

Nascent Transcript Identification Using CHM13

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nascent

Updated to nascent v1

- Updated Multiqc report with v1 metrics
 - Great spot checking for data mining
- Finished up homer transcript identification
- Started on dREG(Going to have to package it up)
 - Kinda makes me want to build our own transcript identification model

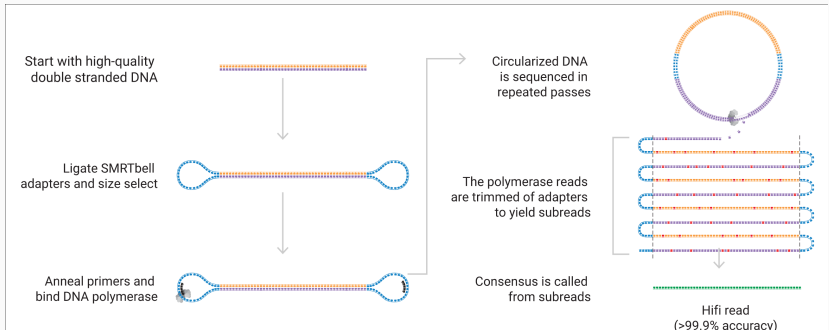
DeepVariant Approach

- Looking Through DeepVariant's Eyes | DeepVariant Blog
- They're trying to identify single bases
- We're just looking at the bigger picture
- CNNs are great at this

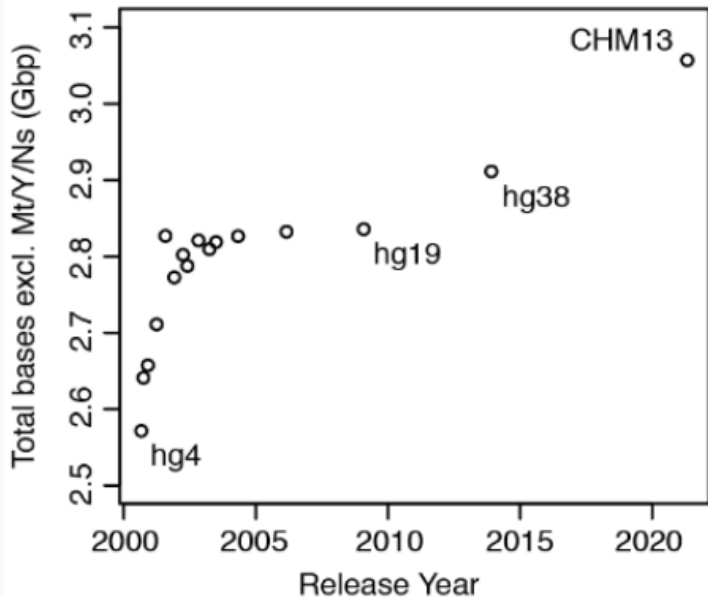
CHM13 Refresher

- Used PacBio's HiFi, and Nanopore's "ultra-long" reads to resolve complex forms of structural variation and gaps in GRCh38
- CHM13 Cell line a complete hydatidiform mole (CHM) cell line, "essentially haploid nature"

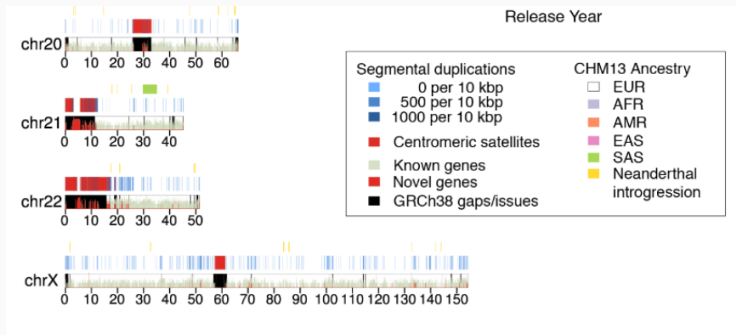
Pacbio HiFi Reads



CHM13



CHM13



CHM13 Results

The intersection of Homer identified peaks and centromeres

- Near identical number of reads mapped(There's only 5% more to align to)
- Had to use hg19 centromeres for the intersection
- There were no hits
- Reviewed the multiqc and found there was an issue with the homer identification in CHM13(18% and 6% efficiency)
- Confirmed with bedtools intersect call to aligned reads for centromeres

Tertiary analysis Best practices

Some Goals

- Reproducibility
- Easy for others to read the code
- Easy for “exploratory analysis”
- Scales (Cluster or cloud submission)
 - Avoid for loops
- Could I teach this to someone in a Summer semester?
(Applied Genomics)

Shiny New Datascience tools popping up

- Kedro
- Ploomber - Build data pipelines. FAST.
- The Julia Programming Language

Computational Biology is a beautiful mess

The first step of any bioinformatics project is to define a new file format, incompatible with all previous ones.

- Those all looked exciting. Until I needed to align some histones to hg38.
- Why waste a bunch of time rewriting scripts that already worked?

Code complexity aside

```
def bar():  
    x = 1  
    if x == 2:  
        print("Success")
```

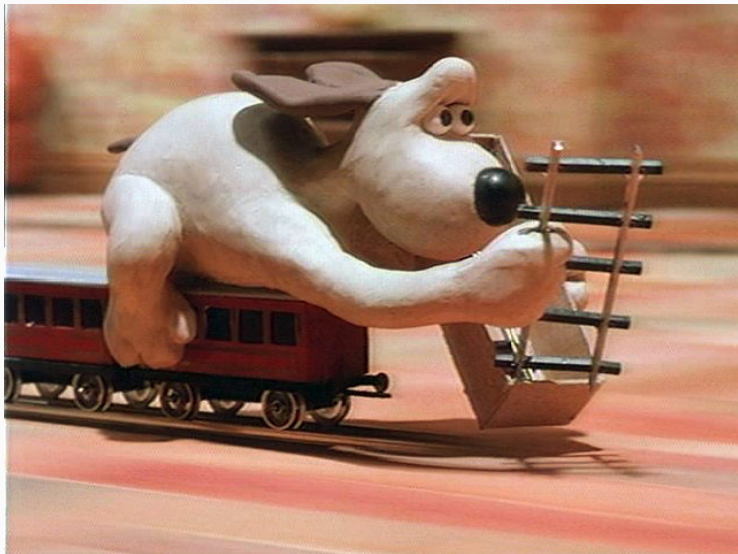
```
def foo():  
    evens = [2, 4, 6, 8, 10]  
    odds = [1, 3, 5, 7, 9]  
    for x in evens:  
        for y in odds:  
            product = x * y  
            if product % 2 == 0:  
                print "Product result is even"  
            if product % 5 == 0:  
                print "Product is divisible by 5"  
            if product % 3 == 0:  
                print "Product is divisible by 3"
```

- McCabe's Cyclomatic Complexity

What if I just used nextflow?

- I was afraid it would be too “heavy”
 - Making container images
 - Boilerplate
- Somewhat Bioinformatics specific

Reality



Some mental shifts

- Just use conda environments for scripts
- Feedback of using the results directory for creating exploratory scripts
- Utilize nf-core modules
- Don't try to make features/job submission one size fits all(Leave that to nf-core)
 - Allowed me to cut down on a lot of the boilerplate
- Could a new grad student reproduce this and pick up where I left off?

How do we compare versions of things?

```
# move to the tag v1
git checkout v1
# store results in v1 directory
nextflow run . --outdir ./results/v1
```

```
# move to the tag v2
git checkout v2
# store results in v2 directory
nextflow run . --outdir ./results/v2
```

Some examples

- [qbic-pipelines/rnadeseq](#): Differential gene expression analysis and pathway analysis of RNAseq data
- [qbic-pipelines/bamtofastq](#)
- [Functional-Genomics-Lab/eRNA-complementarity](#)