

Pathway Analysis - NK cells

Input data: DEGs

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1 Introducción y Objetivo

En este script realizo un pathways análisis con los DEGs obtenidos de los Venn Diagrams, y preparo el resultado en archivos gem para extraer un gráfico posteriormente en Cytoscape (<https://cran.r-project.org/web/packages/gprofiler2/vignettes/gprofiler2.html>).

2 Paquetes y datos

3 Datos

Cargo inicialmente los listados de células NK obtenidas en jVenn.

```
setwd("~/Desktop/ELENA_UOC/TFM")
jVenn_NKcells_vs_GSE22155_02 <- read_csv("DEGs/Venn_diagrams_cell_type_treatments_vs_untreated/jVenn_NK")
head(jVenn_NKcells_vs_GSE22155_02) # sección de los resultados
```

```
## # A tibble: 6 x 3
##   GSE50509 GSE22155_02 `GSE50509|GSE22155_02`
##   <chr>      <chr>      <chr>
## 1 LPIN1     AFP         AUTS2
## 2 TRAPPC2   RNPS1       BCL2
## 3 DKC1      TRIP11      ZNF318
## 4 SLC2A12   PHF1        PUM1
## 5 RCBTB2    CDK6        NABP2
## 6 RPS6KA3   SRD5A3      GNG2
```

```
jVenn_NKcells_vs_GSE22155_47 <- read_csv("DEGs/Venn_diagrams_cell_type_treatments_vs_untreated/jVenn_NKCells_vs_GSE22155_47.csv")
jVenn_NKcells_vs_GSE91061 <- read_csv("DEGs/Venn_diagrams_cell_type_treatments_vs_untreated/jVenn_NKCells_vs_GSE91061.csv")
jVenn_NKcells_vs_GSE35640 <- read_csv("DEGs/Venn_diagrams_cell_type_treatments_vs_untreated/jVenn_NKCells_vs_GSE35640.csv")
jVenn_NKcells_vs_GSE61992 <- read_csv("DEGs/Venn_diagrams_cell_type_treatments_vs_untreated/jVenn_NKCells_vs_GSE61992.csv")
setwd("~/Desktop/ELENA_UOC/TFM/GOBPs") # Directorio de trabajo
jVenn_NKcells <- read_delim("~/Desktop/ELENA_UOC/TFM/DEGs/Venn_diagrams_cell_type_treatments_comparison.csv",
  delim = ",", escape_double = FALSE, trim_ws = TRUE)
```

4 Uncovered

En esta sección contemplo la posibilidad que los DEGs únicos en el dataset sin tratar se traten de mecanismos que se escapan al tratamiento pero que igualmente se ven afectados por la enfermedad.

Primero realizo el pathway multianálisis:

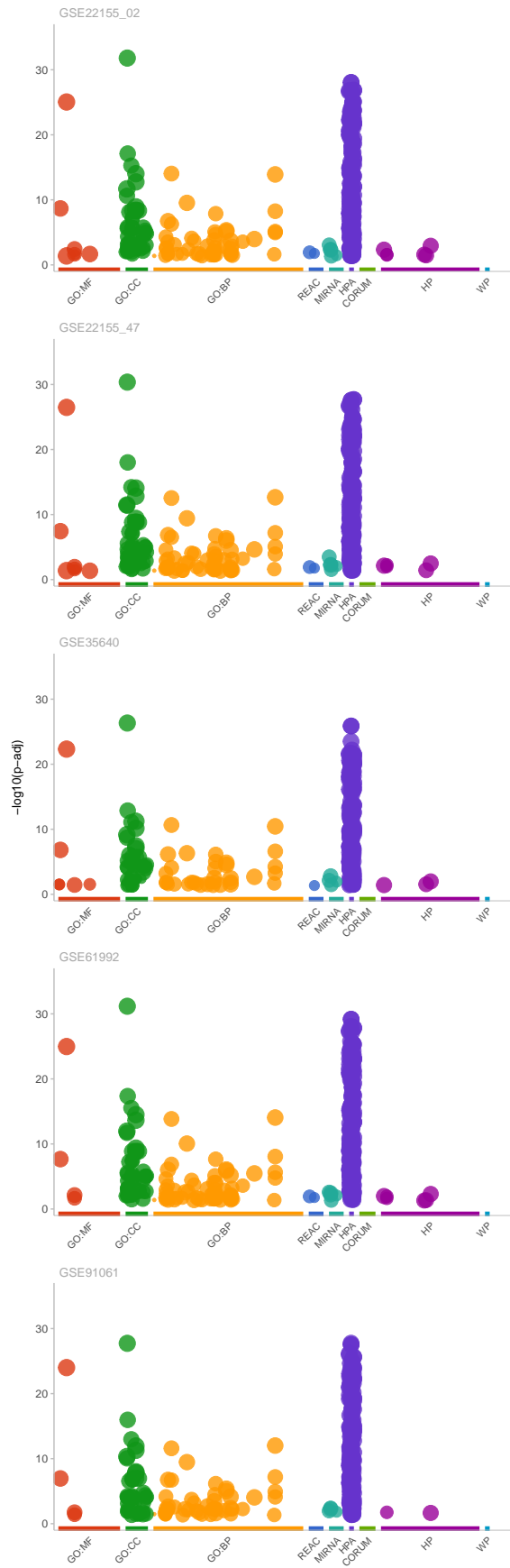
En esta sección contemplo la posibilidad que los DEGs únicos en el dataset sin tratar se traten de mecanismos que se escapan al tratamiento pero que igualmente se ven afectados por la enfermedad.

Primero realizo el pathway multianálisis:

```
multi_gostres <- gost(query = list("GSE22155_02" = unique(jVenn_NKcells_vs_GSE22155_02$GSE50509[which(is.na(jVenn_NKcells_vs_GSE22155_02$GSE50509) == FALSE])),
  "GSE22155_47" = jVenn_NKcells_vs_GSE22155_47$GSE50509[which(is.na(jVenn_NKcells_vs_GSE22155_47$GSE50509) == FALSE)],
  "GSE35640" = jVenn_NKcells_vs_GSE35640$GSE50509[which(is.na(jVenn_NKcells_vs_GSE35640$GSE50509) == FALSE)],
  "GSE91061" = jVenn_NKcells_vs_GSE91061$GSE50509[which(is.na(jVenn_NKcells_vs_GSE91061$GSE50509) == FALSE)],
  "GSE61992" = jVenn_NKcells_vs_GSE61992$GSE50509[which(is.na(jVenn_NKcells_vs_GSE61992$GSE50509) == FALSE)]),
  evcodes = TRUE, multi_query = FALSE,
  sources = c("GO", "REAC", "MIRNA", "CORUM", "HP", "HPA", "WP"))
head(multi_gostres$result[,1:7])
```

##	query	significant	p_value	term_size	query_size	intersection_size
## 1	GSE22155_02	TRUE	9.128453e-15	2556	1325	268
## 2	GSE22155_02	TRUE	1.231616e-14	5986	1325	518
## 3	GSE22155_02	TRUE	2.919690e-10	4912	1325	423
## 4	GSE22155_02	TRUE	5.560949e-09	2074	1325	209
## 5	GSE22155_02	TRUE	1.320218e-08	1600	1325	170
## 6	GSE22155_02	TRUE	1.789227e-07	3613	1325	317
##	precision					
## 1	0.2022642					
## 2	0.3909434					
## 3	0.3192453					
## 4	0.1577358					
## 5	0.1283019					
## 6	0.2392453					

```
p <- gostplot(multi_gostres, capped = FALSE, interactive = F)
p
```



Estos resultados son los que utilizo para generar los archivos gem que archivo para usar más tarde en Cytoscape.

```
# format to GEM
gem <- multi_gostres$result[,c("query", "term_id", "term_name", "p_value", "intersection_size")]
colnames(gem) <- c("query", "GO.ID", "Description", "p.Val", "Genes")
gem$FDR <- gem$p.Val
gem$Phenotype = "+1"

gem %>% group_by(query) %>%
  group_walk(~
    write.table(data.frame(.x[,c("GO.ID", "Description", "p.Val", "FDR", "Phenotype", "Genes")]),
      file = paste0("./Venn_diagrams_cell_type_treatments_vs_untreated/gProfiler_NKcells_Only",
        sep = "\t", quote = F, row.names = F))
```

Proporciones de los pathways para cada estudio:

```
df <- as.data.frame(multi_gostres$result[,1:13])
table(df$query)

##
## GSE22155_02 GSE22155_47 GSE35640 GSE61992 GSE91061
## 335 335 283 338 316

prop.table(table(df$query))*100

##
## GSE22155_02 GSE22155_47 GSE35640 GSE61992 GSE91061
## 20.84630 20.84630 17.61045 21.03298 19.66397

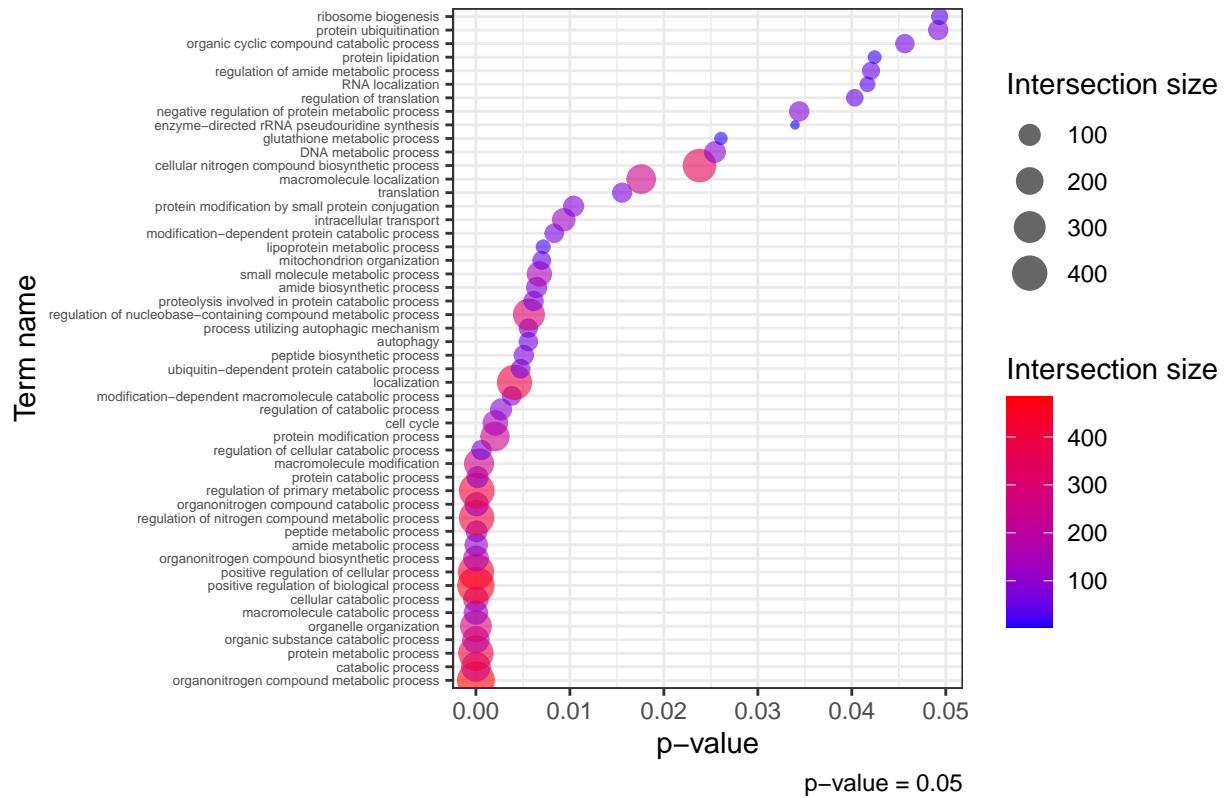
#table(df$term_name)
#prop.table(table(df$term_name))*100
df2 <- as.data.frame(table(df$term_name, df$query))
#length(unique(df2$Var1))
write.table(df, file = "./Venn_diagrams_cell_type_treatments_vs_untreated/NKcells_GOBP.txt", sep = "\t")
write.table(df2, file = "./Venn_diagrams_cell_type_treatments_vs_untreated/NKcells_GOBP_freq.txt", sep = "\t")
#rm(jVenn_NKcells_vs_GSE22155_02, jVenn_NKcells_vs_GSE22155_47, jVenn_NKcells_vs_GSE35640, jVenn_NKcells_vs_GSE61992, jVenn_NKcells_vs_GSE91061)
```

Barplot of the top GO-BPs:

```
plot_gobps <- function(study, n = 50){
df2 <- df[df$source == "GO:BP",]
df2_GSE91061 <- df2[df2$query == study,]
df2_GSE91061 <- df2_GSE91061[1:n,]
ggplot(df2_GSE91061, aes(p_value, reorder(term_name,p_value))) + ggtitle(study) +
  geom_point(aes(p_value, reorder(term_name,p_value), colour = intersection_size, size = intersection_size)) +
  theme_bw() + theme(axis.text.y.left = element_text(size = 5)) +
  labs(x= "p-value", y="Term name", colour = "Intersection size", size = "Intersection size", caption = "Intersection size")
  scale_color_gradient(low="blue", high="red")
}

plot_gobps(study = "GSE91061")
```

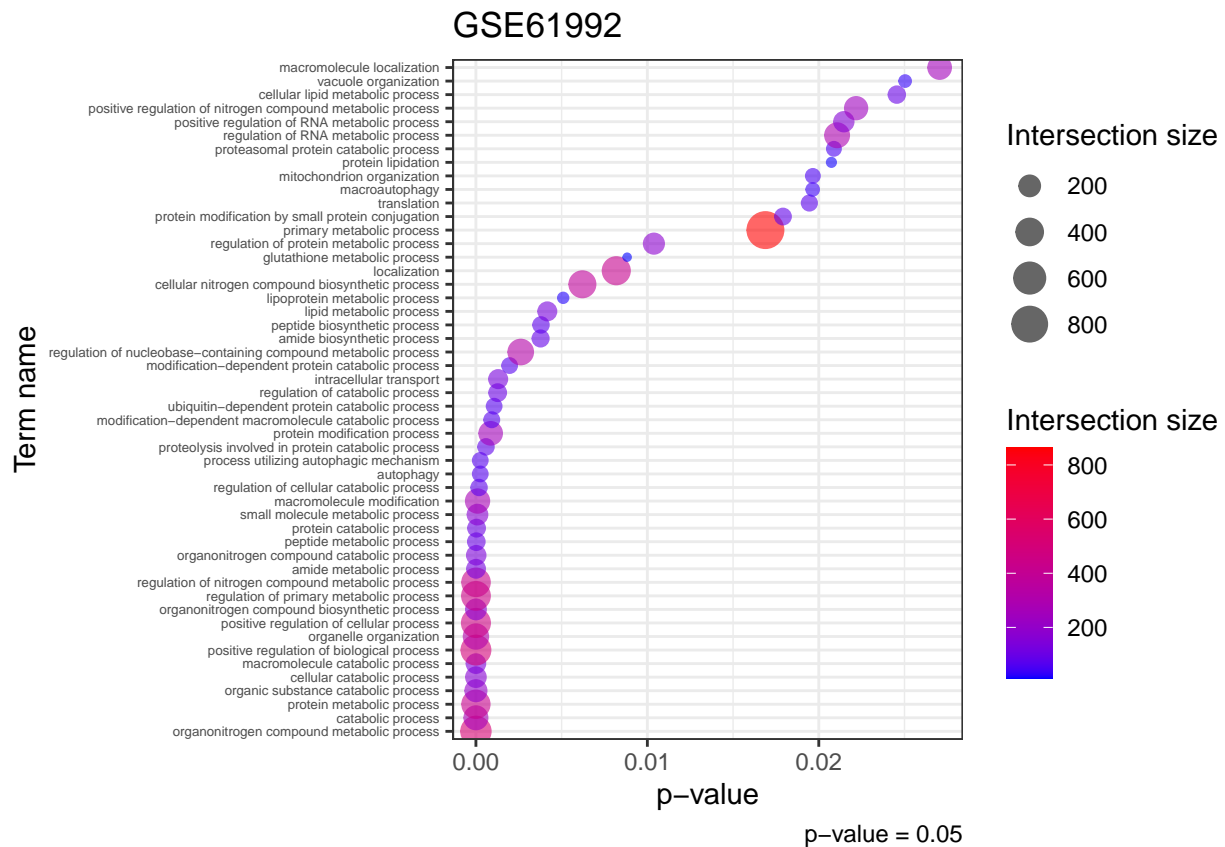
GSE91061



```
print("\n")
```

```
## [1] "\n"
```

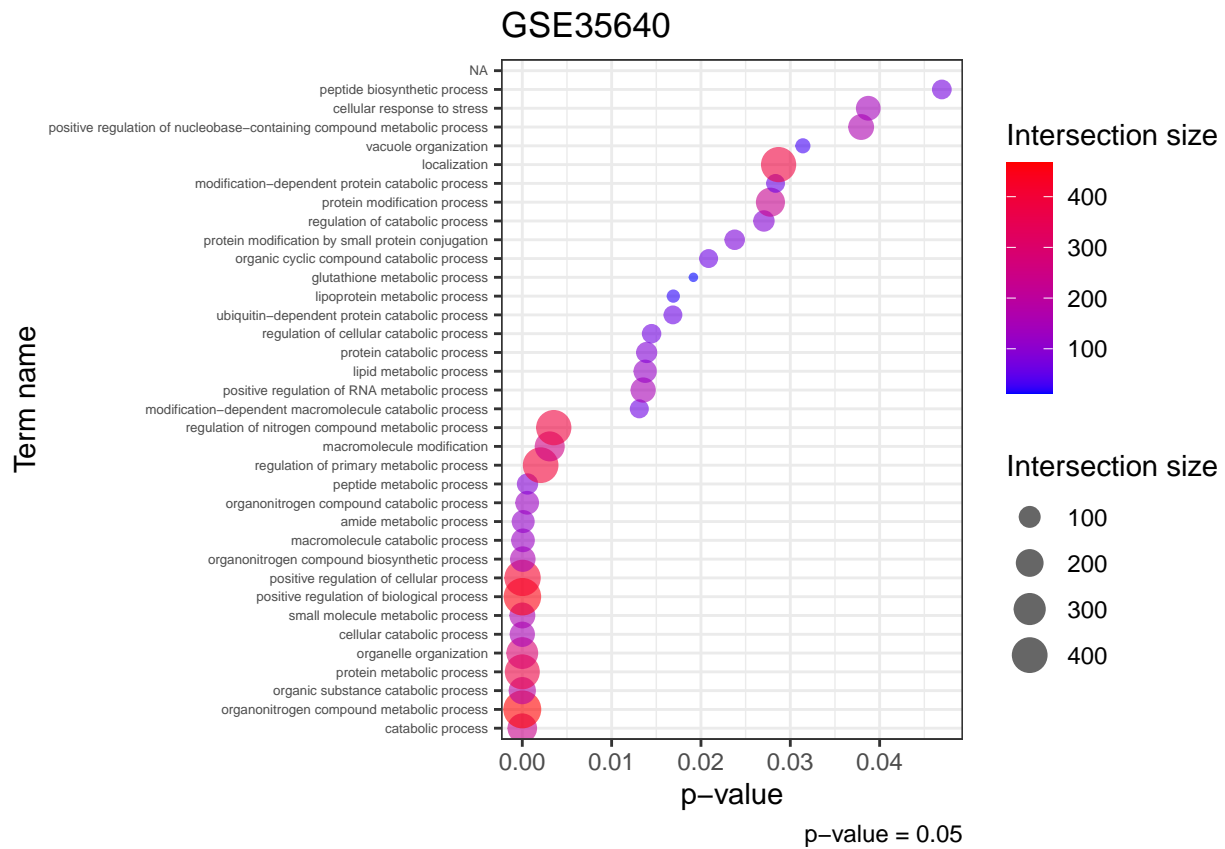
```
plot_gobps(study = "GSE61992")
```



```
print("\n")
```

```
## [1] "\n"
```

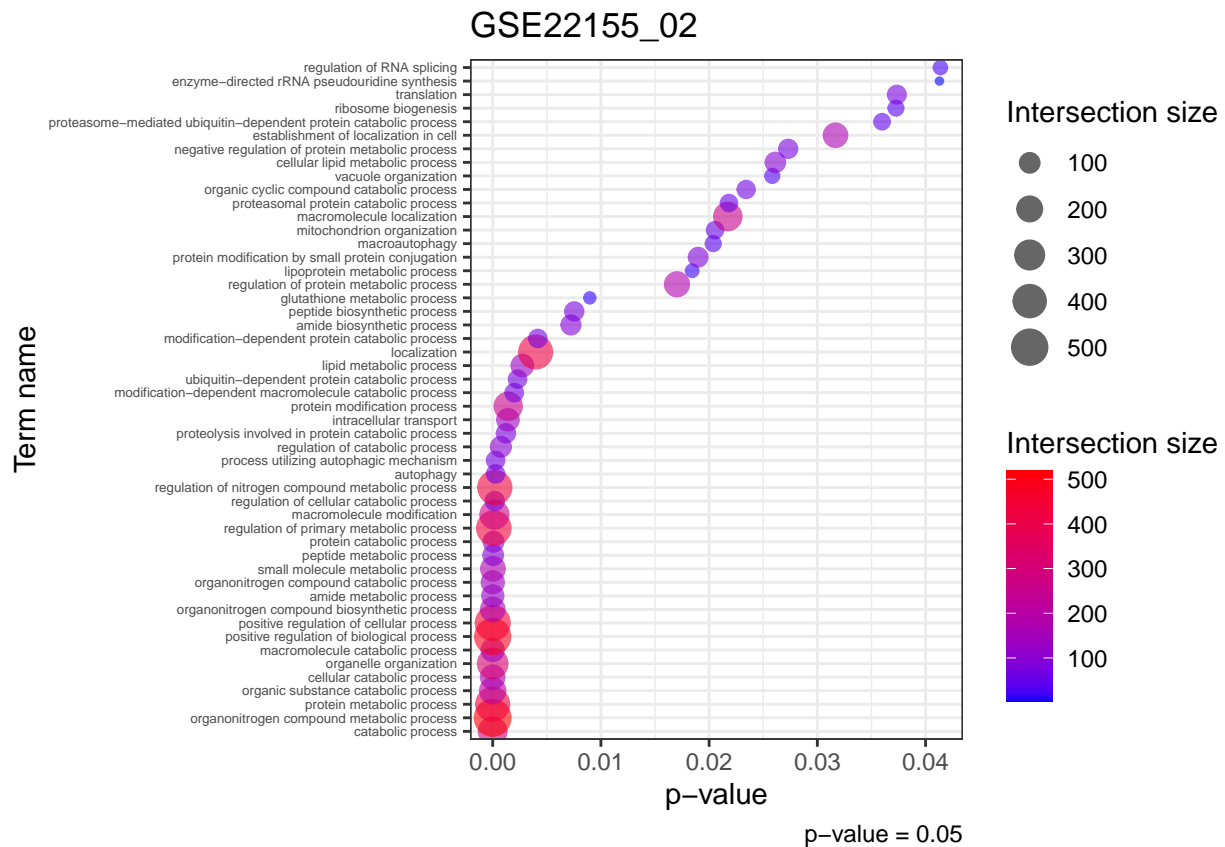
```
plot_gobps(study = "GSE35640")
```



```
print("\n")
```

```
## [1] "\n"
```

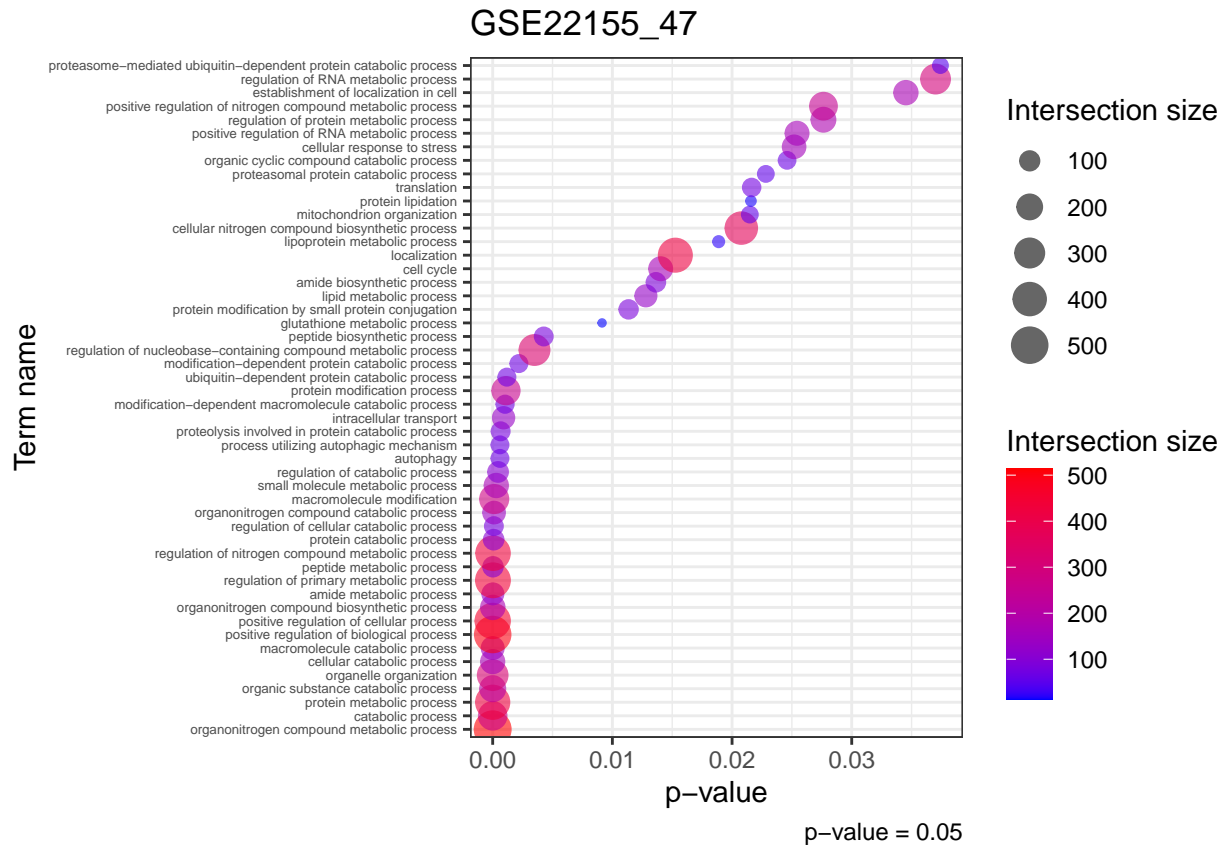
```
plot_gobps(study = "GSE22155_02")
```



```
print("\n")
```

```
## [1] "\n"
```

```
plot_gobps(study = "GSE22155_47")
```

5 Exclusive

En esta sección analizo los DEGs exclusivos para cada estudio, y realizo un multi-análisis para cada estudio. Los resultados los guardo como archivos gem específicos para usar en Cytoscape.

Primero realizo el pathway multianálisis:

```
multi_gostres <- gost(query = list("GSE61992" = jVenn_NKcells$GSE61992[which(is.na(jVenn_NKcells$GSE61992))],
                                   "GSE91061" = jVenn_NKcells$GSE91061[which(is.na(jVenn_NKcells$GSE91061))]),
                      evcodes = TRUE, multi_query = FALSE,
                      sources = c("GO", "REAC", "MIRNA", "CORUM", "HP", "HPA", "WP"))
head(multi_gostres$result[,1:7])
```

##	query	significant	p_value	term_size	query_size	intersection_size
## 1	GSE61992	TRUE	9.193374e-05	3855	163	59
## 2	GSE61992	TRUE	1.217385e-04	2637	163	46
## 3	GSE61992	TRUE	1.980644e-03	1984	163	36
## 4	GSE61992	TRUE	2.077135e-03	1988	163	36
## 5	GSE61992	TRUE	2.343299e-03	220	163	11
## 6	GSE61992	TRUE	6.173393e-03	115	163	8
##	precision					
## 1	0.36196319					
## 2	0.28220859					
## 3	0.22085890					
## 4	0.22085890					
## 5	0.06748466					

```
## 6 0.04907975
```

Estos resultados son los que utilizo para generar los archivos gem que archivo para usar más tarde en Cytoscape.

```
# format to GEM
```

```
gem <- multi_gostres$result[,c("query", "term_id", "term_name", "p_value", "intersection_size")]
```

```
colnames(gem) <- c("query", "GO.ID", "Description", "p.Val", "Genes")
```

```
gem$FDR <- gem$p.Val
```

```
gem$Phenotype = "+1"
```

```
gem %>% group_by(query) %>%
```

```
  group_walk(~
```

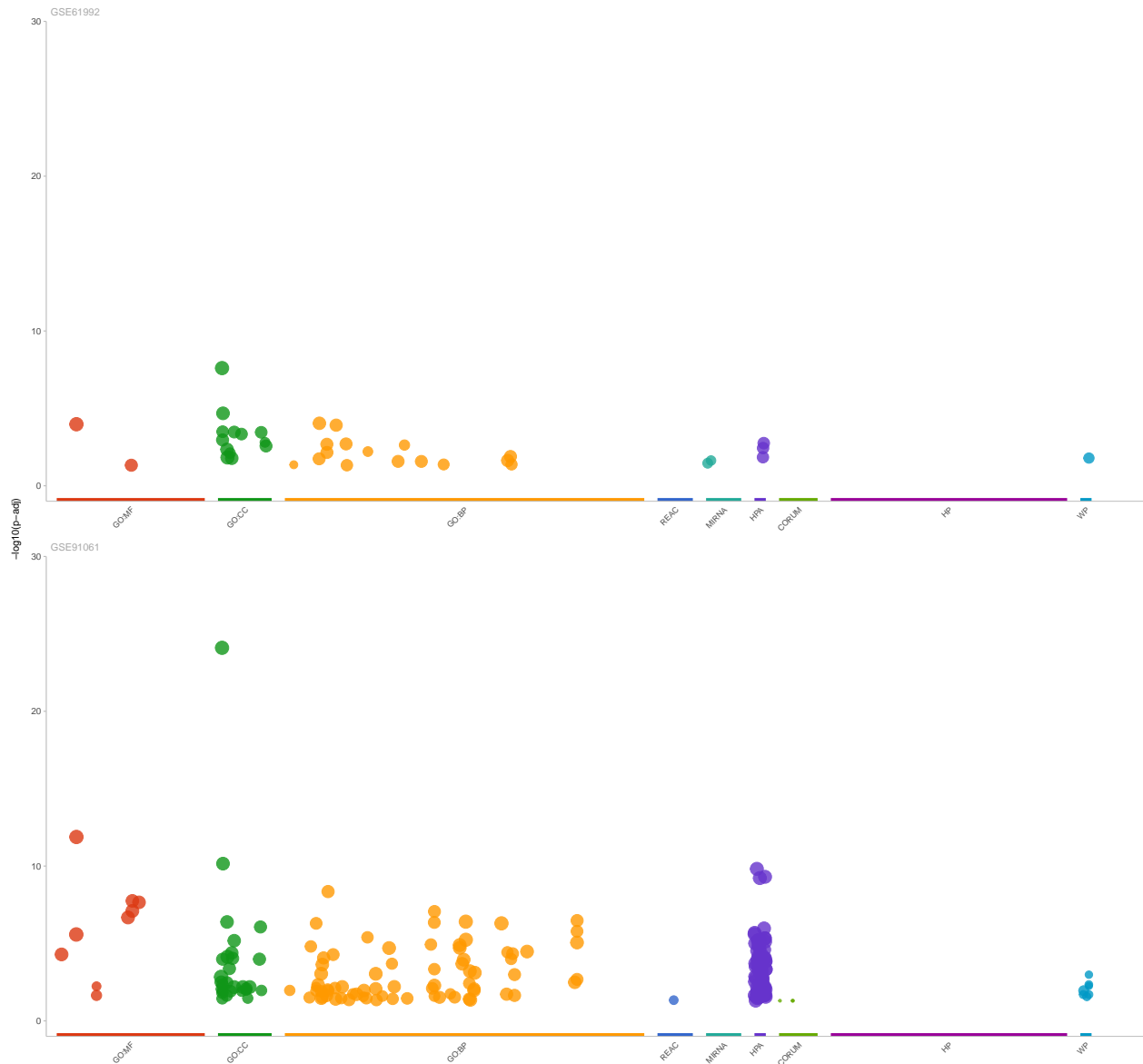
```
    write.table(data.frame(.x[,c("GO.ID", "Description", "p.Val", "FDR", "Phenotype", "Genes")]),
```

```
                  file = paste0("./Treatment_comparisons/gProfiler_NKcells_Only_geneset_", unique(.y$query),
```

```
                  sep = "\t", quote = F, row.names = F))
```

```
p <- gostplot(multi_gostres, capped = FALSE, interactive = F)
```

```
p
```



Proporciones de los pathways para cada estudio:

```
df <- as.data.frame(multi_gostres$result[,1:13])
table(df$query)
```

```
##
## GSE61992 GSE91061
##      37      211
```

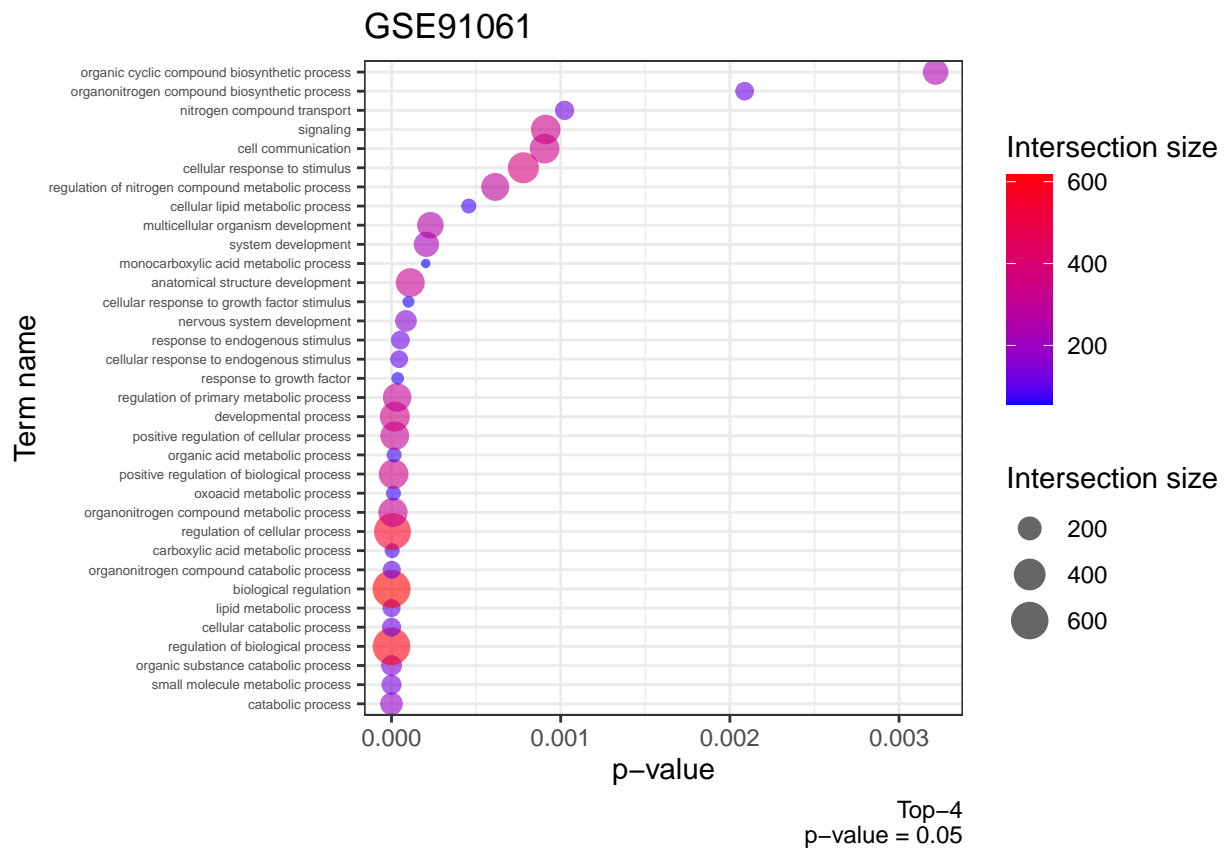
```
write.table(gem, file = "./Treatment_comparisons/gProfiler_NKcells_Only_genesets.txt", sep = "\t", quote = FALSE)
```

```
plot_gobps <- function(study){
df2_GSE91061 <- df2[df2$query == study,]
ggplot(df2_GSE91061, aes(p_value, reorder(term_name,p_value))) + ggtitle(study) +
  geom_point(aes(p_value, reorder(term_name,p_value), colour = intersection_size, size = intersection_size)) +
  theme_bw() + theme(axis.text.y.left = element_text(size = 5)) +
  labs(x= "p-value", y="Term name", colour = "Intersection size", size = "Intersection size", caption = "Intersection size")
  scale_color_gradient(low="blue", high="red")
}
```

```

}
df2 <- df[df$source == "GO:BP",]
df2 <- df2[1:50,]
plot_gobps(study = "GSE91061")

```



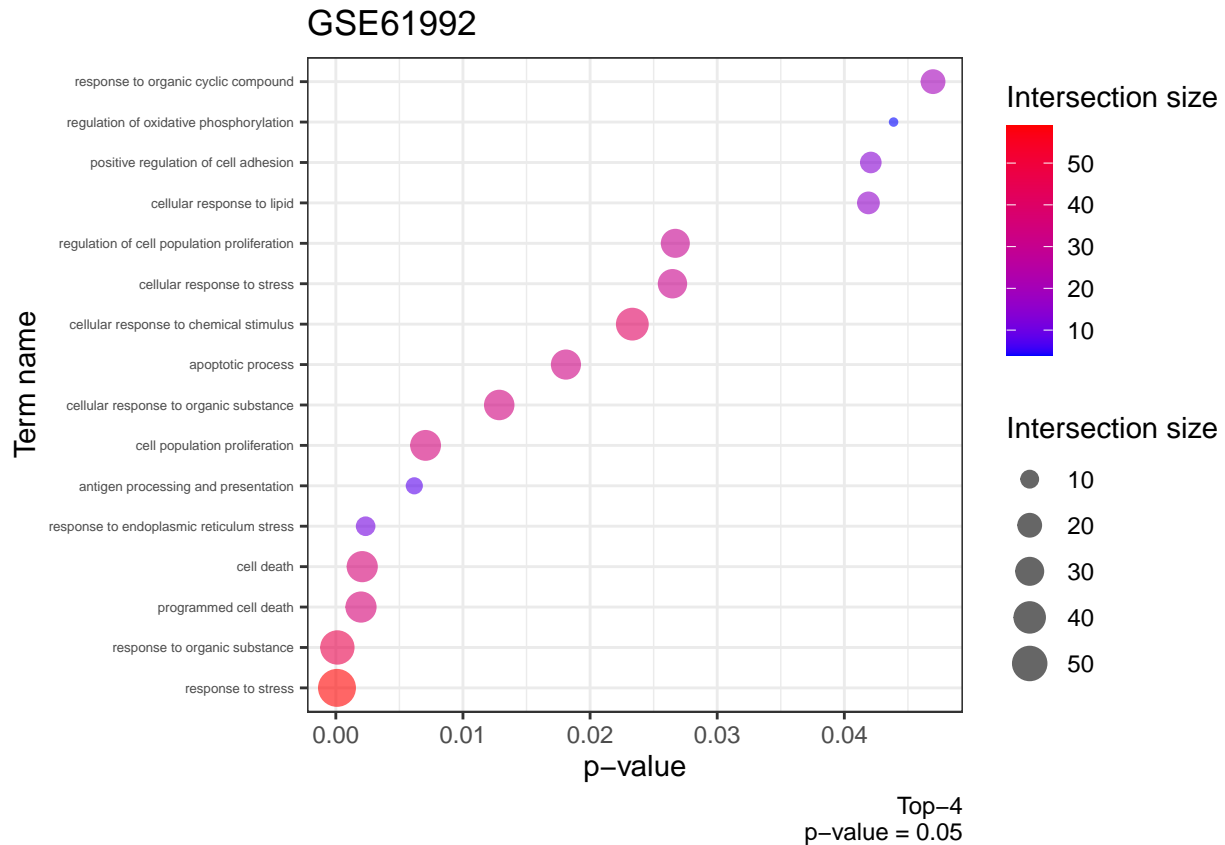
```
print("\n")
```

```
## [1] "\n"
```

```

df2 <- df[df$source == "GO:BP",]
plot_gobps(study = "GSE61992")

```



6 Máximo solapamiento

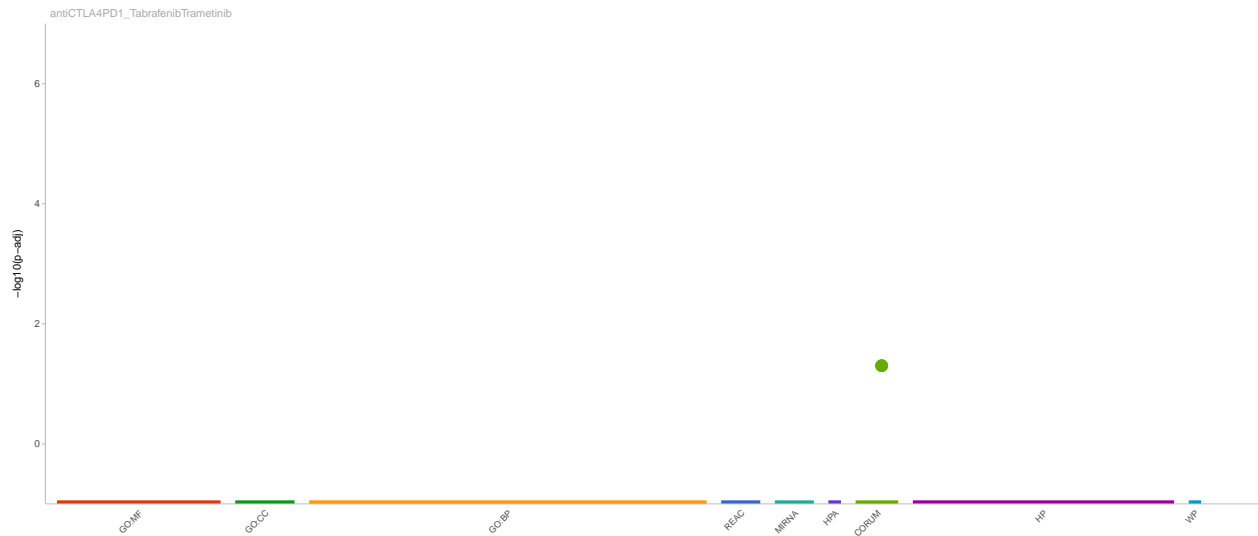
En esta sección analizo los DEGs que más se comparten entre los estudios para observar los mecanismos que pudieran haber en común entre los tratamientos.

Primero realizo el pathway multianálisis:

```
multi_gostres <- gost(query = list(
  "antiCTLA4PD1_TabrafenibTrametinib" = jVenn_NKcells$`GSE91061|GSE61992`
),
  evcodes = TRUE, multi_query = FALSE,
  sources = c("GO", "REAC", "MIRNA", "CORUM", "HP", "HPA", "WP"))
head(multi_gostres$result[,1:7])
```

```
##               query significant    p_value term_size query_size
## 1 antiCTLA4PD1_TabrafenibTrametinib    TRUE 0.04993169         2         1
## 2 antiCTLA4PD1_TabrafenibTrametinib    TRUE 0.04993169         2         1
## 3 antiCTLA4PD1_TabrafenibTrametinib    TRUE 0.04993169         2         1
## 4 antiCTLA4PD1_TabrafenibTrametinib    TRUE 0.04993169         2         1
## intersection_size precision
## 1             1         1
## 2             1         1
## 3             1         1
## 4             1         1
```

```
p <- gostplot(multi_gostres, capped = FALSE, interactive = F)
p
```



Estos resultados son los que utilizo para generar los archivos gem que archivo para usar más tarde en Cytoscape.

```
# format to GEM
gem <- multi_gostres$result[,c("query", "term_id", "term_name", "p_value", "intersection_size")]
colnames(gem) <- c("query", "GO.ID", "Description", "p.Val", "Genes")
gem$FDR <- gem$p.Val
gem$Phenotype = "+1"

gem %>% group_by(query) %>%
  group_walk(~
    write.table(data.frame(.x[,c("GO.ID", "Description", "p.Val", "FDR", "Phenotype", "Genes")],
      file = paste0("./Treatment_comparisons/gProfiler_NKcells_maxOverlap_geneset_", unique(query)),
      sep = "\t", quote = F, row.names = F))
  )
```

Al mirar los procesos por cada comparación salieron demasiados para poder visualizarlos bien, por lo que decidí restringir a los procesos GO-BPs únicamente y guardarlos en pdf.

En este caso no hay resultado de GOBPs.

```
#unique(gem$query)

gem2 <- gem[grep("GO",gem$GO.ID),]
plot_gobps <- function(query, cutoff = 0.05) {
  gem2 <- gem2[gem2$FDR <= cutoff,]
  gem2_1 <- gem2[gem2$query == query, ]
  ggplot(gem2_1, aes(FDR, Description)) + ggtitle(query) +
    geom_point(aes(FDR, Description, colour = Genes, size = Genes), alpha = 0.6)+
    theme_bw() + theme(axis.text.y.left = element_text(size = 5)) +
    labs(caption = paste0("FDR = ",cutoff))+
    scale_color_gradient(low="blue", high="red")
}

plot_gobps("antiCTLA4PD1_TabrafenibTrametinib")
```

antiCTLA4PD1_TabrafenibTrametinib

Description

FDR

FDR = 0.05

```
df <- as.data.frame(multi_gostres$result[,1:13])
df2 <- df[df$source == "GO:BP",]
plot_gobps <- function(study){
df2_GSE91061 <- df2[df2$query == study,]
ggplot(df2_GSE91061, aes(p_value, reorder(term_name,p_value))) + ggtitle(study) +
  geom_point(aes(p_value, reorder(term_name,p_value), colour = intersection_size, size = intersection_size)) +
  theme_bw() + theme(axis.text.y.left = element_text(size = 5)) +
  labs(x= "p-value", y="Term name", colour = "Intersection size", size = "Intersection size", caption = "Intersection size")
  scale_color_gradient(low="blue", high="red")
}
plot_gobps(study = "antiCTLA4PD1_TabrafenibTrametinib")
```

antiCTLA4PD1_TabrafenibTrametinib

Term name

p-value

p-value = 0.05