

Lachancea waltii

RNA-seq analysis

Eric Toro Delgado, Iria Pose Lagoa, Sara Vega Abellaneda

CONTENTS



- 1 Introduction
- 2 Analysis strategy
- 3 Pre-processing
- 4 Transcripts analysis
- 5 Novel transcripts characterization
- 6 Conclusions

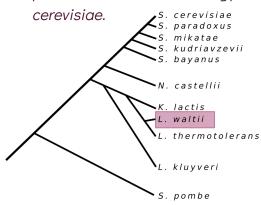
1. INTRODUCTION



Species: Lachancea waltii



- Protoploid budding yeast species.
- Existing annotation based on predicted ORFs and homology with S.

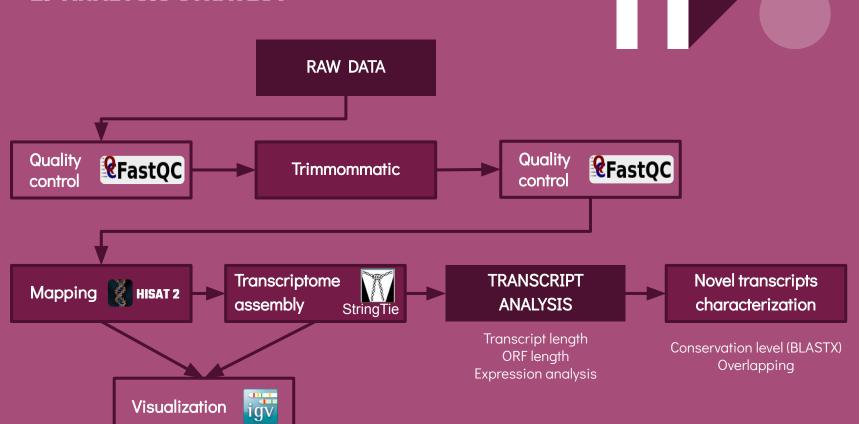


OBJECTIVES

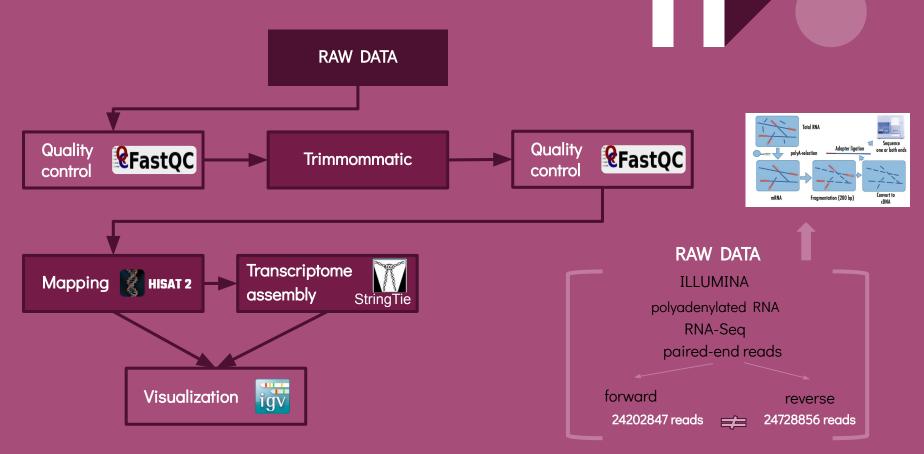
 Understand the difference between novel and annotated transcripts.

 Understand the patterns in the features related to the origin of novel transcripts, following the framework of *de novo* gene formation proposed by *Carvunis et al. (2012).*

2. ANALYSIS STRATEGY



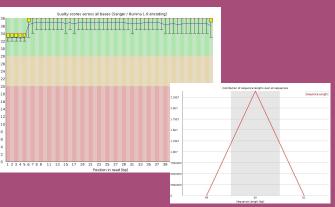
3. PRE-PROCESSING

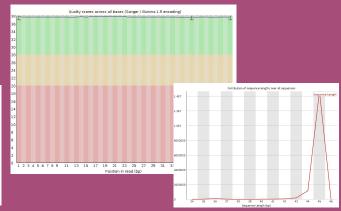




Quality control

BEFORE TRIMMING





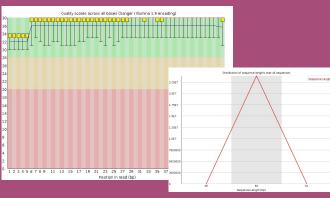
Quality scores across all bases (Sanger / Illumina 1.9 encoding)

THIS INCLUDED THE TAXABLE PARTY OF THE PARTY

1 2 3 4 5 6 7 8 9 11 13 15 17 19 21 23 25 27 29 31



READ 1

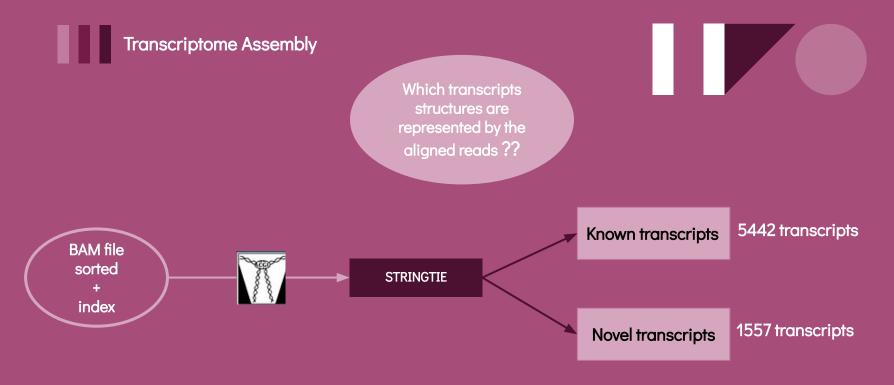




forward 16430915 reads

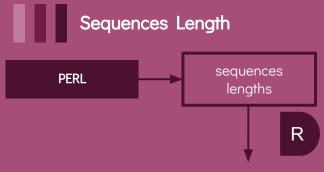


Mapping reads assigned to a FASTA files indexes HISAT 2 specific location in (cleaned reads) the genome 96,44% overall alignment rate SAM file SAMTOOLS 33707563 reads mapped BAM file sorted index





4. TRANSCRIPTS ANALYSIS

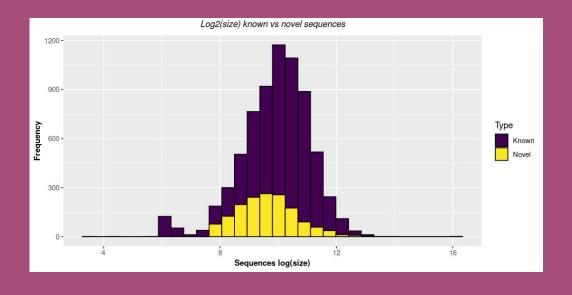


	KNOWN	NOVEL	
Length Range	12 - 14769	201 - 76379	
Mean	1362	1064	
Median	1125	767	

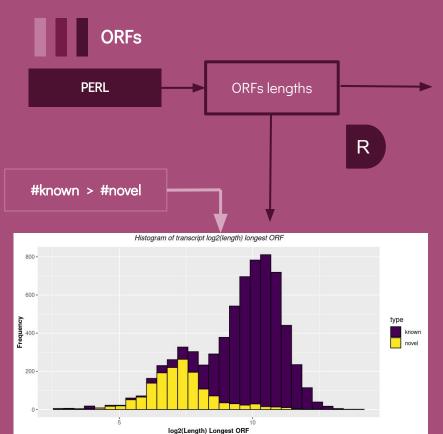
Mann-Whitney-Wilcoxon test

W=5294271 p-value<2.2e-16

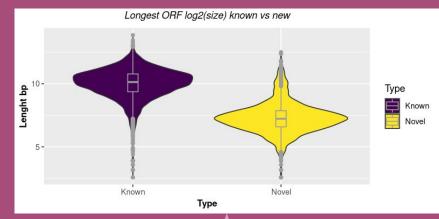




4. TRANSCRIPTS ANALYSIS





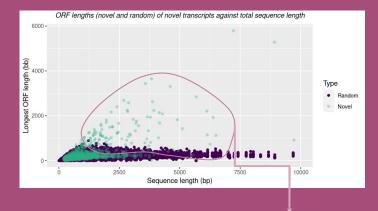


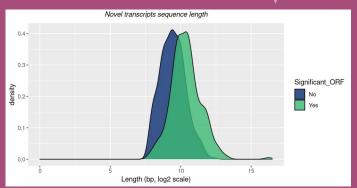
mean: 1318 > 251.8 bp

Mann-Whitney-Wilcoxon test
W=7414911 p-value<2.2e-16

PERL sequences lengths + random sequence length



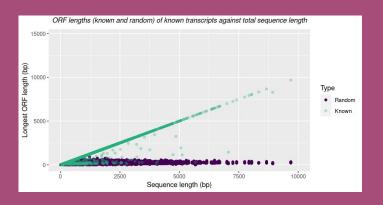




- For random sequences, the length of the ORFs depend of the length of sequences
- 306 out of 1541 (19.86%) novel ORFs of length longer than random ORFs are putative coding sequences

R

• Sequences annotations come from computational predictions based on ORFs, so that is the reason of the "perfect" correlated line on known transcripts.

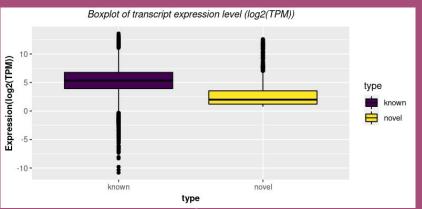


4. TRANSCRIPTS ANALYSIS









mean known > mean novel

Mann-Whitney-Wilcoxon test

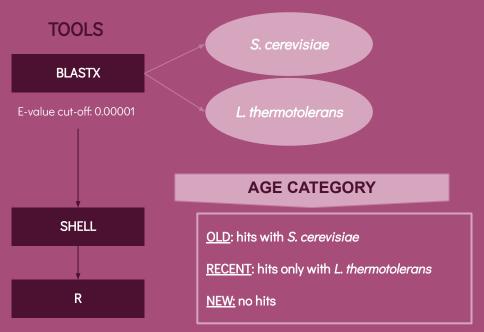
W = 6372692 p-value < 2.2e-16

Mann-Whitney-Wilcoxon test

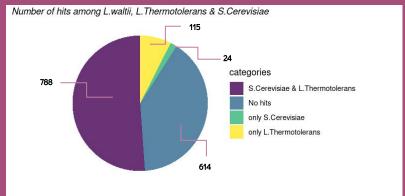
W = 6372692 p-value<2.2e-16

5. Novel transcripts characterization

Homology and conservation analysis



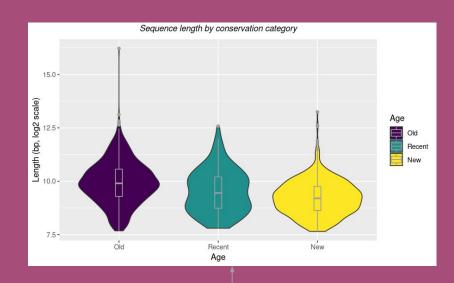


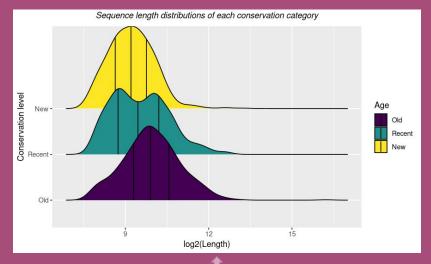


specie	hits	no hits	TOTAL
S.Cerevisiae	812	729	1541
L. Thermotolerans	903	638	1541









Kruskal-Wallis test

X²=191.53 p-value<2.2e-16

Mean ± SD

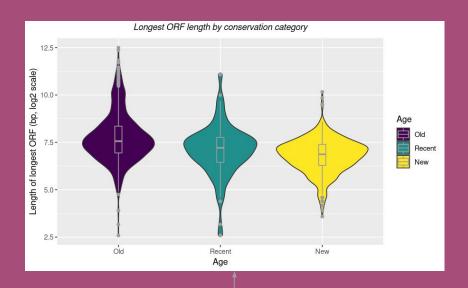
Old: 1340.1 ± 2816.18

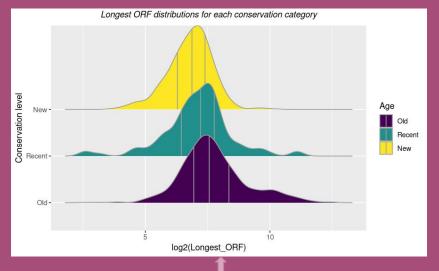
Recent 971.5 ± 865.08

New: 717.4 ± 641.33









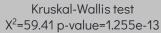
Kruskal-Wallis test

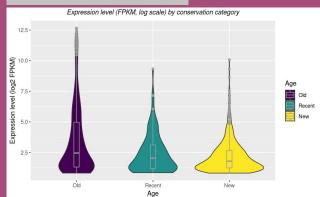
X²=208.37 p-value<2.2e-16

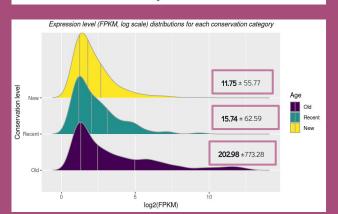
Mean ± SD

Old: 271.6 ± 522.07 Recent: 248.7 ± 309.17 New: 237.1 ± 95.19

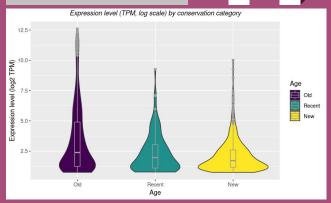
Expression

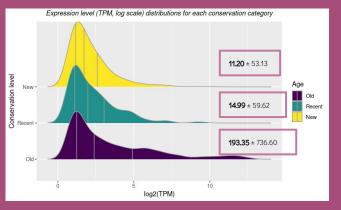






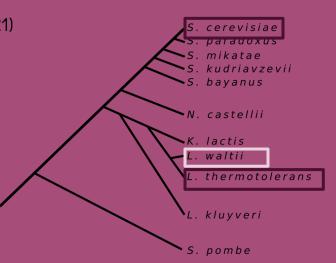
Kruskal-Wallis test X²=59.41 p-value=1.255e-13





Limitations

- Compared with only 2 species
- Compared with protein DB only \rightarrow some transcripts may have homologues that have lost function
- Did not check for paralogs nor for synteny (Blevins et al. 2021)
- No Ribo-Seq data → cannot confirm translational activity
- No expression threshold of correct transcript assembly
- Quality of reference genomes and annotations

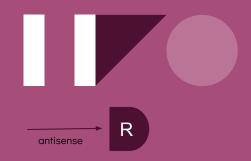


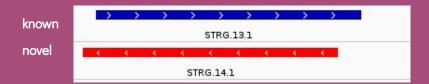
5. Novel transcripts characterization

Opposite strand overlap with known genes

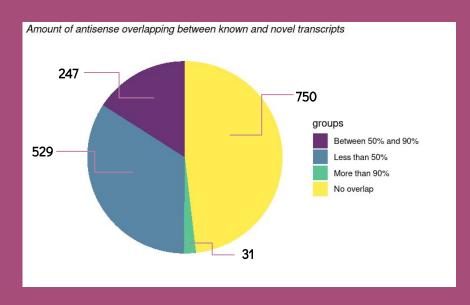
TOOLS SHELL BEDTOOLS

any overlap
≥ 50% of reciprocal overlap
≥ 90% of reciprocal overlap







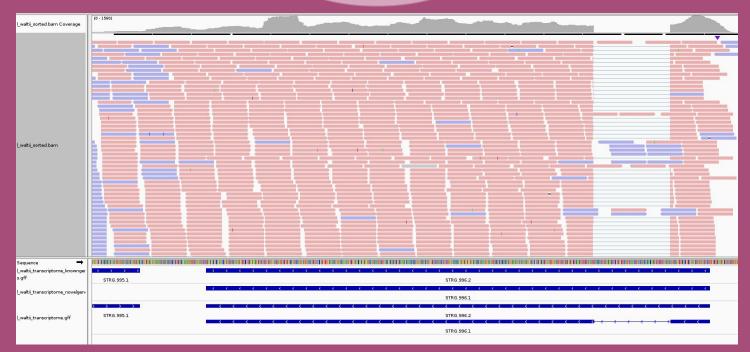


5. Novel transcripts characterization

Same strand overlap

Sequences with more or equal to 90% of reciprocal overlap were nearly identical, why ??





6. Conclusions



→ Identification of 1556 novel transcripts (1541 without rep): 1557/6999 = 22.25% of the total transcriptome catalog for *L. waltii*.

novel vs known

- Novel transcripts are smaller, have lower expression levels and have shorter ORFs than annotated.
- Novel transcripts may not have been previously annotated due to their short ORFs.
- For novel sequences of the same length, ORFs that are longer than ORFs obtained randomly, could be putative coding or truly functional.

novel transcripts features

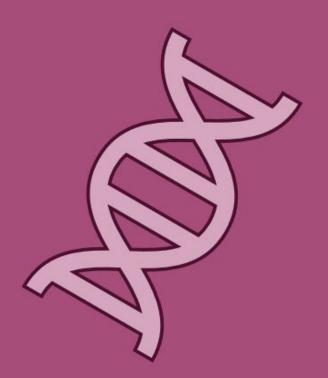
- Despite limitations of our study, 614 non annotated putative *de novo* transcripts.
- Novel transcripts follow a pattern of increasing total length, ORF length and expression with older dates of origin.
- Many novel transcripts overlapping with already annotated ones in an antisense configuration, but with small overlap.

References



- 1. Ares M., J., Grate, L., & Pauling, M. H. (1999). A handful of intron-containing genes produces the lion's share of yeast mRNA [2]. In RNA (Vol. 5, Issue 9, pp. 1138–1139). Cambridge University Press. https://doi.org/10.1017/S1355838299991379
- 2. Blevins, W. R., Carey, L. B., & Albà, M. M. (2019). Transcriptomics data of 11 species of yeast identically grown in rich media and oxidative stress conditions. BMC Research Notes, 12(1), 1–4. https://doi.org/10.1186/s13104-019-4286-0
- 3. Blevins, W. R., Ruiz-Orera, J., Messeguer, X., Blasco-Moreno, B., Villanueva-Cañas, J. L., Espinar, L., Díez, J., Carey, L. B., & Albà, M. M. (2021). Uncovering de novo gene birth in yeast using deep transcriptomics. Nature Communications, 12(1), 1–13. https://doi.org/10.1038/s41467-021-20911-3
- 4. Cai, J., Zhao, R., Jiang, H., & Wang, W. (2008). De Novo Origination of a New Protein-Coding Gene in Saccharomyces cerevisiae. Genetics, 179(1), 487–496. https://doi.org/10.1534/GENETICS.107.084491
- 5. Carvunis, A.-R., Rolland, T., Wapinski, I., Calderwood, M. A., Yildirim, M. A., Simonis, N., Charloteaux, B., Hidalgo, C. A., Barbette, J., Santhanam, B., Brar, G. A., Weissman, J. S., Regev, A., Thierry-Mieg, N., Cusick, M. E., & Vidal, M. (2012). Proto-genes and de novo gene birth. Nature, 487(7407), 370–374. https://doi.org/10.1038/nature11184
- 6. Di Rienzi, S. C., Lindstrom, K. C., Lancaster, R., Rolczynski, L., Raghuraman, M. K., & Brewer, B. J. (2011). Genetic, genomic, and molecular tools for studying the protoploid yeast, L. waltii. Yeast, 28(2), 137–151. https://doi.org/10.1002/YEA.1826
- 7. Kellis, M., Birren, B. W., & Lander, E. S. (2004). Proof and evolutionary analysis of ancient genome duplication in the yeast Saccharomyces cerevisiae. Nature 2007 428:6983, 428(6983), 617–624. https://doi.org/10.1038/nature02424
- 8. Kempken, F. (2013). Alternative splicing in ascomycetes. In Applied Microbiology and Biotechnology (Vol. 97, Issue 10, pp. 4235–4241). Springer. https://doi.org/10.1007/s00253-013-4841-x Knight, R. D., Freeland, S. J., & Landweber, L. F. (2001). A simple model based on mutation and selection explains trends in codon and amino-acid usage and GC composition within and across genomes. Genome Biology, 2(4), 1–13. https://doi.org/10.1186/GB-2001-2-4-RESEARCH0010/TABLES/5
- 9. Ma, C., & Kingsford, C. (2019). Detecting, Categorizing, and Correcting Coverage Anomalies of RNA-Seq Quantification. Cell Systems, 9(6), 589-599.e7. https://doi.org/10.1016/J.CELS.2019.10.005
- 10. Prat, Y., Fromer, M., Linial, N., & Linial, M. (2009). Codon usage is associated with the evolutionary age of genes in metazoan genomes. BMC Evolutionary Biology, 9(1), 285. https://doi.org/10.1186/1471-2148-9-285
- 11. Vakirlis, N., Sarilar, V., Drillon, G., Fleiss, A., Agier, N., Meyniel, J. P., Blanpain, L., Carbone, A., Devillers, H., Dubois, K., Gillet-Markowska, A., Graziani, S., Huu-Vang, N., Poirel, M., Reisser, C., Schott, J., Schacherer, J., Lafontaine, I., Llorente, B., ... Fischer, G. (2016). Reconstruction of ancestral chromosome architecture and gene repertoire reveals principles of genome evolution in a model yeast genus.

 Genome Research, 26(7), 918–932. https://doi.org/10.1101/GR.204420.116



Lachancea waltii

RNA-seq analysis

Thank you very much for your attention