

Biophysical Journal

Supplemental Information

GRPY - program for calculation of hydrodynamic properties of biomacromolecules using generalized Rotne-Prager-Yamakawa approximation

Pawel Jan Zuk, Bogdan Cichocki, Piotr Szymczak

Biophysical Journal 1

GRPY User Manual

1. Introduction and Description

The *GRPY* program is a *FORTRAN* code to calculate the hydrodynamic properties of rigid macromolecules using Generalized Rotne-Prager-Yamakawa approximation of hydrodynamic interactions (1, 2). The shape of a rigid macromolecule has to be represented with a set of beads having different radii that can overlap. The *GRPY* calculates among others, translational and rotational diffusion coefficients, rotational relaxation times, sedimentation coefficient, the position of mobility center and intrinsic viscosity of a macromolecule. For the user convenience the *GRPY* accepts input in GRPY original format, *hydro++* program format (3, 4) and *US-SOMO* generated *bead_model* format (5–7).

2. Availability, permissions and citations

The GRPY open source code is freely available for download and usage under GNU General Public License version 3 from

- http://www.fuw.edu.pl/ piotrek/sofware website;
- GitHub https://github.com/pjzuk/GRPY repository with Zenodo listed DOI ...

When using the program please cite

- this contribution (8);
- generalized Rotne-Prager-Yamakawa method (1);
- generalized Rotne-Prager-Yamakawa method for different sized beads (9);
- intrinsic viscosity of macromolecules with generalized Rotne-Prager-Yamakawa approximation (2).

3. Package content

The GRPY package contains

- user guide;
- source code for the single core GRPY program compilation including LAPACK procedures (10);
- source code for the parallel *plasmaGRPY* program compilation;
- compiled single core GRPY program for Linux and Windows;
- compiled parallel *plasmaGRPY* program for Linux;
- input examples in GRPY, hydro++ and bead_model formats;
- PLASMA library (11) (needed to compile *plasmaGRPY*) installer.

Additional content

• scripts to generate hydrated atom bead model from pdb file using the MSMS program (12).

4. Program compilation and execution

4.1 Precompiled executables

There are four precompiled executables ready to use after download:

1. single core for Linux users:

```
GRPY/bin/GRPYLinux/GRPY.exe
```

2. multi core for Linux users:

```
GRPY/bin/GRPYLinux/plasmaGRPY.exe
```

3. single core 32bit version for Windows users:

```
GRPY/bin/GRPYWindows_x86/GRPY_x86.exe
```

4. single core 64bit version for Windows users:

```
GRPY/bin/GRPYWindows_x64/GRPY_x64.exe
```

4.2 Compilation from source code

A source code for single core version

```
GRPY/src/GRPY/GRPY.f
```

that needs additional BLAS/LAPACK routines, that are attached in the separate files

```
GRPY/src/lapackGRPY/lapack_dgsev.f
GRPY/src/lapackGRPY/lapack_dsyev.f
GRPY/src/lapackGRPY/lapblas_matinv.f.
```

For the Linux users a suitable Makefile

```
Makefile
```

is provided to compile the *GRPY* program from the source using the *gfortran* compiler. To compile the the program inside the GRPY folder run

```
make -f Makefile linux
```

To clean compilation run

```
make -f Makefile clean
```

Also a source code for the multi core version is provided

```
GRPY/src/plasmaGRPY/plasmaGRPY.f
```

In order to compile plasmaGRPY program the PLASMA library has to be installed. The installer is provided in

```
GRPY/ThirdParty/plasma-installer_2.8.0
```

folder. Please proceed with the instructions provided by the package and install the libraries in the default location. After successful installation of *PLASMA* library in order to compile *plasmaGRPY* run

```
make -f Makefile plasmaLinux
```

The users of other operating systems should modify these scripts for their needs.

5. Program input

The input file for *GRPY* program can be prepared in three ways

- GRPY original format;
- *hydro*++ program format by de La Torre et al. (3, 4);
- *US-SOMO* generated *bead_model* format (5–7).

For all the above example input files are attached in

```
GRPY/examples
```

folder.

5.1 GRPY input file

For each model of a rigid molecule a separate input file needs to be prepared. It consists of two blocks. The first block contains the general parameters of the model grouped in two columns. The first column contains the values of the parameters whereas the second column contains a short description. Importantly, the first column should have 32 characters. The first block contains

- 1. Model name a string with model name,
- 2. Temperature solution temperature T in Celsius,

- 3. Solvent viscosity solvent viscosity η_0 in Poise,
- 4. Molecular weight molecular weight M_w in Daltons,
- 5. Specific volume specific volume of macromolecule \bar{v} in $\left[\frac{\mathrm{cm}^3}{\mathrm{g}}\right]$,
- 6. Unit length unit length in which bead coordinates and radii are given in [cm],
- 7. Number of beads number of beads N that program will read from the following list (it cannot be larger than the number of listed beads)

The second block comprises four columns of floating numbers separated by spaces. In ith row the first three columns contain the spatial coordinates (x_i, y_i, z_i) of the center of a bead, whereas the radius of the bead (a_i) is given in the fourth one. All four entries are expressed in length unit specified in the first block. As an example we present input file for a rigid dumbbell (N=2) of two touching spheres of radii $a_1=20$ $[\mathring{A}]$ and $a_2=30$ $[\mathring{A}]$ with the smaller one at the origin of coordinate system and the larger one placed at $x_2=0$ $[\mathring{A}]$, $y_2=50$ $[\mathring{A}]$, $z_2=0$. In the example given below there are three lines specifying bead coordinates, however the third line is not be read by the program because the number of particles has been set to N=2. On the other hand if N is larger than the number of provided lines the program will exit with an error. The particles are suspended in water of temperature 20° [C] and viscosity of 0.01 [P] (Poise). The molecular weight of the particle is 1000 [Da] and the specific volume is 0.9 $\left[\frac{\mathrm{cm}^3}{\mathrm{g}}\right]$. We write the particle positions and radii in the units of $\mathring{A}=10^{-8}$ $[\mathrm{cm}]$.

rigid dumbbell				Model name
20.				Temperature
0.01				Solvent viscosity
1.E+3				Molecular weight
0.9				Specific volume
1.0				Solution density
1.E-8				Unit length [cm]
2				Number of beads
0.0	0.0	0.0	20.0	
0.0	50.0	0.0	30.0	
0.0	0.0	50.0	30.0	

5.2 hydro++ input file

For the details of the structure of the hydro++ input file please refer to the hydro++ user manual (3, 4).

5.3 US-SOMO input file

For the details of the structure of the *bead_model* input file from *US-SOMO* program please refer to the user manual (5–7).

6. Program execution

For both Linux and Windows users the program needs to be executed from the command line. Here we give an example of the the single core *GRPY.exe* program execution

In case of GRPY input format program is executed as

```
<path to GRPY.exe> <path to GRPY inupt file>
```

In case of hydro++ input format the additional flag -d has to be added

```
<path to GRPY.exe> -d <path to hydro++ inupt file>
```

In case of US-SOMO generated bead_model format the additional flag -u has to be added

```
<path to GRPY.exe> -u <path to .bead_model inupt file>
```

In case of multi core *plasmaGRPY.exe* program additionally the number of cores for calculation can be specified with system variable PROC_NUM. If not specified the program will use single core. The usage of different input files is the same as in single core version. For example to run calculations using 5 cores on the *US-SOMO* file one has to type

```
+
```

```
export PROC_NUM=5
<path to plasmaGRPY.exe> -u <path to .bead_model inupt file>
```

The number of cores used in calculation will not exceed the number of available physical cores. For details please refer to the PLASMA library user guide.

7. Program output

All the information presented in the program output are the same for the GRPY, hydro++ and US-SOMO input however the outputs for GRPY and US-SOMO input is displayed to the standard output and for the hydro++ input is written to the file with filename <file name for output file>-GRPY.dat, where <file name for output file> is specified in the hydro++ input file in analogy to original hydro++ program.

Here we present the example of program output from the calculations for the non symmetric dumbbell, for which the input file is presented above and included in the *examples* folder

```
Radius of gyration:
                                                                        3.020E-07 [cm]
                                    Rotational diffusion coefficient:
                                                                        3.514E+06 [s<sup>-1</sup>]
                        Rotational relaxation time for rank 1 tensor:
                                              -> Relaxation time (1):
                                                                        1.284E-07 [s]
                                              -> Relaxation time (2):
                                                                        1.284E-07 [s]
                                              -> Relaxation time (3):
                                                                        1.816E-07 [s]
                        Rotational relaxation time for rank 2 tensor:
                                              -> Relaxation time (1):
                                                                        3.899E-08 [s]
                                              -> Relaxation time (2):
                                                                        6.053E-08 [s]
                                              -> Relaxation time (3):
                                                                        5.319E-08 [s]
                                              -> Relaxation time (4):
                                                                        5.319E-08 [s]
                                              -> Relaxation time (5):
                                                                        3.899E-08 [s]
                                    Harmonic mean (correlation) time:
                                                                        4.743E-08 [s]
       Sedimentation coefficient (Mw Dlt (1/(vbar*rho)-1)/(nA kB T)):
                                                                        2.769E-02 [Svedberg]
                                    Non Brownian intrinsic viscosity:
                                                                        2.584E+02 [cm^3/g]
                                        Brownian intrinsic viscosity:
                                                                        2.837E+02 [cm^3/q]
            calculated using the origin
              of the coordiante system:
 0.000E+00 0.000E+00 0.000E+00
                 as the reference point
       in the standard reference frame:
       e1: 1.000E+00 0.000E+00 0.000E+00
       e2:
            0.000E+00
                      1.000E+00 0.000E+00
       e3: 0.000E+00 0.000E+00 1.000E+00
                                 Translational diffusion coefficient: 8.251E-07 [cm<sup>2</sup>/s]
                 6x6 diffusion matrix:
                                              Dtt Dtr
                                              Drt Drr
                      0.000E+00
                                    -0.000E+00 -0.000E+00 9.479E-01
 9.137E-07 -0.000E+00
                                    -0.000E+00 -0.000E+00 -0.000E+00
-0.000E+00 6.480E-07
                      0.000E+00
 0.000E+00 0.000E+00
                      9.137E-07
                                    -9.479E-01 -0.000E+00 -0.000E+00
-0.000E+00 -0.000E+00 -9.479E-01
                                     2.753E+06 -0.000E+00 -0.000E+00
-0.000E+00 -0.000E+00 -0.000E+00
                                    -0.000E+00 5.035E+06 -0.000E+00
                                    -0.000E+00 -0.000E+00 2.753E+06
 9.479E-01 -0.000E+00 -0.000E+00
```

```
calculated using the mobility center:
          y [cm]
                    z [cm]
0.000E+00 3.443E-07 0.000E+00
               as the reference point
              in the reference frame:
         1.000E+00 0.000E+00 0.000E+00
     e1:
     e2: 0.000E+00 0.000E+00 1.000E+00
     e3: 0.000E+00 1.000E+00 0.000E+00
                               Translational diffusion coefficient: 6.075E-07 [cm<sup>2</sup>/s]
               6x6 diffusion matrix:
                                            Dtt
                                               Dtr
                                            Drt Drr
5.873E-07 0.000E+00 0.000E+00
                                   0.000E+00 1.142E-16 0.000E+00
0.000E+00 5.873E-07 0.000E+00
                                  -1.142E-16 0.000E+00 0.000E+00
0.000E+00 0.000E+00 6.480E-07
                                  0.000E+00 0.000E+00 0.000E+00
0.000E+00 -1.142E-16 0.000E+00
                                   2.753E+06 0.000E+00 0.000E+00
1.142E-16 0.000E+00 0.000E+00
                                   0.000E+00 2.753E+06 0.000E+00
0.000E+00 0.000E+00 0.000E+00
                                   0.000E+00 0.000E+00 5.035E+06
                                   Radius of the sphere with equal:
                               Translational diffusion coefficient:
                                                                     3.534E-07 [cm]
                                  Rotational diffusion coefficient:
                                                                    3.579E-07 [cm]
                      Rotational relaxation time for rank 1 tensor:
                                            -> Relaxation time (1): 4.988E-07 [cm]
                                            -> Relaxation time (2): 4.988E-07 [cm]
                                            -> Relaxation time (3): 5.598E-07 [cm]
                      Rotational relaxation time for rank 2 tensor:
                                            -> Relaxation time (1): 3.352E-07 [cm]
                                            -> Relaxation time (2): 3.882E-07 [cm]
                                            -> Relaxation time (3): 3.718E-07 [cm]
                                            -> Relaxation time (4): 3.718E-07 [cm]
                                            -> Relaxation time (5): 3.352E-07 [cm]
                                              Mean relaxation time: 3.579E-07 [cm]
                                Intrinsic viscosity (non Brownian): 3.447E-07 [cm]
                                    Intrinsic viscosity (Brownian): 3.557E-07 [cm]
```

The output file provides following information

- 1. Radius of gyration r_q calculated according to Ref. (13)
- 2. Rotational diffusion coefficient D^r where $D^r = \frac{1}{3} \text{Tr} \mathbf{D}^r$ with \mathbf{D}^r defined as in Eq. (8) of Ref. (8);
- 3. Rotational relaxation times for rank 1 tensor τ_i calculated in Eq. (18) of Ref. (8);
- 4. Rotational relaxation times for rank 2 tensor f_i^{-1} : calculated as in Eq. (C.1) of Ref. (8);
- 5. Harmonic mean (correlation) time: harmonic mean calculated with 5 rotational relaxations times for rank 2 tensor f_i^{-1} ;
- 6. Sedimentation coefficient S calculated as in Eq. (22) of Ref. (8);
- 7. Non Brownian intrinsic viscosity $[\eta]_{\infty}$ calculated as in Eq. (26) of Ref. (8);
- 8. Brownian intrinsic viscosity $[\eta]_0$ calculated as in Eq. (27) of Ref. (8);
- 9. Translational diffusion coefficient $D_s^t(\mathbf{R}_0)$ calculated as in Eq. (19) of Ref. (8) in the original coordinate system (in which the positions of the beads were given in the input file)
- 10. 6×6 diffusion matrix divided into four submatrices \mathbf{D}^{tt} , \mathbf{D}^{rt} , \mathbf{D}^{rt} , \mathbf{D}^{rt} calculated in the in the original coordinate system

- 11. Position of the mobility center of the particle \mathbf{R}_{mc} see Ref. (8);
- 12. The reference frame $\hat{\bf e}_1, \hat{\bf e}_2, \hat{\bf e}_3$ in which the rotational diffusion matrix D^{rr} is diagonal (given by the eigenvectors of D^{rr})
- 13. Long time diffusion coefficient $D_l^t = D_s^t(\mathbf{R}_{mc})$ equal to the short time diffusion coefficient calculated with mobility center as the reference point $\mathbf{R}_0 = \mathbf{R}_{mc}$;
- 14. 6×6 diffusion matrix divided into four submatrices \mathbf{D}^{tt} , \mathbf{D}^{rt} , \mathbf{D}^{rt} , \mathbf{D}^{rt} calculated in the in the reference frame $\hat{\mathbf{e}}_1$, $\hat{\mathbf{e}}_2$, $\hat{\mathbf{e}}_3$ with mobility center as the reference point $\mathbf{R}_0 = \mathbf{R}_{mc}$;
- 15. List of radii of equivalent spheres that have the same hydrodynamic properties as the macromolecule

8. Hydrated atom bead model

In addition to the *GRPY* program we provide scripts that can be used to construct the hydrated atomic model described in the main text. The scripts are present in the

```
hydrAtom
```

folder. This is not a stand alone code and surface detection has to be done with MSMS program (12). The script

```
hydrAtom/pdbToHydrAtom
```

can be used to convert the pdb file to the .xyzr file containing positions and radii of surface atoms with added hydration layer. Inside this script absolute paths to the hydrAtom folder and folder containing MSMS program have to be specified. The script

```
hydrAtom/pdb_to_xyzrn_mod
```

is a slightly modified version of *pdb_to_xyzrn* script added to *MSMS* program, which allows to execute it correctly from another directory. Python (version 2.7) script

```
hydrAtom/msmsVertNameTohydrAtom.py
```

chooses the surface atoms from .vert file resulting from execution of MSMS program and adds the hydration radius. Additionally if a GRPY header file (first block of of native GRPY input file is present) the complete input file for GRPY is generated. An example for 1znf.pdb file attached in

```
hydrAtom/examples
```

folder. After specifying appropriate paths one can execute (1znf.pdb and 1znf.header files are provided)

```
<path to pdbToHydrAtom script> <path to 1znf.pdb file>
```

For example inside

hydrAtom

folder execute

```
./pdbToHydrAtom ./example/1znf.pdb
```

This will result in output files

- example/1znf_noHET.pdb pdb file without HETATM lines
- example/1znf_noHET.xyzrn x,y,z,radius,name file from pdb_to_xyzrn_mod script
- example/1znf_noHET.xyzrn_msms.face and *lznf_noHET.xyzrn_msms.vert* files from MSMS program
- example/1znf_hydAtom.xyzr-x,y,z,radius file with hydrated radii of surface atoms
- example/1znf_hydAtom_GRPY GRPY program input file

also the file example/1znf_hydAtom_GRPY_out resulting from GRPY program is provided.

+

REFERENCES AND FOOTNOTES

REFERENCES and FOOTNOTES

- 1. Wajnryb, E., K. A. Mizerski, P. J. Zuk, and P. Szymczak, 2013. Generalization of the Rotne–Prager–Yamakawa mobility and shear disturbance tensors. *J. Fluid Mech.* 731:R3.
- 2. Zuk, P. J., B. Cichocki, and P. Szymczak, 2017. Intrinsic viscosity of macromolecules within the generalized Rotne–Prager–Yamakawa approximation. *J. Fluid. Mech.* 822.
- 3. de la Torre, J. G., S. Navarro, M. L. Martinez, F. Diaz, and J. L. Cascales, 1994. HYDRO: a computer program for the prediction of hydrodynamic properties of macromolecules. *Biophys. J.* 67:530–531.
- 4. de la Torre, J. G., G. del Rio Echenique, and A. Ortega, 2007. Improved calculation of rotational diffusion and intrinsic viscosity of bead models for macromolecules and nanoparticles. *J. Phys. Chem. B* 111:955–961.
- 5. Rai, N., M. Nöllmann, B. Spotorno, G. Tassara, O. Byron, and M. Rocco, 2005. SOMO (SOlution MOdeler): differences between X-ray-and NMR-derived bead models suggest a role for side chain flexibility in protein hydrodynamics. *Structure* 13:723–734.
- 6. Brookes, E., B. Demeler, C. Rosano, and M. Rocco, 2010. The implementation of SOMO (SOlution MOdeller) in the UltraScan analytical ultracentrifugation data analysis suite: enhanced capabilities allow the reliable hydrodynamic modeling of virtually any kind of biomacromolecule. *European Biophysics Journal* 39:423–435.
- 7. Brookes, E., B. Demeler, and M. Rocco, 2010. Developments in the US-SOMO Bead Modeling Suite: New Features in the Direct Residue-to-Bead Method, Improved Grid Routines, and Influence of Accessible Surface Area Screening. *Macromolecular bioscience* 10:746–753.
- 8. Zuk, P. J., B. Cichocki, and P. Szymczak, 2017. A fast and effective method for calculation of hydrodynamic properties of rigid molecules of arbitrary shape. *Biophys. J. (submitted)*.
- 9. Zuk, P. J., E. Wajnryb, K. A. Mizerski, and P. Szymczak, 2014. Rotne-Prager-Yamakawa approximation for different-sized particles in application to macromolecular bead models. *J. Fluid Mech.* 741:R5.
- 10. Anderson, E., Z. Bai, C. Bischof, S. Blackford, J. Demmel, J. Dongarra, J. Du Croz, A. Greenbaum, S. Hammarling, A. McKenney, and D. Sorensen, 1999. LAPACK Users' Guide. Society for Industrial and Applied Mathematics, Philadelphia, PA, third edition.
- 11. Agullo, E., J. Dongarra, B. Hadri, J. Kurzak, J. Langou, J. Langou, H. Ltaief, P. Luszczek, and A. YarKhan, 2009. Plasma users guide. Technical report, Technical report, ICL, UTK.
- 12. Sanner, M. F., A. J. Olson, and J.-C. Spehner, 1996. Reduced surface: an efficient way to compute molecular surfaces. *Biopolymers* 38:305–320.
- 13. Solvez, J. A., A. Iniesta, and J. G. la Torre, 1988. Radius of gyration of multisubunit macromolecules: application to myosin heads, myosin rod and whole myosin. *International Journal of Biological Macromolecules* 10:39–43.