Functional biogeography of the thermal thresholds for post-dispersal embryo growth in Conopodium majus

Thank you for agreeing to review this paper for Annals of Botany. The Annals of Botany aims to be among the very top of plant science journals and as we receive over 1000 submissions every year we need to be very selective in deciding which papers we can publish. In making your assessment of the manuscript's suitability for publication in the journal please consider the following points.

Scientific Scope

Annals of Botany welcomes papers in all areas of plant science. Papers may address questions at any level of biological organization ranging from molecular through cells and organs, to whole organisms, species, communities and ecosystems. Its scope extends to all flowering and non-flowering taxa, and to evolutionary and pathology research. Many questions are addressed using comparative studies, genetics, genomics, molecular tools, and modeling.

To merit publication in Annals of Botany, contributions should be substantial, concise, written in clear English and combine originality of content with potential general interest.

- We want to publish papers where our reviewers are enthusiastic about the science: is this a paper that you would keep for reference, or pass on to your colleagues? If the answer is "no" then please enter a low priority score when you submit your report.
- We want to publish papers with novel and original content that move the subject forward, not papers that report incremental advances or
 findings that are already well known in other species. Please consider this when you enter a score for originality when you submit your report.

Notes on categories of papers:

All review-type articles should be novel, rigorous, substantial and "make a difference" to plant science. The purpose is to summarise, clearly and succinctly, the "cutting edge" of the subject and how future research would best be directed. Reviews should be relevant to a broad audience and all should have a strong conclusion and illustrations including diagrams.

- Primary Research articles should report on original research relevant to the scope of the journal, demonstrating an important advance in the
 subject area, and the results should be clearly presented, novel and supported by appropriate experimental approaches. The Introduction
 should clearly set the context for the work and the Discussion should demonstrate the importance of the results within that context. Concise
 speculation, models and hypotheses are encouraged, but must be informed by the results and by the authors' expert knowledge of the subject.
- Reviews should place the subject in context, add significantly to previous reviews in the subject area and moving forward research in the subject area. Reviews should be selective, including the most important and best, up-to-date, references, not a blow-by-blow and exhaustive listing.
- Research in Context should combine a review/overview of a subject area with original research, often leading to new ideas or models; they
 present a hybrid of review and research. Typically a Research in Context article contains an extended Introduction that provides a general
 overview of the topic before incorporating new research results with a Discussion proposing general models and the impact of the research.
- *Viewpoints* are shorter reviews, presenting clear, concise and logical arguments supporting the authors' opinions, and in doing so help to stimulate discussions within the topic.
- Botanical Briefings are concise, perhaps more specialised reviews and usually cover topical issues, maybe involving some controversy.

- 1 Original article: Functional biogeography of the thermal thresholds for
- 2 post-dispersal embryo growth in Conopodium majus
- 3 Cristina Blandino^{1,2}, Brith Natlandsmyr³, Sylvi M. Sandvik⁴, Hugh W.
- 4 Pritchard^{1,5}, Eduardo Fernández-Pascual^{6,*}
- ¹ Royal Botanic Gardens, Kew, Wakehurst, Ardingly, Haywards Heath, West
- 6 Sussex RH17 6TN, UK
- ² Department of Biological, Geological and Environmental Science, University of
- 8 Catania, Catania, Italy
- ³ Department of Natural History, University Museum, University of Bergen,
- 10 Bergen, Norway
- ⁴ Department of Natural Sciences, University of Agder, Kristiansand, Norway
- ⁵ Kunming Institute of Botany, Chinese Academy of Sciences, Kunming,
- 13 Yunnan 650201, PR China
- ⁶ Biodiversity Research Institute (IMIB), University of Oviedo CSIC -
- Principality of Asturias, Mieres, Spain https://orcid.org/0000-0002-4743-9577
- Running title: Functional biogeography of embryo growth
- * Corresponding author: Eduardo Fernández-Pascual, Instituto Mixto de
- 18 Investigación en Biodiversidad, Campus de Mieres, Edificio de Investigación, 5ª
- 19 planta, c/ Gonzalo Gutiérrez Quirós s/n, E-33600 Mieres, Spain. Email:
- fernandezpeduardo@uniovi.es. Telephone: +34985104781.

ABSTRACT

- Background and Aims Plant regeneration by seeds is driven by a set of physiological traits, many of which show functional intraspecific variation along biogeographic gradients. In many species, germination phenology depends on a germination delay imposed by the need for post-dispersal embryo growth (a.k.a. morphological dormancy). Such growth occurs as a function of environmental temperatures and shows base, optimum and ceiling temperatures (i.e. cardinal temperatures or thermal thresholds). However, the biogeographical variation in such thresholds has not been tested.
- Methods We used a thermal time approach and field experiments to assess intraspecific variation at the continental scale in the embryo growth thermal thresholds of the geophyte Conopodium majus (Apiaceae) across its distribution from the Iberian Peninsula to Scandinavia.
- **Key Results** Thermal thresholds for embryo growth varied across the latitudinal gradient, with the estimated optimum temperatures between 2.5 and 5.2 °C, ceiling temperatures between 12 and 20.5 °C and base temperatures between -6.6 and -2.7 °C. Germination in the field peaked in the months of January and February. The limiting factor for embryo growth was the ceiling temperature, which was negatively correlated with latitude and the bioclimatic environment of each population. In contrast, the optimal and base temperature were independent of local climate.

Conclusions These results indicate that thermal thresholds for embryo
growth are functional ecophysiological traits that drive seed germination
phenology and seed responses to soil climatic environment. Therefore,
post-dispersal embryo growth can be a key trait impacting climate
change effects on phenology and species distributions.

KEYWORDS

- 7 Cardinal temperatures for germination, Apiaceae, Conopodium majus,
- 8 embryo:endosperm ratio, morphological dormancy, post-dispersal embryo
- 9 growth, functional seed traits, plant regeneration, intraspecific variation

INTRODUCTION

The three aims of functional biogeography are to describe the distribution of 3 functions along environmental gradients and across spatial scales; to use this 4 5 information to explain the geographic distribution of organisms; and to predict their responses to environmental changes using trait-based predictive models 6 (Violle et al., 2014). A relevant aspect of plant function that has been 7 8 underutilized by biogeographical studies is the physiological thermal control of plant reproduction (Bykova et al., 2012), and especially seed germination 9 (Rosbakh and Poschlod, 2015). The temperature to which imbibed seeds are 10 11 exposed affects their germination rate (Fernández-Pascual et al., 2019). This phenomenon can be numerically described by the "cardinal germination" 12 temperatures", i.e., the optimum temperature (T_0) , at which the germination rate 13 is maximal and the base (T_b) and ceiling (T_c) temperatures that are, 14 respectively, estimated to be the coldest and the warmest temperature at which 15 16 the rate of germination tends to zero. The measurement of these temperatures 17 for a given species enables prediction of its seed germination rate and germination success under different temperature scenarios (Orrù et al., 2012, 18 19 Fernández-Pascual et al., 2015). Therefore, the cardinal temperatures can be key parameters to explain the contribution of regeneration thermal niches on 20 species distributions and responses to climatic changes (Parmesan and Hanley, 21 22 2015, Baskin and Baskin, 2022, Walck et al., 2011, Cochrane et al., 2015). 23 In many species, however, seed dormancy prevents germination even in the presence of suitable conditions, so that the exposure of seedlings to 24 unfavourable environments is avoided and the timing of germination is matched 25

to favourable regeneration windows (Lamont and Pausas, 2023, Pausas et al., 1 2 2022). A particular case of seed germination delay occurs when the embryo, at the time of dispersal, is not completely developed and needs to grow to a critical 3 size before germination can occur; a trait known as post-dispersal embryo 4 growth or morphological dormancy (Baskin and Baskin, 2004, Vandelook et al., 5 2009b). Such embryo growth is a distinct ecophysiological process controlled by 6 7 complex interactions between hormones and the seed tissues (Walker et al., 2021). As happens with germination, temperature is a major environmental 8 driver of post dispersal embryo growth (Baskin et al., 2000, Phartyal et al., 9 10 2009, Vandelook et al., 2007, Vandelook et al., 2009a, Blandino et al., 2019, Vandelook and Van Assche, 2008), influencing the rate of development of the 11 embryo, a mechanism that allows a precise timing of germination (Porceddu et 12 13 al., 2017). 14 Within-species biogeographical variation in several germination parameters is well documented. For example, seeds from warmer sites have been shown to 15 have lower minimum temperatures for germination at different geographical 16 scales (Daws et al., 2004, Rosbakh and Poschlod, 2015). The requirements for 17 cold stratification can also vary according to the local climate: populations from 18 habitats with longer winters require a longer period of cold stratification 19 compared with populations from milder habitats (Allen and Meyer, 1998, 20 Fenner, 1991). However, much less research has been dedicated to traits 21 related to post-dispersal embryo growth. Mondoni et al. (2008) compared post-22 dispersal embryo growth between mountain and lowland populations of the 23 temperate woodland forb *Anemone nemorosa* and found that, although embryo 24 size at dispersal was similar in all the populations, embryo growth at cool 25

temperatures was faster in the mountain population. This suggests a capacity of 1 2 post-dispersal embryo growth to adapt to local conditions, either by local adaptation or phenotypic plasticity, analogous to that shown by other seed traits 3 (Fernández-Pascual et al., 2013). However, Porceddu et al. (2017) found no 4 variation in the thermal thresholds for embryo growth in two closely located 5 6 populations of Aquilegia barbaricina. Further research is warranted, to measure 7 the thermal thresholds for post-dispersal embryo growth across large biogeographical scales and investigate whether they vary in association with 8 environmental gradients. 9 In this study, we assess the functional biogeography of post-dispersal embryo 10 11 growth in the geophyte Conopodium majus (Apiaceae) across its latitudinal 12 distribution, from the Iberian Peninsula to Scandinavia. To our knowledge, this is the first study to quantify the thermal thresholds that regulate embryo growth 13 14 rate across the whole latitudinal distribution of a species. To do this, we develop a model of embryo growth as a function of temperature to describe its cardinal 15 temperatures for embryo growth. It has been shown that Scottish populations of 16 this species require post-dispersal embryo growth, and that embryo growth and 17 germination occur optimally around 5 °C (Blandino et al., 2019). Such narrow 18 thermal control in a species with a relatively wide latitudinal distribution could 19 make the species' regeneration vulnerable to climate change (Walck et al., 20 2011), unless the species shows functional variation in its embryo growth 21 thermal thresholds. Therefore, we hypothesized that (1) the thermal thresholds 22 for embryo growth will show variation across populations sampled over the 23 species' latitudinal gradient; and (2) the variation of thermal thresholds will be 24

related to bioclimatic features along the latitudinal gradient. Specifically, we

- predicted that thermal thresholds for embryo growth would be influenced by
- 2 high maximum temperatures and drought at the lower latitudes, and by low
- 3 minimum temperatures at the higher latitudes.

MATERIALS AND METHODS

5 Study species

4

Conopodium majus is a geophyte with a European Atlantic distribution from 6 Southern Spain to Central Norway (Tutin et al., 1968). Conopodium majus is a 7 8 member of Apiaceae, a family that originated and diversified in the southern hemisphere, but that today is mostly distributed in the northern hemisphere 9 10 (Calviño et al., 2016). As is common in Apiaceae (Walker et al., 2021, Baskin and Baskin, 2014, Vandelook et al., 2012), seeds of Conopodium majus 11 possess undeveloped linear embryos (Martin, 1946) and germinate when they 12 extend to the full length of the endosperm (Blandino et al., 2019). For this 13 reason, in this study, the relative embryo size (i.e. embryo length / endosperm 14 length, hereafter referred to as "E:E ratio") is used to describe embryo 15 development, and germination is defined as the point at which E:E ratio is ≥ 1. 16 In Conopodium majus, germination has been observed to occur both at 0 and 5 17 °C with a similar rate of embryo growth (Blandino et al., 2019). Such low 18 temperature requirements are indicative of germination in winter. For a species 19 adapted to woodlands as well as oligotrophic grasslands, the control of the 20 germination process in this way can allow seedlings to establish before the 21 development of a tree canopy or of competing vegetation. 22

Seed collection

Mericarps of Conopodium majus were collected in the summer of 2016 from 1 2 nine naturally occurring populations sampled across the western European latitudinal range of the species (Table 1). Since the seed cannot be separated 3 from the fruit in this species, the dispersal unit will be hereafter referred to as 4 the "seed". A population was sampled only if it consisted of at least 200 5 individual plants. Seeds were sampled from 50 plants within the population to 6 7 secure a representative sample of the genetic variability of the population. At least 4000 seeds were collected from each population. All seeds were collected 8 between July and August 2016 and the experiments started within three weeks 9 10 of seed collection. Seeds were kept at below full hydration, dry exposed to the surrounding atmosphere, under ambient condition on a laboratory bench until 11 the beginning of the tests. 12

Initial measurements

13

Each collection was cleaned from debris and empty seeds were removed using 14 a gravity seed separator machine. From each population, 10 seeds were 15 selected randomly and allowed to rehydrate overnight at 20 °C and 100% RH. 16 The seeds were then placed on 1% agar-water for 24 hours to become fully 17 imbibed. Thereafter, seeds were prepared for vital staining with 1% aqueous 18 solution of triphenyl tetrazolium chloride (TZ). A slice of seed coat was removed 19 from the dorsal surface of each seed using a scalpel and seeds were incubated 20 in TZ solution at 30 °C in the dark for 24 hours. Each seed was then cut 21 longitudinally, and the embryo was extracted. Embryos and endosperms were 22 photographed using a camera (Carl Zeiss Axiocam Colour) mounted on a Stemi 23 SV 11 Microscope (Carl Zeiss, Welwin Garden City, Herts, UK) microscope and 24 their lengths measured using the software Axiovision 3.1.2.1 (Carl Zeiss Vision 25

- 1 GmbH). The initial relative embryo length was measured only for the seeds that
- stained red with the TZ, i.e., indicating viability; unstained seeds / embryos were
- discarded. Relative embryo size was used because it describes the growth of
- 4 the embryo regardless of the size of each seed. From each population 99 seeds
- were placed in a controlled humidity room at 15% RH and left to dry. The dry
- 6 seed weight of 99 seeds for each population was measured using a precision
- 7 scale.

12

13

14

15

16

18

20

22

25

8 Embryo growth in controlled temperature conditions

9 From each population and treatment, 16 subsamples of 15 seeds each were

randomly taken and sown in separate, 8 cm diameter Petri dishes containing

1% agar-water substrate. Seeds were sown at -2.5 °C, 0 °C, 2.5 °C, 5 °C, 7.5 °C

and 10 °C in incubators with a daily light regime of 12 hours. Every 14 days one

subsample from each population and treatment was retrieved and the 15 seeds

were placed for 24 hours in 1% TZ solution at 30°C in the dark, after a slice of

the seed coat was removed. From this subsample, the embryo and endosperm

length of 10 viable seeds was measured following further dissection of the seed.

17 An E:E value of 1 was assigned to all germinated seeds. Seed measurement

was stopped when the seeds ceased germinating. The experiment continued

for 224 days, until all the 16 subsample assessments were concluded.

Calculation of a thermal model for embryo growth

21 The mean E:E ratio of 10 seeds for each population x temperature x time

combination was calculated. All the temperatures for the same population had

23 the same initial E:E ratio at time = 0, while the maximum value was fixed at 1,

24 after which the seed was able to germinate. Since the data followed a sigmoidal

growth distribution, except the treatments at -2.5 °C, a logistic model was fitted

- to each population x temperature combination using the software OriginLab 9.0.
- The models of each population were bounded between the initial value of E:E
- 3 for that population and 1. A linear model was fitted to the -2.5°C treatments.
- 4 From the equation of the logistic and linear models, it was possible to calculate
- 5 the time expressed in days (tr) at which each temperature x population
- 6 combination would have reached the following deciles of relative embryo size:
- 7 0.3, 0.4, 0.5, 0.6 and 0.7. Deciles < 0.3 could not be calculated because they
- were less than the initial E:E. Deciles > 0.7 were not calculated to keep the
- 9 symmetry of the analyses regarding deciles of the population. For each
- treatment, the embryo growth rate was calculated as 1/tr.

12

13

14

15

16

17

18

19

20

21

22

23

24

25

For each population and decile, embryo growth rate was plotted against temperature. Each dataset was visually divided into sub-optimal and supra-optimal ranges, using the point with the highest value of 1/ t as the dividing point. Linear regressions were fitted separately to the sub- and supra-optimal ranges. The intersection with the temperature axis of the sub-optimal and supra-optimal regression are, respectively, the base (T_b) and the ceiling (T_c) temperatures; these estimates are the temperatures below and above which the embryo growth rate is projected to be zero. The optimal temperature (T_o) , defined as the temperature at which the rate of embryo growth is estimated to be fastest, is the x-coordinate of the intersection point between sub-optimal and supra-optimal regressions. Then, for each population, the cardinal temperatures $(T_b, T_c \text{ and } T_o)$ were averaged across all the deciles calculated to define a mean value of the population (Ellis et al., 1986). The regression lines of each decile were recalculated and forced to pass through a common origin defined by the mean T_b (for the sub-optimal regressions) or the mean T_c (for the supra-optimal

- regressions) (Hardegree, 2006). For the three southernmost populations, only
- the cardinal temperatures calculated for the relative embryo size of 0.4, 0.5 and
- 3 0.6 were used, because it was not possible to fit a supra-optimal regression to
- 4 the 0.3 decile.
- The slopes of these new linear regressions were then taken as a reciprocal to estimate the sub-optimal (θ_b) and supra-optimal (θ_c) thermal times for embryo
- 7 growth. θ, expressed in degree days (°Cd), indicates the cumulative thermal
- 8 time units above T_b (θ_b) or below T_c (θ_c) that the seed must accumulate for the
- 9 embryo to reach successive E:E deciles. For each population, the deciles were
- plotted against θ_b and θ_c , expressed both as their value and as the natural
- logarithm of the value, and linear regressions were fitted to the data. The
- regressions fitted to θ and to $\log(\theta)$ were compared in each case by their R²
- 13 (Hardegree, 2006). The regression models with the highest R² were chosen to
- represent the rate of embryo growth as a function of thermal time for each
- population. The R² of the models obtained fitting embryo growth and log-normal
- (log °Cd) were slightly higher than the R² of the model obtained using normal
- distributed thermal times (°C). The only exception was constituted by the
- Spanish population of Central del Chorro (CHO), for which the best model fit
- was obtained using the non-transformed thermal time values, thus describing a
- 20 linear increase of relative embryo size with accumulated heat.

Embryo growth in natural conditions

- 22 Embryo growth in the soil was recorded for three population representing the
- southern (CHO), middle (Wakehurst Place, UK, "WAK") and northern (Bergen,
- Norway, "BER") distribution of the species. The experiment was replicated in
- two locations where Conopodium majus naturally occurs: at Wakehurst Place,

England (site of collection of the "WAK" population); and in a meadow on the 1 periphery of Bergen, Norway (close to the site of collection of the "BER" 2 population). Sixteen subsamples of 20 seeds for each population and 3 experimental site were mixed with 20 g of soil collected at the site and passed 4 through a 3 mm sieve. Seeds and soil were placed in mesh net bags and buried 5 at a depth of 5 cm. A datalogger that recorded soil temperature every 30 min 6 was placed in each location (Tinytag View 2, Gemini Dataloggers Ltd., 7 Chichester. UK and EasyLog USB-2, Lascar Electronics, in Norway). The seeds 8 were buried in England on 1st September 2016 and in Norway on 14th 9 10 September 2016. Every 14 days a bag for each population was retrieved and the soil washed. Seed bags buried in Norway were shipped to England for 11 measurements within a week of retrieval. All the seeds retrieved were prepared 12 13 for TZ staining and their embryo and endosperm lengths measured. It was easiest to measure the seeds when most of the seeds were not germinated. 14 15 With an increasing number of germinated seeds and seedlings, the number of empty seed coats left in the soil bags made it difficult to distinguish between 16 mouldy or germinated seeds. At this point, the experiment was terminated, 17 representing nine measurements in Norway and thirteen in England. 18

Validation of the thermal time model with field data

19

20

21

22

23

24

25

To compare the embryo growth predicted by the thermal time model with embryo growth in natural conditions, embryo growth in the field sites was plotted against time. A logistic regression was fitted to these curves, and from the equations, the t to reach every decile of relative embryo growth was calculated. The units of thermal time required by each population to reach every t during the field experiment were calculated for both field locations using the

- data recorded by the loggers. To account for every temperature fluctuation
- during the day, the thermal time was expressed in "°C 30 min" and the heat
- accumulated by the seed was calculated for every 30 min temperature record.
- The difference (ΔT) between each temperature record and the population T_0
- 5 (averaged between deciles) was summed. When the temperature was higher
- 6 than the mean T_c or lower than the mean T_b the heat accumulated was
- 7 considered = 0 and the difference (ΔT) between each temperature record and
- 8 the T_o was summed.

- 9 The time necessary in the field to accumulate enough heat to reach the thermal
- time necessary for each t was compared with the t estimated from the embryo
- growth data. The time (in days) needed to sum enough heat to reach the θT_b
- and θT_c calculated in the model, for each t decile (t model) in each population
- was compared with the time needed by each population to reach the same
- decile of relative embryo growth in the field (t field). These estimates were then
- graphically compared expressing the different t in function of E:E.

Relationship between embryo growth and germination

- 17 Germination was scored for each independent sample before measuring the
- relative embryo size and expressed as percentage of germinated seeds vs time.
- 19 For each population, the germination data for the treatments at 2.5 and 5°C
- were fitted with the Boltzmann equation using the software OriginLab9. The
- other temperatures were not used because germination was too slow. For each
- 22 population, from the fitted Boltzmann equation the day to reach 50%
- 23 germination (t_{a50}) was calculated. The t_{a50} was then used to calculate the
- corresponding E:E ratio at the same day using the logistic regression of the E:E
- 25 data for the same treatment. For each population, the mean E:E ratio

- corresponding to the t_{a50} for germination at the two temperatures used was
- 2 displayed as the mean E:E ratio for 50% germination in that population. The
- mean between all the populations represented the mean for the species.

4 Relationship between environmental data and germination parameters

- 5 The relationship between embryo development and seed germination
- 6 parameters and geographical and bioclimatic data was explored for each
- 7 population. A data matrix was built including latitude, altitude, mean annual
- 8 temperature, precipitation of the driest month, mean maximum temperature of
- 9 the hottest month and minimum mean temperature of the coldest month, seed
- dry mass, initial E:E ratio and cardinal temperatures for each population. Climatic
- 11 data for the seed collection sites was extracted from WorldClim (Fick and
- Hijmans, 2017). Data was checked for autocorrelation using the Pearson
- correlation coefficient to exclude the variables with a strong autocorrelation.
- 14 Finally, a PCA was run on the dataset, scaling the axis.

15 **RESULTS**

21

16 Initial embryo length and seed mass

- The initial relative embryo size ranged from a mean value of 0.10 (± 0.03 SD) for
- the population BER to a mean value of 0.19 (±0.03 SD) for the population CHO
- 19 (Table 2). Mean seed dry mass ranged just under two-fold from 1.21 mg (±0.38
- 20 SD) in TRE to 2.03 mg (±0.53 SD) in FLE (Table 2).

Embryo growth in controlled temperature conditions

- The rate of embryo growth was strictly dependent on the temperature and the
- increase in embryo size could be appreciated already after 14 days of sowing
- 24 for germination. For all the populations, the temperature treatments with the

- highest rate of embryo growth were 2.5 and 5 °C. Clearly 0 °C was sub-optimal
- 2 for embryo growth rate, and 7.5 and 10 °C were supra-optimal (Fig.1). The
- 3 seeds survived cooling to -2.5 °C but the embryo did not grow at this
- 4 temperature.

23

Relationship between embryo growth and germination

The first germination was scored after 84 days of imbibition in the four Spanish 6 populations at temperatures of 0, 2.5 and 5 °C. The populations from WAK and 7 BER first germinated after 112 days of imbibition. The last population to begin 8 9 germinating was SCO, after 126 days of imbibition. Germination occurred when the embryo reached the same length of the endosperm (E:E=1) and a mean 10 11 E:E = 1 corresponded to 100% germination in the sample. The treatments that 12 had the highest mean germination across all the populations after 32 weeks of imbibition were 2.5 °C, and 5 °C with, respectively, 97.7 and 98.4 % of seeds 13 germinated at the last sampling. The lowest germination was observed at -2.5 14 15 and 10 °C. The population that reached, across all the treatments, the highest mean germination at week 32 (the end of the experiment), was TRE (80% ± 32) 16 SD) while the lowest was achieved by SCO (59%, ± 42 SD). The time to reach 17 50 % germination (T_{q50}), interpolated with the Boltzmann equation ranged 18 between 111 (BAS) and 147 days (FLE) at 2.5 °C and between 116 (LEO) and 19 20 150 days (SCO) at 5 °C. The values of E:E corresponding to the estimated T₅₀ in these two treatments were averaged between population and temperatures to 21 describe a value of 0.89 (± 0.02 SD) for the species. 22

Cardinal temperatures for embryo growth

- Between populations, T_b estimates varied between -2.63 (SCO) and -6.65 °C 1
- 2 (BER). In addition, To varied from 2.54 (LEO) and 5.23 °C (CHO). Finally, Tc
- was between 12.08 (BER) and 20.54 °C (TRE) (Table 2). 3

Embryo growth in natural conditions 4

The minimum temperature recorded in Norway in winter was -2 °C in mid-5 November while the highest (18.5 °C) was recorded at the beginning of the 6 experiment, on 15th September 2016. In England the minimum temperature 7 recorded was 1.6 °C at the end of January and the maximum 17.0 °C, recorded 8 9 on the same day as the Norwegian site, during an autumn heat wave. Embryo growth in natural condition was faster, for all the population tested, in the 10 11 northern most location of Bergen (BER) where daily mean temperatures were 12 lower than at Wakehurst (WAK), UK. However, in both sites the rate of embryo growth started to increase when the temperatures fell below 10 °C (Fig. 2). 13 Even if the southern population (CHO) had the greater initial E:E ratio, its 14 15 growth rate was not different from the other populations tested. Eventually, the three growth curves tended to converge when a mean E:E ratio approached 0.8 16 (Fig. 2). Germination in nature tended to peak in the months of January and 17 February. Fitting a logistic regression to the curves permitted an estimation of 18 the time, in days, to reach different deciles of relative embryo size.

Comparison of the model with field data

19

20

The comparison of the thermal models against estimates of embryo growth in 21 the field gave different results between the three populations but was consistent 22 between experimental sites (Fig. 3). Estimates of time to reach successive 23 deciles of E:E ratio were similar if calculated using the ΘT_b and ΘT_c of the WAK 24

- population for both sites but higher than the Tr_q estimated from the logistic
- 2 regression of embryo growth in the field. The BER population showed a rate of
- 3 embryo growth that could be better predicted by the ΘT_c rather than by ΘT_b
- 4 while both models diverged from the observed pattern of embryo growth in the
- 5 southern population CHO.

6 Environmental correlates of embryo growth traits

A PCA ordered the populations according to their seed and germination 7 8 parameters and to the climate of the collection site (Fig. 4). The first axis, that explained 50% of the variability in the data, separated the southern populations 9 from the northern ones. The axis was described mostly by geographic and 10 11 bioclimatic variables and the only seed traits that had a significant correlation with it were seed dry mass and the ceiling temperature for embryo growth (T_c). 12 In particular, there was a strong negative correlation between precipitation of 13 the driest month and T_c. Mountain populations of Conopodium majus were in 14 the southern portion of the distribution range of the species and were 15 16 characterized by higher maximum temperatures and more severe drought stress. The seeds from these populations had a lower dry mass but a greater 17 initial relative embryo size than the northern, lowland populations. The second 18 axis explained 18.8 % of the variability in the data and had a significant 19 correlation only with T_b and T_o. The two cardinal temperatures showed opposite 20 trends, such that a higher optimum corresponded to a lower T_b. The two 21 22 southern most populations, CHO and TRE, remained separated from the others: they came from the highest altitude and are exposed to the strongest 23 heat and drought stress. The remaining populations can be separated in three 24

- groups: SCO, BAS and LEO had the highest T_b, HER and FLE the biggest
- 2 seeds and BER and WAK had the higher T_o.

DISCUSSION

3

4 Conopodium majus shows considerable intraspecific variability in the thermal thresholds for post-dispersal embryo growth along its European distribution. 5 Additionally, variability in post-dispersal embryo growth thresholds appears 6 related to the climate of the collection sites, with warmer and drier sites 7 correlating with a capacity for embryos to grow at warmer temperatures. These 8 results indicate that there is functional variation in the embryo growth thermal 9 thresholds, and therefore that these thresholds might be functional 10 11 ecophysiological traits. To our knowledge, this is the first work showing 12 biogeographical variation in the thermal thresholds for embryo growth. A previous study had found no variation in embryo thresholds between closely 13 located populations of the Ranunculaceae species Aquilegia barbaricina, even if 14 the same populations showed variation in the thermal thresholds for 15 germination (Porceddu et al., 2017). 16 The ceiling temperature for embryo growth varied between 12 and 21 °C and 17 had a strong negative correlation with latitude and precipitation. Populations 18 from northern and wetter locations had lower ceilings; while the two 19 southernmost populations (CHO and TRE) had a ceiling above 20 °C. 20 Interestingly, the two southernmost populations (CHO and TRE) belong to the 21 subspecies Conopodium majus subsp. marizianum, while the others belong to 22 the typical subspecies Conopodium majus subsp. majus (the LEO population 23 grows in the contact area between the two subspecies). Conopodium majus 24 subsp. marizianum and all the other species in the genus are restricted to the 25

Iberian Peninsula and northern Africa, with the only exception of Conopodium majus subsp. majus, which is the only taxon to be widely distributed in western and northern Europe (Mateo and López Udias, 2003). This may suggest that the lower ceiling temperature for embryo growth in Conopodium majus subsp. majus played a key role in allowing the subsp. to colonize colder regions, a hypothesis that should be explored with broader comparative studies in the genus. From a functional point of view, the correlation between the ceiling temperature and precipitation reflects the general role of drought and frost as the major drivers of regeneration strategies (Jurado and Flores, 2005). A higher ceiling temperature in Conopodium majus subsp. marizianum would allow embryo growth during late summer and autumn, and therefore seedling emergence during winter, the season of lowest water stress in the Mediterranean mountains where the subspecies lives. Conversely, the lower ceiling temperature in Conopodium majus subsp. majus would retard embryo growth until winter, and lead to seedling emergence in early spring, a more favourable regeneration environment in cold-limited regions. Moreover, embryo growth (and the potential to germinate) under cold temperatures (close to 0°C) would enable the start of growth during winter and emergence around the time of snowmelt, a behaviour that has been described in sub-alpine meadows where Conopodium majus can be found (Fernández-Pascual et al., 2017, Shimono and Kudo, 2005). This could give the seedling an early start that can result in a competitive edge, making full use of a short growing season. In another frequent habitat of Conopodium majus, forest understoreys, early seedling emergence is a common trait in herbaceous species because it allows the young plant to start photosynthesis before the tree canopy closes.

1

2

3

4

5

6

7

8

9

10

11

12

13

14

15

16

17

18

19

20

21

22

23

24

The optimum temperature for embryo growth ranged between 2.5 and 5 °C 1 2 (Table 2). This is a very low optimal temperature when compared to the optimal temperatures for germination found in other species: in the review by Dürr et al. 3 (2015), the mean optimal temperature for germination across species was 27 4 °C, with the lowest value being 7 °C in the tree Acer saccharum (McCarragher 5 6 et al., 2011). In the case of the Apiaceae crops Apium graveolens and Daucus 7 carota, optimal embryo growth or germination occurred at 20-25 °C (Finch-Savage et al., 1998, Rowse and Finch-Savage, 2003, Walker et al., 2021), but it 8 has been shown that crops tend to have higher thermal threshold for 9 10 germination than wild species (Dürr et al., 2015). In the future, it would be interesting to explore whether Conopodium majus is an outlier, or embryo 11 growth temperatures tend to be lower than the temperatures for seed 12 13 germination across species. It is also worth noting that, in our case, To had a negative correlation with T_b, a phenomenon already observed for germination 14 15 temperatures by Dürr et al. (2015). In other words, the populations with the higher T_b (BAS, LEO and SCO) also have the lower T_o, and therefore they have 16 17 a narrower window of suboptimal conditions for embryo growth. In practical 18 terms, these populations are at greater risk from climate change, which could produce a mismatch between their narrow germination niche and soil 19 temperatures (Walck et al., 2011, Orrù et al., 2012, Maleki et al., 2024). 20 All the populations considered are estimated to have a negative base 21 temperature for embryo growth, ranging from -6.7 °C in BER to -2.7 in BAS. 22 Although it cannot be discarded that such low values are an artefact of the 23 thermal time modelling approach, values of T_b lower than zero have been 24 reported for some temperate trees, crops (mainly legumes) and wild plants but 25

are not common (Dürr et al., 2015). However, to our knowledge, no values as 1 2 low as -6.7 °C have been reported previously, the lowest being a T_b of -3.9 °C for Cryptantha minima (Boraginaceae) (Wei et al., 2009) and -4.5 °C for 3 Krascheninnikovia lanata (Amaranthaceae) (Wang et al., 2006). The 4 germination of Cryptantha minima at negative temperatures was explained as 5 an adaptation to take advantage of the water of the snowmelt in early spring 6 7 and develop its annual cycle before the summer drought (Wang et al., 2006). In the case of Conopodium majus, that is a perennial, this strategy could, 8 however, offer some advantage at the southern range of its distribution, where 9 10 summer drought can be a recurrent issue, as already observed for 11 Mediterranean subalpine species (Fernández-Pascual et al., Krascheninnikovia lanata seeds show a positive effect of seed size on the ability 12 13 to germinate at sub-zero temperatures (Wang et al., 2006). The authors demonstrate that bigger seeds had a higher concentration of sugars (glucose, 14 15 raffinose and sucrose) that probably lower the freezing point of the seed tissues. There are no reports on sub-zero germination in Apiaceae but an 16 optimal temperature for embryo growth of 2 °C has been described for 17 18 Heracleum sphondylium (Stokes, 1953) and it is not unlikely that this species, or others from the same family, could present equally low T_b for embryo growth. 19 However, in practical terms, it is unlikely that such low temperatures do have a 20 21 functional ecological role in embryo growth in the field: our field collected data and averaged climatic data from 1970-2000 for the collection sites show that 22 such low mean temperatures are rare in the natural environment of 23 Conopodium majus. Therefore, in the field it is likely that embryo growth is 24 possible throughout the winter season and is limited only by the higher 25

temperatures (and the ceiling threshold) in autumn. In fact, results from the

ordination analysis, showed that T_b is independent from climatic and geographic

factors and is not even correlated to seed size or initial E:E ratio. Therefore, we

can conclude that the limiting factor for embryo growth in Conopodium majus is

the ceiling temperature and its interaction with warmer temperatures during the

6 annual cycle.

2

3

4

5

9

10

11

12

13

14

15

16

17

18

20

21

22

23

24

25

7 Our thermal model outputs are supported by the results of the field sowings,

8 although these should be taken with caution since we were not able to perform

field sowing at the southernmost distribution of our experiment, in central Spain.

The responses of different populations to the warmer, drier sites in central

Spain could provide useful data for predicting possible effects of a warming

climate on seed regeneration in Conopodium majus. A most interesting follow

up study would involve reciprocal transplants of populations across the

latitudinal gradient of the species, to establish how much of the variation

measured and modelled is genetic and how much is due to phenotypic plasticity

(Franks et al., 2014). This would inform on the role for adaptation in any

species' response to environmental change through seed regeneration.

CONCLUSION

19 Species responses to climate determine how plants cope with ongoing global

change. Parmesan and Hanley (2015) identified three key issues for climate

change research on plants: changing phenology, changing distributions and the

role of plasticity and adaptation. Our study suggests that the thermal thresholds

for post-dispersal embryo growth may have functional relevance for these three

issues. The dependence of embryo growth on a relatively low ceiling

temperature means that warmer winter temperatures could slow down post

- dispersal embryo growth and potentially shift the emergence phenology from 1 2 one season to another, as has been described in alpine systems in relation to another seed process, physiological dormancy release (Mondoni et al., 2012). 3 Regarding distribution, we have found that two vicariant subspecies show 4 diverging thermal thresholds for embryo growth. This supports the concept that 5 seed ecophysiology can be a major driver of the distribution of taxa (Bykova et 6 7 al., 2012), and of species migrations as a response to climate change (Walck et al., 2011, Baskin and Baskin, 2022). At the same time, our study also shows 8 intraspecific variation in the embryo thermal thresholds, and therefore the 9 10 potential for plasticity and/or adaptation to changes in the climatic environment (Cochrane et al., 2015, Nicotra et al., 2010, Franks et al., 2014). 11
- 12 To date, most studies on the responses of seeds to climate have focused on germination. To complete the picture, our study highlights the importance of 13 14 post-dispersal embryo growth, showing intraspecific variation in this key ecophysiological trait across the latitudinal distribution of a species. Thus, 15 embryo growth seems to be one of the many ways by which plants interact with 16 changing soil temperatures (Amstutz et al., 2024). We make the case for more 17 ecological, evolutionary and comparative studies on post-dispersal embryo 18 19 growth, fully incorporating this trait into the research agenda on plant regeneration (Saatkamp et al., 2019) and plant responses to climate change 20 (Parmesan and Hanley, 2015). 21

AKNOWLEDGEMENTS

- 23 Álvaro Bueno Sánchez, Joseba Garmendia, Luis Carlón, Giles Laverack and
- 24 Maria Marin helped with seed collection.

FUNDING

22

- 1 This research received funding from the People Programme (Marie Curie
- 2 Actions) of the European Union's Seventh Framework Programme FP7/2007-
- 3 2013/ under REA grant agreement n°607785.

4 CONFLICT OF INTEREST STATEMENT

5 The authors declare no conflict of interest.

6 AUTHOR CONTRIBUTIONS

- 7 Cristina Blandino: Conceptualization; Methodology; Investigation; Data
- 8 Curation; Formal Analysis; Visualization; Writing Original Draft Preparation;
- 9 Writing Review & Editing. **Brith Natlandsmyr**: Investigation; Writing Review
- 10 & Editing. Sylvi M. Sandvik: Investigation; Writing Review & Editing. Hugh
- 11 W. Pritchard: Funding acquisition; Conceptualization; Methodology; Writing -
- 12 Review & Editing. Eduardo Fernández-Pascual: Conceptualization;
- 13 Methodology; Investigation; Writing Review & Editing.

14 DATA AVAILABILITY STATEMENT

15 Upon acceptance, all data will be deposited into Zenodo.

16 LITERATURE CITED

27

- Allen PS, Meyer SE. 1998. Ecological aspects of seed dormancy loss. *Seed Science Research*, 8: 183-191.
- Amstutz A, Firth LB, Spicer JI, De Frenne P, Gómez-Aparicio L, Graae BJ, Kuś S, Lindmo S,
 Orczewska A, Rodríguez-Sánchez F, Vangansbeke P, Vanneste T, Hanley ME. 2024.
 Taking sides? Aspect has limited influence on soil environment or litter decomposition in pan-European study of roadside verges. *Pedobiologia*, 102: 150927.
- Baskin CC, Baskin JM. 2014. Seeds. Ecology, Biogeography and Evolution of Dormancy and
 Germination. Second Edition. San Diego: Academic Press.
- Baskin CC, Baskin JM. 2022. Plant regeneration from seeds: A global warming perspective. San
 Diego: Academic Press.
 - **Baskin CC, Milberg P, Andersson L, Baskin JM. 2000**. Deep complex morphophysiological dormancy in seeds of *Anthriscus sylvestris* (Apiaceae). *Flora*, **195**: 245-251.
- Baskin JM, Baskin CC. 2004. A classification system for seed dormancy. Seed Science Research,
 14: 1-16.
- 31 **Blandino C, Fernández-Pascual E, Marin M, Vernet A, Pritchard HW. 2019**. Seed ecology of 32 the geophyte *Conopodium majus* (Apiaceae), indicator species of ancient woodland 33 understories and oligotrophic meadows. *Plant Biology*, **21**: 487-497.
- 34 **Bykova O, Chuine I, Morin X, Higgins SI. 2012**. Temperature dependence of the reproduction niche and its relevance for plant species distributions. *Journal of Biogeography,* **39**: 2191-2200.
- 37 **Calviño CI, Teruel FE, Downie SR. 2016**. The role of the Southern Hemisphere in the evolutionary history of Apiaceae, a mostly north temperate plant family. *Journal of Biogeography*, **43**: 398-409.

Cochrane A, Yates CJ, Hoyle GL, Nicotra AB. 2015. Will among-population variation in seed
 traits improve the chance of species persistence under climate change? Global Ecology
 and Biogeography, 24: 12-24.

- Daws MI, Lydall E, Chmielarz P, Leprince O, Matthews S, Thanos CA, Pritchard HW. 2004.

 Developmental heat sum influences recalcitrant seed traits in *Aesculus hippocastanum* across Europe. *New Phytologist*, **162**: 157-166.
- **Dürr C, Dickie JB, Yang XY, Pritchard HW. 2015**. Ranges of critical temperature and water potential values for the germination of species worldwide: Contribution to a seed trait database. *Agricultural and Forest Meteorology*, **200**: 222-232.
- Ellis RH, Covell S, Roberts EH, Summerfield RJ. 1986. The influence of temperature on seed germination rate in grain legumes: II. Intraspecific variation in chickpea (*Cicer arietinum* L.) at constant temperatures. *Journal of Experimental Botany*, 37: 1503-1515
- **Fenner M. 1991**. The effects of the parent environment on seed germinability. *Seed Science Research*, **1**: 75-84.
 - **Fernández-Pascual E, Jiménez-Alfaro B, Bueno A. 2017.** Comparative seed germination traits in alpine and subalpine grasslands: Higher elevations are associated with warmer germination temperatures. *Plant Biology,* **19**: 32-40.
 - Fernández-Pascual E, Jiménez-Alfaro B, Caujapé-Castells J, Jaén-Molina R, Díaz TE. 2013. A local dormancy cline is related to the seed maturation environment, population genetic composition and climate. *Annals of Botany*, **112**: 937-945.
 - **Fernández-Pascual E, Mattana E, Pritchard HW. 2019**. Seeds of future past: Climate change and the thermal memory of plant reproductive traits. *Biological Reviews*, **94**: 439-456.
 - **Fernández-Pascual E, Seal CE, Pritchard HW. 2015.** Simulating the germination response to diurnally alternating temperatures under climate change scenarios: Comparative studies on *Carex diandra* seeds. *Annals of Botany*, **115**: 201-209.
 - **Fick SE, Hijmans RJ. 2017**. WorldClim 2: New 1-km spatial resolution climate surfaces for global land areas. *International Journal of Climatology,* **37**: 4302-4315.
 - **Finch-Savage WE, Steckel JRA, Phelps K. 1998.** Germination and post-germination growth to carrot seedling emergence: predictive threshold models and sources of variation between sowing occasions. *New Phytologist,* **139**: 505-516.
 - **Franks SJ, Weber JJ, Aitken SN. 2014**. Evolutionary and plastic responses to climate change in terrestrial plant populations. *Evolutionary Applications*, **7**: 123-139.
 - **Hardegree SP. 2006**. Predicting germination response to temperature. I. Cardinal-temperature models and subpopulation-specific regression. *Annals of Botany*, **97**: 1115-1125.
 - **Jurado E, Flores J. 2005**. Is seed dormancy under environmental control or bound to plant traits? *Journal of Vegetation Science*, **16**: 559-564.
 - **Lamont BB, Pausas JG. 2023**. Seed dormancy revisited: Dormancy-release pathways and environmental interactions. *Functional Ecology,* **37**: 1106-1125.
 - Maleki K, Soltani E, Seal CE, Colville L, Pritchard HW, Lamichhane JR. 2024. The seed germination spectrum of 486 plant species: A global meta-regression and phylogenetic pattern in relation to temperature and water potential. *Agricultural and Forest Meteorology*, 346: 109865.
 - **Martin AC. 1946**. The comparative internal morphology of seeds. *The American Midland Naturalist*, **36**: 513-660.
- Mateo G, López Udias S. 2003. Conopodium W.D.J. Koch. In: Nieto Feliner G, Jury SL, Herrero
 Nieto A, eds. Flora iberica Vol. X. Araliaceae-Umbelliferae: 168-181. Madrid: Real
 Jardín Botánico, CSIC.
 - **McCarragher SR, Goldblum D, Rigg LS. 2011**. Geographic variation of germination, growth, and mortality in sugar maple (*Acer saccharum*): Common garden and reciprocal dispersal experiments. *Physical Geography*, **32**: 1-21.

- Mondoni A, Probert R, Rossi G, Hay F, Bonomi C. 2008. Habitat-correlated seed germination
 behaviour in populations of wood anemone (*Anemone nemorosa* L.) from northern
 Italy. Seed Science Research, 18: 213-222.
- **Mondoni A, Rossi G, Orsenigo S, Probert RJ. 2012**. Climate warming could shift the timing of seed germination in alpine plants. *Annals of Botany,* **110**: 155-164.

- Nicotra AB, Atkin OK, Bonser SP, Davidson AM, Finnegan EJ, Mathesius U, Poot P, Purugganan MD, Richards CL, Valladares F. 2010. Plant phenotypic plasticity in a changing climate. *Trends in Plant Science*, **15**: 684-692.
- **Orrù M, Mattana E, Pritchard HW, Bacchetta G. 2012**. Thermal thresholds as predictors of seed dormancy release and germination timing: Altitude-related risks from climate warming for the wild grapevine *Vitis vinifera* subsp. *sylvestris*. *Annals of Botany*, **110**: 1651-1660.
- **Parmesan C, Hanley ME. 2015**. Plants and climate change: complexities and surprises. *Annals of Botany*, **116**: 849-864.
 - **Pausas JG, Lamont BB, Keeley JE, Bond WJ. 2022**. Bet-hedging and best-bet strategies shape seed dormancy. *New Phytologist,* **236**: 1232-1236.
 - Phartyal SS, Kondo T, Baskin JM, Baskin CC. 2009. Temperature requirements differ for the two stages of seed dormancy break in *Aegopodium podagraria* (Apiaceae), a species with deep complex morphophysiological dormancy. *American Journal of Botany*, **96**: 1086-95.
- **Porceddu M, Mattana E, Pritchard HW, Bacchetta G. 2017**. Dissecting seed dormancy and germination in *Aquilegia barbaricina*, through thermal kinetics of embryo growth. *Plant Biology*, **19**: 983-993.
 - **Rosbakh S, Poschlod P. 2015**. Initial temperature of seed germination as related to species occurrence along a temperature gradient. *Functional Ecology*, **29**: 5-14.
 - **Rowse HR, Finch-Savage WE. 2003**. Hydrothermal threshold models can describe the germination response of carrot (*Daucus carota*) and onion (*Allium cepa*) seed populations across both sub- and supra-optimal temperatures. *New Phytologist,* **158**: 101-108.
 - Saatkamp A, Cochrane A, Commander L, Guja LK, Jimenez-Alfaro B, Larson J, Nicotra A, Poschlod P, Silveira FAO, Cross AT, Dalziell EL, Dickie J, Erickson TE, Fidelis A, Fuchs A, Golos PJ, Hope M, Lewandrowski W, Merritt DJ, Miller BP, Miller RG, Offord CA, Ooi MKJ, Satyanti A, Sommerville KD, Tangney R, Tomlinson S, Turner S, Walck JL. 2019. A research agenda for seed-trait functional ecology. New Phytologist, 221: 1764-1775.
 - **Shimono Y, Kudo G. 2005**. Comparisons of germination traits of alpine plants between fellfield and snowbed habitats. *Ecological Research*, **20**: 189-197.
- **Stokes P. 1953**. A physiological study of embryo development in *Heracleum sphondylium* L.: III. The effect of temperature on metabolism. *Annals of Botany*, **17**: 157-174.
- Tutin TG, Heywood VH, Burges NA, Moore DM, Valentine DH, Walters SM, Webb DA. 1968.

 Flora Europaea. Volume 2. Rosaceae to Umbelliferae. Cambridge: Cambridge University Press.
- Vandelook F, Bolle N, Van Assche JA. 2007. Seed dormancy and germination of the european Chaerophyllum temulum (Apiaceae), a member of a trans-atlantic genus. Annals of Botany, 100: 233-239.
 - Vandelook F, Bolle N, Van Assche JA. 2009a. Morphological and physiological dormancy in seeds of *Aegopodium podagraria* (Apiaceae) broken successively during cold stratification. *Seed Science Research*, 19: 115-123.
- Vandelook F, Janssens SB, Probert RJ. 2012. Relative embryo length as an adaptation to habitat and life cycle in Apiaceae. *New Phytologist*, 195: 479-487.
- Vandelook F, Lenaerts J, Van Assche Jozef A. 2009b. The role of temperature in post-dispersal
 embryo growth and dormancy break in seeds of *Aconitum lycoctonum L. Flora Morphology, Distribution, Functional Ecology of Plants,* 204: 536-542.

- Vandelook F, Van Assche JA. 2008. Deep complex morphophysiological dormancy in Sanicula
 europaea (Apiaceae) fits a recurring pattern of dormancy types in genera with an
 Arcto-Tertiary distribution. Botany, 86: 1370-1377.
 - **Violle C, Reich PB, Pacala SW, Enquist BJ, Kattge J. 2014**. The emergence and promise of functional biogeography. *Proceedings of the National Academy of Sciences,* **111**: 13690-13696.
 - Walck JL, Hidayati SN, Dixon KW, Thompson K, Poschlod P. 2011. Climate change and plant regeneration from seed. *Global Change Biology*, 17: 2145-2161.
 - Walker M, Pérez M, Steinbrecher T, Gawthrop F, Pavlović I, Novák O, Tarkowská D, Strnad M, Marone F, Nakabayashi K, Leubner-Metzger G. 2021. Molecular mechanisms and hormonal regulation underpinning morphological dormancy: a case study using *Apium graveolens* (Apiaceae). *The Plant Journal*, 108: 1020-1036.
 - Wang R, Bai Y, Low NH, Tanino K. 2006. Seed size variation in cold and freezing tolerance during seed germination of winterfat (*Krascheninnikovia lanata*) (Chenopodiaceae). *Canadian Journal of Botany*, 84: 49-59.
- Wei Y, Bai Y, Henderson DC. 2009. Critical conditions for successful regeneration of an endangered annual plant, *Cryptantha minima*: A modeling approach. *Journal of Arid Environments*, 73: 872-875.

TABLES

4

5

6

7

8

9

10

11

12

13

14

15

19

20

Table 1: Provenance of seeds used in the experiments.

Population	Country	Location	Latitude	Longitude	Elevation (m a.s.l.)
HER	Norway	Herdla	60° 34'29.784" N	4º 56' 53.627" E	37
BER	Norway	Bergen	60° 20′ 7.35 N	5º 22' 17.79" E	97
FLE	Norway	Flekkeroya	58º 4'5.34" N	7º 59' 53.56" E	19
SCO	UK	Dalreoch Farm	56° 44' 47.36" N	3º 32' 25.03" W	252
WAK	UK	Wakehurst Place	51º 04' 12.79" N	0° 05' 28.28" W	114
BAS	Spain	Ondarre	43° 01' 42.8" N	2º 03' 55.7" W	809
LEO	Spain	El Tendero	42° 54′ 26,62″ N	5º 49' 25,87" W	1426
CHO	Spain	Central del Chorro	40° 18' 26.17" N	5º 40' 09.39" W	1398
TRE	Spain	Tremedal	40° 22' 00.5" N	5º 37' 57.20" W	1555

23

22

Table 2: Initial E:E, seed dry mass and cardinal temperatures averaged

- between deciles (all as mean ± SD) in seeds of all populations of Conopodium
- 26 majus studied. To have a symmetric result around the middle value, when the
- lower deciles were excluded for being too close to the initial embryo size, the
- 28 higher ones were excluded too.

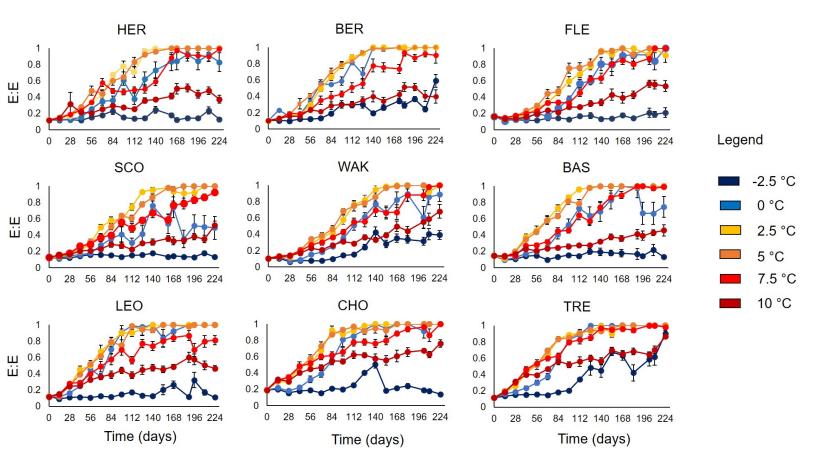
Population Initial E:E Seed dry	T _b (°C)	T _o (°C)	T _c (°C)	Deciles
---------------------------------	---------------------	---------------------	---------------------	---------

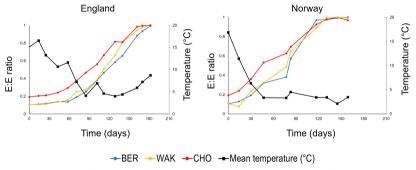
		used		
HER	0.12 ±0.03	1.94 ±0.58 -4.01 ± 0.57 4.26 ± 0.5	30 12.90 ± 1.86	0.3 - 0.7
BER	0.10 ±0.03	1.80 ±0.55 -6.65 ± 0.62 4.58 ± 0.0	02 12.08 ± 1.32	0.3 - 0.7
FLE	0.17 ±0.04	2.03 ±0.53 -3.90 ± 0.14 4.50 ± 0.	07 13.70 ± 0.71	0.3 - 0.7
SCO	0.13 ±0.02	1.63 ±0.54 -2.63 ± 0.38 2.80 ± 0.3	25 14.42 ± 2.47	0.3 - 0.7
WAK	0.10 ±0.02	1.75 ±0.42 -6.20 ± 0.89 4.59 ± 0.	14.44 ± 1.72	0.3 - 0.7
BAS	0.15 ±0.02	1.43 ±0.59 -2.75 ± 0.10 2.69 ± 0.	10 13.07 ± 0.93	0.3 - 0.7
LEO	0.12 ±0.03	1.66 ±0.52-3.17 ± 0.06 2.54 ± 0.0	03 14.64 ± 2.23	0.4 - 0.6
CHO	0.19 ±0.03	1.24 ±0.38 -4.09 ± 0.59 5.23 ± 1.0	05 20.48 ± 9.09	0.4 - 0.6
TRE	0.11 ±0.02	1.21 ±0.38 -6.47 ± 0.41 4.86 ± 0.41	04 20.54 ± 7.25	0.4 - 0.6

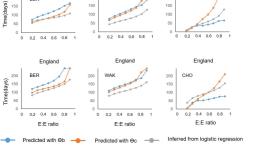
2

FIGURES

- Fig. 1: Patterns of embryo growth (E:E ratio) for all the seed populations of
- 4 Conopodium majus and all temperatures tested. Each data point represents the
- 5 mean of ten replicates (± SE).
- 6 **Fig. 2:** Embryo growth in the field for buried seeds of *Conopodium majus*. Each
- 7 data point represents the mean E:E ratio for 20 seeds of three representative
- 8 populations: CHO for the southern edge of the distribution range, WAK for the
- 9 middle and BER for the northern; soil temperature is also shown. The burial
- 10 experiment was performed in England (on the collection site of the WAK
- population) and in Norway (on the collection site of BER population). The
- experiment started on 1st September 2016 in England and on 15th September
- 2016 in Norway. For each site, the experiment finished when all populations
- reached 100% radicle emergence (corresponding to E:E =1).
- Fig. 3: Time (in days) required by each population of *Conopodium majus* seeds
- in each field location to reach different deciles of E:E ratio according to: 1)
- interpolation from the logistic regression of embryo growth in the field (grey
- line); 2) ΘT_b (blue line); and 3) ΘT_c (orange line) obtained from the model.
- 19 Fig. 4: Principal component analysis of seed traits (in red) and geographic and
- 20 bioclimatic variables (in blue) across the latitudinal distribution of *Conopodium*
- 21 majus.







Norway

200 WAK

150

Norway

BER

200

150

100

Norway

200 CHO

150

