosa", ]
versicolor <- iris[iris$Species == "versicolor", ]

```

OK, a lot happened with those two lines. Let’s take a look:

- `iris` is the `data.frame` we worked with before.

- `iris$Species` refers to one column in `iris`, that is, the column with the name of the species (setosa, versicolor, or virginica).
- The square brackets [`<position 1>`, `<position 2>`] are used to indicate a subset of the `iris` data. A `data.frame` is effectively a two-dimensional structure - it has some number of rows (the first dimension) and some number of columns (the second dimension). We can see how many rows and columns are in a `data.frame` with the `dim` command. `dim(iris)` prints out the number of rows (150) and the number of columns (5):

```
dim(iris)
```

```
## [1] 150 5
```

We use the square brackets to essentially give an address for the data we are interested in. We tell R which rows we want in the first position and which columns we want in the second position. If a dimension is left blank, then all rows/columns are returned. For example, this returns all columns for the third row of data in `iris`:

```
iris[3, ]
```

```
## Sepal.Length Sepal.Width Petal.Length Petal.Width Species
## 3          4.7          3.2          1.3          0.2 setosa
```

So the code

```
setosa <- iris[iris$Species == "setosa", ]
```

will extract all columns (because there is nothing after the comma) in the `iris` data for those rows where the value in the `Species` column is “setosa” *and* assign that information to a variable called `setosa`.

Comparing the `iris` data and the `setosa` data, we see that there are indeed fewer rows in the `setosa` data:

```
nrow(iris)
```

```
## [1] 150
```

```
nrow(setosa)
```

```
## [1] 50
```

Now to compare the two species, we call the `t.test` function in R, passing each set of data to `x` and `y`.

```
# Compare Petal.Length of these two species
```

```
setosa.v.versicolor <- t.test(x = setosa$Petal.Length, y = versicolor$Petal.Length)
```

The output of a *t*-test is a little different than an ANOVA; we only have to enter the name of the variable to see the results (in contrast, we had to use `summary` to see the significance of our ANOVA).

```
setosa.v.versicolor
```

```
##
## Welch Two Sample t-test
##
## data: setosa$Petal.Length and versicolor$Petal.Length
## t = -39.493, df = 62.14, p-value < 2.2e-16
## alternative hypothesis: true difference in means is not equal to 0
## 95 percent confidence interval:
## -2.939618 -2.656382
## sample estimates:
## mean of x mean of y
## 1.462 4.260
```

The results include:

- Test statistic, degrees of freedom, and p-value
- The confidence interval for the difference in means between the two data sets
- The means of each data set

So we reject the hypothesis that these species have the same petal lengths. As before, though, if we want to save these results to a file, we use `sink`:

```
sink(file = "output/petal-length-setosa-versicolor-t-test.txt")
setosa.v.versicolor
sink()
```

The final script should be:

```
# T-test on iris petal lengths
# Jeff Oliver
# jcoliver@email.arizona.edu
# 2016-09-09

# Compare setosa and versicolor

# Subset data
setosa <- iris[iris$Species == "setosa", ]
versicolor <- iris[iris$Species == "versicolor", ]

# Run t-test
setosa.v.versicolor <- t.test(x = setosa$Petal.Length, y = versicolor$Petal.Length)

# Save results to file
sink(file = "output/petal-length-setosa-versicolor-t-test.txt")
setosa.v.versicolor
sink()
```

## Challenge 2

Test for significant differences in petal lengths between *I. setosa* and *I. virginica* and between *I. versicolor* and *I. virginica*.

---

## Linear regression

For this final section, we will test for a relationship between life expectancy and per capita gross domestic product (GDP). Start by downloading the data from <http://tinyurl.com/qb83k3z> (right-click or Ctrl-click on link and Save As...). Save this to the ‘data’ directory you created in the Setup section. The file has comma-separated values for 142 countries at twelve different years; the data can be loaded in R with the `read.csv` function:

```
# Test relationship between life expectancy and GDP
# Jeff Oliver
# jcoliver@email.arizona.edu
# 2016-07-29

gapminder <- read.csv(file = "data/gapminder-FiveYearData.csv")
```

This reads the file into memory and stores the data in a data frame called `gapminder`.

Recall you can see the first few rows with the `head` function.

```
head(gapminder)
```

```
##      country year      pop continent lifeExp gdpPercap
## 1 Afghanistan 1952  8425333      Asia  28.801  779.4453
## 2 Afghanistan 1957  9240934      Asia  30.332  820.8530
## 3 Afghanistan 1962 10267083      Asia  31.997  853.1007
## 4 Afghanistan 1967 11537966      Asia  34.020  836.1971
## 5 Afghanistan 1972 13079460      Asia  36.088  739.9811
## 6 Afghanistan 1977 14880372      Asia  38.438  786.1134
```

Another useful quality assurance tool is `summary`, which provides a basic description for each column in the data frame.

```
summary(gapminder)
```

```
##      country      year      pop      continent
## Afghanistan: 12  Min.   :1952  Min.   :6.001e+04  Africa :624
## Albania      : 12  1st Qu.:1966  1st Qu.:2.794e+06  Americas:300
## Algeria      : 12  Median :1980  Median :7.024e+06  Asia   :396
## Angola       : 12  Mean    :1980  Mean    :2.960e+07  Europe :360
## Argentina    : 12  3rd Qu.:1993  3rd Qu.:1.959e+07  Oceania : 24
## Australia    : 12  Max.    :2007  Max.    :1.319e+09
## (Other)      :1632
##      lifeExp      gdpPercap
## Min.   :23.60  Min.   : 241.2
## 1st Qu.:48.20  1st Qu.: 1202.1
## Median :60.71  Median : 3531.8
## Mean    :59.47  Mean    : 7215.3
## 3rd Qu.:70.85  3rd Qu.: 9325.5
## Max.    :82.60  Max.    :113523.1
##
```

For the four numeric columns (`year`, `pop`, `lifeExp`, and `gdpPercap`), some descriptive statistics are shown. For the `country` and `continent` columns the first few values and frequencies of each value are shown (i.e. there are 12 records for Afghanistan and 624 records for Africa).

For this analysis, we only want the data from 2007, so we start by subsetting those data. This creates a new variable and stores only those rows in the original `gapminder` data frame where the value in the `year` column is 2007.

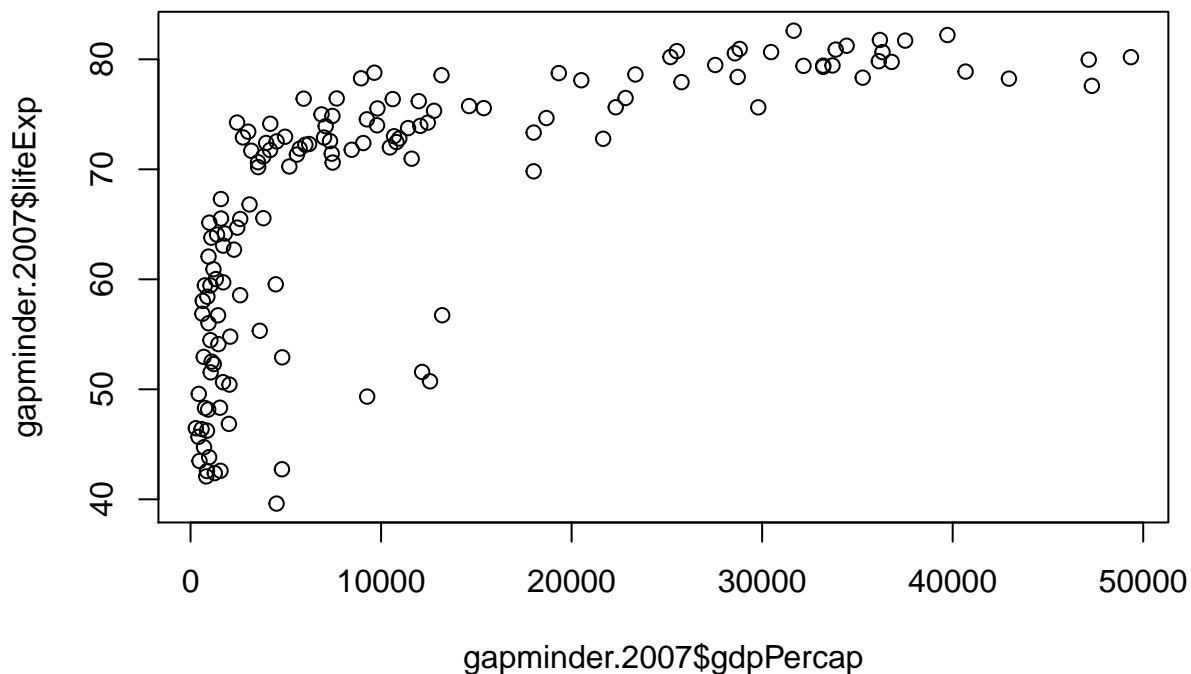
```
# Subset 2007 data
```

```
gapminder.2007 <- gapminder[gapminder$year == 2007, ]
```

As we did for the ANOVA analyses, it is usually a good idea to visually inspect the data when possible. Here we can use the `plot` function to create a scatterplot of the two columns of interest, `lifeExp` and `gdpPercap`.

```
# Plot to look at data
```

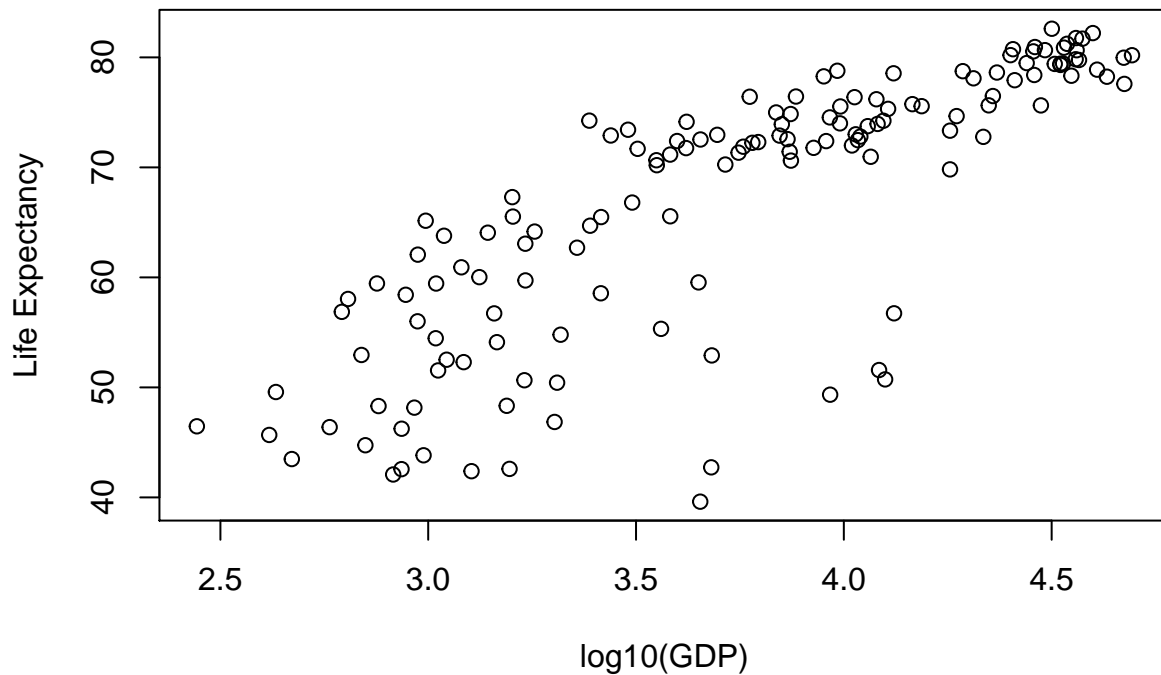
```
plot(x = gapminder.2007$gdpPercap, y = gapminder.2007$lifeExp)
```



We can see immediately that this is unlikely a linear relationship. For our purposes, we will need to log-transform the GDP data. Create a new column in the `gapminder.2007` data frame with the  $\log_{10}$ -transformed GDP and plot this transformed data.

```
# Create log-transformed GDP
gapminder.2007$logGDP <- log10(gapminder.2007$gdpPerCap)

# Plot again, with log-transformed GDP on the x-axis
plot(x = gapminder.2007$logGDP, y = gapminder.2007$lifeExp, xlab = "log10(GDP)", ylab = "Life Expectancy")
```



Notice also that we passed two additional arguments to the `plot` command: `xlab` and `ylab`. These are used to label the x- and y-axis, respectively (try the `plot` function without passing `xlab` and `ylab` arguments to



see what happens without them).

Now that the data are properly transformed, we can create the linear model for the predictability of life expectancy based on gross domestic product.

```
# Run a linear model
lifeExp.v.gdp <- lm(lifeExp ~ logGDP, data = gapminder.2007)

# Investigate results of the model
summary(lifeExp.v.gdp)

##
## Call:
## lm(formula = lifeExp ~ logGDP, data = gapminder.2007)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -25.947  -2.661   1.215   4.469  13.115
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)    4.950      3.858   1.283   0.202
## logGDP         16.585      1.019  16.283 <2e-16 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 7.122 on 140 degrees of freedom
## Multiple R-squared:  0.6544, Adjusted R-squared:  0.652
## F-statistic: 265.2 on 1 and 140 DF,  p-value: < 2.2e-16
```

For our question, the relationship between life expectancy and GDP, focus on the *coefficients* section, specifically the line for *logGDP*:

```
## logGDP          16.585      1.019  16.283 < 2e-16 ***
```

First of all, there *is* a significant relationship between these two variables ( $p < 2 \times 10^{-16}$ , or, as R reports in the `Pr(>|t|)` column,  $p < 2e-16$ ). The `Estimate` column of the results lists a value of 16.585, which means that for every 10-fold increase in per capita GDP (remember we  $\log_{10}$ -transformed GDP), life expectancy increases by almost 17 years.

As before, if we want to instead save the results to a file instead of printing them to the screen, we use the `sink` function.

```
sink(file = "output/lifeExp-gdp-regression.txt")
summary(lifeExp.v.gdp)
sink()
```

The final script should be:

```
# Test relationship between life expectancy and GDP
# Jeff Oliver
# jcoliver@email.arizona.edu
# 2016-07-29

# Read data from comma-separated values file
gapminder <- read.csv(file = "data/gapminder-FiveYearData.csv")

# Subset 2007 data
gapminder.2007 <- gapminder[gapminder$year == 2007, ]
```

```

# Plot to look at data
plot(x = gapminder.2007$gdpPercap, y = gapminder.2007$lifeExp)

# Create log-transformed GDP
gapminder.2007$logGDP <- log10(gapminder.2007$gdpPercap)

# Plot new variable
plot(x = gapminder.2007$logGDP, y = gapminder.2007$lifeExp, xlab = "log10(GDP)", ylab = "Life Expectancy")

# Run linear model
lifeExp.v.gdp <- lm(lifeExp ~ logGDP, data = gapminder.2007)

# Save results to file
sink(file = "output/lifeExp-gdp-regression.txt")
summary(lifeExp.v.gdp)
sink()

```

### Challenge 3

Test for a relationship between life expectancy and log base 2 of GDP for the 1982 data. How does life expectancy change with a four-fold increase in GDP?

---

## Answers to challenges

### Challenge 1

Use ANOVA to test for differences in sepal width among the three species. What is the value of the  $F$ -statistic?

### Solution

```

sepal.width.aov <- aov(formula = Sepal.Width ~ Species, data = iris)
summary(object = sepal.width.aov)

```

```

##              Df Sum Sq Mean Sq F value Pr(>F)
## Species      2  11.35   5.672   49.16 <2e-16 ***
## Residuals    147  16.96   0.115
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

```

The  $F$ -statistic = 49.16, and the p-value is quite small, so there are significant sepal width differences among species.

---

### Challenge 2

Test for significant differences in petal lengths between *I. setosa* and *I. virginica* and between *I. versicolor* and *I. virginica*.

## Solution

### First comparison: *I. setosa* vs. *I. virginica*

```
# Subset setosa data
setosa <- iris[iris$Species == "setosa", ]
# Subset virginica data
virginica <- iris[iris$Species == "virginica", ]
# Run t-test
setosa.v.virginica <- t.test(x = setosa$Petal.Length, y = virginica$Petal.Length)
# Print the results
setosa.v.virginica
```

```
##
## Welch Two Sample t-test
##
## data: setosa$Petal.Length and virginica$Petal.Length
## t = -49.986, df = 58.609, p-value < 2.2e-16
## alternative hypothesis: true difference in means is not equal to 0
## 95 percent confidence interval:
## -4.253749 -3.926251
## sample estimates:
## mean of x mean of y
## 1.462 5.552
```

*I. setosa* and *I. virginica* have significantly different petal lengths.

---

### Second comparison: *I. versicolor* and *I. virginica*

```
# Subset versicolor data
versicolor <- iris[iris$Species == "versicolor", ]
# Subset virginica data
virginica <- iris[iris$Species == "virginica", ]
# Run t-test
versicolor.v.virginica <- t.test(x = versicolor$Petal.Length, y = virginica$Petal.Length)
# Print the results
versicolor.v.virginica
```

```
##
## Welch Two Sample t-test
##
## data: versicolor$Petal.Length and virginica$Petal.Length
## t = -12.604, df = 95.57, p-value < 2.2e-16
## alternative hypothesis: true difference in means is not equal to 0
## 95 percent confidence interval:
## -1.49549 -1.08851
## sample estimates:
## mean of x mean of y
## 4.260 5.552
```

*I. versicolor* and *I. virginica* also have different significantly different petal lengths.

---

### Challenge 3

Test for a relationship between life expectancy and log base 2 of GDP for the 1982 data. How does life expectancy change with a four-fold increase in GDP?

### Solution

```
# Read data from comma-separated values file
gapminder <- read.csv(file = "data/gapminder-FiveYearData.csv")

# Subset 1982 data
gapminder.1982 <- gapminder[gapminder$year == 1982, ]

# Create log2-transformed GDP
gapminder.1982$log2GDP <- log2(gapminder.1982$gdpPercap)

# Run linear model
lifeExp.v.gdp <- lm(lifeExp ~ log2GDP, data = gapminder.1982)
summary(lifeExp.v.gdp)

##
## Call:
## lm(formula = lifeExp ~ log2GDP, data = gapminder.1982)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -18.7709  -2.8743   0.4812   3.6039  14.6986
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)  -0.6505     3.3463  -0.194   0.846
## log2GDP       5.1942     0.2766  18.780 <2e-16 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 5.762 on 140 degrees of freedom
## Multiple R-squared:  0.7158, Adjusted R-squared:  0.7138
## F-statistic: 352.7 on 1 and 140 DF, p-value: < 2.2e-16
```

The line to focus on is the log2GDP line in the coefficients section:

```
## log2GDP      5.1942      0.2766  18.780  <2e-16 ***
```

The coefficient for  $\log_2$  GDP in the model is positive, with increases in GDP correlating with increased life expectancy. The estimated coefficient for the relationship is 5.19. Remember that we  $\log_2$ -transformed the GDP data, so this coefficient indicates the change in life expectancy for every two-fold increase in per capita GDP. For a four-fold increase in GDP, we multiply this coefficient by two (because four is two two-fold changes) to conclude that a four-fold increase in GDP results in an increase of 10.39 years in life expectancy.

---

Questions? e-mail me at [jcoliver@email.arizona.edu](mailto:jcoliver@email.arizona.edu).