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ki, J. J., Kaden, D. A.,
 ' R., Slapikoff, S. A.,
 man, B. W. (1980) in
 ciples and Methods for
 der, A., and de Serres,
 331-364.
Spring Harbor Symp.
 3.
 i., Spector, J. F. S.,
 wn, M. M. M. (1979)
 r's Thesis M.I.T. De-
 d Food Science.
 . M., Mankovitz, R.,
 ore, G. F., Siminovitch,
 Cell 1, 9-21.
 M., and Baker, R. M.
 . Res. 44, 401-412.
 H. L., and Buckanan,
 72, 285-294.

A Nonlinear Regression Program for Small Computers

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A BASIC computer program for performing weighted nonlinear regression is described and a listing of the program is given. The program, which is small and simple to use, has been designed to be run by users with little knowledge of mathematics or computers. Robust methods of analysis are described which may be applied to data in which experimental errors are not normally distributed, and the program incorporates one such method. It is shown that the program is useful for the analysis of data conforming to the Michaelis-Menten equation, a single exponential, and to binding equations, and other applications are discussed.

The quantitative analysis of experimental data frequently requires comparison with some sort of mathematical equation or model. All too often this analysis involves a transformation of the data which are then plotted and a straight line is drawn through them. The slope and intercept may then be transformed or combined in various ways to obtain the parameters of the original equation. For example Eq. [1] describes a

$$\hat{y} = y_0 e^{-kt} \quad [1]$$

first-order decay curve,¹ and the classical method of analysis is to plot

In (y) against t . From the slope (S) and the intercept (I) the parameters are obtained using $k = -S$ and $y_0 = e^I$. A much better method for the analysis of first-order decay curves is to fit Eq. [1] directly to the data by nonlinear regression. Although this type of analysis is relatively straightforward, nonlinear regression has made very little impact in biochemistry with the notable exception of enzyme-kinetic studies.

A great many packaged computer pro-

¹ Throughout the paper a distinction will be drawn between y , the observed value of the dependent variable; \hat{y} , a value calculated for a particular set of parameter values; and \bar{y} the true, but usually unknown, value.

grams are available for performing nonlinear regression analysis but these are, without exception, long and sophisticated programs designed to be run on large computers. In this paper, a simple BASIC nonlinear regression program is presented which can be run on mini- or even microcomputers. Some of the underlying theory is presented but a understanding of this theory is not a prerequisite for using the program. The program has been deliberately limited to the situation in which the equation to be fitted has two parameters, as such equations occur quite commonly.

THEORY

Transforming experimental data into a form which may be plotted as a straight line is a useful method of displaying the data but it is not a reliable method for its analysis. Experimental errors can be grossly magnified as is the case with the Lineweaver-Burk plot of enzyme-kinetic data and fitting of a straight line to the transformed data will not, in general, yield the "best" values for the parameters. This is true, regardless of whether the fit is performed "by eye" or by linear least-squares analysis. Careful weighting of the transformed data may compensate for the distortion in certain in-

stances but in others, such as a Scatchard plot in which the observed variable appears on both axes, distortion is unavoidable. If transformation of the data is to be eliminated it is necessary to fit the mathematical equation to the data directly. The form of the equation is not usually a matter of choice, but rather it depends on some underlying theoretical model. Generally, these theoretical considerations lead to an equation which is nonlinear in the parameters and the fitting procedure will involve nonlinear regression.

On the whole, biochemists regard nonlinear regression with a mixture of awe and suspicion, as something which is beyond their capacity to comprehend. In fact it is quite simple, requiring little more than a knowledge of elementary algebra and in this section the basic principles are set out. Later, a simple and flexible computer program, which embodies these principles, will be described.

Classical Methods

Nonlinear regression. The fundamental ideas underlying the Gauss-Newton method of nonlinear regression have been described by Wilkinson (1). These ideas are best understood against a background of the principles of linear regression which will be described briefly. Consider the case where we wish to fit Eq. [2] to a set of N observations, where a_1 and a_2 are parameters to

$$\hat{y} = a_1 x_1 + a_2 x_2 \quad [2]$$

be estimated and x_1 and x_2 are independent variables. (This is not intended to imply that x_1 and x_2 are necessarily independent of one another; for example, x_2 may equal x_1^2). To estimate a_1 and a_2 we first form the sums

$$s_1 = \sum w x_1^2; s_2 = \sum w x_1 x_2; s_3 = \sum w x_2^2;$$

$$s_4 = \sum w x_1 y; s_5 = \sum w x_2 y,$$

where w is a nonnegative "weight" attached to each observation, and which is discussed in detail below. The parameters may

be calculated using

$$a_1 = (s_3 s_4 - s_2 s_5) / \Delta$$

$$a_2 = (s_1 s_5 - s_3 s_4) / \Delta$$

where

$$\Delta = s_1 s_3 - s_2^2.$$

To calculate the standard errors of a_1 and a_2 , we calculate the sum of squares of residuals (s_6) and the residual standard error (r_s) using

$$s_6 = \sum w(y - \hat{y})^2$$

$$r_s = [s_6/(N - 2)]^{1/2}$$

The standard errors of a_1 and a_2 are given by

$$SE(a_1) = r_s(s_3/\Delta)^{1/2}$$

$$SE(a_2) = r_s(s_1/\Delta)^{1/2}$$

The values of a_1 and a_2 calculated above are "best-fit" values in the sense that they minimize the weighted sum of squares, s_6 . This is necessarily so because the formulae for a_1 and a_2 are found by differentiating s_6 with respect to a_1 and a_2 , setting these derivatives equal to zero and solving the resultant simultaneous equations.

For nonlinear equations, a similar procedure does not lead to a simple solution and we cannot calculate the best-fit values for the parameters in a single step. What can be done in a single step is to take some estimates of these values and correct them to give better estimates.

Suppose we are trying to fit a nonlinear equation in which there is a single parameter, b :

$$\hat{y} = f(b)$$

An estimate, \bar{b} , will differ from the best-fit value, \hat{b} , by an unknown amount q :

$$\bar{b} = \hat{b} + q.$$

From the Taylor series we may write

$$\hat{y} = f(\bar{b} + q)$$

$$= f(\bar{b}) + qf'(\bar{b}) + \frac{q^2}{2!}f''(\bar{b})$$

$$+ \frac{q^3}{3!}f'''(\bar{b}) + \dots$$

where f' , f'' , an differentiation w all the terms in $f(\bar{b})$ as \bar{y} , and r the experimenta

In other words q can be found by the difference between the calculated dependent vari is treated as the value for q so correct due to the high-order term ever, the newl be refined by a cedure repeat "iteration."

This concept in which the parameter. Co described by 1

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These new es then be refin q_1 and q_2 ar the standard to those of q

where f' , f'' , and so on denote successive differentiation with respect to b . If we ignore all the terms in q^2 and beyond, represent $f(b)$ as \bar{y} , and replace the unknown \bar{y} with the experimental y , we can rearrange to get

$$y - \bar{y} = qf'(b).$$

In other words, an approximate value for q can be found by linear regression in which the difference between the experimental and the calculated value of y is treated as the dependent variable while the derivative $f'(b)$ is treated as the independent variable. The value for q so derived will not be exactly correct due to the approximation in ignoring high-order terms of the Taylor series. However, the newly calculated value of b may be refined by applying this correction procedure repeatedly, a process known as "iteration."

This concept may be generalized to cases in which there are more than a single parameter. Consider the arbitrary function described by Eq. [3], where b_1 and b_2 are

$$\bar{y} = f(b_1, b_2; X), \quad [3]$$

nonlinear parameters and X represents the values of one or more independent variables. If the initial estimates of the parameters are $b_1^{(0)}$ and $b_2^{(0)}$, we may calculate corrections (q_1 and q_2) to these parameters by fitting the equation

$$z = q_1 p_1 + q_2 p_2$$

in which z is the residual $(y - \bar{y})$ while p_1 and p_2 are the partial derivatives, $\delta\bar{y}/\delta b_1$ and $\delta\bar{y}/\delta b_2$. The coefficients, q_1 and q_2 , are estimated as described above for the linear case and are used to correct the values of the nonlinear parameters

$$b_1^{(1)} = b_1^{(0)} + q_1$$

$$b_2^{(1)} = b_2^{(0)} + q_2.$$

These new estimates of the parameters may then be refined in further iterations. When q_1 and q_2 are negligible ("convergence"), the standard errors of b_1 and b_2 are equal to those of q_1 and q_2 , respectively (1), and

are calculated as described earlier.

Partial derivatives. We saw above that in nonlinear regression the calculation of the corrections (q) requires values of the partial derivatives (p) which are treated as independent variables. Ideally, these derivatives should be obtained by analytical differentiation of the nonlinear function (Eq. [3]), which may involve some tedious algebra. In practice, the derivatives can be calculated to the required precision by numerical differentiation which avoids the algebra. The function (Eq. [3]) is evaluated after the parameter b_1 is perturbed by an amount d_1 :

$$\bar{y}' = f(b_1 + d_1, b_2; X)$$

and a first-order approximation of p_1 is given by

$$p_1 \approx (\bar{y}' - \bar{y})/d_1.$$

A more accurate value may be found using a second-order approximation if the function is evaluated at a second point:

$$\bar{y}'' = f(b_1 - d_1, b_2; X)$$

$$p_1 \approx (\bar{y}' - \bar{y}'')/2d_1.$$

A value for p_2 is found by applying this same procedure to b_2 . In the computer program to be described later, d_1 and d_2 are chosen to be 2% of b_1 and b_2 , respectively.

Weighting. It may happen that we have advance knowledge that some observations are more accurate than others and this information should be incorporated in the analysis. This is achieved by weighting each observation by an amount (w) which is inversely proportional to its variance, so that fitting involves minimizing the weighted sum of squares, s_6 . This same weight must also be applied in forming the sums $s_1 - s_5$ which are used to calculate regression coefficients. Frequently, these *a priori* weights are calculated from some simple weighting function. For example, the standard deviation of y may be approximately proportional to y in which case $w = 1/y^2$ will be used as the weighting function. One of the

methods of robust regression described below is based on weighting.

Robust Methods

Robust regression. The classical method for fitting a function to experimental data involves minimizing the sum of squares of residuals. Since each residual (z) is squared in the summation and as the worst observations will have the largest residuals, the fit tends to be dominated by these observations. A drastic solution to this difficulty is to discard the worst observations but to do this it is necessary to introduce an essentially arbitrary division between acceptable and unacceptable observations. A gentler procedure is to use a "robust" method in which the residuals are modified so that less emphasis is placed on the larger ones.

Wahrendorf (2) has described a robust method which he has applied to the analysis of Scatchard plot data. Briefly, the residual sum of squares is replaced with the function $\sum \rho(z)$:

$$\rho(z) = \begin{cases} z^2 & \text{if } |z| < c \\ 2c|z| - c^2 & \text{if } |z| \geq c \end{cases}$$

where c is a "robustness constant." The value of $\rho(z)$ increases as the square of z when z is numerically less than c , but thereafter increases as the absolute value of z . There is a smooth transition at $z = c$. If c is chosen to be very large, this method is indistinguishable from the normal least-squares method.

A somewhat different method has been described by Mosteller and Tukey (3) in which each squared residual is multiplied by a "bisquare weight," b_w :

$$b_w = \begin{cases} (1 - u^2)^2 & \text{if } |u| \leq 1 \\ 0 & \text{if } |u| > 1, \end{cases}$$

where $u = z/c$ and c is the robustness constant. If $z > c$, that particular observation is "weighted-out" of the analysis (i.e., ignored), while moderate-sized residuals acquire a fractional weight. Observations

which agree well with the fitted function have a small residual and are given close to a full weight of 1.0. As with *a priori* weights, the bisquare weight is applied in calculating the sums s_1-s_6 from which the regression coefficients are calculated.

If c is chosen to be very large, b_w will equal 1.0 for all observations and we have the usual least-squares method. Usually we will want to choose a value of c which is large enough that $|u| < 1$ for the great majority of observations. In the computer program to be described later, a value equal to six times the mean absolute residual ($c = 6 \sum |z|/N$) has been utilized but Mosteller and Tukey have pointed out that many other values will also work well. Bisquare weighting can be used in conjunction with *a priori* weights in which case the final weight applied will be the product of b_w and w . In the calculation of b_w and c for this latter case, we must use the weighted residual $zw^{1/2}$ in place of z alone.

Median methods. If experimental data were free of error, values for the two parameters of Eq. [3] could be obtained by measuring y at two points and solving the resultant nonlinear simultaneous equations. In practice, of course, data do contain some variability and more than two measurements are made. The purpose of the additional measurements is to increase the reliability of the parameter estimates and, more importantly, to permit the calculation of a measure of this reliability. Cornish-Bowden and Eisenthal (4) have suggested a robust method of analysis for the case where Eq. [3] represents the Michaelis-Menten equation, and this may be adapted to any two-parameter, nonlinear equation. Values for the parameters are calculated from each possible pair of measurements and these $N(N-1)/2$ values for b_1 and b_2 are used to determine the best estimates of the values. It was originally proposed (4) that the median values of b_1 and b_2 should be taken as the best estimates but it was subsequently pointed out (5) that the median values for b_1 and b_2 may be biased. It is usually

possible to find which are more preferred procedures of measurement and these are unbiased for the latter values of their magnitudes. The median values are measures of b_1 , a median value transformation involved is used. For example, the median of the Michaelis-Menten equation, whereas $c = b_1 + b_2$ is the median value, is simple: $V = 1$.

Confidence intervals found by an exact method (6). Kendall's method finds the ranks of the data interval at any point. The values of the ranks are determined by the number of observations falling between the rank and the next rank.

An alternative method described by Cornish-Bowden (7) on a special case of the determination of the parameters of each set of data. The estimate of \hat{y} is calculated from the simultaneous equations of the Michaelis-Menten equation. The values of the parameters are calculated from each possible pair of measurements and these $N(N-1)/2$ values for b_1 and b_2 are used to determine the best estimates of the values. It has the advantage of being able to handle transformations of the data and its applications to various types of data have been demonstrated.

All median methods are solutions of a system of linear equations. The solutions are obtained by an iterative process. The equations are solved simultaneously until the results are consistent with the data.

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possible to find combinations of b_1 and b_2 which are median-unbiased and the preferred procedure is as follows. For each pair of measurements, b_1 and b_2 are calculated and these are transformed to the median-unbiased combinations, c_1 and c_2 . These latter values are separately ranked in order of their magnitude and the centrally ranked values are located. Finally, the best estimates of b_1 and b_2 are calculated from the median values of c_1 and c_2 by reversing the transformation. The type of transformation involved is usually quite simple and an example will serve to illustrate this point. The median values of $b_1 = V$ and $b_2 = K$ of the Michaelis-Menten equation are biased whereas $c_1 = 1/V$ and $c_2 = K/V$ are median-unbiased. The reverse transformations which are used to calculate V and K from the median values of c_1 and c_2 are equally simple: $V = 1/c_1$ and $K = c_2/c_1$.

Confidence intervals for b_1 and b_2 may be found by an extension of this median method (6). Kendall's S^* statistic is calculated to find the ranks which enclose the confidence interval at any desired probability level, and the values of c_1 and c_2 which occupy these ranks are determined. Limits on the parameters are found by transforming c_1 and c_2 to b_1 and b_2 .

An alternative median method has been described by Duggleby (7,8) which is based on a special experimental design. Multiple determinations of y are made under two sets of experimental conditions and the median of each set of replicates is taken as an estimate of \bar{y} . Values for b_1 and b_2 , which are calculated by solving the resulting two simultaneous equations, will be the best estimates of these parameters. This method has the advantage that it avoids the necessity of transforming into median-unbiased combinations of the parameters. Other advantages have been described previously (7,8).

All median methods require the algebraic solution of a set of nonlinear equations. The solutions will depend on the form of the equations and for this reason it is difficult (but by no means impossible) to in-

corporate a median method into a general computer program. Thus, in the program described below, robustness has been approached by the bisquare weighting method. For the sake of completeness, the solutions required for median methods are also given for the specific models considered below. These solutions may be useful for calculating initial estimates of the parameters.

RESULTS

A computer program embodying the nonlinear regression principles outlined under the Theory section has been written in BASIC. The program was developed using BASIC-11, a version of this language which is used in the PDP-11 series of computers. Exploitation of special features of this version of the language was deliberately avoided to facilitate transfer of the program to other computers which will support the BASIC language.

A listing of the program is shown in Fig. 1 and while it might appear that the program is quite long, this impression is largely illusory. Of the 176 lines in the program, 68 are REM statements which contribute nothing to the operation of the program but serve solely to document it. Of the remaining 108 lines, 26 print either blank lines or headings. Thus, the heart of the program is less than one-half of the total and compression of the source code may be achieved readily, an important consideration for microcomputers where storage limitations are critical.

The only statement which depends on the equation to be fitted is line 2650 which, in Fig. 1, describes the Michaelis-Menten equation. Other models may be fitted by replacing this line with the appropriate expression. For example, the first-order decay curve described by Eq. [1] might be written:

$$2650 G = B(1)*\exp(-B(2)*X).$$

Partial derivatives are calculated by nu-

FIG. 1. BASIC computer program for weighted nonlinear regression analysis. In general, the variable names used in the program correspond to those used in the Theory section. In general, the major exceptions are quantities used in calculating bisquare weights (here named R1-R5) and $\hat{\beta}$ (here named G). The only library functions used are the square root function (SQR) and the absolute value function (ABS). The circumflex ($\hat{}$) which appears in several places (e.g., line 1580) indicated exponentiation.

merical differentiation (lines 1940-2020) and this method has been found to be satisfactory for all the models which have been

tested. This group of statements can be replaced with the appropriate expressions for analytical derivatives if this is felt to

be desirable. I parameter value zero, the perturbed derivative may universal remed program user s viously, a value used as an initial. A number of weightings including equal, $1/y^2$, $1/y^4$ weighting included. Alteration of y (or a factor) be specified expiring options is available bisquare weighting until the sum of relative change (i.e., $\sum |q/b|$) is the program converged. If converges, a war values of the parameters.

The program of models and scribed. These flexibility of the the equations methods descript are given as w combinations o

Substrate saturation of an enzyme obeys the family of equations [4] and the

will fit this equation. The accuracy was assessed using the values calculated by this method and $\delta\bar{y}/\delta K = -$. Values from zero to one were found to be a

be desirable. In rare instances when a parameter value happens to fall close to zero, the perturbation used to calculate the derivative may be too small. There is no universal remedy to this situation but the program user should be aware of it. Obviously, a value of zero should never be used as an initial estimate of a parameter. A number of weighting options are available including equal weighting, $1/y$ weighting and $1/y^2$ weighting and others may be easily included. Alternatively, the standard deviation of y (or factor proportional to it) can be specified explicitly. Each of these weighting options is available both with and without bisquare weighting. Iteration is continued until the sum of the absolute values of the relative changes in the parameter values (i.e., $\sum |q/b|$) is less than 10^{-5} at which time the program is considered to have converged. If convergence is not reached in 10 iterations, a warning is issued and the current values of the parameters are printed.

The program has been tested with a variety of models and three of these will be described. These do not represent the limit of flexibility of the program. For each model, the equations necessary for the median methods described under the Theory section are given as well as the median-unbiased combinations of the parameters.

Substrate saturation kinetics. Saturation of an enzyme by its substrate frequently obeys the familiar Michaelis-Menten equation [4] and the program shown in Fig. 1

$$\dot{y} = \frac{V \cdot x}{K + x}, \quad [4]$$

will fit this equation to experimental data. The accuracy of numerical differentiation was assessed using this model by comparing the values of derivatives calculated by this method with those obtained using the analytical derivatives $\dot{y}/\delta V = \delta \dot{y}/V$ and $\delta \dot{y}/\delta K = -\dot{y}^2/Vx$. Over a range of x values from zero to 800 K , the derivative for V calculated by the numerical method was found to be accurate to within 0.0003%,

a not unexpected finding since the model is exactly linear in V . The derivative for K is somewhat less accurate but in no case did the error exceed 0.05% of the value. Such errors are of no consequence when fitting to experimental data as is indicated using some data for the enzyme prephenate dehydratase (9), assuming constant standard deviation in y . The fit obtained using analytical derivatives gave² $V = 18.1554 \pm 0.4877$ U/mg and $K = 491.075 \pm 30.793 \mu\text{M}$ while the corresponding values using numerical differentiation were 18.1555 ± 0.4876 U/mg and $491.078 \pm 30.787 \mu\text{M}$. For all practical purposes, the figures obtained by the two methods are identical.

If median methods are used to estimate the kinetic parameters, solutions for the resultant simultaneous equation are

$$K = (y_2 - y_1)/(y_1/x_1 - y_2/x_2)$$

$$V = (K + x_1)y_1/x_1.$$

The median-unbiased combinations are $1/V$ and K/V (5).

First-order decay. The equation for a first-order decay curve has been given previously (Eq. [1]) while the solutions required for median methods are

$$k = (\ln y_2 - \ln y_1)/(t_1 - t_2)$$

$$y_0 = y_1 e^{kt_1}.$$

Both k and y_0 are median-unbiased. Nimmo and Atkins (11) have compared various methods for analyzing this type of data and have observed that their computer program, which uses a rather sophisticated nonlinear regression method, failed to converge with 30-40 out of 500 sets of simulated data. This simulation was repeated using data containing normally distributed errors with a standard deviation equal to $\dot{y}^{1/2}$ (their case

In general, the major exception (here named G), its value function is exponential.

statements can be appropriate expressions if this is felt to

² The absurd number of decimal places given is necessary to illustrate that analytical and numerical derivatives do, in fact, give different results. There is no suggestion intended that V and K are determined to an accuracy of six significant figures.

TABLE I
STABILIZATION BY BISQUARE WEIGHTING OF THE FIT
TO BINDING DATA WHEN AN OUTLIER IS PRESENT^a

Value of y at x = 3.0	Without bisquare weighting		With bisquare weighting	
	K	N	K	N _~
2.8	1.701	1.061	0.955	0.996
2.6	1.311	1.021	0.955	0.996
2.5	1.170	1.009	0.958	0.996
2.4	1.054	1.001	0.994	0.998
2.3	0.957	0.996	0.963	0.998
2.2	0.876	0.993	0.919	0.996
2.1	0.808	0.993	0.952	0.996
2.0	0.749	0.994	0.955	0.996
1.8	0.653	1.001	0.955	0.996

^a A simulated set of data was obtained by solving Eq. [8] for \hat{y} at x values of 0.5, 1.0, 1.5, 2.0, 3.0, 4.0, 5.0, 6.0, 8.0, and 10.0, assuming $K = N = 1$. These theoretical data were rounded to one decimal place and the point at $x = 3.0$ changed from this simulated value of 2.3 to the value indicated. These data were then fitted to Eq. [8] assuming constant variance both with and without bisquare weighting to obtain the values of K and N . Due to the errors introduced by rounding, the fit obtained using the value of 2.3 does not give values for K and N of exactly 1.

$N/2$) and using weights³ of $1/y^2$ (equivalent to their method WNL). The program described here failed to converge within 10 iterations for 30 of these sets of simulated data, so while the program is no better than that used by Nimmo and Atkins, it is no worse either. Ten of these failures were selected for further study and in each case satisfactory convergence could be achieved by allowing more than 10 iterations⁴ or by adjusting the initial estimates of y_0 and k .

Binding equations. The binding of a ligand to a comparable concentration of an acceptor is described by Eq. [5] in which F and B represent the concentrations of free and

$$B = \frac{N \cdot F}{K + F} \quad [5]$$

bound ligand, respectively, N is the total concentration of binding sites and K is the dissociation constant of the ligand-acceptor complex. This equation is similar in form to the Michaelis-Menten equation but now we must take account of the fact that there is significant depletion of free ligand by complex formation. Usually, only one of B or F will be measured while the other is calculated from the fact that B plus F equals the total ligand concentration (N). Thus we may consider two different cases depending on whether B or F is measured.

If B is the measured quantity (y), Eq. [5] is rewritten as Eq. [6] which, upon rearrangement, gives Eq. [7], a quadratic which

$$\hat{y} = \frac{N(x - \hat{y})}{K + x - \hat{y}} \quad [6]$$

$$\hat{y}^2 - \hat{y}(K + N + x) + Nx = 0 \quad [7]$$

may be solved for \hat{y} by the usual methods. Solutions for the simultaneous equations generated by median methods are

$$K = \frac{(y_2 - y_1)(x_1 - y_1)(x_2 - y_2)}{y_1 x_2 - y_2 x_1}.$$

$$N = \frac{y_1(K + x_1 - y_1)}{x_1 - y_1}.$$

The median-unbiased combinations of K and N are $1/N$ and K/N .

In the situation where unbound ligand is the measured quantity we again get a quadratic (Eq. [8]). Note that this equation may

$$\hat{y}^2 + \hat{y}(K + N - x) - Kx = 0 \quad [8]$$

be obtained from Eq. [7] by interchanging K and N and reversing their signs and these same substitutions may be used to obtain the equations required for median methods. The median-unbiased combinations are, as before, $1/N$ and K/N .

The usefulness of bisquare weighting was

³ These are not the correct weights for the error distribution that is being simulated. These incorrect weights are used so that the results could be compared with those of Nimmo and Atkins (11).

⁴ The iteration limit may be changed by modifying line 1730. This has often been found to be necessary when the bisquare weighting option is selected.

assessed using the case where free ligand results obtained without bisquare weighting are sensitive to the point outlier. K changing by 8% for a change of 10% of 10 data points introduced, though to small deviations, a point but larger ignored. A similar effect is seen for the insensitivity of the outlier ascribed to data points occurring in a data set with a value fitting to a data set with a value giving rise to a change larger than those

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The analysis of biochemistry, sciences, frequently be compared with which describes model. In many nonlinear in the appropriate method fitting the equation linear regression native of transformation retains its p regression component been designed with in mind.

This paper presents computer program features. It language which puts and which languages for them. Those who are should be able to use BASIC in a few been deliberately implementation and microcom-

[5]

y , N is the total number of sites and K is the ligand concentration. The equation is similar to the Hanes-Menten equation except of the fact that B plus depletion of free ion. Usually, only F is measured while the fact that B plus concentration (x) are two different cases or F is measured. Linearity (y), Eq. [5] which, upon rearranging, quadratic which

$$\frac{-y}{-y} = \frac{N}{x} + Nx = 0 \quad [6]$$

x) + $Nx = 0$ [7]
the usual methods. Itaneous equations methods are

$$\begin{aligned} -y_1(x_2 - y_2) \\ -y_2x_1 \\ y_1 \end{aligned}$$

combinations of K . If unbound ligand is we again get a quadratic at this equation may

$-x) - Kx = 0$ [8]
[7] by interchanging K and x signs and these same ed to obtain the equation methods. The inations are, as before,

square weighting was

assessed using some simulated data for the case where free ligand is measured and the results obtained are shown in Table 1. Without bisquare weighting, the fit is very sensitive to the presence of an outlier with K changing by 80% from its "true" value for a change of only 20% in the value of 1 of 10 data points. With bisquare weighting introduced, the fitted value of K responds to small deviations in the aberrant data point but larger deviations are essentially ignored. A similar but less pronounced effect is seen for the values of N . The relative insensitivity of N to the presence of an outlier is ascribed to the fact that the spurious data point occurs at a moderately small x value. Fitting (without bisquare weighting) to a data set with an outlier at a high x value gives rise to changes in N which are much larger than those seen in Table 1.

DISCUSSION

The analysis of experimental data in biochemistry, as in other quantitative sciences, frequently requires that the data be compared with a mathematical equation which describes an underlying theoretical model. In many instances this equation is nonlinear in the parameters and the appropriate method of analysis will involve fitting the equation to the data by nonlinear regression. The frequently used alternative of transforming the data into a linear form retains its popularity because nonlinear regression computer programs have not been designed with a laboratory environment in mind.

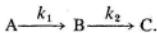
This paper presents a nonlinear regression computer program with a number of desirable features. It is written in BASIC, a language which is available on most computers and which is one of the simplest languages for the novice to understand. Those who are proficient in FORTRAN should be able to learn the elements of BASIC in a few hours. The program has been deliberately kept short to encourage implementation and to facilitate use on mini and microcomputers for which program

size can impose limitations. The equation to be fitted to the data is specified in a single statement so changing to other equations is extremely easy. Partial derivatives are calculated by a numerical method to relieve the user of the sometimes onerous task of deriving these functions algebraically. A variety of weighting options are available which should cover most commonly encountered situations and others can be added easily. Finally, a bisquare weighting option is available which detects and reduces the effects of observations which deviate markedly from the fitted equation.

The output from the program consists of best-fit values of the parameters, standard errors of these values, and a comparison of the experimental data with the fitted equation. For a nonlinear equation these standard errors are only an approximate guide to the precision of the parameters and should be interpreted with this in mind. More reliable methods for estimating precision have been described (10) but these cannot be incorporated into the present program without a substantial increase in complexity. The aim was not to produce a program which could cope with any contingency, but rather to produce one which would be useful in many situations, and which is short and simple to use. Restricting the size of the program makes it inevitable that there will be limitations of capability but these are not excessive. The main limitation (and this is not fundamental) is that the program will only fit equations in which there are two parameters to be estimated and one independent variable. Clearly there will be some models for which the program cannot be used without substantial modification. The second limitation is that the program uses the Gauss-Newton method of nonlinear regression which is known to be ineffective when initial estimates of the parameters are not reasonably close to the best fit values. In spite of this, the program was found to perform as well as a much more sophisticated program on a test problem (11).

Results have been presented from fitting

three equations but these do not represent the limits of applicability. Other models which have been successfully fitted include the analysis of C_d curves (12) and the three compartment model



Further applications could include the determination of pK 's and stability constants, the analysis of ultracentrifugation data and of the effects of temperature on enzymatic and chemical reactions. This list is by no means exhaustive; the only limit is the imagination of the user.

REFERENCES

1. Wilkinson, G. N. (1961) *Biochem. J.* **80**, 324-332.
2. Wahrendorf, J. (1979) *Int. J. Bio-Med. Comput.* **10**, 75-87.
3. Mosteller, F., and Tukey, J. W. (1977) in *Data Analysis and Regression*, pp. 353-365, Addison-Wesley, Reading, Mass.
4. Cornish-Bowden, A., and Eisenthal, R. (1974) *Biochem. J.* **139**, 721-730.
5. Cornish-Bowden, A., and Eisenthal, R. (1978) *Biochim. Biophys. Acta* **523**, 268-272.
6. Porter, W. R., and Trager, W. F. (1977) *Biochem. J.* **161**, 293-302.
7. Duggleby, R. G. (1979) *J. Theor. Biol.* **81**, 671-684.
8. Duggleby, R. G. (1980) in *Design and Analysis of Enzyme and Pharmacokinetic Experiments* (Endrenyi, L., ed.), Plenum Press, in press.
9. Duggleby, R. G., Sneddon, M. K., and Morrison, J. F. (1978) *Biochemistry* **17**, 1548-1554.
10. Duggleby, R. G. (1980) *Eur. J. Biochem.* **109**, 93-96.
11. Nimmo, I. A., and Atkins, G. L. (1979) *Anal. Biochem.* **94**, 270-273.
12. Britten, R. J., and Kohne, D. E. (1968) *Science* **161**, 529-540.

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