1. Since we have
$$E[f(8n)] = \sum_{k=1}^{L} w_n^{(k)} f(2n^k)$$

let $\{2n^{(k)}\}$ is a sed of $p(2n|X_{n-1})$
 $w_n^{(k)} = \frac{p(X_n(2n^k))}{\sum_{k=1}^{L} p(X_n(2n^k))}$
2. $E(\hat{f}) = E[\hat{f}] \sum_{k=1}^{L} f(2^{(k)})]$
 $= \frac{1}{L^2} E[\hat{f}] = E[\hat{f}]^2 = E[\hat{f}]^2 = E[\hat{f}]^2$
 $= \frac{1}{L^2} E[(\sum_{k=1}^{L} f(2^{(k)}))^2] - E[\hat{f}]^2$
 $= \frac{1}{L^2} E[\sum_{k=1}^{L} f(2^{(k)}))^2] - E[\hat{f}]^2$
 $= \frac{1}{L^2} E[\sum_{k=1}^{L} f(2^{(k)})] + \sum_{k=1}^{L} f(2^{(k)}) f(2^{(k)})] - E[\hat{f}]^2$
 $= \frac{1}{L^2} E[\sum_{k=1}^{L} f(2^{(k)})] + \sum_{k=1}^{L} E[\hat{f}]^2 - E[\hat{f}]^2$
 $= \frac{1}{L^2} \cdot L \cdot E[\hat{f}^2] - \frac{1}{L} E[\hat{f}]^2$

= 1 E(f) - 1 E(f) = 1 E[(f-E(f))2]

3. Since we have $g(8) = p(y) \cdot \left[\frac{dy}{d8}\right]$ $y = arc \tan \frac{2 \cdot c}{b} \Rightarrow \frac{dy}{de} = \frac{1}{b} \cdot \frac{1}{1 + \left[\frac{(2 \cdot c)}{b}\right]^2}$ $g(2) = 1 \cdot \frac{1}{b} \cdot \frac{(1 \cdot 1)^2}{1 + \left[\frac{(2 \cdot c)}{b}\right]^2}$ is a pdf.

By (11.16) $k = \frac{1}{b}$.

Application Questions

Since there's some compelling was too slow, I can only do the screen shot for the problems, but not knit it to pdf.

```
1 - ---
 2 title: "cs539 hw4 Application Questions"
 3 author: "Enbo Tian"
 4 date: '2022-03-25'
 5 output: pdf_document
 6 - --
7 → # Viterbi Decoding Algorithm
8 * ## a)
9 * ``{r}
10 library(HMM)
11 # Viterbi algorithm
12 * Viterbi=function(v,a,b,initial_distribution) {
13
14
      T = length(v)
15
      M = nrow(a)
16
      prev = matrix(0, T-1, M)
      omega = matrix(0, M, T)
17
18
19
      omega[, 1] = log(initial_distribution * b[, v[1]])
20 +
      for(t in 2:T){
21 -
        for(s in 1:M) {
          probs = omega[, t - 1] + log(a[, s]) + log(b[s, v[t]])
22
23
          prev[t - 1, s] = which.max(probs)
24
          omega[s, t] = max(probs)
25 ^
26 -
      }
27
28
      S = rep(0, T)
29
      last_state=which.max(omega[,ncol(omega)])
30
      S[1]=last\_state
31
32
      i=2
33 +
      for(i in (T-1):1){
34
        S[j]=prev[i,last_state]
        last_state=prev[i,last_state]
35
36
        j=j+1
37 🔺
      }
38
39
      S[which(S==1)]='A'
40
      S[which(S==2)]='B'
41
```

```
S[which(S==1)]='A'
39
      S[which(S==2)]='B'
40
41
42
     S=rev(S)
43
44
      return(S)
45
46 ^ }
47
48
   #repeat the result presented in the pdf file
49
   51
52
   observations = c("G", "G", "C", "A", "C", "T", "G", "A", "A")
53
    viterbi = viterbi(hmm, observations)
54
55 viterbi
56 *
     [1] "H" "H" "H" "L" "L" "L" "I" "I" "I"
57
58 + ## b)
59 + ``{r}
#the observed sequence of: AGTCGTA
observations = c("A","G","T","C","G","T","A")
   viterbi = viterbi(hmm,observations)
63 viterbi
64 -
     [1] "L" "L" "L" "H" "H" "L" "L"
```

```
66 → # Bayesian Filtering
67 * ## a)
68 * ``{r}
69 # create X and Y
70 x0 < rnorm(1,0,1)
71 \quad x = x0
72 X \leftarrow rep(0,100)
73 Y \leftarrow rep(0,100)
74 - for(i in 1:100){
75
     X[i] \leftarrow rnorm(1,0.99*x+0.1,0.1)
76
     x \leftarrow X[i]
77
      Y[i] < rnorm(1, -2*x+1, 0.4)
78 4 }
79 X <- c(x0,X)
80 head(X)
81 head(Y)
82 -
      [1] 1.082742 1.113829 1.268545 1.397311 1.623790 1.670091
      [1] -1.596619 -1.715995 -1.250474 -2.215364 -2.754681 -2.480468
83
84 * ## b)
85 * ``{r}
86 pygx <- pnorm(Y, -2*X[-1], 0.4)
87 pxgx1 <- pnorm(X[-1], 0.99*X[1:100]+0.1, 0.1)
88 pxkgy1k1 <- pygx*pxgx1
89 z \leftarrow pygx[100]*pxkgy1k1[100]
90 pxkgy1k \leftarrow 1/z * pygx *pxkgy1k1
91 head(pxkgy1k)
92 -
      [1] 0.2707539 0.7765434 0.7172082 0.9888070 0.3311852 0.6504082
0.2
```

```
94 + ## c)
95 + ```{r}
 96 post < qnorm(pxkgy1k, 0.99*X[1:101]+0.1, 0.1)
 97 post = post[-100]

98 mean(post)

99 conf <- post/X[-1]
100 alpha <- 1-conf
101 length(alpha[abs(alpha)<0.05])
102 -
       [1] 2.51459
       [1] 1.0160477 0.9991755 0.9986455 0.9975084 1.0124402 0.9994709
       [1] -0.0160477017 0.0008244506 0.0013545006 0.0024916007 -0.0124402129
       0.0005290819
       [1] 95
103
104 + ## d)
105 + ``{r
         {r}
106 library("readxl")
     data<-read_excel("filter_problem.xlsx", col_names = c("index","Xk","Yk"))
107
108 Y <- data$Yk[-1]
109
     X <- data$Xk
     pygx < -pnorm(Y, -2*X[-1], 0.4)
110
     pxgx1 \leftarrow pnorm(X[-1], 0.99*X[1:100]+0.1, 0.1)
111
112 pxkgy1k1 <- pygx*pxgx1
113 z <- pygx[100]*pxkgy1k1[100]
114
     pxkgy1k <- 1/z * pygx *pxkgy1k1
115 head(pxkgy1k)
116 post <- qnorm(pxkgy1k,0.99*X[1:101]+0.1,0.1)
117
     post = post[-100]
118 mean(post)
119 conf <- post/X[-1]
120 alpha <- 1-conf
121 length(alpha[abs(alpha)<0.05])
122 -
       [1] 58.38498 73.13255 74.34234 73.62419 69.11370 67.46193
```

Alpha = [1] 96

```
124 + ## e)
125 + ```{r}
                                                                                                               ∰ ¥ ▶
126 x0 <- rnorm(1,0,1)
127 x = x0
128 X <- rep(0,100)
129 • for(i in 1:100){
130    X[i] <- rnorm(1,x,0.2)
131
         x \leftarrow X[i]
132 * }
133 X <- c(x0,X)
134 pygx <- pnorm(Y, -2*X[-1], 0.4)

135 pxgx1 <- pnorm(X[-1], X[1:100], 0.2)

136 pxkgy1k1 <- pygx*pxgx1

137 z <- pygx[100]*pxkgy1k1[100]
138 pxkgy1k <- 1/z * pygx *pxkgy1k1
139 head(pxkgy1k)
140 post <- qnorm(pxkgylk,X[1:100],0.2)
141 post = post[-100]
141 post = post
142 mean(post)
143 conf <- post/X[-1]
144 alpha <- 1-conf
145
       length(alpha[abs(alpha)<0.05])</pre>
146 -
         [1] 3.752094e+89 7.791352e+88 4.688298e+88 4.846695e+86 2.044845e+79 3.585929e+80
         Warning in qnorm(pxkgy1k, X[1:100], 0.2): 产生了NaNs
         [1] NaN
         Warning in post/X[-1]:
           longer object length is not a multiple of shorter object length
         [1] 33
148 + ## f)
149 Since there is NA value in d and e, y may not follow the distribution which we
       provided.
150 In addition,c d have a good confidence but e doesn't.
151
152 * # Sequential MCMC
153 * ```{r}
154 Y <- data$Yk[-1]
155 X <- data$Xk
                                                                                                                     £ 1
156 pygx <- pnorm(Y, -2*X[-1], 0.4)
157 pxgx1 <- pnorm(X[-1],0.99*X[1:100]+0.1,0.1)

158 pxkgy1k1 <- pygx*pxgx1

159 z <- pygx[100]*pxkgy1k1[100]
160 pxkgy1k <- 1/z * pygx *pxkgy1k1
161 #a)
162
       mean(sample(pxkgy1k,100))
163 #b)
164 mean(sample(pxkgy1k,1000,replace = TRUE))
165 -
         [1] 31.32629
         [1] 32.46146
```

```
# GP and Linear Regression
##a)
 ``{r}
library(GPfit)
library(brms)
data <- read.csv("synchronous_machine.csv")</pre>
### It is too slow for my computer to do the chain with sampling in the model,
### so I can only pick one parameter for Gp.(nearly 10 min once) fitgp <- brm(qIy~gp(PF)+e+dIf+If,chain = 4, data = data)
summary(fitgp)
 Warning: Parts of the model have not converged (some Rhats are > 1.05). Be careful
 when analysing the results! We recommend running more iterations and/or setting
 stronger priors.
 Family: gaussian
  Links: mu = identity; sigma = identity
 Formula: qIy \sim gp(PF) + e + dIf + If
    Data: data (Number of observations: 557)
   Draws: 4 chains, each with iter = 2000; warmup = 1000; thin = 1;
          total post-warmup draws = 4000
 Gaussian Process Terms:
              Estimate Est.Error 1-95% CI u-95% CI Rhat Bulk_ESS Tail_ESS
 sdgp(gpPF)
                                      0.38
                  0.73
                             0.33
                                                1.73 1.63
                                                                  8
                                                                          18
 lscale(gpPF)
                  0.11
                             0.10
                                      0.01
                                                0.30 2.56
                                                                          19
 Population-Level Effects:
           Estimate Est.Error 1-95% CI u-95% CI Rhat Bulk_ESS Tail_ESS
 Intercept -1091.42
                        544.66 -2392.03 -350.81 1.75
                                                               6
                                                                       15
                                                               7
              -7.07
                         3.61 -10.51
                                           2.58 1.60
                                                                       13
                        460.75 -2020.75
                                         -293.37 1.75
 dIf
            -920.50
                                                               6
                                                                       15
                        461.14 300.54 2028.76 1.75
 Ιf
             927.63
                                                                       15
 Family Specific Parameters:
       Estimate Est.Error 1-95% CI u-95% CI Rhat Bulk_ESS Tail_ESS
                               0.34
                                       0.57 1.84
                     0.09
 Draws were sampled using sampling(NUTS). For each parameter, Bulk_ESS
 and Tail_ESS are effective sample size measures, and Rhat is the potential
 scale reduction factor on split chains (at convergence, Rhat = 1).
```

```
## b)
```{r}
fit <- lm(qIy~PF+e+dIf+If,data = data)</pre>
summary(fit)
 Call:
 lm(formula = qIy \sim PF + e + dIf + If, data = data)
 Residuals:
 Min
 10 Median
 3Q
 -1.70820 -0.39040 -0.03484 0.31406 2.14232
 Coefficients: (2 not defined because of singularities)
 Estimate Std. Error t value Pr(>|t|)
 0.4654 -15.13 <2e-16 ***
 (Intercept) -7.0398
 <2e-16 ***
 10.8102
 0.4628 23.36
 PF
 е
 NA
 NA
 NA
 NA
 <2e-16 ***
 dIf
 7.4658
 0.2663
 28.03
 Ιf
 NA
 NA
 NA
 NA
 Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' '1
Residual standard error: 0.5767 on 554 degrees of freedom
Multiple R-squared: 0.5872, Adjusted R-squared: 0.5857
F-statistic: 394 on 2 and 554 DF, p-value: < 2.2e-16
c)
MSE of gp is much less than linear regression.
```