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Compositional novelty of plant, fungal and bacterial communities across urban habitats --Manuscript Draft--

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Abstract:	We tested the hypothesis that cities are composed of different degrees of ecological novelty by studying four urban habitats with differing degrees of management and human legacy: park lawns, roadsides, residential vacant lots, and industrial vacant lots. We focused on community compositional novelty, by comparing the plant, fungal and bacterial species composition between urban habitats and two reference pre-urban habitats: forest remnants and hay meadows. We used a compositional novelty index based on multidimensional ordination, which is straightforward to calculate and only requires species co-occurrence data for urban and reference habitats. As expected, (1) plant communities displayed the highest novelty and soil bacteria the lowest; (2) urban communities were markedly different from forest communities and relatively more similar to meadow communities; and (3) the degree of compositional novelty was highest in industrial vacant lots. On the contrary, managed park lawns, which we had expected to be highly novel, were relatively close to hay meadows. The lowest novelty among urban habitats was recorded in residential vacant lots, which had biological communities that more closely resembled those of pre-urban habitats. Our results highlight the effect of habitat type as a major driver of urban community composition and compositional novelty. These results suggest that city biodiversity can be enhanced by an integrative approach to the urban landscape that favors habitat heterogeneity by passive rewilding of managed park lawns, non-intervention on residential vacant lots, direct restoration of industrial vacant lots, and conservation of natural and agricultural habitat remnants as sources of native species.

- A compositional novelty index based on multidimensional ordination is straightforward to calculate and only requires species co-occurrence data for urban and reference pre-urban habitats.
- Novelty was the highest for plant communities and the lowest for soil bacteria.
- Urban communities were markedly different from forest communities and relatively more like meadow communities.
- Compositional novelty was highest in industrial vacant lots, and lowest in residential vacant lots.
- Park lawns were relatively close in composition to hay meadows.

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1 Compositional novelty of plant, fungal and bacterial communities across

2 urban habitats

3 Running title: Compositional novelty across urban habitats

4 Abstract (250 words)

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Keywords (7)

- 6 Urban biodiversity, vegetation, soil fungi, soil bacteria, DNA metabarcoding, soil pollution,
- 7 novel ecosystems

8 Highlights (3 to 5, 85 words each)

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- more like meadow communities.
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- vacant lots.
- Park lawns were relatively close in composition to hay meadows.

Introduction

Urban development affects landscape composition and related biodiversity patterns worldwide (Grimm et al., 2008). Urbanization converts natural and agricultural habitats into a fine-scale mosaic of diverse urban habitats varying in size, fragmentation, past and present land uses, and degrees of human intervention (Pauleit and Breuste, 2011). At the same time, novel urban habitats experience climatic differences from the surrounding landscape, as well as high within-city microclimatic variation (Forman, 2014; Parlow, 2011). Urban soils likewise vary in permeability, compaction, organic matter content and human-concentrated chemicals (Forman, 2014; Sauerwein, 2011). Urban habitats are colonized by organisms from the regional species pool that are capable of living in urban environments (Johnson et al., 2017). The process of colonization requires species to pass through a series of hierarchical filters (biogeographical, abiotic, biotic and human) resulting in the formation of an urban species pool and the assembly of specialized urban communities (Aronson et al., 2016; Swan et al., 2021). Urbanization promotes specialized ruderal species and traits (Di Giulio et al., 2024; Raposo et al., 2024; Sotillo et al., 2024), and thus urban communities are rich in species that can tolerate urban disturbance, drought and pollution (Forman, 2014). For this reason, the urban biota can be a source of nature-based solutions, such as the bioremediation of degraded soils (Matanzas et al., 2021; Song et al., 2019) and ecosystem design (Klaus and Kiehl, 2021; Perrelet et al., 2024). Urban communities also host a high proportion of the regional biodiversity (Fernández-Pascual et al., 2025; Gentili et al., 2024) and make up a major part of the supply of natural elements in densely populated landscapes (Kowarik, 2018). Proper management of urban communities

can turn them into assets with high ecological and social returns (Anderson and Minor, 2017; Fekete et al., 2024; Itescu and Jeschke, 2024). The diversity of urban communities can be characterized along two axes: naturalness and novelty (Kowarik, 2018). Naturalness refers to the degree of self-organization, i.e., the extent to which community composition and functions are self-assembled or dependent on direct human inputs (Kowarik, 2018). Novelty refers to the degree of differentiation from the historical reference communities that existed in the area before urbanization (Teixeira et al., 2021; Teixeira and Fernandes, 2020). For example, urban communities under direct human management (e.g., park lawns) should have low naturalness and high novelty. When management stops, naturalness increases, and the community can either transition back to a state resembling the historical reference (e.g., a low novelty hay meadow) or into a novel assemblage of species (e.g., a high novelty urban grassland with non-native species) (Teixeira et al., 2021; Teixeira and Fernandes, 2020). The fate of a specific urban site will depend on the interaction between the legacy of human intervention and inherent site characteristics (Cramer et al., 2008). Intense human legacies usually push communities into novel states, often dominated by non-native and otherwise unwanted taxa (Dunn and Heneghan, 2011; Forman, 2014), which usually require more intensive and costly actions to restore the land for conservation purposes (Cramer et al., 2008). Understanding the patterns and drivers of urban community novelty can help to predict the restoration and rewilding potential of vacant urban spaces, facilitating their integration into the management of biodiversity in cities.

The goal of this article was to test the hypothesis that urban habitats are comprised of different degrees of ecological novelty (Teixeira and Fernandes, 2020). We addressed this hypothesis

by studying four urban habitats representing different land uses and human legacies, from managed park lawns to relatively unmanaged strips of land along transportation corridors, physically altered soils in residential vacant lots, and highly contaminated industrial brownfields. To address community compositional novelty, we compared the plant, fungal and bacterial species composition between urban habitats and two reference pre-urban habitats: forest remnants and hay meadows. We expected novelty to be the highest for plants, which in many cases are the object of direct management and introduction (Lázaro-Lobo et al., 2024); and lowest for bacteria, which should be less affected than fungi by the usually alkaline urban soils (Forman, 2014). We expected novelty to be higher when urban habitats are compared to natural habitats (forests), and lower when compared to agricultural habitats (meadows). We also expected novelty to be higher in the actively managed habitat (park lawns) and in the unmanaged habitat with the highest human legacy (industrial brownfields). Finally, we expected that compositional novelty would be related to soil physicochemical properties, reflecting the major aspect of human legacy in urban habitats. To test these expectations, we tackled three main research goals: (1) to characterize the variation in soil properties across urban and pre-urban habitats; (2) to characterize the variation in plant, fungal and bacterial communities across urban and pre-urban habitats; and (3) to measure the compositional novelty across habitats and taxonomic groups.

Materials and methods

20 Study city

Gijón/Xixón is a medium-sized coastal city (c. 270,000 inhabitants; Instituto Nacional de Estadística) located in the Bay of Biscay, within the central Asturias metropolitan area (c. 800,000 inhabitants) of northern Spain. The city belongs to the Cantabrian Mixed Forests ecoregion (Fernández-Pascual et al., 2025; Lázaro-Lobo et al., 2024), at the border between the temperate and Mediterranean climatic zones of Europe. Climate is temperate oceanic with cool summers and wet and mostly frost-free winters (mean annual temperature = 13.5 °C, annual precipitation = 1050 mm; Agencia Estatal de Meteorología). Most of the city is built on alluvial and estuarine deposits, sand or limestone bedrock. The potential pre-human vegetation was likely dominated by deciduous mixed forests, together with riparian forests, wetlands and coastal dunes. The area has a long history of human habitation dating back to the Cantabrian Upper Paleolithic (Straus, 2005). The city itself has been inhabited at least since Roman times (Fernández Ochoa and Martínez Díaz, 1987). During the medieval and early modern times, the urban center was limited to the original walled city, with fishing being the main economic activity. Expansion beyond the original core started in the 18th and 19th centuries. Exposure to global trade networks and the warm temperate climate turned the region into a hotspot for biological invasions, especially from the Americas and Asia (Lázaro-Lobo et al., 2024). In 1852, the city was connected by railway to regional mining and industrial centers and became a major shipping port (Gómez-Martín and Cañal-Fernández, 2024). During the following century the city experienced significant industrial development (e.g., canning, coal, steel, shipbuilding) and rapid population growth, becoming the most populated city in the Principality of Asturias (Obeso Muñiz and Fernández García, 2017). Industrial and urban growth brought environmental problems including loss of natural wetlands and air and watercourse pollution (Utanda Moreno and Feo Parrondo, 1995).

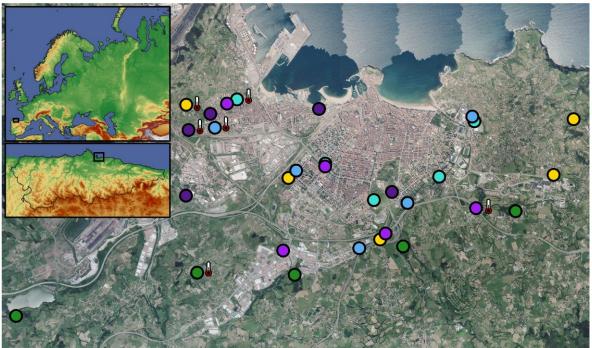
- 1 Starting in the 1970s, the city and the metropolitan area have been affected by post-industrial
- 2 land-use changes leaving large abandoned industrial sites (i.e., brownfields) (Gallego et al.,
- 3 2016; Matanzas et al., 2021).
- 4 Definition of urban habitats
- 5 To survey urban biodiversity across habitats, we defined six habitat types (Figure 1A)
- 6 representing different theoretical states of naturalness and novelty (Kowarik, 2018): (1) forest
- 7 patches < 10 km from the city center, i.e., remnants of pre-urban natural habitats; (2) late
- 8 mown hay meadows, i.e., remnants of pre-urban agricultural habitats; (3) park lawns, i.e.,
- 9 green spaces created and maintained by humans; (4) roadsides, i.e., relatively unmanaged
- strips of land alongside roads and train tracks; (5) residential vacant lots, i.e., unmanaged
- land which has suffered some degree of soil physical disturbance and is waiting to be
- urbanized; and (6) industrial vacant lots, i.e., unmanaged land with an intense legacy of soil
- disturbance and contamination.
- 14 Sampling plots
- During June 2023, we surveyed the city to establish five sampling plots (4 x 4 m) per each
- of the six habitat types (n = 30 plots). The plots were distributed across the city (**Figure 1B**)
- and represent the variation of vegetation types found within each habitat type. In one plot per
- habitat (**Figure 1B**), we buried a datalogger (M-Log5W, GeoPrecision, Ettlingen, Germany;
- accuracy: +/- 0.1 °C (at 0 °C), resolution: 0.01 °C) at a depth of 5 cm below the upper layer
- of the soil. Dataloggers recorded temperature hourly for one year (1 Jan to 31 Dec 2024),

- 1 after which we downloaded their records to describe the thermal environment of the city and
- 2 the habitats.

(A) Habitat types



(B) Distribution of plots across the study area



- 1 Figure 1: Pre-urban and urban habitats included in this study. (A) Representative pictures
- 2 of each habitat, indicating their level of naturalness (i.e., degree of ecosystem self-
- 3 organization) and novelty (i.e., degree of divergence from pre-human reference ecosystems).
- 4 (B) Location of the sampling plots within the city of Gijón/Xixón, Asturias, Spain, Western
- 5 Europe. Each plot (dots) is colored according to habitat type. A thermometer to the right of
- 6 the sampling plot indicates that a soil temperature datalogger was installed there.
- 7 Soil physicochemical sampling
- 8 From each of the 30 plots, we collected 500 g of soil to analyse soil physicochemical
- 9 properties. We took the samples from 5 points in each plot (center + 4 cardinal points), down
- to a depth of 10 cm. We removed stones and other debris, sieved the soils in a 5 mm mesh
- and homogenized the samples in a plastic bag. Samples were analyzed at INEA Laboratory
- 12 (Valladolid, Spain), following the standard methodology of the Spanish Ministry of
- 13 Agriculture (Vallejo Acevedo, 1994). Soil analysis included pH, texture, bulk density,
- electrical conductivity, organic matter content, total N, C/N ratio, active limestone, total
- carbonates, assimilable P, K, Ca, Mg, Na and heavy metals (Zn, Cr, Cu, Cd, Hg, Pb, Ni).
- 16 Plant, fungal and bacterial diversity sampling
- 17 In each plot, we recorded all the spontaneous vascular plant species, as well as the cover (%)
- of each species in the plot. Plant species names follow Euro+Med (Euro+Med, 2006).
- 19 Additionally, from each of the 30 plots, we collected soil samples for DNA metabarcoding
- 20 of fungal and bacterial (including Archaea) communities. We performed the sampling in
- sterile conditions, and always before entering the plot for any of the other tasks. We cleaned

and sterilized all tools with water and ethanol between samples. In each plot, we took samples from the four cardinal points, 0.5 m inwards from the edge, at a depth of 5 cm. Before taking the samples, we removed the vegetation with a sterile tool. Then, we collected the soil samples in a 50 ml Falcon tube, mixed them, homogenized them with a lancet and finally stored the sample in a 2 ml Eppendorf tube which we immediately froze. The frozen tubes were sent to AllGenetics & Biology SL (A Coruña, Spain) for DNA metabarcoding analysis. DNA analysis included DNA extraction; library preparation with 2 pairs of different primers specific for barcoding (ITS for fungi, and 16S for bacteria; sequencing on an Illumina NovaSeq PE250 platform (assuming an amplicon size < 450 bp); and standard bioinformatic analysis including demultiplexing, quality control and sample preprocessing, inference of Amplicon Sequence Variants (ASVs), taxonomic assignment, number of reads per taxon and generation of rarefaction curves (see Appendix S1 for extended DNA metabarcoding methodology).

Statistical analysis and compositional novelty index

We performed all analyses using R version 4.3.1 (R Core Team, 2023) and the package *tidyverse* (Wickham et al., 2019) for data manipulation and visualization. To describe soil temperatures as recorded by the dataloggers, we calculated the mean annual temperature, the mean diurnal range and the annual range. To assess the variability in soil physicochemical properties, we conducted a Principal Component Analysis (PCA) as implemented in the R package *FactoMineR* (Lê et al., 2008) and a correlation analysis to detect auto-correlated soil variables. Multivariate analyses of community composition were performed in *vegan* (Oksanen et al., 2022): we used PERMANOVA to test differences between habitat types in

 their soil variables and their composition of plant species, fungal ASVs and bacterial ASVs;

2 Non-metric Multi-Dimensional Scaling (NMDS) ordination to assess variation in taxonomic

composition of plants (abundance values of species), fungi (presence/absence of ASVs) and

bacteria (presence/absence of ASVs); and environmental fitting to test the correlation

5 between community ordination axes and soil physicochemical properties.

6 To calculate a compositional novelty index, we used the NMDS multidimensional distance

from each urban plot (i.e., park, roadside, residential or industrial) to the centroid of the two

reference pre-urban habitats (i.e., forests and meadows). We calculated compositional

novelty separately for plant, fungal and bacterial communities. We tested the effect of

taxonomic group, pre-urban reference habitat (forest or meadow), urban habitat (park,

roadside, residential or industrial) and soil physicochemical variables on compositional

novelty, by fitting a Generalized Mixed Model with the package *nlme* (Pinheiro et al., 2020).

The model included sampling plot as a random factor. We checked and confirmed model

assumptions using the package *performance* (Lüdecke et al., 2021).

Results

16 Soil properties across habitats

Soil temperature records (**Figure 2A**) indicated that forest soils were relatively buffered in

their thermal range of variation, with a mean annual temperature of 13.9 °C, a mean diurnal

range of 1.8 °C and an annual range of 10.8 °C. Meadow and urban plots had more thermal

oscillation, with a mean annual temperature between 12.4 °C (roadsides) and 16.8 °C

- 1 (meadows), a mean diurnal range between 3.8 °C (roadsides) and 4.6 °C (residential lots),
 2 and an annual range between 16.5 °C (residential lots) and 19.5 °C (meadows).
- PCA ordination of the soil physicochemical properties (**Appendix S2**) indicated that the first axis of variation, explaining 23% of the variance, ordered soils according to the concentration of heavy metals. The variables with the highest contribution to this axis (> 10%) were the soil contents in Pb, Cu, Zn, Cd and Cr, which accounted for 66% of the variability of the first axis. This first axis did not separate urban from pre-urban habitats: for example, while some industrial lots had a very high concentration of Pb and other metals, the median Pb value of

the industrial habitat was below the median value of forests (Figure 2B; Appendix S2).

- The second PCA axis (**Appendix S2**) explained 21% of the variance and ordered soils according to their compaction and water retention capacities, with the highest contributing variables (> 10%) being organic matter content, bulk density, N concentration and pH, which made up 59% of the axis' variability. This axis separated soils according to their habitat: for example, forests had high organic matter and N content, while industrial vacant lots had higher bulk density and pH (**Figure 2B**; **Appendix S2**).
- The third PCA axis (**Appendix S2**) explained 10% of the variability and ordered soils according to their content in Mg, with the highest contributing variables (>10%) being Mg content, Mg/K ratio and conductivity, together explaining 50% of the axis. In this case, like for the first axis, there was no clear separation between habitats, with industrial sites having some of the highest and lowest values (**Figure 2B**; **Appendix S2**).

- 1 The analysis showed strong correlations within the groups of variables contributing to the
- 2 first, second and third PCA axes (**Appendix S2**). To avoid autocorrelation issues, for further
- analyses we kept three variables which taken together provide a straightforward biological
- 4 interpretation of patterns in soil composition: Pb content, organic matter content and Mg
- 5 content (**Figure 2B**). As previously indicated by PCA, organic matter was the only one with
- 6 a clear pattern of variation: a decrease from pre-urban to urban habitats (Figure 2B).
- 7 PERMANOVA supported that there was not a significant difference between habitats in their
- 8 soil physicochemical properties (100,000 permutations, $R^2 = 0.19$, F = 1.15, p = 0.31).

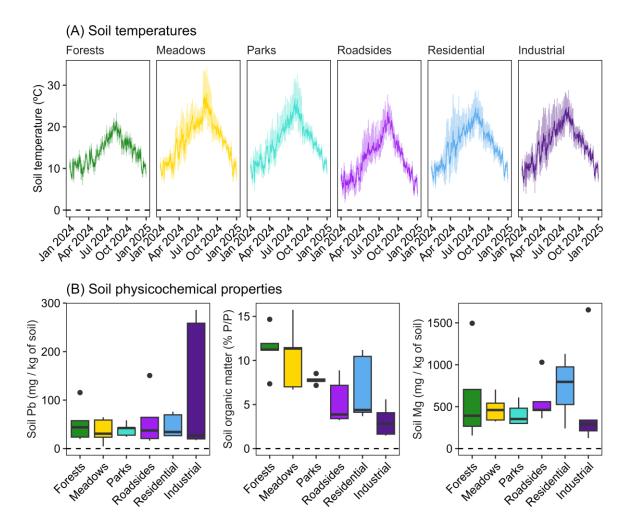


Figure 2: Soil properties across habitats. (A) Temperature recorded every hour in one sampling plot per habitat from 1 Jan 2024 to 31 Dec 2024, indicating the hourly records (shaded colors) and the daily averages (full colors). (B) Soil physicochemical properties. Values are shown for the three most explanatory and uncorrelated variables: soil Pb content, soil organic matter content, and soil Mg content.

- Plant, fungal and bacterial communities across habitats
- 8 The survey of spontaneous plant diversity yielded 216 species, with the average richness per
- 9 plot being 26 (min = 10, max = 38). Of these species, 48% were found only in urban habitats

(parks, roadsides, residential lots and industrial lots), 27% occurred only in pre-urban habitats (forests and meadows) and 25% were shared by both urban and pre-urban habitats (Fig3A). On average, the habitat with the highest richness per plot was industrial lots (29 species), followed by residential lots (27), roadsides (24), meadows (23), parks (20) and forests (18). The most frequent species were Holcus lanatus (22 plots), Plantago lanceolata (20), Helminthotheca echioides (16), Dactylis glomerata (15) and Lotus corniculatus (14). PERMANOVA supported that the plant species composition was significantly different across habitats (100,000 permutations, $R^2 = 0.42$, F = 3.48, p = < 0.001). NMDS ordination of the plant species composition (Fig. 3D) indicated that the plots belonging to the same habitat were relatively grouped together. The major gradient in vegetation composition along the first axis was a separation between forest plots and the other habitats. Environmental fitting of the Pb, organic matter and Mg vectors onto the ordination indicated that none of the soil factors had a significant correlation ($R^2 < 0.14$, p > 0.05) with the composition of the plant communities. DNA metabarcoding of soil fungal communities resulted in 4,689 amplicon sequence variants (ASVs). Of these, 46% were only found in urban habitats (parks, roadsides, residential lots and industrial lots), 36% occurred only in pre-urban habitats (forests and meadows) and 18% were shared by both urban and pre-urban habitats (Fig3B). The average number of ASVs per plot was 340 (min = 174, max = 587), with the habitat averages decreasing from residential vacant lots (415) to forests (393), meadows (336), parks (323), industrial vacant lots (300) and roadsides (276). The majority of ASVs were assigned to Ascomycota (56%), Basidiomycota (24%), Rozellomycota (10%) and Glomeromycota (4%).

Forest habitats had the highest proportion of Basidiomycota (25% vs. < 17% in the other habitats) and the lowest proportion of Ascomycota (57% vs. > 64%). The proportion of Rozellomycota was similar across habitats (from 6% in meadows to 11% in parks). The proportion of Glomeromycota varied between 5% in meadows to 0.5% in forests. PERMANOVA performed on the presence/absence of ASVs indicated significant differences in composition between habitats (100,000 permutations, $R^2 = 0.33$, F = 2.39, p =< 0.001). NMDS ordination of the fungal ASV composition (Fig. 3E) behaved similarly to the ordination of plant communities, with plots tending to group by habitat, and the major compositional gradient reflecting the separation between forest plots and the rest. Environmental fitting of the Pb, organic matter and Mg vectors onto the ordination indicated that only organic matter had a significant correlation ($R^2 = 0.46$, p < 0.001) with the composition of the fungal communities. Low organic matter content was associated with the fungal communities from industrial vacant lots and roadsides (Fig. 3E). For bacterial communities, DNA metabarcoding yielded 16,306 ASVs. Of these, 44% were found only in urban habitats (parks, roadsides, residential lots and industrial lots), 31% occurred only in pre-urban habitats (forests and meadows) and 25% were shared by both urban and pre-urban habitats (Fig3C). The average number of ASVs per plot was 1769 (min = 1478, max = 2074), with the habitat averages decreasing from roadsides (1848) to industrial lots (1794), residential lots (1792), meadows (1775), parks (1706) and forests (1697). The majority of ASVs were assigned to Proteobacteria (22%), Planctomycetota (15%), Actinobacteriota (15%) and Acidobacteriota (10%). Compared to other habitats, forests had higher proportions of Proteobacteria and Acidobacteriota, a lower proportion of

vacant lots and roadsides (Fig. 3F).

Actinobacteriota, and a similar proportion of Planctomycetota. PERMANOVA performed on the presence/absence of ASVs indicated significant differences in composition between habitats (100,000 permutations, $R^2 = 0.32$, F = 2.51, p = < 0.001). NMDS ordination of the bacterial ASV composition (Fig. 3F) resulted in less grouping by habitat than for plants and fungi, but still reflected a main compositional gradient from forests to industrial vacant plots. Like in the case of the fungal communities, environmental fitting of the Pb, organic matter and Mg vectors onto the ordination indicated that only organic matter content had a significant correlation ($R^2 = 0.39$, p = 0.001) with the composition of the bacterial communities. Low organic matter content correlated with the communities from industrial

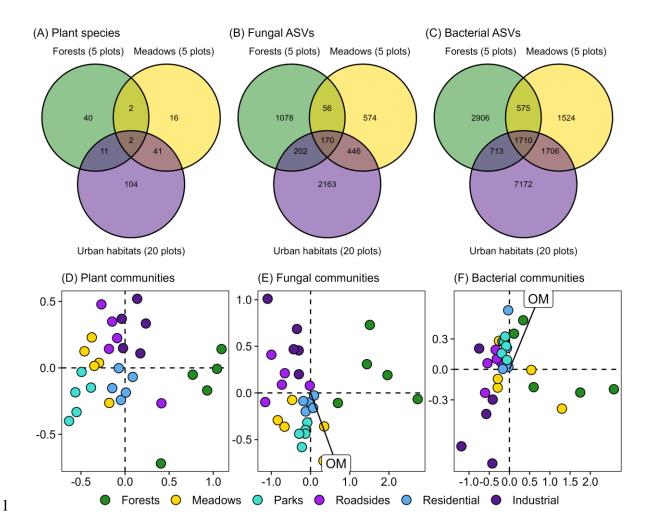


Figure 3: Plant, fungal and bacterial diversity across habitats. Top panels show Venn plots for the shared numbers of (A) plant species, (B) fungal ASVs and (C) bacterial ASVs between urban (parks, roadsides, residential lots and industrial lots) and pre-urban (forests, meadows) habitats. Bottom panels show biplots of community composition ordination produced by Non-metric Multi-Dimensional Scaling (NMDS). NMDs were conducted for (D) the plant survey (by species abundance), (E) the soil fungal survey (by ASV presence/absence) and (F) the soil bacterial survey (by ASV presence/absence). Colors indicate the habitat. In the biplots, arrows and labels indicate soil variables with a significant

- 1 correlation with the composition of the communities. Only soil organic matter content and
- 2 fungal-bacterial communities had a significant correlation.
- 3 Compositional novelty across habitats and taxonomic groups
- 4 We used the NMDS multidimensional compositional distance (calculated from the
- ordinations in **Fig. 3D-F**) from each of the urban plots (i.e., park, roadside, residential or
- 6 industrial) to the centroid of the two reference pre-urban habitats (i.e., forests and meadows)
- 7 to calculate a compositional novelty index (**Fig. 4**). Novelty was significantly different
- between taxonomic groups (F = 45.88, p < 0.001), being higher for plants (2.5 \pm 0.3 S.E.)
- 9 than for fungi (2.2 \pm 0.4 S.E.) and bacteria (1.6 \pm 0.3 S.E.). Novelty was also significantly
- different depending on the pre-urban habitat that was used as a reference (F = 199.08, p <
- 11 0.001), being higher compared to forests (2.6 \pm 0.3 S.E.) than to meadows (1.5 \pm 0.3 S.E.).
- Nonetheless, for each taxonomic group, there was a high positive correlation between the
- 13 novelty compared against forests and meadows (r = 0.5 for plants; 0.7 for fungi; 0.9 for
- bacteria). Different urban habitats also differed significantly in their novelty (F = 9.51, p =
- (0.001), being higher in industrial vacant lots $(2.5 \pm 0.3 \text{ S.E.})$ than in roadsides (2.2 ± 0.4)
- 16 S.E.), parks (2.0 \pm 0.4 S.E.) and residential vacant lots (1.6 \pm 0.3 S.E.). The soil
- 17 physicochemical properties (Pb, organic matter, Mg) had no significant effect on
- 18 compositional novelty (p > 0.05).



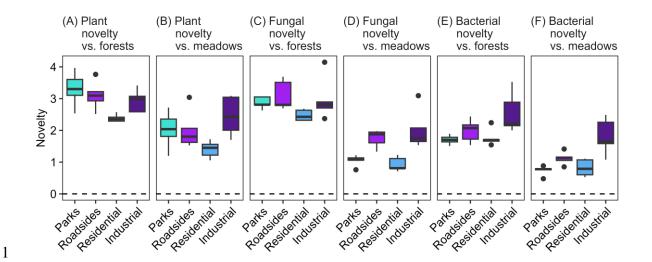


Figure 4: Compositional novelty for plants, fungi and bacteria. Compositional novelty was calculated as the NMDS multidimensional distance from each urban plot to the centroid of two reference pre-urban habitats (forests, panels A-C-E; and meadows, panels B-D-F). Novelty was calculated separately for plants (panels A-B), fungi (panels C-D) and bacteria (panels E-F).

Discussion

This study shows the compositional novelty of biological communities across urban habitats when they are compared with two reference pre-urban habitats: forest remnants and agricultural hay meadows. As expected, (1) novelty was the highest for plant communities and the lowest for soil bacteria; (2) urban communities were markedly different from forest communities and relatively more similar to meadow communities; and (3) the degree of compositional novelty was the highest among industrial vacant lots. However, managed park lawns, which we had expected to be highly novel, were relatively close in composition to hay meadows. Among urban habitats, the residential vacant lots had the lowest novelty, with

- biological communities that more closely resembled those of the reference pre-urban
- 2 habitats.
- 3 Widespread soil contamination across urban habitats
- 4 Our results partly agreed with the expected effects of urbanization on soils (Forman, 2014;
- 5 Sauerwein, 2011): we found more microclimatic variation, less organic matter content and
- 6 more compacted and alkaline soils than their pre-urban counterparts. But, contrary to our
- 7 expectations, urban and pre-urban soils did not show a consistent pattern of contamination
- 8 by heavy metals, possibly because pollution is widespread across the city. Indeed, soil
- 9 contamination had a weak but positive correlation (r = 0.25) with the geographical proximity
- to the hotspot of heavy industry in the western end of the city (Obeso Muñiz and Fernández)
- García, 2017). This geographical correlation was especially strong for the residential lots
- (0.89), parks (0.68) and forests (0.68); and weak for the roadsides (0.10), industrial lots (0.03)
- and meadows (-0.14). Overall, the measured soil properties had a minor effect on the
- 14 composition of the biological communities, as organic matter content was the only variable
- to influence the composition of soil fungal and bacterial communities.
- 16 Contrasting diversity patterns in plant-bacterial vs. fungal communities
- Our survey found 216 vascular plant species growing in the study plots, representing 9 % of
- the regional flora (Fernández Prieto et al., 2014). The highest plant species richness was
- recorded in vacant industrial and residential plots, while forests displayed the lowest. It must
- 20 be considered that vacant lots, especially those of industrial origin, had a high number of
- 21 non-native species, including five that are classified as highly impactful invasives in the

ecoregion: Buddleja davidii, Cortaderia selloana, Dittrichia viscosa, Paspalum dilatatum and Sporobolus indicus (Lázaro-Lobo et al., 2024). Still, vacant lots had more species than forests even considering only native plants. Contrary to plants, fungi had higher richness in our forest plots than in the roadsides or industrial vacant lots, with forests being especially rich in Basidiomycota, as found in other studies (Hui et al., 2017; Marczylo et al., 2021). This agrees with previous reports of fungal communities decreasing with urbanization (Abrego et al., 2020). One potential explanation for this pattern is the tendency of urban soils to be alkaline, favoring bacteria over fungi (Delgado-Baquerizo et al., 2021; Forman, 2014). Indeed, in our study, organic matter content (highly and inversely correlated with pH) was the soil variable that had an effect on the composition of fungal and bacterial communities; and bacterial richness was higher in urban habitats versus forests (i.e., the opposite to fungi). Relatively low novelty in parks lawns and residential vacant lots We used a compositional novelty index based on multidimensional ordination, which is straightforward to calculate and only requires data on species co-occurrence for urban and reference habitats. As such, this index can help to characterize and compare novelty across urban habitats, which is a major research goal for novel ecosystem research (Teixeira et al., 2021; Teixeira and Fernandes, 2020). Contrary to our expectations, managed park lawns showed a relatively low novelty when compared to agricultural meadows. This indicates that park lawns have a high potential for low-cost urban rewilding policies, such as low-frequency and late mowing, which could convert lawns into urban grasslands rich in native biota and resembling traditional hay meadows. Urban grasslands are able to hold a high portion of the plant diversity found in their rural counterparts (Swacha et al., 2024), especially when

seminatural habitat remnants are available in the neighboring landscape to serve as a source of species (Gros et al., 2023). We also found that residential vacant lots have a relatively low level of novelty, including a high richness of fungi and native plants. These temporarily unmanaged sites are therefore a highly valuable asset to integrate biodiversity in cities (Anderson and Minor, 2017; Kowarik, 2018). Additionally, identifying synanthropic and native species able to thrive in these vacant lots can help to find candidate species for urban landscaping that require low levels of watering, fertilization, weeding or mowing (Bretzel et al., 2024; Fekete et al., 2024). Such native and hardy seed mixes are important tools for urban greening, given the increasing awareness of the importance of using locally sourced seed for restoration and rewilding (Hancock et al., 2023).

Conclusions

Our results highlight the effect of habitat type as a major driver of urban community composition and compositional novelty, supporting the findings of previous studies (Gill et al., 2020; Hui et al., 2017; Marczylo et al., 2021). The high impact of habitat types on shaping urban communities and their compositional novelty may be related to the fact that anthropogenic vegetation is highly responsive to habitat-specific disturbance regimes (Fernández-Pascual et al., 2025), while soil microbial communities are in turn highly conditioned by plant inputs (Hui et al., 2017). These results suggest that city biodiversity can be enhanced by an integrative approach to the urban landscape that favors habitat heterogeneity by passive rewilding of managed park lawns, non-intervention on residential vacant lots, direct restoration of industrial vacant lots, and conservation of natural and agricultural habitat remnants as sources of native species (Dylewski et al., 2023; Fekete et

- 1 al., 2024; Labadessa and Ancillotto, 2023). In this way, the potential of urban habitats can be
- 2 mobilized so they continue to support ecosystem functioning in anthropogenic city
- 3 landscapes (Fan et al., 2023; Torija et al., 2025; Weiskopf et al., 2024).

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- **Supplementary**
- **Appendix S1** Extended methodology for the soil DNA metabarcoding analysis.
- 14 Appendix S2 Principal Component Analysis and correlation analysis of the soil
- 15 physicochemical properties.

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