

Lihua Wu¹, Justin S. Williams², Ning Wang¹, Wasi A. Khatri², Daniele San Román², Teh-hui Kao^{1,2,*}

¹Intercollege Graduate Degree Program in Plant Biology, The Pennsylvania State University, University Park, Pennsylvania 16802 USA

²Department of Biochemistry and Molecular Biology, The Pennsylvania State University, University Park, Pennsylvania 16802 USA

Background Information

Self/non-self recognition is regulated by the polymorphic S-locus; matching of the pollen S-haplotype with one of the two pistil S-haplotypes results in inhibition of pollen tube growth. The S-locus houses S-RNase for pistil specificity, and, for both S₂- and S₃-haplotypes, 17 S-locus F-box (SLF) genes for pollen specificity. All SLFs are assembled into similar SCF complexes, containing Rbx1, pollen-specific Cullin1 and Skp1-like protein. According to the collaborative non-self recognition model, for a given S-haplotype, each SCF complex interacts with a subset of non-self S-RNases to mediate their ubiquitination and degradation by the 26S proteasome. Our lab has used a transgenic assay (Fig. 1) to determine interaction relationships of SLF proteins and S-RNases. Among those determined, S₂-SLF1 and S₃-SLF1 (an allelic pair of SLF1, differing in 44 amino acids) show differential interactions with several S-RNases, i.e., S₂-SLF1, but not S₃-SLF1, interacts with S₃⁻, S₇⁻ and S₁₃⁻RNases (Table 1).

	S ₂ ⁻ RNase	S ₇ ⁻ RNase	S ₁₃ ⁻ RNase	S _{6a} ⁻ RNase	S ₇ ⁻ RNase	S ₁₁ ⁻ RNase	S ₁₂ ⁻ RNase	S ₁₃ ⁻ RNase	S ₁₆ ⁻ RNase	S ₂ ⁻ RNase	S ₁₃ ⁻ RNase
S ₂ -SLF1	—	+	—	—	+	—	+	+	—	—	—
S ₃ -SLF1	—	—	—	—	—	—	+	—	—	—	—

Table 1. S₂-SLF1 and S₃-SLF1 show differential interactions with three S-RNases.

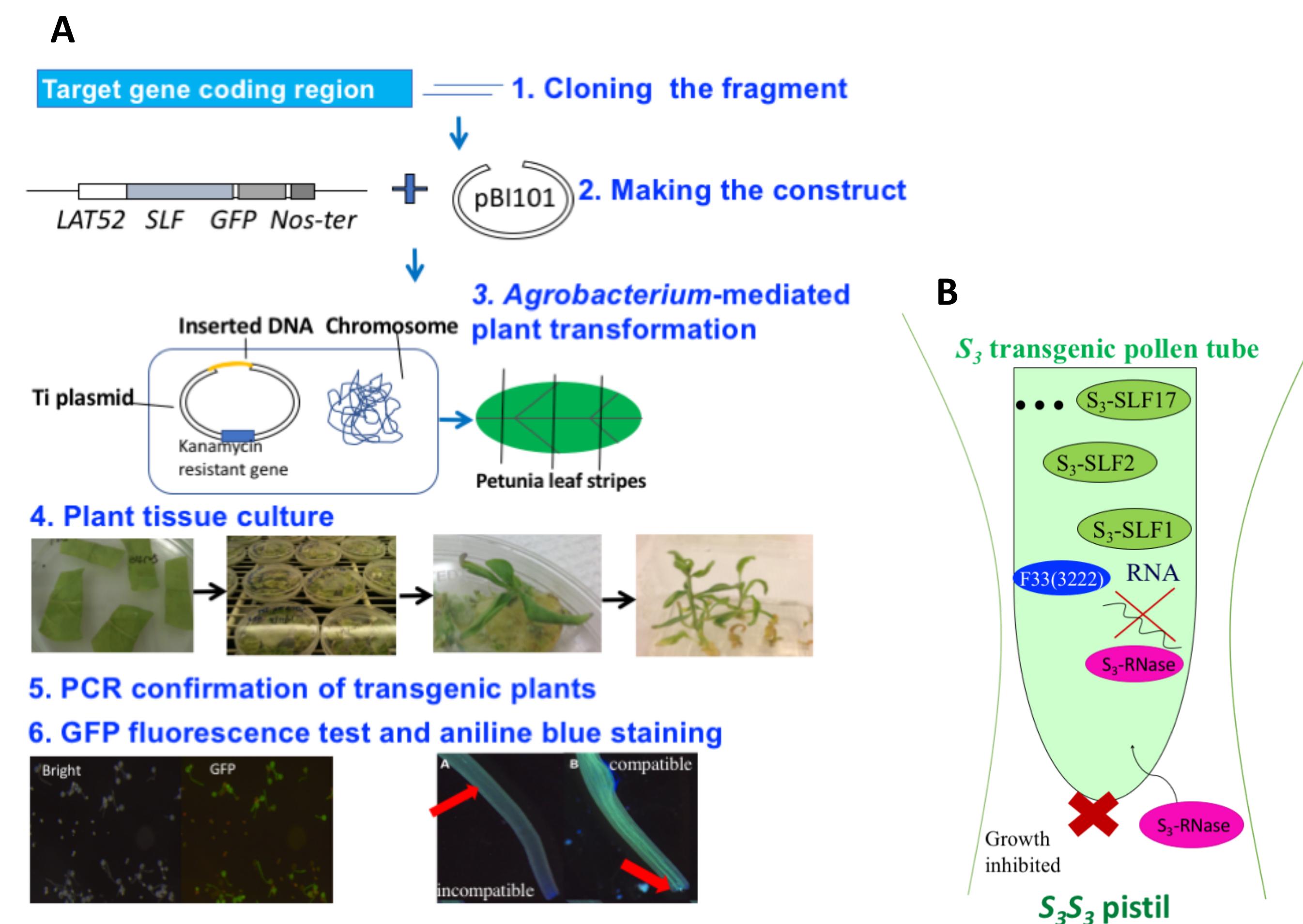


Figure 1. A. Standard procedure for generating transgenic plants expressing an SLF gene. B. *in vivo* functional assay

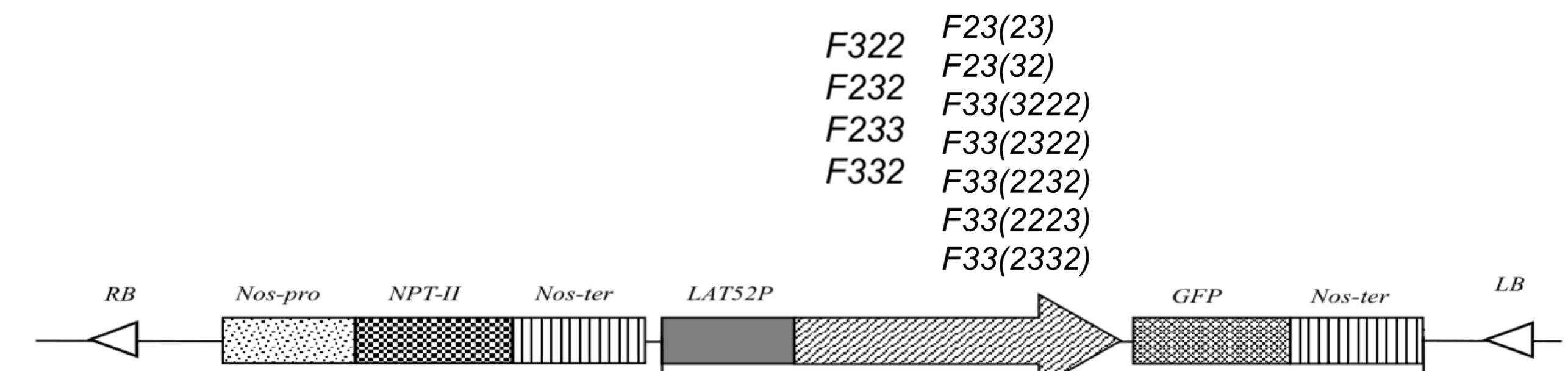


Figure 2. Transgene constructs for expressing 11 chimeric proteins of S₂-SLF1 and S₃-SLF1

Methods

To determine the biochemical basis for differential interactions of S₂-SLF1 and S₃-SLF1 with S₃-RNase, we first divided SLF1 into 3 functional domains (FD1, FD2 and FD3); generated 4 chimeric genes (F322, F232, F233, and F332) (Fig. 2); and used a transgenic assay (Fig. 1) to determine whether each encoded chimeric protein of S₂-SLF1 and S₃-SLF1 interacts with S₃-RNase. Based on the results (Fig. 3A), we further divided FD3 into 2 subdomains and then into 4 mini-domains (A, B, C, and D); generated 7 chimeric genes (Fig. 2); and similarly examined the ability of the resulting 7 chimeric proteins to interact with S₃-RNase (Fig. 3A,B). Protein structures were modeled using the I-TASSER server, and protein-protein docking analysis was performed by ClusPro (Fig. 4). FD3s of 4 SLF1s were aligned using MEGA 6 and ClustalW to identify amino acids conserved among the three that interact with S₃-RNase (Fig. 5).

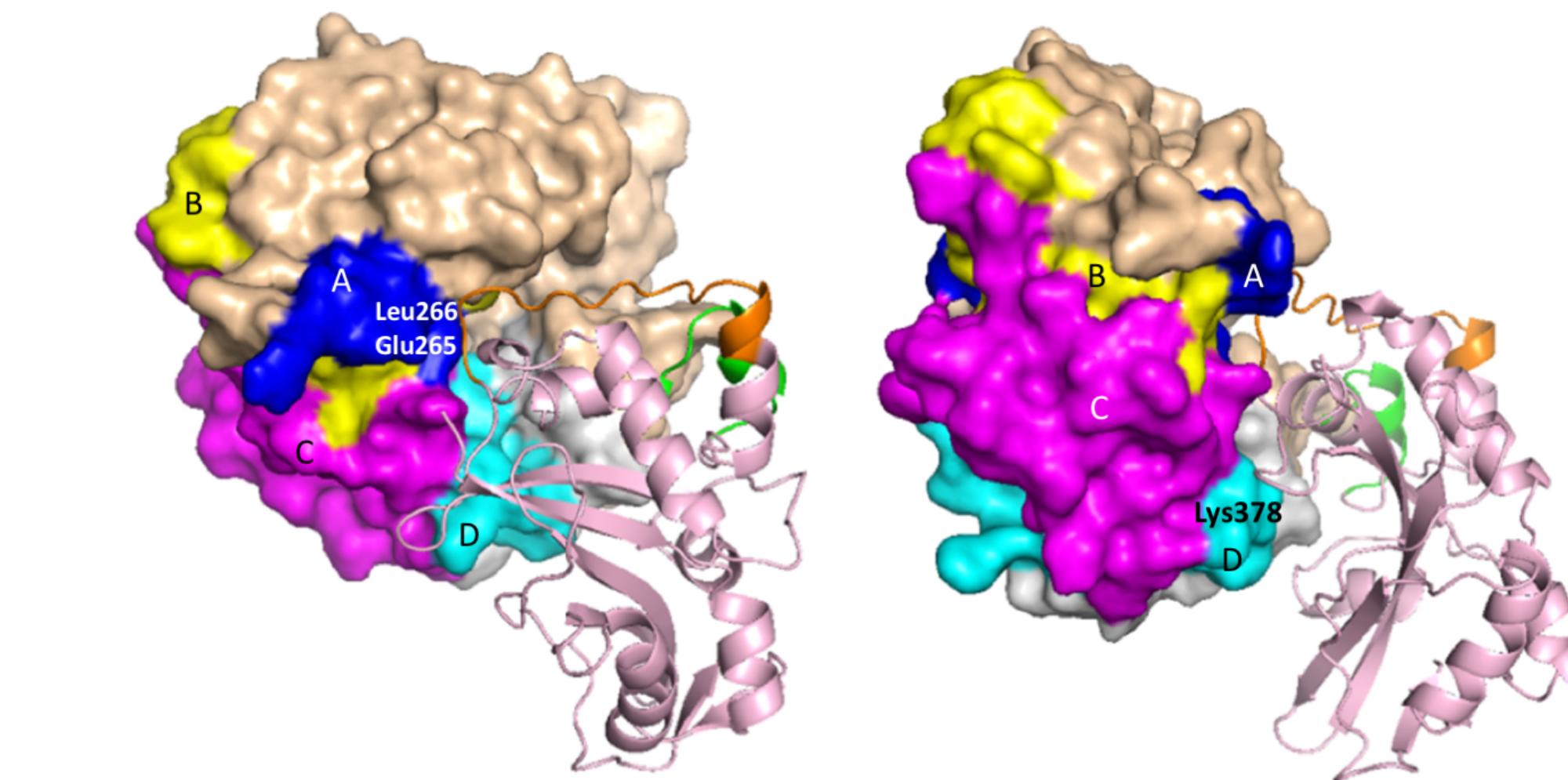


Figure 4. Computational modeling of S₂-SLF1 and molecular docking of S₃-RNase onto S₂-SLF1, as visualized in PyMOL.

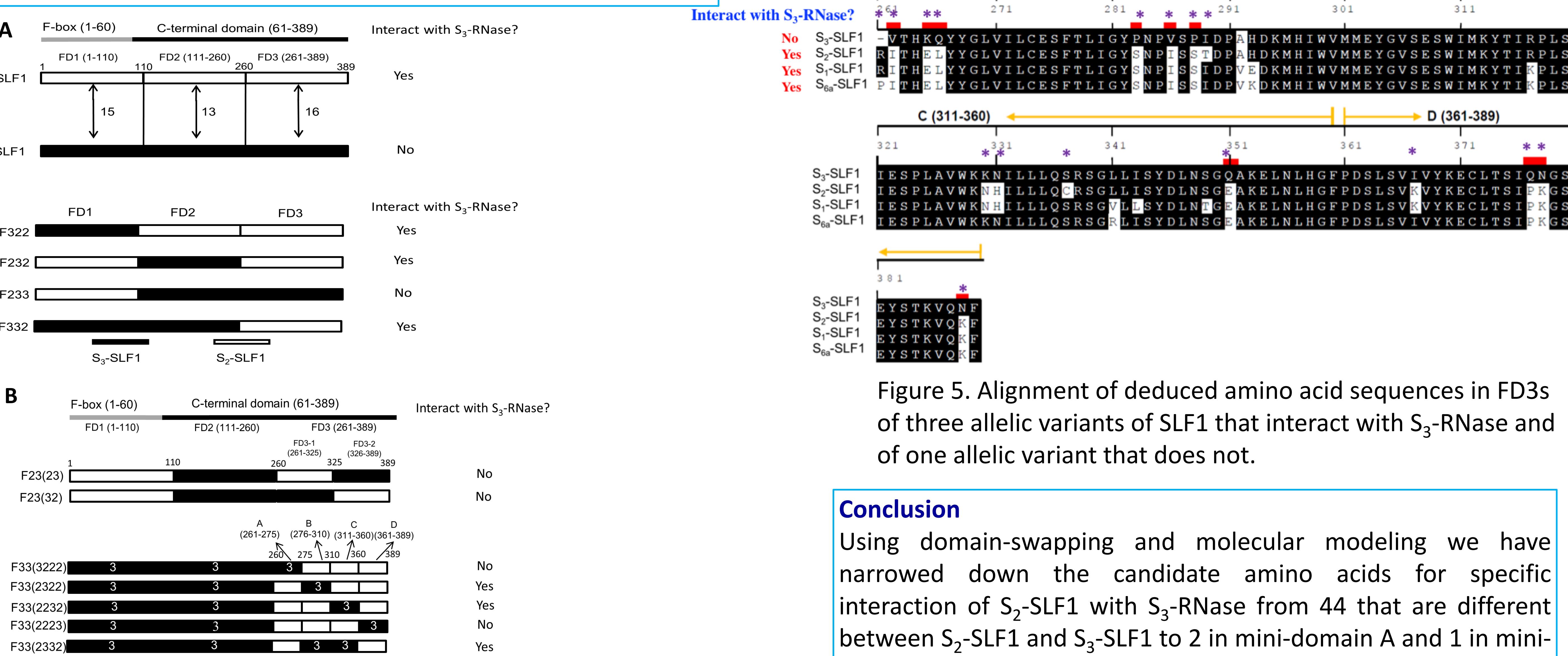


Figure 3. 11 chimeric proteins of S₂-SLF1 and S₃-SLF1, and their ability to interact with S₃-RNase.

Results

The results shown in Fig. 3 allowed us to first narrow the candidate amino acids for specific interaction of S₂-SLF1 with S₃-RNase to the 16 in FD3, and then to 4 in mini-domain A and 4 in mini-domain D. Molecular modeling of interactions between S₃-RNase and S₂-SLF1 revealed that 3 of these 8 are at the interaction surface (Fig. 4), and all 3 are conserved in S₁-SLF1 and S_{6a}-SLF1 that also interact with S₃-RNase (Fig. 5). Three of the chimeric proteins were used to determine whether FD3 alone contains the amino acids required for specific interaction of S₂-SLF1 with S₇-RNase and S₁₃-RNase. The results revealed that, unlike the case of S₂-SLF1's interaction with S₃-RNase, FD2 of S₂-SLF1 is required for interaction with S₇-RNase, and both FD1 and FD2 are required for interaction with S₁₃-RNase.

Conclusion

Using domain-swapping and molecular modeling we have narrowed down the candidate amino acids for specific interaction of S₂-SLF1 with S₃-RNase from 44 that are different between S₂-SLF1 and S₃-SLF1 to 2 in mini-domain A and 1 in mini-domain D of FD3. In contrast, FD1, or both FD1 and FD2, contain(s) amino acids required for interactions with S₇-RNase or S₁₃-RNase, suggesting diversity and complexity of interactions between SLF proteins and S-RNases.

Acknowledgements

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References

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