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| [Home](http://docs.google.com/home.html)  [Goals](http://docs.google.com/goals.html)  [Abstract](http://docs.google.com/abstract.html)  [Introduction](http://docs.google.com/intro.html)  [Literature Review](http://docs.google.com/review.html)  [Hypothesis/Prediction](http://docs.google.com/hypo.html)  [Procedure](http://docs.google.com/proced.html)  [Data](http://docs.google.com/data.html)  [Images](http://docs.google.com/images.html)  [Conclusions](http://docs.google.com/conc.html)  [Recommendations](http://docs.google.com/recom.html)  [Bibliography](http://docs.google.com/biblio.html)  [Journal](http://docs.google.com/journal.html)  [Web Resources](http://docs.google.com/resource.html)  [Acknowledgements](http://docs.google.com/acknow.html)  [Project Creek Watch](http://www.pleasanton.k12.ca.us/amador/creek/index.html) | Materials:   1. micropipettes that hold 1000 *u*L, 30 *u*L, 200 *u*L, 60 *u*L, 20 *u*L, 10 *u*L, 5*u*L 2. cups for each sample 3. salt 4. distilled water 5. 1.5 *u*L microfuge tube 6. centrifuge 7. 99 degrees C heat block or hot water bath (for hot bath, need floater for tubes) 8. 5% Chelex 9. microfuge rack 10. 200 *u*L PCR tube 11. Master Mix1 12. Primer Mix2 13. thermal cycler 14. loading dye 15. 2% agarose gel (agarose powder and buffer) 16. 100 bp ladder 17. power supply / gel box 18. ethidium bromide (0.5 *u*g/mL) 19. UV light box 20. safety glasses 21. gloves 22. apron / lab coat 23. +Control: 20 *u*L Master Mix, 20 *u*L Primer Mix, 10 *u*L +Control DNA 24. -Control: 10 *u*L sterile water, 20 *u*L Master Mix, 20 *u*L Primer Mix   **1) Alu Master Mix:**  5 µL 10X PCR Buffer II X 50 = 250 µL  3 µL (25 mM) MgCl2 X 50 = 150 µL  1 µL (10 mM) dATP X 50 = 50 µL  1 µL (10 mM) dCTP X 50 = 50 µL  1 µL (10 mM) dGTP X 50 = 50 µL  1 µL (10 mM) dTTP X 50 = 50 µL  0.3 µL AmpliTaq Gold (5 Units/µL) X 50 = 15 µL  7.7 µL H2O X 50 = 385 µL  **2) Alu Primer Mix:**  0.67 µL (15µM) PV92 forward primer X 50 = 33.5 µL  0.67 µL (15µM) PV92 reverse primer X 50 = 33.5 µL  18.66 µL H2O X 50 = 933 µL |

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