

# Deconvolución de datos de melanoma con quantiseqr: Datos bulk RNA-seq: GSE54467

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## Contents

|   |                         |   |
|---|-------------------------|---|
| 1 | Introducción y Objetivo | 1 |
| 2 | Paquetes y datos        | 1 |

## 1 Introducción y Objetivo

## 2 Paquetes y datos

Repositorio GitHub de : [https://github.com/Danko-Lab/quantiseqr/blob/main/tutorial\\_deconvolution.pdf](https://github.com/Danko-Lab/quantiseqr/blob/main/tutorial_deconvolution.pdf)

```
knitr::opts_chunk$set(warning=FALSE)
#if (!requireNamespace("BiocManager", quietly = TRUE)) {
#  install.packages("BiocManager")
#}
#BiocManager::install("quantiseqr")
package_to_load <- c("readr", "dplyr", "ggplot2", "tidyr", "dplyr", "RColorBrewer",
                     "Biobase", "quantiseqr", "gplots")
for (package in package_to_load) {
  require(package, character.only = T); packageVersion(package)
}
extra_to_load <- c("knitr", "stringr", "stringi", "ggrepel", "ggpubr", "ggbreak",
                  "reshape2", "ggfortify", "cowplot", "GEOquery", "Seurat", "data.table",
                  "limma", "illuminaHumanv4.db", "SummarizedExperiment", "tibble")
for (package in extra_to_load) {
  require(package, character.only = T); packageVersion(package)
}
rm(package_to_load, extra_to_load)
```

#Datos

#Deconvolución

En este análisis utilizo los datos del estudio GSE54467 descargados mediante la función `getGEO` des de la base de datos GEO, del NCBI: <https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE54467>.

Las muestras consisten en 65 muestras analizadas con la plataformas GPL570, y con varios tratamientos: dabrafenib + trametinib.

Este estudio es de especial interés para el TFM debido a que los autores también proporcionan metadata la respuesta de los pacientes, cosa que permitirá estudiar posibles correlaciones con las poblaciones obtenidas de

la deconvolución.

```
setwd("~/Desktop/ELENA_UOC/TFM")

gset <- getGEO("GSE22155", GSEMatrix =TRUE, getGPL=FALSE)
if (length(gset) > 1) idx <- grep("GPL6102", attr(gset, "names")) else idx <- 1
gset_GPL6102 <- gset[[idx]]
#table(gset_GPL6102$characteristics_ch1.1) # OS (days)
#table(gset_GPL6102$characteristics_ch1.2) # event (0=alive, 1=dead)
#table(gset_GPL6102$characteristics_ch1.3) # sex
#table(gset_GPL6102$characteristics_ch1.4) # age at metastases
#table(gset_GPL6102$characteristics_ch1.5) # type of metastases: Lymohnode
#table(gset_GPL6102$characteristics_ch1.6) # age at primary diagnosis
#table(gset_GPL6102$characteristics_ch1.7) # localization of primary melanoma
#table(gset_GPL6102$characteristics_ch1.8) # type
#table(gset_GPL6102$characteristics_ch1.9) # breslow
#table(gset_GPL6102$characteristics_ch1.10) # clark
#table(gset_GPL6102$characteristics_ch1.11) # Stage (III and IV)
#table(gset_GPL6102$characteristics_ch1.12) # braf/nras
#table(gset_GPL6102$characteristics_ch1.13) # cdkn2a (hd=homozygous deletion, *=germline)
#table(gset_GPL6102$characteristics_ch1.14) # molecular subtype
#table(gset_GPL6102$characteristics_ch1.15) # cd3 immunohistochemistry
#table(gset_GPL6102$characteristics_ch1.16) # cd20 immunohistochemistry
#table(gset_GPL6102$characteristics_ch1.17) # ki67 (0=<30%, 1=>30%)
#table(gset_GPL6102$`age at metastases:ch1`) # Age at metastases
#table(gset_GPL6102$`age at primary diagnosis:ch1`) # age at primary diagnosis
#table(gset_GPL6102$`localization of primary melanoma:ch1`) # localization of primary melanoma
#table(gset_GPL6102$`molecular subtype:ch1`) # molecular subtype

if (length(gset) > 1) idx <- grep("GPL6947", attr(gset, "names")) else idx <- 1
gset_GPL6947 <- gset[[idx]]
#table(gset_GPL6947$characteristics_ch1.1)# os (days)
#table(gset_GPL6947$characteristics_ch1.2) # event (0=alive, 1=dead):
#table(gset_GPL6947$characteristics_ch1.3) # sex
#table(gset_GPL6947$characteristics_ch1.4) # age at metastases
#table(gset_GPL6947$characteristics_ch1.5) # type of metastases
#table(gset_GPL6947$characteristics_ch1.6) # age at primary diagnosis
#table(gset_GPL6947$characteristics_ch1.7) # localization of primary melanoma
#table(gset_GPL6947$characteristics_ch1.8) # type
#table(gset_GPL6947$characteristics_ch1.9) # breslow
#table(gset_GPL6947$characteristics_ch1.10) # clark
#table(gset_GPL6947$characteristics_ch1.11) # stage
#table(gset_GPL6947$characteristics_ch1.12) # braf/nras
#table(gset_GPL6947$characteristics_ch1.13) # cdkn2a (hd=homozygous deletion, *=germline)
#table(gset_GPL6947$characteristics_ch1.14) # cdkn2a (hd=homozygous deletion, *=germline)
#table(gset_GPL6947$characteristics_ch1.15) # cd3 immunohistochemistry = NAs
#table(gset_GPL6947$characteristics_ch1.16) # cd20 immunohistochemistry = NAs
#table(gset_GPL6947$characteristics_ch1.17) # ki67 (0=<30%, 1=>30%) = NAs
#table(gset_GPL6947$`localization of primary melanoma:ch1`) # localization of primary melanoma
#table(gset_GPL6947$`molecular subtype:ch1`) # molecular subtype
#table(gset_GPL6947$`tissue:ch1`) # tissue
#table(gset_GPL6947$`stage:ch1`) # All IV
#table(gset_GPL6947$`type of metastases:ch1`)# Type if metastases
```

```

# Debido a que los autores proporcionan los genes con la nomenclatura de Illumina, lo convierto a símbolo
x <- illuminaHumanv4SYMBOL # cargado con el paquete illuminaHumanv4.db
mapped_probes <- mappedkeys(x) # Para sacar los símbolos
xx <- as.list(x[mapped_probes]) # Lo paso a listado
my_genes_GPL6102 <- as.data.frame(unlist(xx[(rownames(gset_GPL6102@assayData$exprs))])) # Lo convierto
my_genes_GPL6947 <- as.data.frame(unlist(xx[(rownames(gset_GPL6947@assayData$exprs))])) # Lo convierto
my_genes_GPL6102$gene <- rownames(my_genes_GPL6102)
my_genes_GPL6947$gene <- rownames(my_genes_GPL6947)

bulk_metadata_GPL6102 <- as.data.frame(gset_GPL6102@phenoData@data) # Paso la metadata disponible a una
bulk_metadata_GPL6947 <- as.data.frame(gset_GPL6947@phenoData@data) # Paso la metadata disponible a una

# Para usar los símbolos en lugar de nombres de ilumina, extraigo los datos de expresión:
bulk.mtx_GPL6102 <- as.data.frame(gset_GPL6102@assayData$exprs) # Los datos de expresión
bulk.mtx_GPL6947 <- as.data.frame(gset_GPL6947@assayData$exprs) # Los datos de expresión
bulk.mtx_GPL6102$gene <- rownames(bulk.mtx_GPL6102) # La columna que usaré para integrar
bulk.mtx_GPL6947$gene <- rownames(bulk.mtx_GPL6947) # La columna que usaré para integrar
bulk.mtx_GPL6102 <- inner_join(my_genes_GPL6102, bulk.mtx_GPL6102, by = "gene") # Integración de ambas
bulk.mtx_GPL6947 <- inner_join(my_genes_GPL6947, bulk.mtx_GPL6947, by = "gene") # Integración de ambas
bulk.mtx_GPL6102$gene <- NULL # Elimino la columna con nombres de Illumina
bulk.mtx_GPL6947$gene <- NULL # Elimino la columna con nombres de Illumina
colnames(bulk.mtx_GPL6102)[1] <- "symbols" # Nombro la columna de símbolos de los genes
colnames(bulk.mtx_GPL6947)[1] <- "symbols" # Nombro la columna de símbolos de los genes

# Agrego los posibles duplicados calculando la media:
bulk.mtx_GPL6102 <- aggregate(bulk.mtx_GPL6102, by = list(c(bulk.mtx_GPL6102$symbols)), mean) # Agregar
bulk.mtx_GPL6947 <- aggregate(bulk.mtx_GPL6947, by = list(c(bulk.mtx_GPL6947$symbols)), mean) # Agregar
rownames(bulk.mtx_GPL6102) <- bulk.mtx_GPL6102$Group.1 # Los nombres de genes únicos sin duplicados sir
rownames(bulk.mtx_GPL6947) <- bulk.mtx_GPL6947$Group.1 # Los nombres de genes únicos sin duplicados sir
bulk.mtx_GPL6102 <- bulk.mtx_GPL6102[, -c(1:2)] # Elimino las columnas usadas para conseguir los nombres
bulk.mtx_GPL6947 <- bulk.mtx_GPL6947[, -c(1:2)] # Elimino las columnas usadas para conseguir los nombres

# Convertir los datos de expresión del bulk RNA-seq a objeto ExpressionSet:
bulk.eset_GPL6102 <- Biobase::ExpressionSet(assayData = as.matrix(as.data.frame(bulk.mtx_GPL6102)))
bulk.eset_GPL6947 <- Biobase::ExpressionSet(assayData = as.matrix(as.data.frame(bulk.mtx_GPL6947)))
print("Object associated to platform GPL6102:")

## [1] "Object associated to platform GPL6102:"
bulk.eset_GPL6102

## ExpressionSet (storageMode: lockedEnvironment)
## assayData: 16269 features, 57 samples
## element names: exprs
## protocolData: none
## phenoData: none
## featureData: none
## experimentData: use 'experimentData(object)'
## Annotation:

print("Object associated to platform GPL6947:")

## [1] "Object associated to platform GPL6947:"
bulk.eset_GPL6947

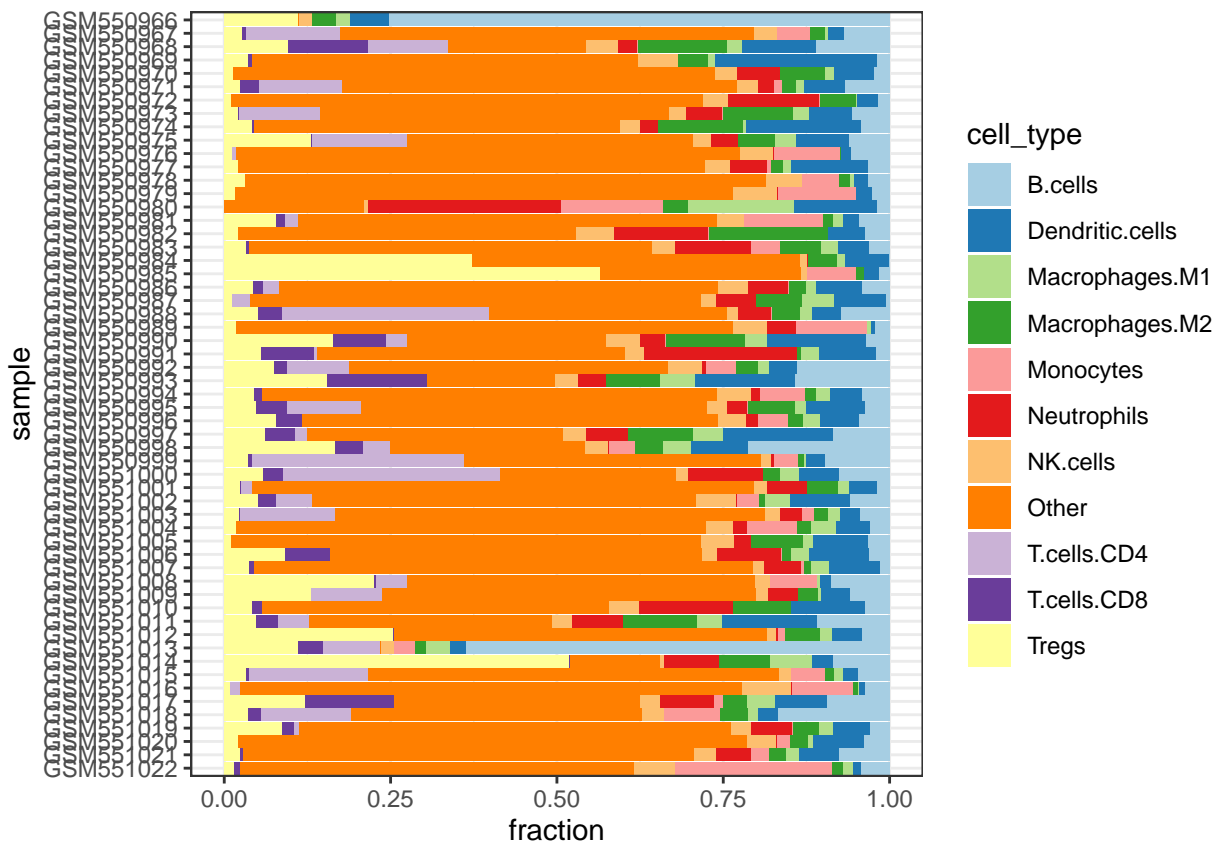
```

```
## ExpressionSet (storageMode: lockedEnvironment)
## assayData: 19130 features, 22 samples
##   element names: exprs
## protocolData: none
## phenoData: none
## featureData: none
## experimentData: use 'experimentData(object)'
## Annotation:

ti_racleGPL6102 <- quantiseqr::run_quantiseqr(
  expression_data = bulk.eset_GPL6102@assayData$exprs,
  signature_matrix = "TIL10",
  is_arraydata = FALSE,
  is_tumordata = TRUE,
  scale_mRNA = TRUE
)
ti_racleGPL6947 <- quantiseqr::run_quantiseqr(
  expression_data = bulk.eset_GPL6947@assayData$exprs,
  signature_matrix = "TIL10",
  is_arraydata = FALSE,
  is_tumordata = TRUE,
  scale_mRNA = TRUE
)
print("Plataforma GPL6102:")

## [1] "Plataforma GPL6102:"

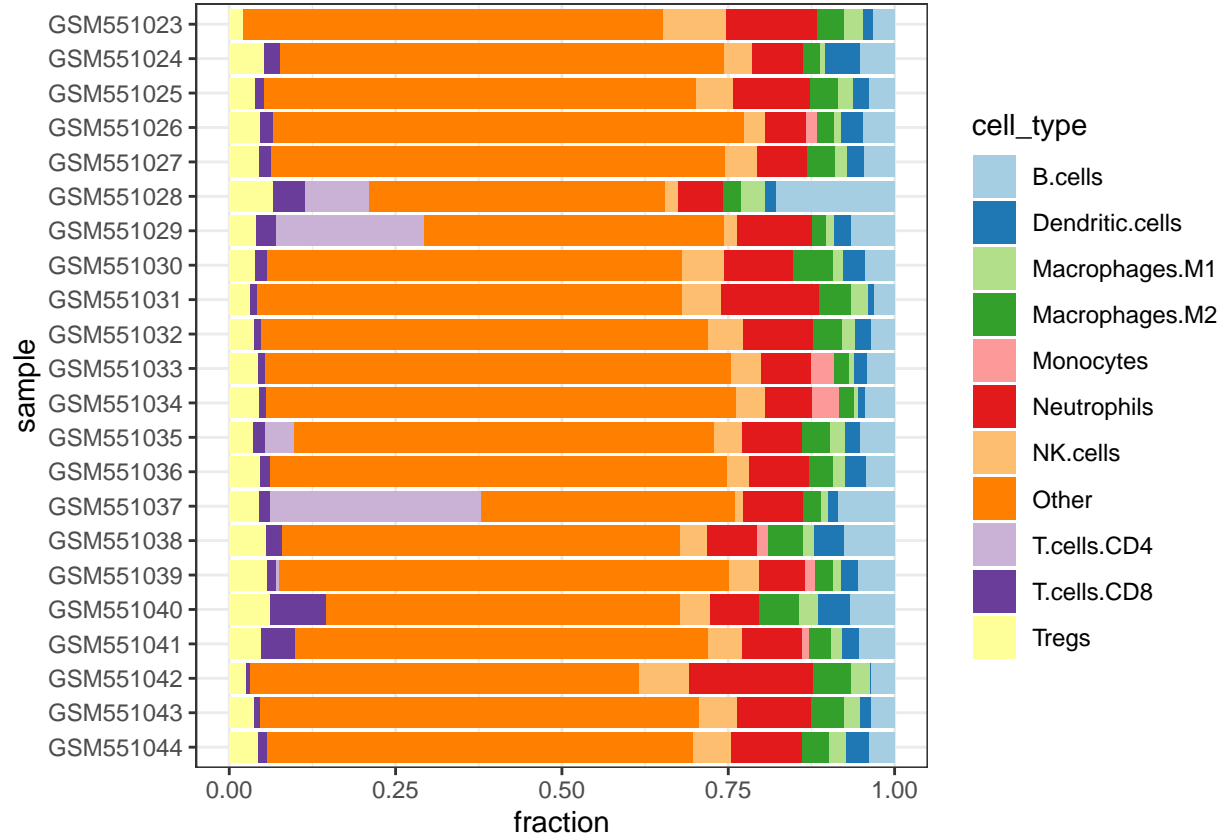
quantipLOT(ti_racleGPL6102)
```



```
print("Plataforma GPL6947:")
```

```
## [1] "Plataforma GPL6947:"
```

```
quantipLOT(ti_racleGPL6947)
```



Encontramos las proporciones del bulk RNA-seq en el apartado `ti_racle`, el cual puedo integrar en la metadata que ya tenía y almacenar en un archivo para posteriores análisis.

```
ref.based.estimates_GPL6102 <- as.data.frame(ti_racleGPL6102)
ref.based.estimates_GPL6947 <- as.data.frame(ti_racleGPL6947)
ref.based.estimates_GPL6102$geo_accession <- rownames(ref.based.estimates_GPL6102)
ref.based.estimates_GPL6947$geo_accession <- rownames(ref.based.estimates_GPL6947)
ref.based.estimates_GPL6102 <- inner_join(ref.based.estimates_GPL6102, bulk_metadata_GPL6102, by = "geo_accession")
ref.based.estimates_GPL6947 <- inner_join(ref.based.estimates_GPL6947, bulk_metadata_GPL6947, by = "geo_accession")
knitr::kable(head(ref.based.estimates_GPL6102[,1:7]), digits=2, caption = "Sección de las primeras muestras")
```

Table 1: Sección de las primeras muestras como ejemplo del resultado con la plataforma GPL6102

| Sample    | B.cells | Macrophages.M1 | Macrophages.M2 | Monocytes | Neutrophils | NK.cells |
|-----------|---------|----------------|----------------|-----------|-------------|----------|
| GSM550966 | 0.75    | 0.02           | 0.04           | 0.00      | 0.00        | 0.02     |
| GSM550967 | 0.07    | 0.01           | 0.02           | 0.05      | 0.00        | 0.04     |
| GSM550968 | 0.11    | 0.02           | 0.14           | 0.00      | 0.03        | 0.05     |
| GSM550969 | 0.02    | 0.01           | 0.05           | 0.00      | 0.00        | 0.06     |
| GSM550970 | 0.02    | 0.01           | 0.07           | 0.00      | 0.06        | 0.03     |
| GSM550971 | 0.07    | 0.01           | 0.02           | 0.01      | 0.02        | 0.03     |

```
knitr::kable(head(ref.based.estimates_GPL6947[,1:7]), digits=2, caption = "Sección de las primeras mues
```

Table 2: Sección de las primeras muestras como ejemplo del resultado con la plataforma GPL6947

| Sample    | B.cells | Macrophages.M1 | Macrophages.M2 | Monocytes | Neutrophils | NK.cells |
|-----------|---------|----------------|----------------|-----------|-------------|----------|
| GSM551023 | 0.03    | 0.03           | 0.04           | 0.00      | 0.14        | 0.09     |
| GSM551024 | 0.05    | 0.01           | 0.03           | 0.00      | 0.08        | 0.04     |
| GSM551025 | 0.04    | 0.02           | 0.04           | 0.00      | 0.12        | 0.05     |
| GSM551026 | 0.05    | 0.01           | 0.03           | 0.02      | 0.06        | 0.03     |
| GSM551027 | 0.05    | 0.02           | 0.04           | 0.00      | 0.08        | 0.05     |
| GSM551028 | 0.18    | 0.04           | 0.03           | 0.00      | 0.07        | 0.02     |

```
write.csv(ref.based.estimates_GPL6102,"./quantiseqr_GSE22155_GPL6102.csv", row.names = FALSE)
write.csv(ref.based.estimates_GPL6947,"./quantiseqr_GSE22155_GPL6947.csv", row.names = FALSE)
```