

Pathway Analysis - Macrophages M1

Input data: DEGs

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1 Introducción y Objetivo

En este script realizo un pathways análisis con los DEGs obtenidos de los Venn Diagrams, y preparo el resultado en archivos gem para extraer un gráfico posteriormente en Cytoscape (<https://cran.r-project.org/web/packages/gprofiler2/vignettes/gprofiler2.html>).

2 Paquetes y datos

3 Datos

Cargo inicialmente los listados de Macrófagos tipo M1 obtenidos en jVenn.

```
setwd("~/Desktop/ELENA_UOC/TFM")
jVenn_MacrophagesM1_vs_GSE22155_02 <- read_csv("DEGs/Venn_diagrams_cell_type_treatments_vs_untreated/jVenn_MacrophagesM1_vs_GSE22155_02.csv")
head(jVenn_MacrophagesM1_vs_GSE22155_02) # sección de los resultados
```

```
## # A tibble: 6 x 3
##   GSE50509 GSE22155_02 `GSE50509|GSE22155_02`
##   <chr>    <chr>    <chr>
## 1 LIF     C5AR1     HAVCR2
## 2 MIR155HG CLEC7A    HCK
## 3 PTPRE    KCNJ15    RAB31
## 4 SOD2     OSM       CD163
## 5 SRSF10   APOBR     ITGAX
## 6 SH2B3    PILRA     Clorf162
```

```
jVenn_MacrophagesM1_vs_GSE22155_47 <- read_csv("DEGs/Venn_diagrams_cell_type_treatments_vs_untreated/jVenn_MacrophagesM1_vs_GSE22155_47.csv")
jVenn_MacrophagesM1_vs_tcgaskcm <- read_csv("DEGs/Venn_diagrams_cell_type_treatments_vs_untreated/jVenn_MacrophagesM1_vs_tcgaskcm.csv")
jVenn_MacrophagesM1_vs_GSE91061 <- read_csv("DEGs/Venn_diagrams_cell_type_treatments_vs_untreated/jVenn_MacrophagesM1_vs_GSE91061.csv")
jVenn_MacrophagesM1_vs_GSE35640 <- read_csv("DEGs/Venn_diagrams_cell_type_treatments_vs_untreated/jVenn_MacrophagesM1_vs_GSE35640.csv")
jVenn_MacrophagesM1_vs_GSE61992 <- read_csv("DEGs/Venn_diagrams_cell_type_treatments_vs_untreated/jVenn_MacrophagesM1_vs_GSE61992.csv")
setwd("~/Desktop/ELENA_UOC/TFM/GOBPs") # Directorio de trabajo
jVenn_MacrophagesM1 <- read_delim("~/Desktop/ELENA_UOC/TFM/DEGs/Venn_diagrams_cell_type_treatments_comparison.csv",
delim = ",", escape_double = FALSE, trim_ws = TRUE)
```

4 Uncovered

En esta sección contemplo la posibilidad que los DEGs únicos en el dataset sin tratar se traten de mecanismos que se escapan al tratamiento pero que igualmente se ven afectados por la enfermedad.

Primero realizo el pathway multianálisis:

En esta sección contemplo la posibilidad que los DEGs únicos en el dataset sin tratar se traten de mecanismos que se escapan al tratamiento pero que igualmente se ven afectados por la enfermedad.

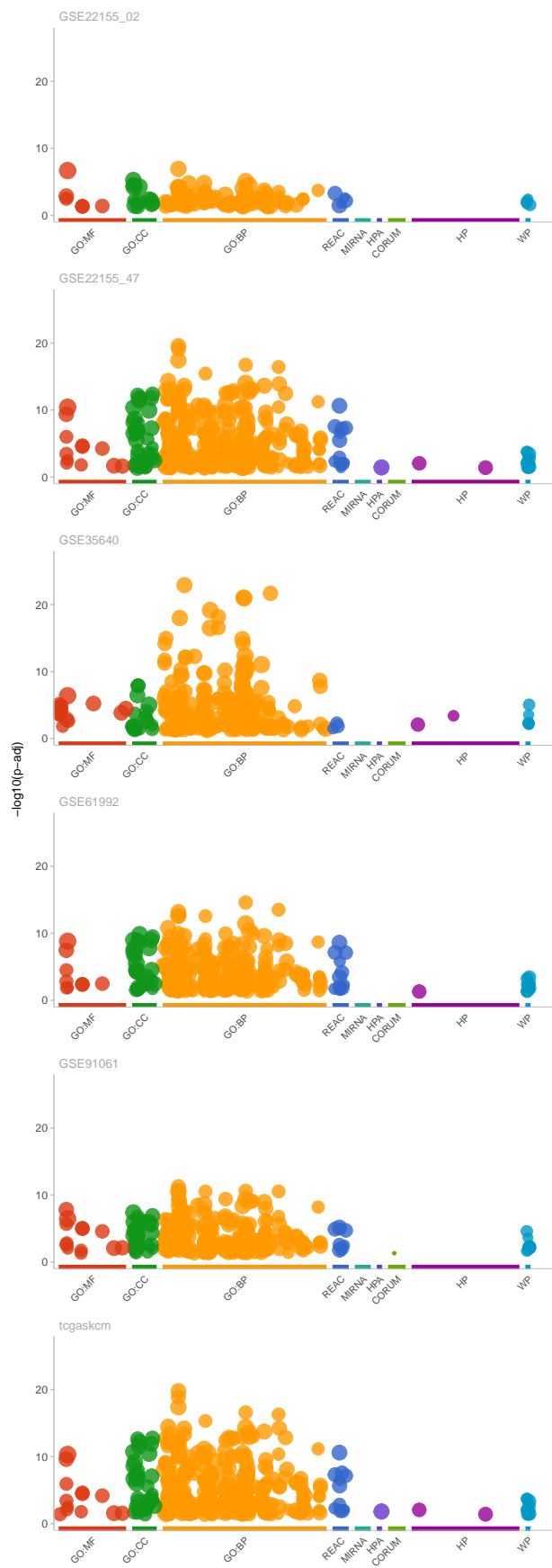
Primero realizo el pathway multianálisis:

```
multi_gostres <- gost(query = list("GSE22155_02" = unique(jVenn_MacrophagesM1_vs_GSE22155_02$GSE50509[which(is.na(jVenn_MacrophagesM1_vs_GSE22155_02$GSE50509) == FALSE])),
"GSE22155_47" = jVenn_MacrophagesM1_vs_GSE22155_47$GSE50509[which(is.na(jVenn_MacrophagesM1_vs_GSE22155_47$GSE50509) == FALSE)],
"GSE35640" = jVenn_MacrophagesM1_vs_GSE35640$GSE50509[which(is.na(jVenn_MacrophagesM1_vs_GSE35640$GSE50509) == FALSE)],
"GSE91061" = jVenn_MacrophagesM1_vs_GSE91061$GSE50509[which(is.na(jVenn_MacrophagesM1_vs_GSE91061$GSE50509) == FALSE)],
"GSE61992" = jVenn_MacrophagesM1_vs_GSE61992$GSE50509[which(is.na(jVenn_MacrophagesM1_vs_GSE61992$GSE50509) == FALSE)],
"tcgaskcm" = jVenn_MacrophagesM1_vs_tcgaskcm$GSE50509[which(is.na(jVenn_MacrophagesM1_vs_tcgaskcm$GSE50509) == FALSE)]),
evcodes = TRUE, multi_query = FALSE,
sources = c("GO", "REAC", "MIRNA", "CORUM", "HP", "HPA", "WP"))
head(multi_gostres$result[,1:7])
```

##	query	significant	p_value	term_size	query_size	intersection_size
## 1	GSE22155_02	TRUE	1.164604e-07	3855	320	107
## 2	GSE22155_02	TRUE	8.748051e-06	8976	320	188
## 3	GSE22155_02	TRUE	1.455333e-05	3437	320	93
## 4	GSE22155_02	TRUE	1.589636e-05	3443	320	93
## 5	GSE22155_02	TRUE	3.189032e-05	7394	320	161
## 6	GSE22155_02	TRUE	6.050687e-05	1791	320	58

```
## precision
## 1 0.334375
## 2 0.587500
## 3 0.290625
## 4 0.290625
## 5 0.503125
## 6 0.181250
```

```
p <- gostplot(multi_gostres, capped = FALSE, interactive = F)
p
```



Estos resultados son los que utilizo para generar los archivos gem que archivo para usar más tarde en Cytoscape.

```
# format to GEM
gem <- multi_gostres$result[,c("query", "term_id", "term_name", "p_value", "intersection_size")]
colnames(gem) <- c("query", "GO.ID", "Description", "p.Val", "Genes")
gem$FDR <- gem$p.Val
gem$Phenotype = "+1"

gem %>% group_by(query) %>%
  group_walk(~
    write.table(data.frame(.x[,c("GO.ID", "Description", "p.Val", "FDR", "Phenotype", "Genes")]),
      file = paste0("./Venn_diagrams_cell_type_treatments_vs_untreated/gProfiler_MacrophagesM",
        sep = "\t", quote = F, row.names = F))
```

Proporciones de los pathways para cada estudio:

```
df <- as.data.frame(multi_gostres$result[,1:13])
table(df$query)

##
## GSE22155_02 GSE22155_47 GSE35640 GSE61992 GSE91061 tcgaskcm
## 122 424 295 303 265 422

prop.table(table(df$query))*100

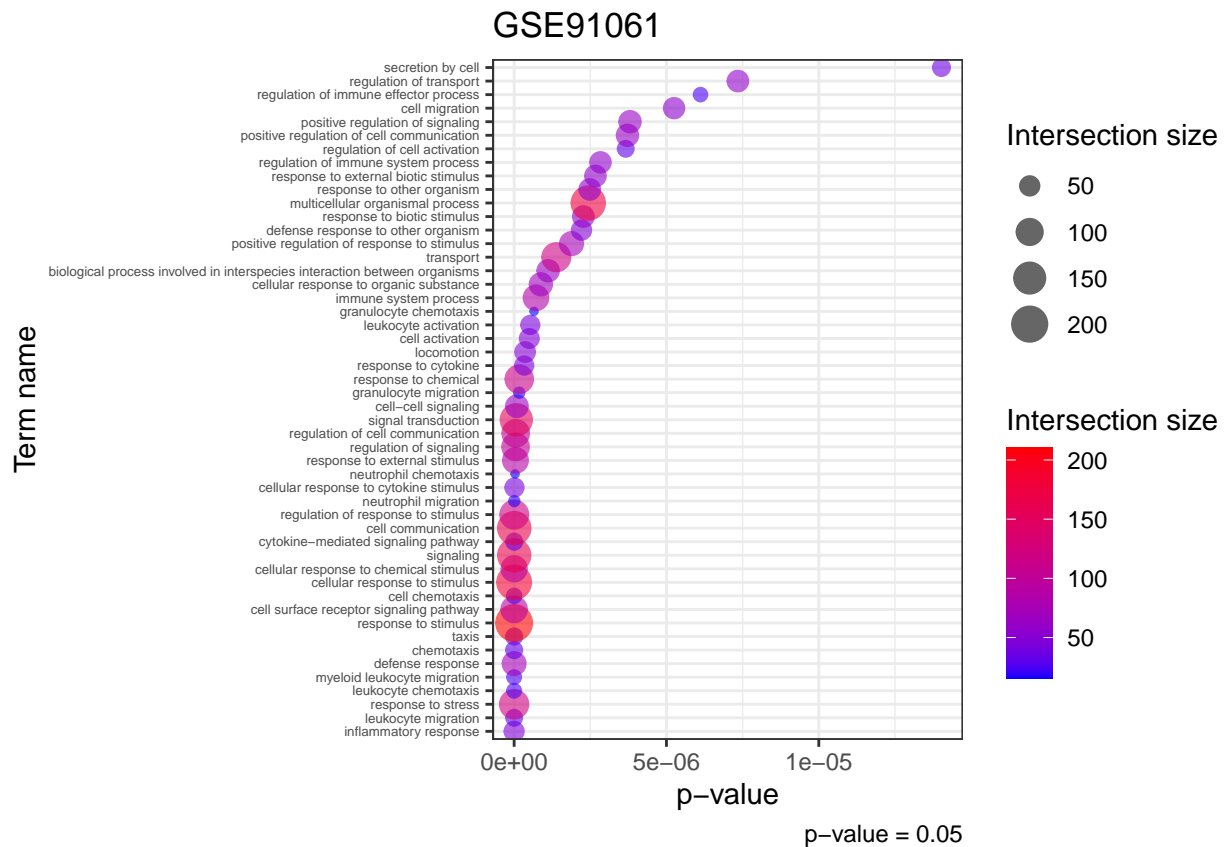
##
## GSE22155_02 GSE22155_47 GSE35640 GSE61992 GSE91061 tcgaskcm
## 6.663026 23.156745 16.111415 16.548334 14.472966 23.047515

#table(df$term_name)
#prop.table(table(df$term_name))*100
df2 <- as.data.frame(table(df$term_name, df$query))
#length(unique(df2$Var1))
write.table(df, file = "./Venn_diagrams_cell_type_treatments_vs_untreated/MacrM1_GOBP.txt", sep = "\t")
write.table(df2, file = "./Venn_diagrams_cell_type_treatments_vs_untreated/MacrM1_GOBP_freq.txt", sep = "\t")
#rm(jVenn_MacrophagesM1_vs_GSE22155_02, jVenn_MacrophagesM1_vs_GSE22155_47, jVenn_MacrophagesM1_vs_GSE91061)
```

Barplot of the top GO-BPs:

```
plot_gobps <- function(study, n = 50){
df2 <- df[df$source == "GO:BP",]
df2_GSE91061 <- df2[df2$query == study,]
df2_GSE91061 <- df2_GSE91061[1:n,]
ggplot(df2_GSE91061, aes(p_value, reorder(term_name,p_value))) + ggtitle(study) +
  geom_point(aes(p_value, reorder(term_name,p_value), colour = intersection_size, size = intersection_size)) +
  theme_bw() + theme(axis.text.y.left = element_text(size = 5)) +
  labs(x= "p-value", y="Term name", colour = "Intersection size", size = "Intersection size", caption = "Intersection size")
  scale_color_gradient(low="blue", high="red")
}

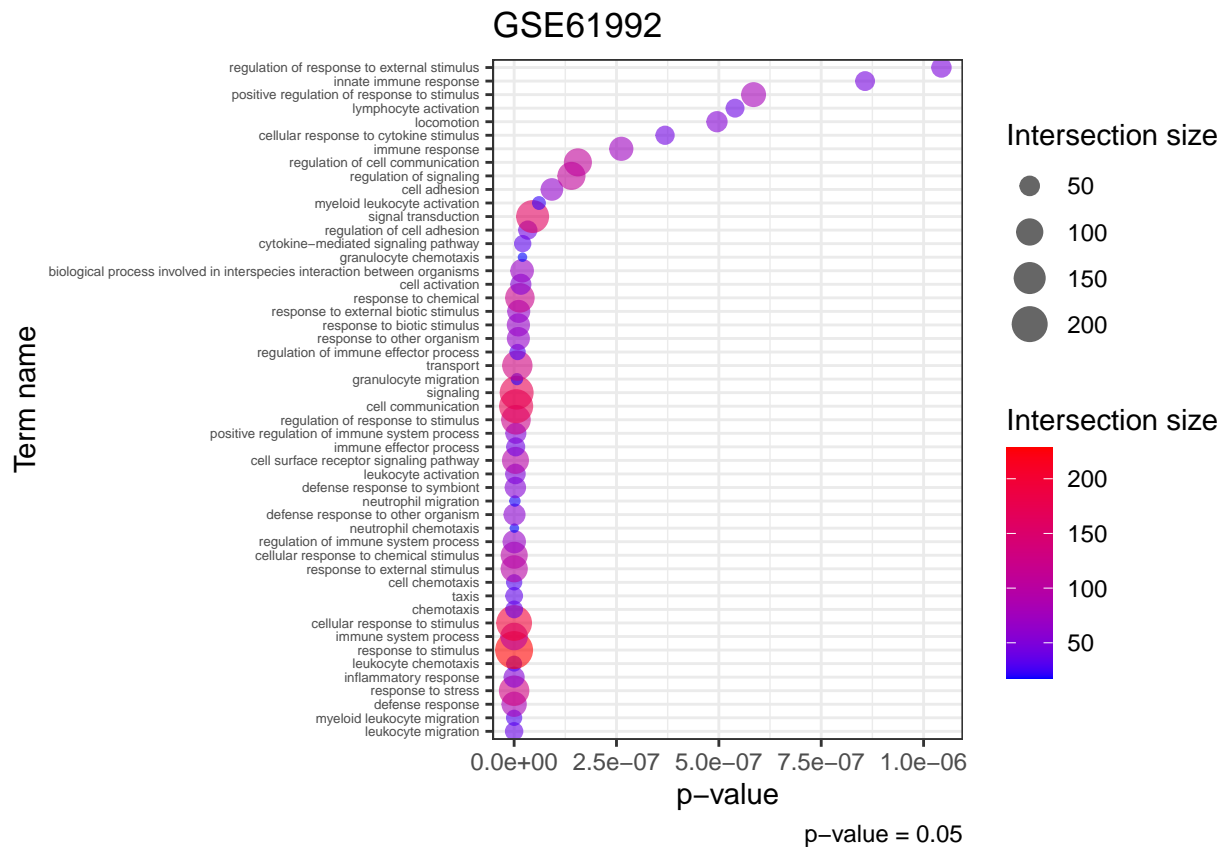
plot_gobps(study = "GSE91061")
```



```
print("\n")
```

```
## [1] "\n"
```

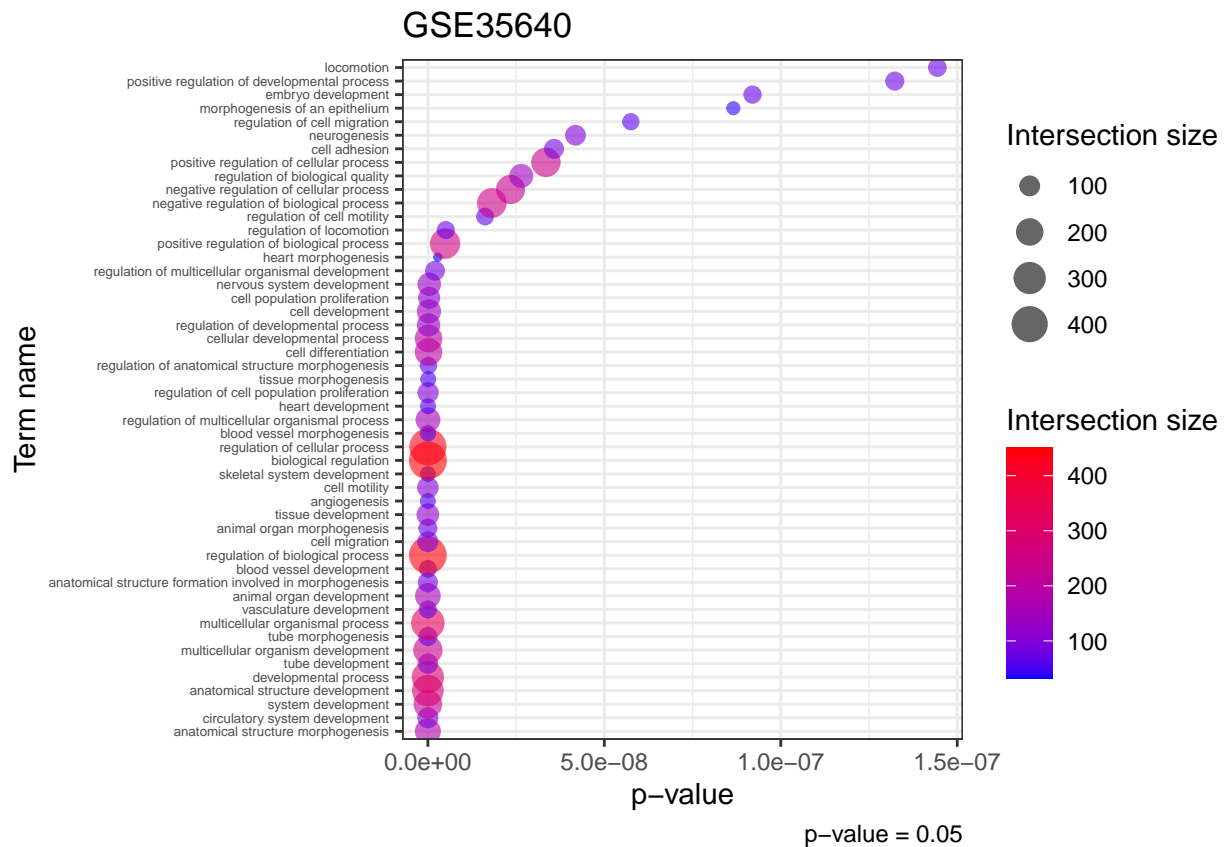
```
plot_gobps(study = "GSE61992")
```



```
print("\n")
```

```
## [1] "\n"
```

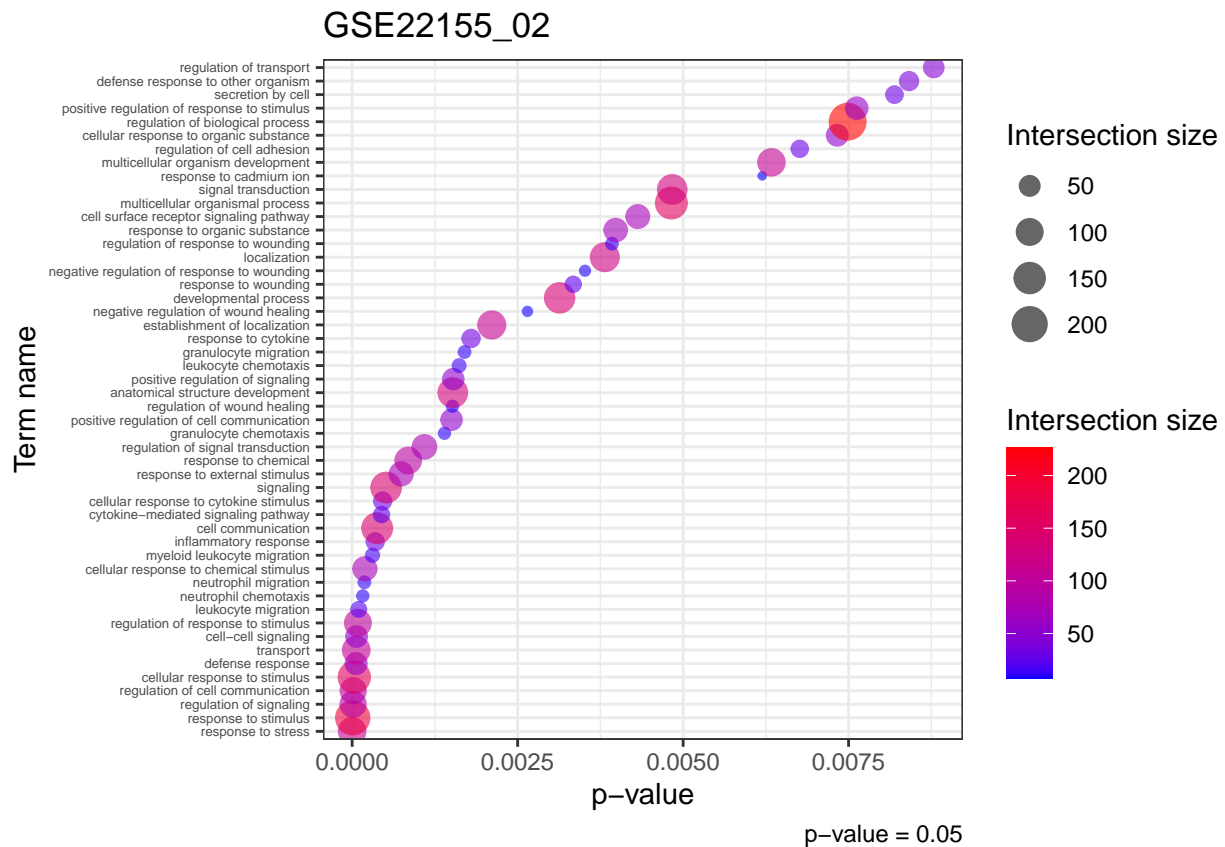
```
plot_gobps(study = "GSE35640")
```



```
print("\n")
```

```
## [1] "\n"
```

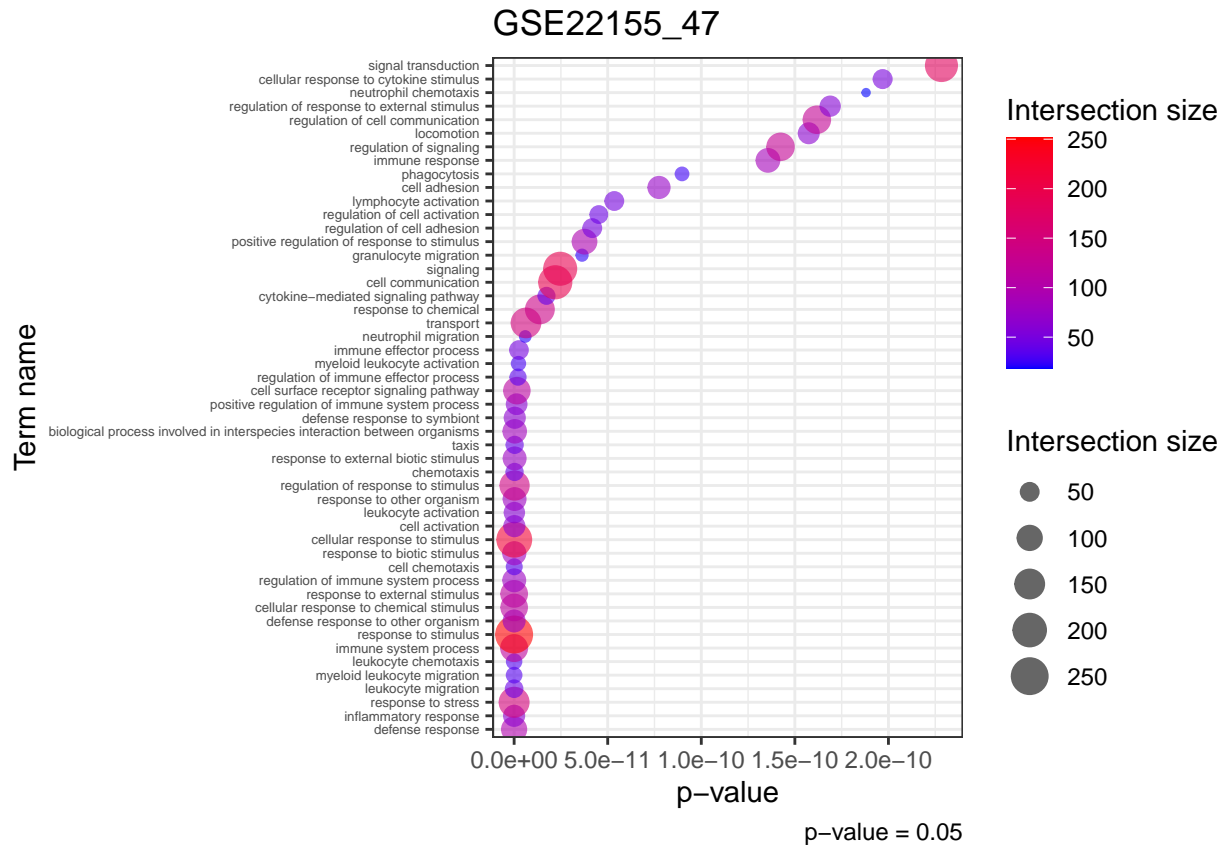
```
plot_gobps(study = "GSE22155_02")
```



```
print("\n")
```

```
## [1] "\n"
```

```
plot_gobps(study = "GSE22155_47")
```

5 Exclusive

En esta sección analizo los DEGs exclusivos para cada estudio, y realizo un multi-análisis para cada estudio. Los resultados los guardo como archivos gem específicos para usar en Cytoscape.

Primero realizo el pathway multianálisis:

```
multi_gostres <- gost(query = list("GSE61992" = jVenn_MacrophagesM1$GSE61992[which(is.na(jVenn_MacrophagesM1$GSE61992))],
                                   "GSE91061" = jVenn_MacrophagesM1$GSE91061[which(is.na(jVenn_MacrophagesM1$GSE91061))],
                                   "tcgaskcm" = jVenn_MacrophagesM1$TCGA-SKCM[which(is.na(jVenn_MacrophagesM1$TCGA-SKCM))]),
                      evcodes = TRUE, multi_query = FALSE,
                      sources = c("GO", "REAC", "MIRNA", "CORUM", "HP", "HPA", "WP"))
head(multi_gostres$result[,1:7])
```

##	query	significant	p_value	term_size	query_size	intersection_size
## 1	GSE61992	TRUE	4.773516e-02	69	20	3
## 2	GSE91061	TRUE	7.384767e-05	33	320	14
## 3	GSE91061	TRUE	5.343907e-04	20	320	10
## 4	GSE91061	TRUE	8.141399e-04	10	320	7
## 5	GSE91061	TRUE	8.141399e-04	10	320	7
## 6	GSE91061	TRUE	1.456767e-03	14	320	8
##	precision					
## 1	0.150000					
## 2	0.043750					
## 3	0.031250					
## 4	0.021875					

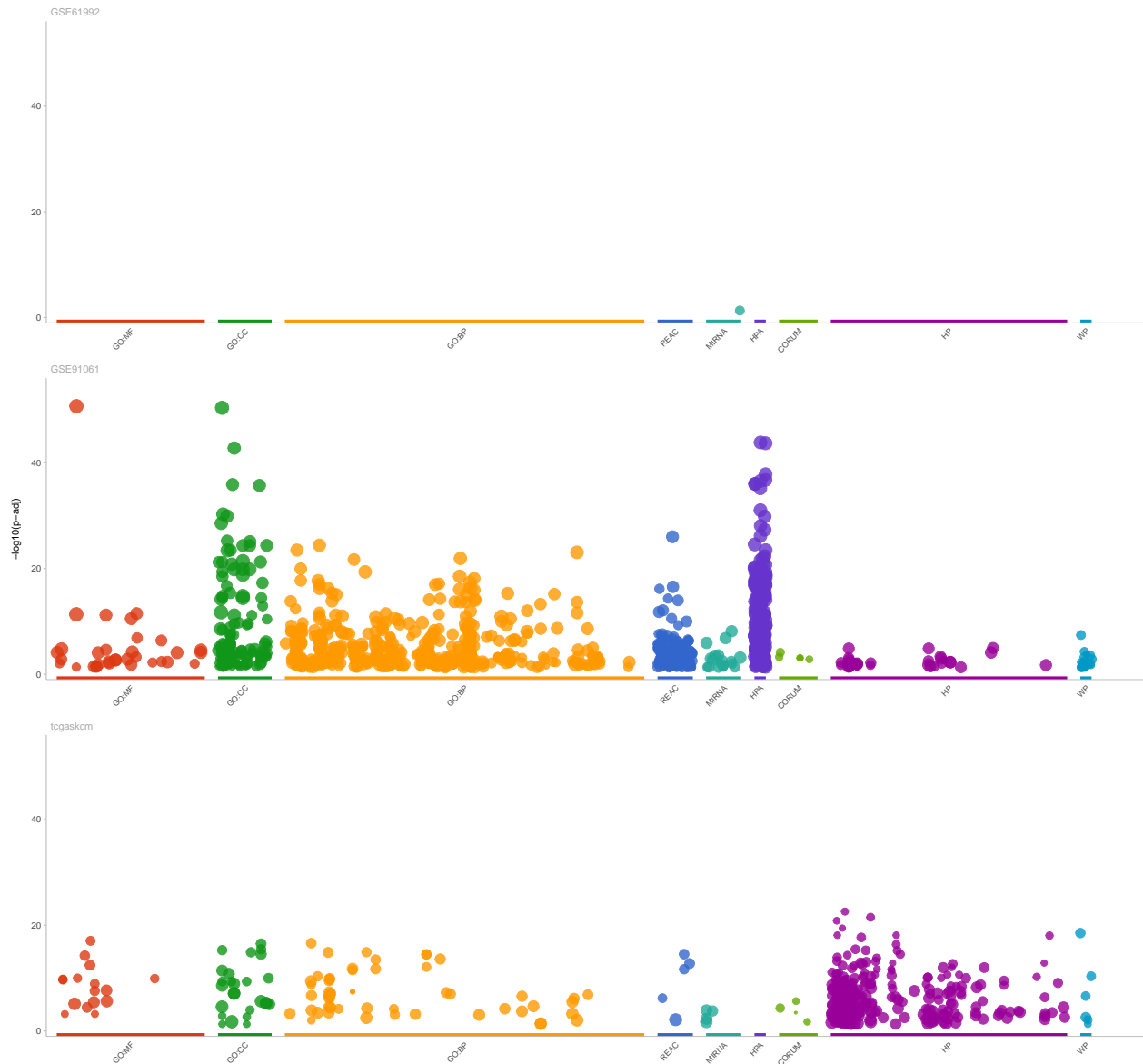
```
## 5 0.021875
## 6 0.025000
```

Estos resultados son los que utilizo para generar los archivos gem que archivo para usar más tarde en Cytoscape.

```
# format to GEM
# format to GEM
gem <- multi_gostres$result[,c("query", "term_id", "term_name", "p_value", "intersection_size")]
colnames(gem) <- c("query", "GO.ID", "Description", "p.Val", "Genes")
gem$FDR <- gem$p.Val
gem$Phenotype = "+1"

gem %>% group_by(query) %>%
  group_walk(~
    write.table(data.frame(.x[,c("GO.ID", "Description", "p.Val", "FDR", "Phenotype", "Genes")]),
      file = paste0("./Treatment_comparisons/gProfiler_MacrophagesM1_Only_geneset_", unique(.x$query)),
      sep = "\t", quote = F, row.names = F))

p <- gostplot(multi_gostres, capped = FALSE, interactive = F)
p
```



Proporciones de los pathways para cada estudio:

```
df <- as.data.frame(multi_gostres$result[,1:13])
table(df$query)
```

```
##
## GSE61992 GSE91061 tcgaskcm
##      1      1039      371
```

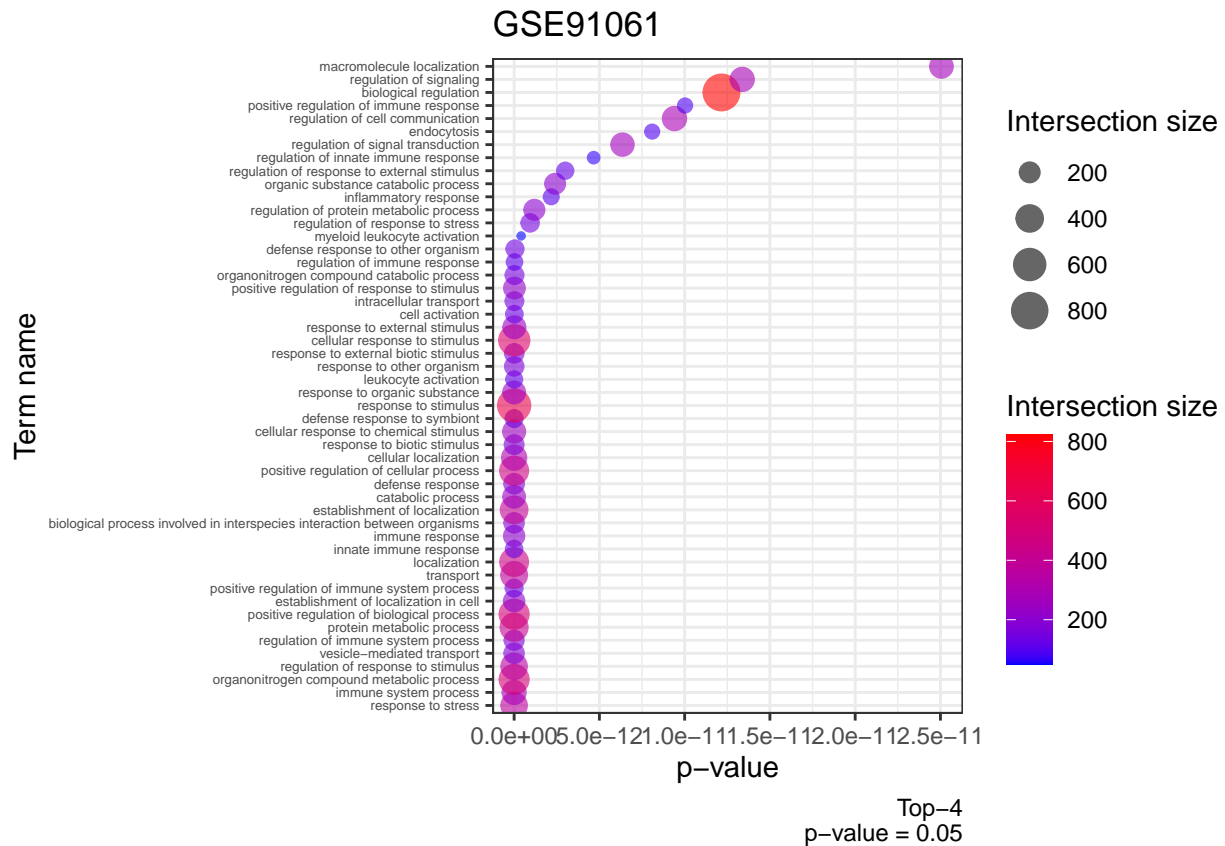
```
write.table(gem, file = "./Treatment_comparisons/gProfiler_MacrophagesM1_Only_genesets.txt", sep = "\t"
```

```
plot_gobps <- function(study){
df2_GSE91061 <- df2[df2$query == study,]
ggplot(df2_GSE91061, aes(p_value, reorder(term_name,p_value))) + ggtitle(study) +
  geom_point(aes(p_value, reorder(term_name,p_value), colour = intersection_size, size = intersection_size)) +
  theme_bw() + theme(axis.text.y.left = element_text(size = 5)) +
  labs(x= "p-value", y="Term name", colour = "Intersection size", size = "Intersection size", caption =
  scale_color_gradient(low="blue", high="red")
```

```

}
df2 <- df[df$source == "GO:BP",]
df2 <- df2[1:50,]
plot_gobps(study = "GSE91061")

```



```
print("\n")
```

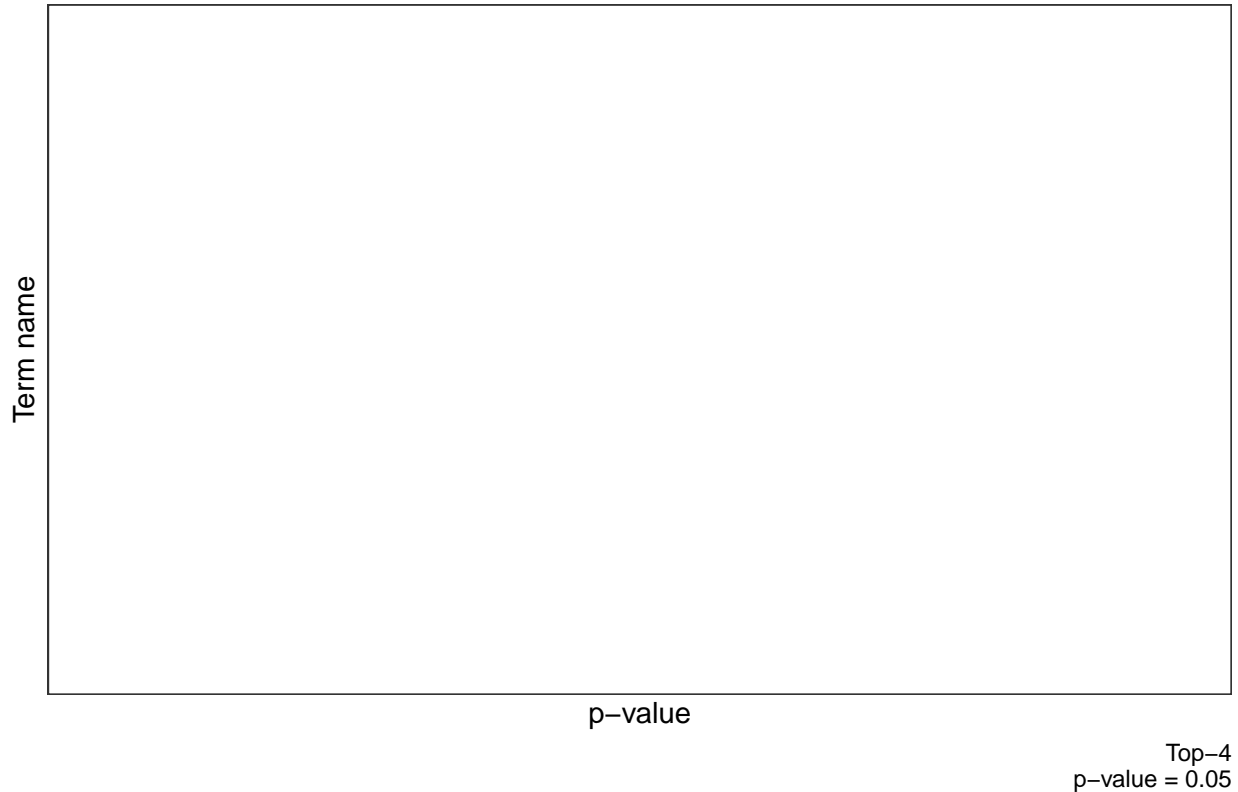
```
## [1] "\n"
```

```

df2 <- df[df$source == "GO:BP",]
plot_gobps(study = "GSE61992")

```

GSE61992



6 Máximo solapamiento

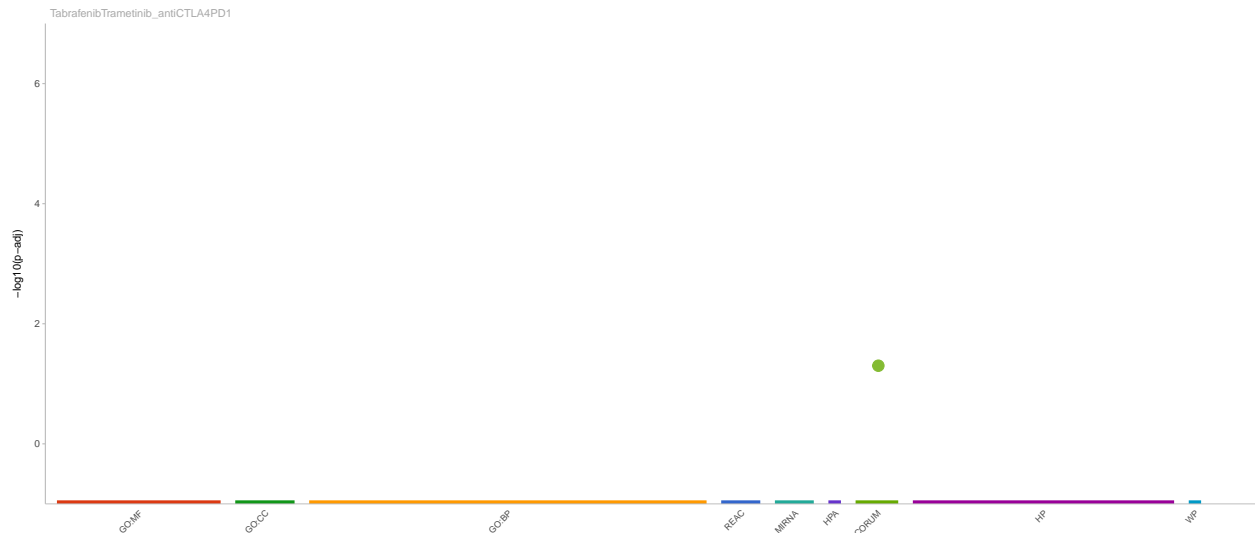
En esta sección analizo los DEGs que más se comparten entre los estudios para observar los mecanismos que pudieran haber en común entre los tratamientos.

Primero realizo el pathway multianálisis:

```
multi_gostres <- gost(query = list(
  "TabrafenibTrametinib_antiCTLA4PD1" = jVenn_MacrophagesM1$`GSE91061|GSE61992`[which(is.na(jVenn_MacrophagesM1$`GSE91061|GSE61992`))],
  evcodes = TRUE, multi_query = FALSE,
  sources = c("GO", "REAC", "MIRNA", "CORUM", "HP", "HPA", "WP"))
head(multi_gostres$result[,1:7])

##               query significant    p_value term_size query_size
## 1 TabrafenibTrametinib_antiCTLA4PD1      TRUE 0.04998799         1         3
##   intersection_size precision
## 1                   1 0.3333333

p <- gostplot(multi_gostres, capped = FALSE, interactive = F)
p
```



Estos resultados son los que utilizo para generar los archivos gem que archivo para usar más tarde en Cytoscape.

```
# format to GEM
# format to GEM
gem <- multi_gostres$result[,c("query", "term_id", "term_name", "p_value", "intersection_size")]
colnames(gem) <- c("query", "GO.ID", "Description", "p.Val", "Genes")
gem$FDR <- gem$p.Val
gem$Phenotype = "+1"

gem %>% group_by(query) %>%
  group_walk(~
    write.table(data.frame(.x[,c("GO.ID", "Description", "p.Val", "FDR", "Phenotype", "Genes")],
      file = paste0("./Treatment_comparisons/gProfiler_MacrophagesM1_maxOverlap_geneset_", query, ".txt"),
      sep = "\t", quote = F, row.names = F))
  )
```

Al mirar los procesos por cada comparación salieron demasiados para poder visualizarlos bien, por lo que decidí restringir a los procesos GO-BPs únicamente y guardarlos en pdf.

```
#unique(gem$query)

gem2 <- gem[grepl("GO",gem$GO.ID),]
plot_gobps <- function(query, cutoff = 0.05) {
  gem2 <- gem2[gem2$FDR <= cutoff,]
  gem2_1 <- gem2[gem2$query == query, ]
  ggplot(gem2_1, aes(FDR, Description)) + ggtitle(query) +
    geom_point(aes(FDR, Description, colour = Genes, size = Genes), alpha = 0.6) +
    theme_bw() + theme(axis.text.y.left = element_text(size = 5)) +
    labs(caption = paste0("FDR = ",cutoff))+
    scale_color_gradient(low="blue", high="red")
}

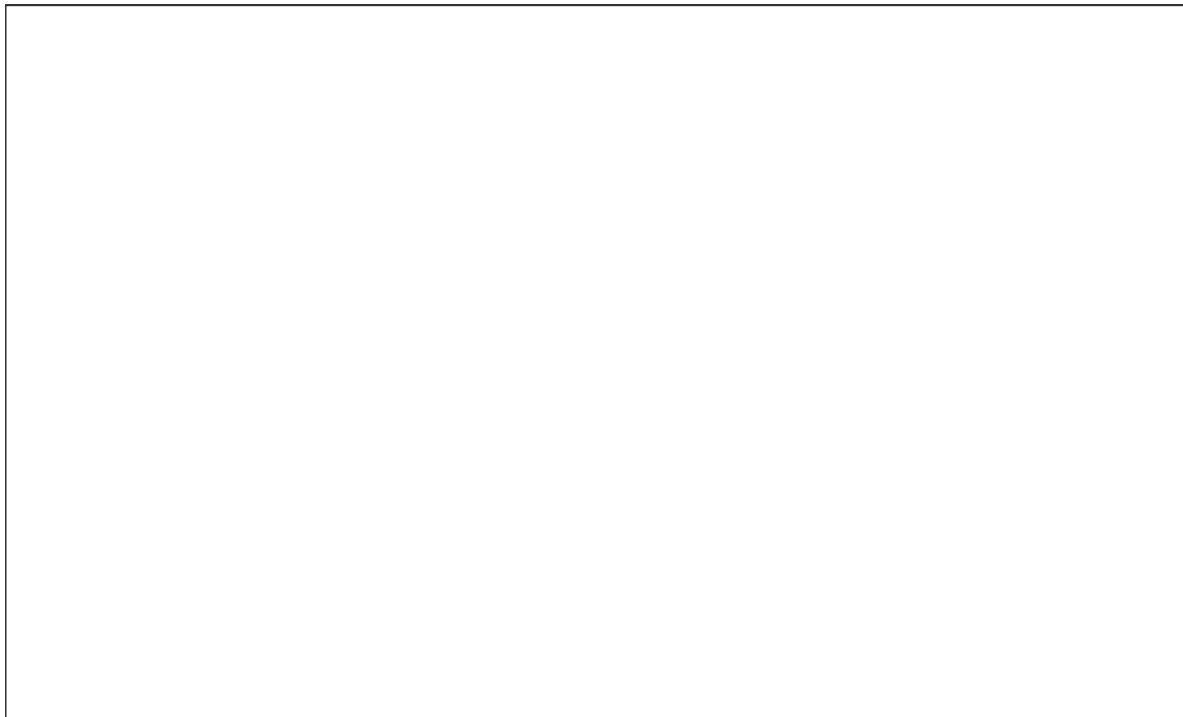
pdf(file = paste0("./Treatment_comparisons/gProfiler_MacrophagesM1_maxOverlap_geneset_TabrafenibTrametinib_antiCTLA4PD1.pdf"))
plot_gobps("TabrafenibTrametinib_antiCTLA4PD1")
dev.off()
```

```
## pdf
## 2
```

```
plot_gobps("TabrafenibTrametinib_antiCTLA4PD1")
```

TabrafenibTrametinib_antiCTLA4PD1

Description



FDR

FDR = 0.05

```
df <- as.data.frame(multi_gostres$result[,1:13])
df2 <- df[df$source == "GO:BP",]
plot_gobps <- function(study){
df2_GSE91061 <- df2[df2$query == study,]
ggplot(df2_GSE91061, aes(p_value, reorder(term_name,p_value))) + ggtitle(study) +
  geom_point(aes(p_value, reorder(term_name,p_value), colour = intersection_size, size = intersection_size)) +
  theme_bw() + theme(axis.text.y.left = element_text(size = 5)) +
  labs(x= "p-value", y="Term name", colour = "Intersection size", size = "Intersection size", caption = "Intersection size")
  scale_color_gradient(low="blue", high="red")
}
plot_gobps(study = "TabrafenibTrametinib_antiCTLA4PD1")
```

TabrafenibTrametinib_antiCTLA4PD1

Term name

p-value

p-value = 0.05