# Background

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# 1 Why R?

#### 1.1 What is R?

R is a number of things, simultaneously. Depending on who is being asked, R is:

- A software package
- A programming language
- A toolkit for developing statistical and analytical tools
- An extensive library of statistical and mathematical software and algorithms
- A scripting language
- much, much more

# 1.2 Why use R?

- R is cross-platform and runs on Windows, Mac, and Linux (as well as more obscure systems).
- R provides a vast number of useful statistical tools, many of which have been painstakingly tested.
- R produces publication-quality graphics in a variety of formats.
- R plays well with FORTRAN, C, and scripts in many languages.
- R scales, making it useful for small and large projects. It is NOT Excel.
- R does not have a meaningfully useful graphical user interface (GUI).

I can develop code for analysis on my Mac laptop. I can then install the *same* code on our 20k core cluster and run it in parallel on 100 samples, monitor the process, and then update a database (for example) with R when complete.

# 1.3 Why not use R?

- R cannot do everything.
- R is not always the "best" tool for the job.
- R will not hold your hand. Often, it will slap your hand instead.
- The documentation can be opaque (but there is documentation).
- R can drive you crazy (on a good day) or age you prematurely (on a bad one).
- Finding the right package to do the job you want to do can be challenging; worse, some contributed packages are unreliable.]{}
- R does not have a meaningfully useful graphical user interface (GUI).

# 1.4 R License and the Open Source Ideal

R is free (yes, totally free!) and distributed under GNU license. In particular, this license allows one to:

- Download the source code
- Modify the source code to your heart's content
- Distribute the modified source code and even charge money for it, but you must distribute the modified source code under the original GNU license]{}

This license means that R will always be available, will always be open source, and can grow organically without constraint.

# 2 R Mechanics

# 2.1 Installing R

The home page for R is called the Comprehensive R Archive Network (CRAN). The website is not pretty (see figure 1), but it has quite a bit of information on it. It is not the best place to find help on R, although it is one of the best places to get R-related software, tools, and updates.

```
knitr::include_graphics('images/CRAN-screenshot.png')
```

Detailed installation instructions are readily available, but here are abbreviated instructions for convenience.

#### 2.1.1 Windows

NOTE: See Windows installation instructions for more detail. Install R and RStudio as regular users.

To install R, visit the Windows base distribution page. Click on the Download R-3.4.0 for Windows link (or use the latest version available). Click on the installer and make the default selection for each option.

To install *RStudio*, visit the RStudio download page. Click on the current RStudio release for Windows link. Click on the installer and follow default instructions.

#### 2.1.2 Mac

NOTE: See R for Mac OS X for more detail.

To install R, visit the R for Mac OS X. Click on the R-3.4.0.pkg link (or use the latest version available). Click on the installer and follow default instructions.

To install *RStudio*, visit the RStudio download page. Click on the current RStudio release for Windows link. Click on the installer and follow default instructions.

#### 2.1.3 Linux

NOTE: See distribution-specific instructions for additional detail.

On debian-based systems, the easiest way to install R is through a package manager manager, run under an administrator account. On Linux one usually needs to install R packages from source, and R package source often contains C, C++, or Fortran code requiring a compiler and  $\neg dev$  versions of various system libraries. It is therefore convenient to install the  $\neg dev$  version of R.

sudo apt-get install r-base r-base-dev

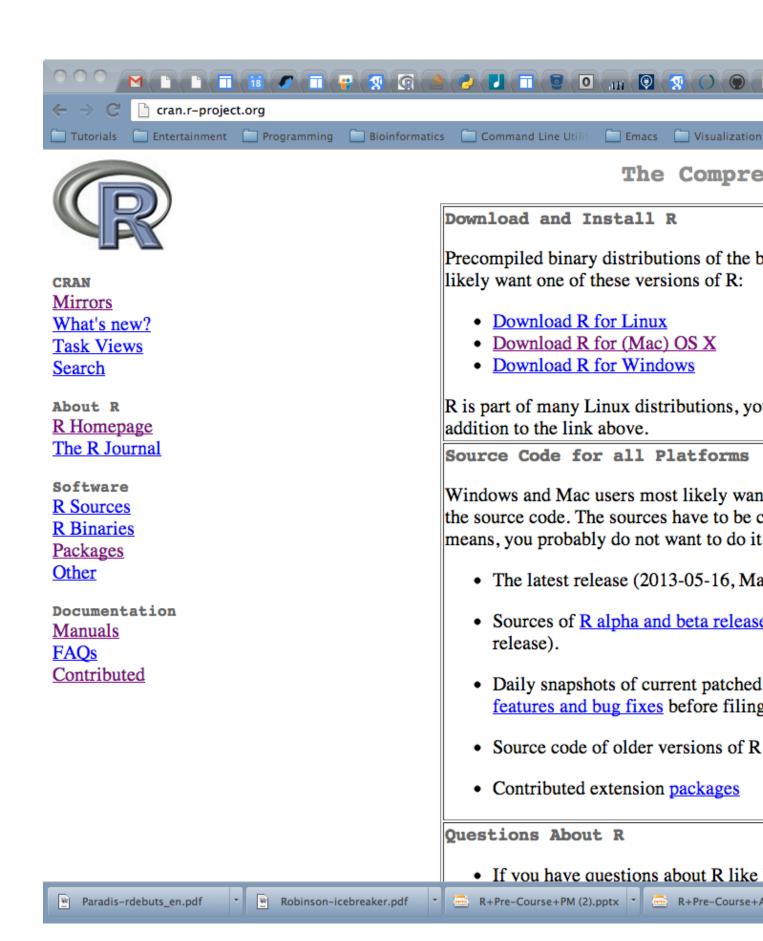


Figure 1: The Comprehensive R Archive Network (CRAN) website

When installing source packages, it may be necessary to have access to the <code>-dev</code> version of various system libraries. Many of these are installed as dependencies of <code>r-base-dev</code>; other common examples include the xml and curl libraries

```
sudo apt-get install libxml2-dev
sudo apt-get install libcurl-dev
```

Note in particular the use specification of libraries (the lib prefix) and the use of the -dev version.

To install *RStudio*, visit the RStudio download page. Download the appropriate archive for your OS. On Ubuntu, install the .deb installer with

```
sudo dpkg -i rstudio-1.0.136-amd64.deb
```

# 2.2 Starting R

How to start R depends a bit on the operating system (Mac, Windows, Linux) and interface. In this course, we will largely be using an Integrated Development Environment (IDE) called *RStudio*, but there is nothing to prohibit using R at the command line or in some other interface (and there are a few). A screenshot of the interface is shown in figure 2.

# 3 First steps

#### 3.1 Interacting with R

The only meaningful way of interacting with R is by typing into the R console. At the most basic level, anything that we type at the command line will fall into one of two categories:

1. Assignments

```
x = 1
y \leftarrow 2
```

2. Expressions

```
1 + pi + sin(42)
```

```
## [1] 3.225071
```

The assignment type is obvious because either the The "<-" or "=" are used. Note that when we type expressions, R will return a result. In this case, the result of R evaluating 1 + pi + sin(42) is 3.2250711.

The standard R prompt is a ">" sign. When present, R is waiting for the next expression or assignment. If a line is not a complete R command, R will continue the next line with a "+". For example, typing the fillowing with a "Return" after the second "+" will result in R giving back a "+" on the next line, a prompt to keep typing.

```
1 + pi + sin(3.7)
```

```
## [1] 3.611757
```

#### 3.2 Rules for Names in R

R allows users to assign names to objects such as variables, functions, and even dimensions of data. However, these names must follow a few rules.

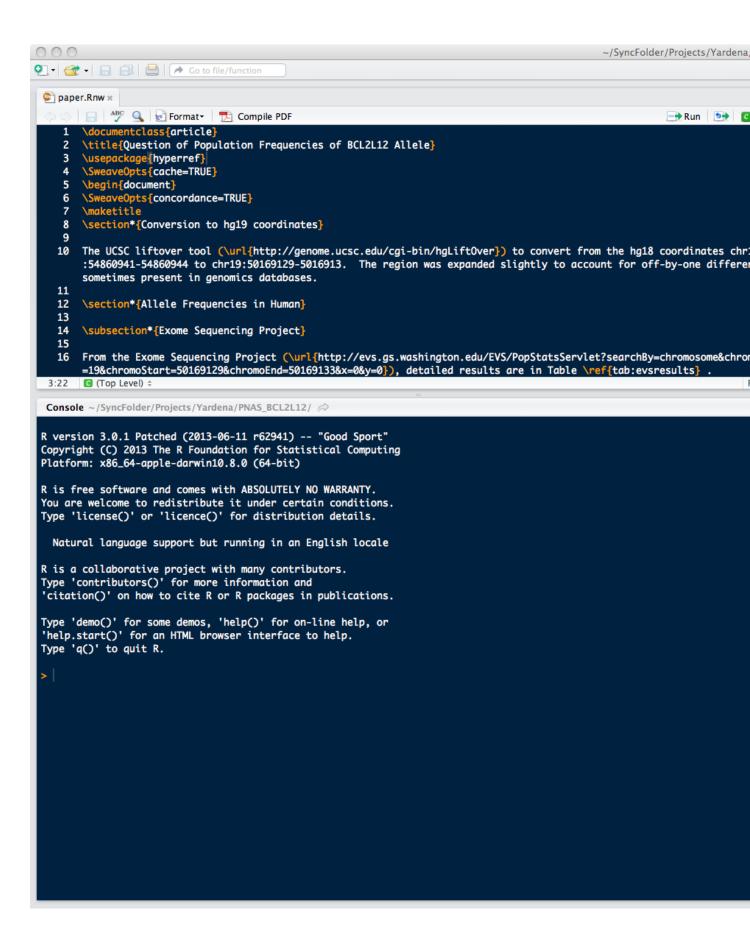


Figure 2: The Rstudio interface

- Names may contain any combination of letters, numbers, underscore, and "."
- Names may not start with numbers, underscore.
- R names are case-sensitive.

Examples of valid R names include:

```
pi
x
camelCaps
my_stuff
MY_Stuff
this.is.the.name.of.the.man
ABC123
abc1234asdf
.hi
```

# 3.3 Resources for Getting Help

There is extensive built-in help and documentation within R.

If the name of the function or object on which help is sought is known, the following approaches with the name of the function or object will be helpful. For a concrete example, examine the help for the print method.

```
help(print)
help('print')
?print
```

If the name of the function or object on which help is sought is not known, the following from within R will be helpful.

```
help.search('microarray')
RSiteSearch('microarray')
```

There are also tons of online resources that Google will include in searches if online searching feels more appropriate.

I strongly recommend using help(newfunction) for all functions that are new or unfamiliar to you.

#### 4 Introduction to R data structures

As in many programming languages, understanding how data are stored and manipulated is important to getting the most out of the experience. In these next few sections, we will introduce some basic R data types and structures as well as some general approaches for working with them.

#### 4.1 Vectors

In R, even a single value is a vector with length=1.

```
z = 1
z
## [1] 1
length(z)
```

#### ## [1] 1

In the code above, we "assigned" the value 1 to the variable named z. Typing z by itself is an "expression" that returns a result which is, in this case, the value that we just assigned. The length method takes an R object and returns the R length. There are numerous ways of asking R about what an object represents, and length is one of them.

Vectors can contain numbers, strings (character data), or logical values (TRUE and FALSE) or other "atomic" data types (table 1). Vectors cannot contain a mix of types! We will introduce another data structure, the R list for situations when we need to store a mix of base R data types.

Data type	Stores
numeric	floating point numbers
integer	integers
complex	complex numbers
factor	categorical data
character	strings
logical	TRUE or FALSE
NA	missing
NULL	empty
function	function type

#### 4.1.1 Creating vectors

# create a vector of integers from 1 to 10

x = 1:10

Character vectors (also sometimes called "string" vectors) are entered with each value surrounded by single or double quotes; either is acceptable, but they must match. They are always displayed by R with double quotes. Here are some examples of creating vectors:

```
# examples of vectors
c('hello','world')

## [1] "hello" "world"
c(1,3,4,5,1,2)

## [1] 1 3 4 5 1 2
c(1.12341e7,78234.126)

## [1] 11234100.00 78234.13
c(TRUE,FALSE,TRUE,TRUE)

## [1] TRUE FALSE TRUE TRUE
# note how in the next case the TRUE is converted to "TRUE"
# with quotes around it.
c(TRUE,'hello')

## [1] "TRUE" "hello"

We can also create vectors as "regular sequences" of numbers. For example:
```

```
# and backwards
x = 10:1
```

The seq function can create more flexible regular sequences.

```
# create a vector of numbers from 1 to 4 skipping by 0.3 y = seq(1,4,0.3)
```

And creating a new vector by concatenating existing vectors is possible, as well.

```
# create a sequence by concatenating two other sequences
z = c(y,x)
z
## [1] 1.0 1.3 1.6 1.9 2.2 2.5 2.8 3.1 3.4 3.7 4.0 10.0 9.0 8.0
```

#### 4.1.2 Vector Operations

Operations on a single vector are typically done element-by-element. For example, we can add 2 to a vector, 2 is added to each element of the vector and a new vector of the same length is returned.

```
x = 1:10

x + 2
```

```
## [1] 3 4 5 6 7 8 9 10 11 12
```

**##** [15] 7.0 6.0 5.0 4.0 3.0 2.0

If the operation involves two vectors, the following rules apply. If the vectors are the same length: R simply applies the operation to each pair of elements.

```
x + x
```

```
## [1] 2 4 6 8 10 12 14 16 18 20
```

If the vectors are different lengths, but one length a multiple of the other, R reuses the shorter vector as needed.

```
x = 1:10

y = c(1,2)

x * y
```

```
## [1] 1 4 3 8 5 12 7 16 9 20
```

## [1] 2 6 12 8 15 24 14 24 36 20

If the vectors are different lengths, but one length not a multiple of the other, R reuses the shorter vector as needed and delivers a warning.

```
x = 1:10

y = c(2,3,4)

x * y
```

```
## Warning in x * y: longer object length is not a multiple of shorter object ## length
```

Typical operations include multiplication ("\*"), addition, subtraction, division, exponentiation ("^"), but many operations in R operate on vectors and are then called "vectorized".

#### 4.1.3 Logical Vectors

Logical vectors are vectors composed on only the values TRUE and FALSE. Note the all-upper-case and no quotation marks.

```
a = c(TRUE, FALSE, TRUE)

# we can also create a logical vector from a numeric vector

# 0 = false, everything else is 1

b = c(1,0,217)

d = as.logical(b)

d

## [1] TRUE FALSE TRUE

# test if a and d are the same at every element
all.equal(a,d)

## [1] TRUE

# We can also convert from logical to numeric
as.numeric(a)

## [1] 1 0 1
```

#### 4.1.4 Logical Operators

Some operators like  $\langle , \rangle$ , ==,  $\rangle$ =,  $\langle =, !=$ can be used to create logical vectors.

```
# create a numeric vector
x = 1:10
# testing whether x > 5 creates a logical vector
x > 5
## [1] FALSE FALSE FALSE FALSE TRUE TRUE TRUE TRUE TRUE
x <= 5
## [1] TRUE TRUE TRUE TRUE TRUE FALSE FALSE FALSE FALSE FALSE</pre>
x != 5
```

## [1] TRUE TRUE TRUE FALSE TRUE TRUE TRUE TRUE TRUE

x == 5

## [1] FALSE FALSE FALSE TRUE FALSE FALSE FALSE FALSE

We can also assign the results to a variable:

```
y = (x == 5)
y
```

## [1] FALSE FALSE FALSE FALSE TRUE FALSE FALSE FALSE FALSE

#### 4.1.5 Indexing Vectors

In R, an index is used to refer to a specific element or set of elements in an vector (or other data structure). [R uses [ and ] to perform indexing, although other approaches to getting subsets of larger data structures are common in R.

```
x = seq(0,1,0.1)
# create a new vector from the 4th element of x
x[4]
## [1] 0.3
We can even use other vectors to perform the "indexing".
x[c(3,5,6)]
## [1] 0.2 0.4 0.5
y = 3:6
x[y]
## [1] 0.2 0.3 0.4 0.5
Combining the concept of indexing with the concept of logical vectors results in a very power combination.
# use help('rnorm') to figure out what is happening next
myvec = rnorm(10)
# create logical vector that is TRUE where myvec is >0.25
gt1 = (myvec > 0.25)
sum(gt1)
## [1] 6
# and use our logical vector to create a vector of myvec values that are >0.25
myvec[gt1]
## [1] 0.7177777 1.7385063 1.6556000 0.5456691 0.5706619 1.0628216
# or <=0.25 using the logical "not" operator, "!"
myvec[!gt1]
## [1] -0.2141878 -0.7771291 -1.0870944 -1.7378650
# shorter, one line approach
myvec[myvec > 0.25]
## [1] 0.7177777 1.7385063 1.6556000 0.5456691 0.5706619 1.0628216
4.1.6 Character Vectors, A.K.A. Strings
R uses the paste function to concatenate strings.
paste("abc","def")
## [1] "abc def"
paste("abc","def",sep="THISSEP")
## [1] "abcTHISSEPdef"
paste0("abc","def")
## [1] "abcdef"
## [1] "abcdef"
paste(c("X","Y"),1:10)
```

```
## [1] "X 1" "Y 2" "X 3" "Y 4" "X 5" "Y 6" "X 7" "Y 8" "X 9" "Y 10"
paste(c("X","Y"),1:10,sep="_")
## [1] "X_1" "Y_2" "X_3" "Y_4" "X_5" "Y_6" "X_7" "Y_8" "X_9" "Y_10"
We can count the number of characters in a string.
nchar('abc')
## [1] 3
nchar(c('abc','d',123456))
## [1] 3 1 6
Pulling out parts of strings is also sometimes useful.
substr('This is a good sentence.',start=10,stop=15)
## [1] " good "
Another common operation is to replace something in a string with something (a find-and-replace).
sub('This','That','This is a good sentence.')
## [1] "That is a good sentence."
When we want to find all strings that match some other string, we can use grep, or "grab regular expression".
grep('bcd',c('abcdef','abcd','bcde','cdef','defg'))
## [1] 1 2 3
grep('bcd',c('abcdef','abcd','bcde','cdef','defg'),value=TRUE)
## [1] "abcdef" "abcd"
                          "bcde"
4.1.7
      Missing Values, AKA "NA"
R has a special value, "NA", that represents a "missing" value, or Not Available, in a vector or other data
structure. Here, we just create a vector to experiment.
x = 1:5
## [1] 1 2 3 4 5
length(x)
## [1] 5
is.na(x)
## [1] FALSE FALSE FALSE FALSE
x[2] = NA
## [1] 1 NA 3 4 5
The length of x is unchanged, but there is one value that is marked as "missing" by virtue of being NA.
length(x)
```

```
## [1] 5
is.na(x)
```

#### ## [1] FALSE TRUE FALSE FALSE

We can remove NA values by using indexing. In the following, is.na(x) returns a logical vector the length of x. The! is the logical NOT operator and converts TRUE to FALSE and vice-versa.

```
x[!is.na(x)]
## [1] 1 3 4 5
```

#### 4.1.8 Factors

## [1] "factor"

A factor is a special type of vector, normally used to hold a categorical variable—such as smoker/nonsmoker, state of residency, zipcode—in many statistical functions. Such vectors have class "factor". Factors are primarily used in Analysis of Variance (ANOVA) or other situations when "categories" are needed. When a factor is used as a predictor variable, the corresponding indicator variables are created (more later).

Note of caution that factors in R often *appear* to be character vectors when printed, but you will notice that they do not have double quotes around them. They are stored in R as numbers with a key name, so sometimes you will note that the factor *behaves* like a numeric vector.

```
# create the character vector
citizen<-c("uk", "us", "no", "au", "uk", "us", "us", "no", "au")
# convert to factor
citizenf<-factor(citizen)</pre>
## [1] "uk" "us" "no" "au" "uk" "us" "us" "no" "au"
citizenf
## [1] uk us no au uk us us no au
## Levels: au no uk us
# convert factor back to character vector
as.character(citizenf)
## [1] "uk" "us" "no" "au" "uk" "us" "us" "no" "au"
# convert to numeric vector
as.numeric(citizenf)
## [1] 3 4 2 1 3 4 4 2 1
R stores many data structures as vectors with "attributes" and "class" (just so you have seen this).
attributes(citizenf)
## $levels
## [1] "au" "no" "uk" "us"
##
## $class
## [1] "factor"
class(citizenf)
```

```
# note that after unclassing, we can see the
# underlying numeric structure again
unclass(citizenf)

## [1] 3 4 2 1 3 4 4 2 1
## attr(,"levels")
## [1] "au" "no" "uk" "us"

Tabulating factors is a useful way to get a sense of the "sample" set available.

table(citizenf)

## citizenf
## au no uk us
## 2 2 2 3
```

# 4.2 Rectangular Data

#### 4.2.1 Matrices and Data Frames

- A matrix is a rectangular array. It can be viewed as a collection of column vectors all of the same length and the same type (i.e. numeric, character or logical).
- A data frame is *also* a rectangular array. All of the columns must be the same length, but they may be of *different* types.
- The rows and columns of a matrix or data frame can be given names.
- However these are implemented differently in R; many operations will work for one but not both.

#### 4.2.2 Matrix Operations

```
x<-1:10
y<-rnorm(10)
# make a matrix by column binding two numeric vectors
mat<-cbind(x,y)</pre>
mat
##
          Х
##
    [1,]
         1 0.3842634
   [2,]
          2 -1.2536470
##
    [3,]
          3 -1.4098608
##
   [4,]
          4 1.7139083
##
   [5,]
          5 -0.1876347
##
   [6,]
          6 -0.3343492
##
    [7,]
          7
             1.6553040
##
   [8,]
          8 0.5974976
  [9,]
         9 -1.8443792
## [10,] 10 1.4203497
# And the names of the rows and columns
rownames (mat)
## NULL
colnames(mat)
## [1] "x" "y"
```

#### 4.2.3 Matrix Operations

Indexing for matrices works as for vectors except that we now need to include both the row and column (in that order).

#### 4.2.4 Matrix Operations

```
# create a matrix with 2 columns and 10 rows
# filled with random normal deviates
m = matrix(rnorm(20),nrow=10)
\# multiply all values in the matrix by 20
m = m*20
\# and add 100 to the first column of m
m[,1] = m[,1] + 100
# summarize m
summary(m)
##
                          ٧2
         ۷1
## Min. : 44.58
                    Min.
                           :-42.4628
## 1st Qu.: 93.26
                    1st Qu.:-19.8712
## Median :100.72
                    Median: 3.6691
## Mean : 95.27
                    Mean : -0.4271
## 3rd Qu.:106.33
                    3rd Qu.: 17.3559
## Max. :111.29
                    Max. : 31.2418
```

#### 4.3 Data Frames

#### 4.3.1 Matrices Versus Data Frames

```
mat<-cbind(x,y)
class(mat[,1])
## [1] "numeric"
z = paste0('a',1:10)
tab<-cbind(x,y,z)
class(tab)
## [1] "matrix"
mode(tab[,1])
## [1] "character"
head(tab,4)</pre>
```

```
## x y z

## [1,] "1" "0.384263423333191" "a1"

## [2,] "2" "-1.25364703104426" "a2"

## [3,] "3" "-1.40986082624299" "a3"

## [4,] "4" "1.7139082579506" "a4"
```

#### 4.3.2 Matrices Versus Data Frames

```
tab<-data.frame(x,y,z)</pre>
class(tab)
## [1] "data.frame"
head(tab)
               y z
## 1 1 0.3842634 a1
## 2 2 -1.2536470 a2
## 3 3 -1.4098608 a3
## 4 4 1.7139083 a4
## 5 5 -0.1876347 a5
## 6 6 -0.3343492 a6
mode(tab[,1])
## [1] "numeric"
class(tab[,3])
## [1] "factor"
rownames(tab)
## [1] "1" "2" "3" "4" "5" "6" "7" "8" "9" "10"
rownames(tab)<-paste0("row",1:10)</pre>
rownames(tab)
                                                              "row8"
## [1] "row1" "row2" "row3" "row4" "row5"
                                                "row6" "row7"
   [9] "row9" "row10"
```

#### 4.3.3 Data Frames, Continued

• Data frame columns can be refered to by name using the "dollar sign" operator

```
tab\$x
## [1] 1 2 3 4 5 6 7 8 9 10
tab\$y
## [1] 0.3842634 -1.2536470 -1.4098608 1.7139083 -0.1876347 -0.3343492
## [7] 1.6553040 0.5974976 -1.8443792 1.4203497
```

• Column names can be set, which can be useful for referring to data later

```
colnames(tab)
## [1] "x" "y" "z"
colnames(tab) = paste0('col',1:3)
```

#### 4.3.4 Exercise: Subsetting Data Frames

```
Try these
ncol(tab)
nrow(tab)
dim(tab)
summary(tab)
```

```
tab[1:3,]
tab[,2:3]
tab[,1]>7
tab[tab[,1]>7,]
tab[tab[,1]>7,3]
tab[tab[,1]>7,2:3]
tab[tab\$x>7,3]
tab\$z[tab\$x>3]
```

# 4.4 Basic Textual Input and Output

#### 4.4.1 Reading and Writing Data Frames to Disk

• The write.table function and friends write a data frame or matrix to disk as a text file.

```
write.table(tab,file='tab.txt',sep="\t",col.names=TRUE)
# remove tab from the workspace
rm(tab)
# make sure it is gone
ls(pattern="tab")
## character(0)
```

• The read.table function and friends read a data frame or matrix from a text file.

```
tab = read.table('tab.txt',sep="\t",header=TRUE)
head(tab,3)
## col1 col2 col3
## row1 1 0.3842634 a1
## row2 2 -1.2536470 a2
## row3 3 -1.4098608 a3
```

#### 4.5 Lists and Objects

#### 4.5.1 Lists

- A list is a collection of objects that may be the same or different types.
- [The objects generally have names, and may be indexed either by name (e.g. my.list\$name3) or component number (e.g. my.list[[3]])
- A data frame is a list of matched column vectors.

#### 4.5.2 Lists in Practice

• Create a list, noting the different data types involved.

```
a = list(1,"b",c(1,2,3))
a
## [[1]]
## [1] 1
##
## [[2]]
## [1] "b"
##
```

```
## [[3]]
## [1] 1 2 3
length(a)
## [1] 3
class(a)
## [1] "list"
a[[3]]
## [1] 1 2 3
```

#### 4.5.3 Lists in Practice

• A data frame is a list.

```
# test if our friend "tab" is a list
is.list(tab)
## [1] TRUE
tab[[2]]
## [1] 0.3842634 -1.2536470 -1.4098608 1.7139083 -0.1876347 -0.3343492
## [7] 1.6553040 0.5974976 -1.8443792 1.4203497
names(tab)
## [1] "col1" "col2" "col3"
```

Table Summary of basic R data structures.

Data type S	tores
vector	one-dimensional data, single data type
matrix	two-dimensional data, single data type
data frame	two-dimensional data, multiple data types
list	list of data types, not all need to be the same type
object	a list with attributes and potentially slots and methods

# 5 Plotting and Graphics

## 5.1 Basic Plot Functions

- The command plot(x,y) will plot vector x as the independent variable and vector y as the dependent variable.
- Within the command line, you can specify the title of the graph, the name of the x-axis, and the name of the y-axis.
  - main='title'
  - xlab='name of x axis'
  - ylab='name of y axis'
- The command lines(x,y) adds a line segment to the plot.
- The command points(x,y) adds points to the plot.
- A legend can be created using legend.

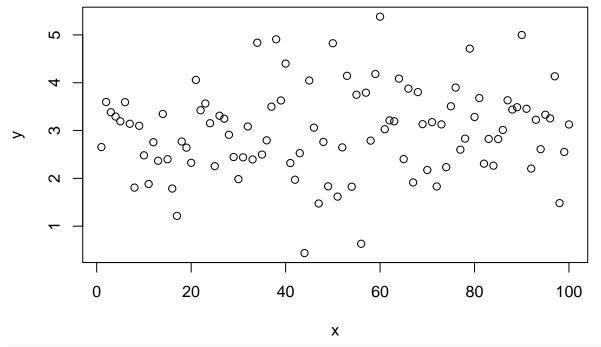
demo

demo(graphics)

## 5.1.1 Simple Plotting Example

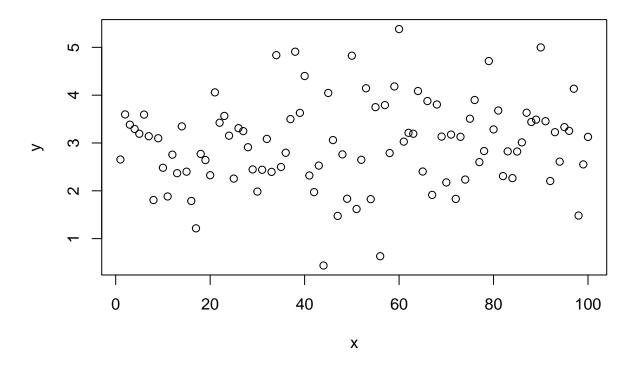
Try this yourself:

```
x = 1:100
y = rnorm(100,3,1) # 100 random normal deviates with mean=3, sd=1
plot(x,y)
```

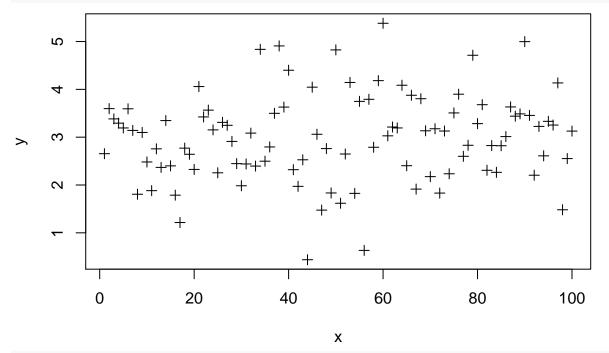


plot(x,y,main='My First Plot')

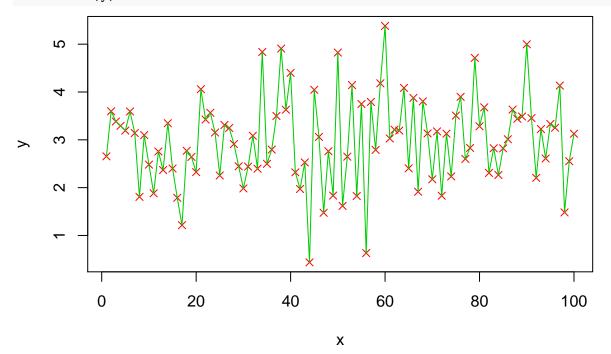
# **My First Plot**



# # change point type plot(x,y,pch=3)



# change color
plot(x,y,pch=4,col=2)
# draw lines between points
lines(x,y,col=3)



#### 5.1.2 More Plotting

```
z=sort(y)
# plot a sorted variable vs x
plot(x,z,main='Random Normal Numbers',
xlab='Index',ylab='Random Number')
# another example
plot(-4:4,-4:4)
# and add a point at (0,2) to the plot
points(0,2,pch=6,col=12)
5.1.3 More Plotting
# check margin and outer margin settings
par(c("mar", "oma"))
plot(x,y)
par(oma=c(1,1,1,1)) # set outer margin
plot(x,y)
par(mar=c(2.5,2.1,2.1,1)) # set margin
plot(x,y)
# A basic histogram
hist(z, main="Histogram",
    sub="Random normal")
# A "density" plot
plot(density(z), main="Density plot",
    sub="Random normal")
# A smaller "bandwidth" to capture more detail
plot(density(z, adjust=0.5),
  sub="smaller bandwidth")
```

## 5.1.4 Graphics Devices and Saving Plots

- to make a plot directly to a file use: png(), postscript(), etc.
- R can have multiple graphics "devices" open.
  - To see a list of active devices: dev.list()
  - To close the most recent device: dev.off()
  - To close device 5: dev.off(5)
  - To use device 5: dev.set(5)

#### 5.1.5 More Plotting

• Save a png image to a file

```
png(file="myplot.png",width=480,height=480)
plot(density(z,adjust=2.0),sub="larger bandwidth")
dev.off()
```

- On your own, save a pdf to a file. NOTE: The dimensions in pdf() are in inches
- Multiple plots on the same page:

```
par(mfrow=c(2,1))
plot(density(z,adjust=2.0),sub="larger bandwidth")
hist(z)

# use dev.off() to turn off the two-row plotting
```

# 5.1.6 R Graphics Galleries and Resources

Visit these sites for some ideas.

- http://www.sr.bham.ac.uk/~ajrs/R/r-gallery.html
- http://gallery.r-enthusiasts.com/
- http://cran.r-project.org/web/views/Graphics.html

# 6 Control Structures, Looping, and Applying

#### 6.1 Control Structures and Looping

#### 6.1.1 Control Structures in R

- R has multiple types of control structures that allows for sequential evaluation of statements.
- For loops

```
for (x in set) {operations}
```

• while loops

```
while (x in condition){operations}
```

• If statements (conditional)

```
if (condition) {
some operations
} else { other operations }
```

#### 6.1.2 Control Structure and Looping Examples

```
x<-1:9
length(x)
# a simple conditional then two expressions
if (length(x)<=10) {</pre>
```

```
x<-c(x,10:20);print(x)}
# more complex
if (length(x)<5) {
    print(x)
} else {
    print(x[5:20])
}
# print the values of x, one at a time
for (i in x) print(i)
for(i in x) i  # note R will not echo in a loop</pre>
```

#### 6.1.3 Control Structure and Looping Examples

```
# loop over a character vector
y<-c('a','b','hi there')
for (i in y) print(i)

# and a while loop
j<-1
while(j<10) { # do this while j<10
    print(j)
    j<-j+2} # at each iteration, increase j by 2</pre>
```

# 6.2 Applying

#### 6.2.1 Why Does R Have Apply Functions

- Often we want to apply the same function to all the rows or columns of a matrix, or all the elements of a list.
- We could do this in a loop, but loops take a lot of time in an interpreted language like R.
- R has more efficient built-in operators, the apply functions.

example If mat is a matrix and fun is a function (such as mean, var, lm ...) that takes a vector as its argument, then you can:

```
apply(mat,1,fun) # over rows--second argument is 1
apply(mat,2,fun) # over columns--second argument is 2
```

In either case, the output is a vector.

#### 6.2.2 Apply Function Exercise

- 1. Using the matrix and rnorm functions, create a matrix with 20 rows and 10 columns (200 values total) of random normal deviates.
- 2. Compute the mean for each row of the matrix.
- 3. Compute the median for each column.

# 6.2.3 Related Apply Functions

• lapply(list, function) applies the function to every element of list

- sapply(list or vector, function) applies the function to every element of list or vector, and returns a vector, when possible (easier to process)
- tapply(x, factor, fun) uses the factor to split vector x into groups, and then applies fun to each group

#### 6.2.4 Related Apply Function Examples

```
# create a list
my.list <- list(a=1:3,b=5:10,c=11:20)
my.list
# Get the mean for each member of the list
# return a vector
sapply( my.list, mean)
# Get the full summary for each member of
# the list, returned as a list
lapply( my.list, summary)
# Find the mean for each group defined by a factor
my.vector <- 1:10
my.factor <- factor(
    c(1,1,1,2,2,2,3,3,3,3))
tapply(my.vector, my.factor, mean)</pre>
```

## 7 Functions

#### 7.0.5 Function Overview

• Functions are objects and are assigned to names, just like data.

```
myFunction = function(argument1, argument2) {
  expression1
  expression2
}
```

- We write functions for anything we need to do again and again.
- You may test your commands interactively at first, and then use the history() feature and an editor to create the function.
- It is wise to include a comment at the start of each function to say what it does and to document functions of more than a few lines.

#### 7.0.6 Example Functions

```
add1 = function(x) {
    # this function adds one to the first argument and returns it
    x + 1
}
add1(17)
## [1] 18
add1(c(17,18,19,20))
## [1] 18 19 20 21
```

You can use the edit() function to make changes to a function. The following command will open a window, allow you to make changes, and assign the result to a new function, add2.

```
add2 = edit(add1)
```

#### 7.0.7 Further Reading

The amount of learning material for R is simply astonishing!

- Thomas Girke's R and Bioconductor Manual
- A HUGE collection of contributed R documentation and tutorials
- Bioconductor course materials
- Sean Davis' website
- The Official R Manuals

# 8 RStudio: A Quick Tour

Panes

Options

Help

Environment, History, and Files

# 9 R: First Impressions

Type values and mathematical formulas into R's command prompt

```
1 + 1 ## [1] 2
```

Assign values to symbols (variables)

```
x = 1
x + x
```

```
## [1] 2
```

Invoke functions such as c(), which takes any number of values and returns a single vector

```
x = c(1, 2, 3)
x
```

```
## [1] 1 2 3
```

R functions, such as sqrt(), often operate efficiently on vectors

```
y = sqrt(x)
y
```

```
## [1] 1.000000 1.414214 1.732051
```

There are often several ways to accomplish a task in R

```
x = c(1, 2, 3)
x
## [1] 1 2 3
x <- c(4, 5, 6)
x
## [1] 4 5 6
x <- 7:9
x
## [1] 7 8 9
10:12 -> x
x
## [1] 10 11 12
Sometimes R does 'surprising' things that can be fun to figure out
x <- c(1, 2, 3) -> y
## [1] 1 2 3
y
## [1] 1 2 3
9.1 R Data types: vector and list
```

'Atomic' vectors

```
• Types include integer, numeric (float-point; real), complex, logical, character, raw (bytes)
```

```
people <- c("Lori", "Nitesh", "Valerie", "Herve")
people</pre>
```

```
## [1] "Lori" "Nitesh" "Valerie" "Herve"
```

• Atomic vectors can be named

```
population <- c(Buffalo=259000, Rochester=210000, `New York`=8400000)
population</pre>
```

```
## Buffalo Rochester New York
## 259000 210000 8400000
```

log10(population)

```
## Buffalo Rochester New York
## 5.413300 5.322219 6.924279
```

• Statistical concepts like NA ("not available")

```
truthiness <- c(TRUE, FALSE, NA)
truthiness
```

```
## [1] TRUE FALSE NA
```

• Logical concepts like 'and' (&), 'or' (|), and 'not' (!)

```
!truthiness
  ## [1] FALSE TRUE
  truthiness | !truthiness
  ## [1] TRUE TRUE
  truthiness & !truthiness
  ## [1] FALSE FALSE
                         NA
• Numerical concepts like infinity (Inf) or not-a-number (NaN, e.g., 0 / 0)
  undefined_numeric_values <- c(NA, 0/0, NaN, Inf, -Inf)
  undefined_numeric_values
           NA NaN NaN Inf -Inf
  ## [1]
  sqrt(undefined_numeric_values)
  ## Warning in sqrt(undefined_numeric_values): NaNs produced
  ## [1] NA NaN NaN Inf NaN
• Common string manipulations
  toupper(people)
  ## [1] "LORI"
                               "VALERIE" "HERVE"
                    "NITESH"
  substr(people, 1, 3)
  ## [1] "Lor" "Nit" "Val" "Her"
• R is a green consumer – recycling short vectors to align with long vectors
  x < -1:3
  x * 2
                    # '2' (vector of length 1) recycled to c(2, 2, 2)
  ## [1] 2 4 6
  truthiness | NA
  ## [1] TRUE
                      NA
  truthiness & NA
  ## [1]
            NA FALSE
                         NA
• It's very common to nest operations, which can be simultaneously compact, confusing, and expressive
  ([: subset; <: less than)
  substr(tolower(people), 1, 3)
  ## [1] "lor" "nit" "val" "her"
  population[population < 1000000]</pre>
  ##
       Buffalo Rochester
  ##
        259000
                   210000
```

Lists

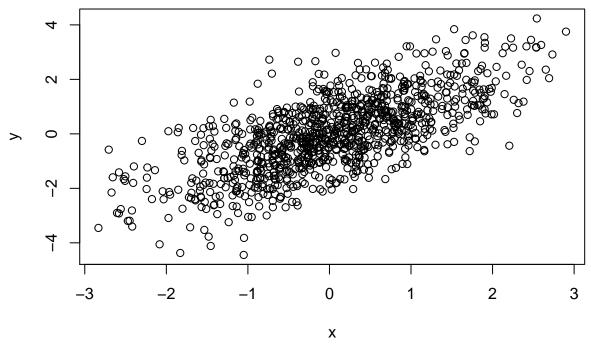
• The list type can contain other vectors, including other lists

```
frenemies = list(
         friends=c("Larry", "Richard", "Vivian"),
         enemies=c("Dick", "Mike")
     frenemies
     ## $friends
     ## [1] "Larry"
                     "Richard" "Vivian"
     ##
     ## $enemies
     ## [1] "Dick" "Mike"
  • [ subsets one list to create another list, [[ extracts a list element
     frenemies[1]
     ## $friends
     ## [1] "Larry"
                      "Richard" "Vivian"
     frenemies[c("enemies", "friends")]
     ## $enemies
     ## [1] "Dick" "Mike"
     ##
     ## $friends
     ## [1] "Larry"
                     "Richard" "Vivian"
     frenemies[["enemies"]]
     ## [1] "Dick" "Mike"
Factors
  • Character-like vectors, but with values restricted to specific levels
     sex = factor(c("Male", "Male", "Female"),
                  levels=c("Female", "Male", "Hermaphrodite"))
     sex
     ## [1] Male
                   Male
                          Female
     ## Levels: Female Male Hermaphrodite
     sex == "Female"
     ## [1] FALSE FALSE TRUE
     table(sex)
     ## sex
     ##
               Female
                                Male Hermaphrodite
     ##
     sex[sex == "Female"]
     ## [1] Female
     ## Levels: Female Male Hermaphrodite
```

# 9.2 Classes: data.frame and beyond

Variables are often related to one another in a highly structured way, e.g., two 'columns' of data in a spreadsheet

```
x = rnorm(1000)  # 1000 random normal deviates
y = x + rnorm(1000)  # another 1000 deviates, as a function of x
plot(y ~ x)  # relationship between x and y
```



Convenient to manipulate them together

• data.frame(): like columns in a spreadsheet

```
df = data.frame(X=x, Y=y)
head(df)  # first 6 rows
```

```
## X Y

## 1 -0.72437726 -0.1684765

## 2 0.40709567 0.4559514

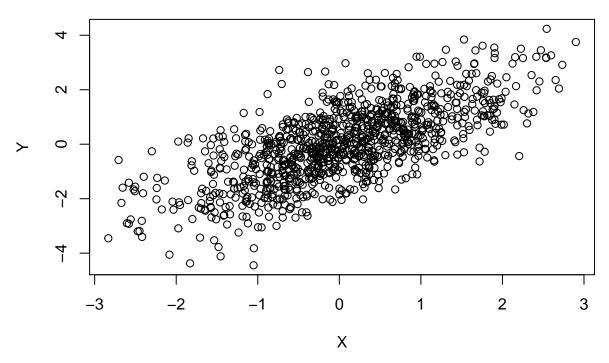
## 3 -0.44450808 -0.1445829

## 4 -0.15238336 1.5561891

## 5 -0.03677902 0.3178355

## 6 0.99614929 0.3245457

plot(Y ~ X, df) # same as above
```



• See all data with View(df). Summarize data with summary(df)

#### summary(df)

```
##
          Х
##
   Min.
           :-2.8326987
                          Min.
                                 :-4.444125
    1st Qu.:-0.6591657
                          1st Qu.:-0.923660
   Median : 0.0003174
                          Median : 0.043338
##
##
    Mean
           : 0.0398925
                          Mean
                                 : 0.006076
    3rd Qu.: 0.7176888
##
                          3rd Qu.: 0.925764
   Max.
           : 2.9001701
                          Max.
                                 : 4.235091
```

• Easy to manipulate data in a coordinated way, e.g., access column X with \$ and subset for just those values greater than 0

```
positiveX = df[df$X > 0,]
head(positiveX)
```

```
## X Y

## 2 0.4070957 0.45595139

## 6 0.9961493 0.32454569

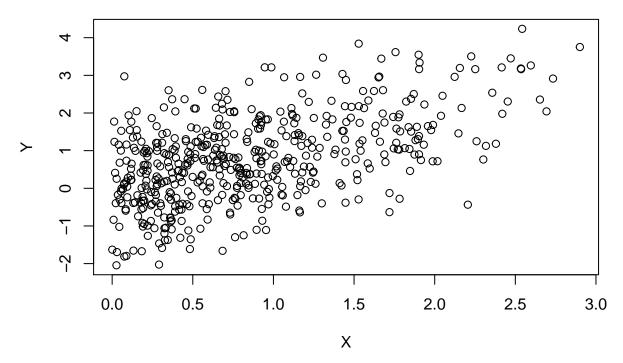
## 8 0.4530330 0.63267547

## 11 0.2187124 0.84211192

## 12 0.2648313 -0.10204721

## 20 0.5874987 -0.08449245

plot(Y ~ X, positiveX)
```



 $\bullet$  R is introspective – ask it about itself

• matrix() a related class, where all elements have the same type (a data.frame() requires elements within a column to be the same type, but elements between columns can be different types).

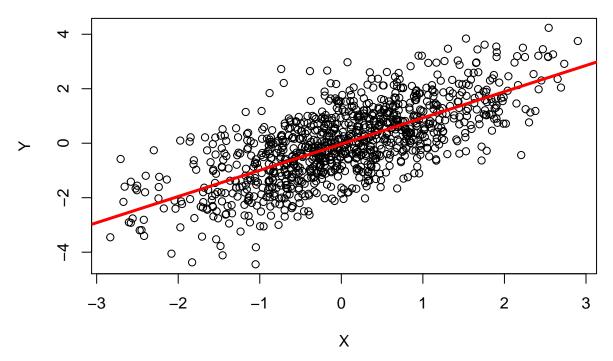
A scatterplot makes one want to fit a linear model (do a regression analysis)

- Use a formula to describe the relationship between variables
- Variables found in the second argument

```
fit \leftarrow lm(Y \sim X, df)
```

• Visualize the points, and add the regression line

```
plot(Y ~ X, df)
abline(fit, col="red", lwd=3)
```



• Summarize the fit as an ANOVA table

```
anova(fit)
```

- N.B. 'Type I' sums-of-squares, so order of independent variables matters; use drop1() for 'Type III'. See DataCamp Quick-R
- Introspection what class is fit? What methods can I apply to an object of that class?

#### class(fit)

## [1] "lm"

#### methods(class=class(fit))

```
##
    [1] add1
                        alias
                                       anova
                                                       case.names
    [5] coerce
                        confint
                                       cooks.distance deviance
                        dfbetas
##
    [9] dfbeta
                                       drop1
                                                       dummy.coef
## [13] effects
                        extractAIC
                                       family
                                                       formula
## [17] hatvalues
                        influence
                                       initialize
                                                       kappa
## [21] labels
                        logLik
                                       model.frame
                                                       model.matrix
## [25] nobs
                        plot
                                       predict
                                                       print
## [29] proj
                        qr
                                       residuals
                                                       rstandard
## [33] rstudent
                        show
                                       simulate
                                                       slotsFromS3
## [37] summary
                        variable.names vcov
## see '?methods' for accessing help and source code
```

# 9.3 Help!

Help available in Rstudio or interactively

• Check out the help page for rnorm()

?rnorm

• 'Usage' section describes how the function can be used

```
rnorm(n, mean = 0, sd = 1)
```

- Arguments, some with default values. Arguments matched first by name, then position
- 'Arguments' section describes what the arguments are supposed to be
- 'Value' section describes return value
- 'Examples' section illustrates use
- Often include citations to relevant technical documentation, reference to related functions, obscure details
- Can be intimidating, but in the end actually very useful

# 10 Exercise 1: BRFSS Survey Data

We will explore a subset of data collected by the CDC through its extensive Behavioral Risk Factor Surveillance System (BRFSS) telephone survey. Check out the link for more information. We'll look at a subset of the data.

1. Use file.choose() to find the path to the file 'BRFSS-subset.csv'

```
path <- file.choose()</pre>
```

2. Input the data using read.csv(), assigning to a variable brfss

```
brfss <- read.csv(path)</pre>
```

- 3. Use command like class(), head(), dim(), summary() to explore the data.
  - What variables have been measured?
    - Can you guess at the units used for, e.g., Weight and Height?

```
class(brfss)
head(brfss)
dim(brfss)
summary(brfss)
```

4. Use the \$ operator to extract the 'Sex' column, and summarize the number of males and females in the survey using table(). Do the same for 'Year', and for both Sex and Year

```
table(brfss$Sex)

##
## Female Male
## 12039 7961

table(brfss$Year)
```

```
## 1990 2010
  ## 10000 10000
  table(brfss$Sex, brfss$Year)
  ##
  ##
               1990 2010
  ##
       Female 5718 6321
       Male
               4282 3679
  with(brfss, table(Sex, Year))
                                                 # same, but easier
  ##
             Year
  ## Sex
               1990 2010
  ##
       Female 5718 6321
  ##
       Male
               4282 3679
5. Use aggregate() to summarize the mean weight of each group. What about the median weight of each
  group? What about the number of observations in each group?
  with(brfss, aggregate(Weight, list(Year, Sex), mean, na.rm=TRUE))
  ##
       Group.1 Group.2
  ## 1
          1990 Female 64.81838
  ## 2
          2010 Female 72.95424
  ## 3
          1990
                   Male 81.17999
  ## 4
          2010
                   Male 88.84657
  with(brfss, aggregate(Weight, list(Year=Year, Sex=Sex), mean, na.rm=TRUE))
  ##
       Year
                Sex
                           x
  ## 1 1990 Female 64.81838
  ## 2 2010 Female 72.95424
  ## 3 1990
              Male 81.17999
  ## 4 2010
              Male 88.84657
6. Use a formula and the aggregate() function to describe the relationship between Year, Sex, and
  Weight
  aggregate(Weight ~ Year + Sex, brfss, mean) # same, but more informative
       Year
                Sex
                      Weight
  ## 1 1990 Female 64.81838
  ## 2 2010 Female 72.95424
  ## 3 1990
              Male 81.17999
  ## 4 2010
              Male 88.84657
  aggregate(. ~ Year + Sex, brfss, mean)
                                                 # all variables
       Year
                Sex
                         Age
                               Weight
                                         Height
  ## 1 1990 Female 46.09153 64.84333 163.2914
  ## 2 2010 Female 57.07807 73.03178 163.2469
  ## 3 1990
               Male 43.87574 81.19496 178.2242
  ## 4 2010
               Male 56.25465 88.91136 178.0139
```

##

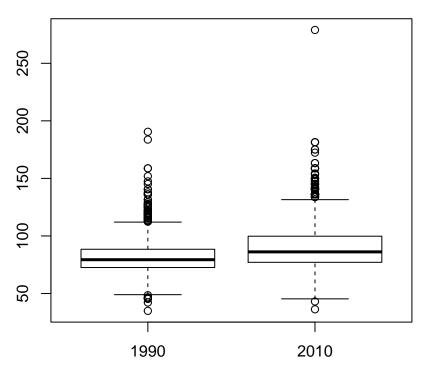
7. Create a subset of the data consisting of only the 1990 observations. Perform a t-test comparing the weight of males and females ("'Weight' as a function of 'Sex'", Weight ~ Sex)

```
brfss_1990 = brfss[brfss$Year == 1990,]
t.test(Weight ~ Sex, brfss_1990)
##
## Welch Two Sample t-test
##
## data: Weight by Sex
## t = -58.734, df = 9214, p-value < 2.2e-16
## alternative hypothesis: true difference in means is not equal to 0
## 95 percent confidence interval:
## -16.90767 -15.81554
## sample estimates:
## mean in group Female
                          mean in group Male
##
               64.81838
                                    81.17999
t.test(Weight ~ Sex, brfss, subset = Year == 1990)
##
## Welch Two Sample t-test
##
## data: Weight by Sex
## t = -58.734, df = 9214, p-value < 2.2e-16
## alternative hypothesis: true difference in means is not equal to 0
## 95 percent confidence interval:
## -16.90767 -15.81554
## sample estimates:
## mean in group Female
                          mean in group Male
               64.81838
                                    81.17999
```

What about differences between weights of males (or females) in 1990 versus 2010? Check out the help page ?t.test.formula. Is there a way of performing a t-test on brfss without explicitly creating the object brfss\_1990?

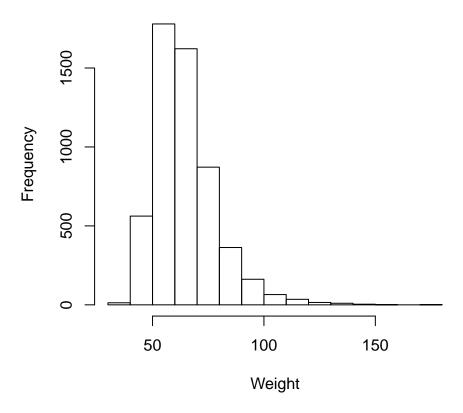
8. Use boxplot() to plot the weights of the Male individuals. Can you transform weight, e.g., sqrt(Weight) ~ Year? Interpret the results. Do similar boxplots for the t-tests of the previous question.

# Males



9. Use hist() to plot a histogram of weights of the 1990 Female individuals.

## Females, 1990



# 11 Exercise 2: ALL Phenotypic Data

This data comes from an (old) Acute Lymphoid Leukemia microarray data set.

Choose the file that contains ALL (acute lymphoblastic leukemia) patient information and input the date using read.csv(); for read.csv(), use row.names=1 to indicate that the first column contains row names.

```
path <- file.choose() # look for ALL-phenoData.csv

stopifnot(file.exists(path))
pdata <- read.csv(path, row.names=1)</pre>
```

Check out the help page ?read.delim for input options. The exercises use ?read.csv; Can you guess why? Explore basic properties of the object you've created, for instance...

```
class(pdata)
```

```
## [1] "data.frame"
colnames (pdata)
    [1] "cod"
                                              "sex"
                                                                 "age"
                           "diagnosis"
                                              "CR"
##
    [5] "BT"
                           "remission"
                                                                 "date.cr"
    [9]
        "t.4.11."
                           "t.9.22."
                                              "cyto.normal"
                                                                 "citog"
                                              "mdr"
                                                                 "kinet"
   [13]
        "mol.biol"
                           "fusion.protein"
                                                                 "f.u"
        "ccr"
                           "relapse"
                                              "transplant"
   [17]
   [21] "date.last.seen"
```

```
dim(pdata)
## [1] 128
            21
head(pdata)
          cod diagnosis sex age BT remission CR
                                                    date.cr t.4.11. t.9.22.
## 01005 1005 5/21/1997
                              53 B2
                                                   8/6/1997
                                                               FALSE
                                                                        TRUE
                           Μ
                                            CR CR
## 01010 1010 3/29/2000
                           Μ
                              19 B2
                                            CR CR 6/27/2000
                                                               FALSE
                                                                       FALSE
## 03002 3002 6/24/1998
                           F
                              52 B4
                                            CR CR 8/17/1998
                                                                  NA
                                                                          NA
## 04006 4006 7/17/1997
                              38 B1
                                            CR CR 9/8/1997
                                                                TRUE
                                                                       FALSE
                           М
## 04007 4007 7/22/1997
                                                               FALSE
                              57 B2
                                            CR CR 9/17/1997
                                                                       FALSE
                           М
                                            CR CR 9/27/1997
## 04008 4008 7/30/1997
                              17 B1
                                                               FALSE
                                                                       FALSE
##
         cyto.normal
                             citog mol.biol fusion.protein mdr
                                                                   kinet
## 01005
               FALSE
                           t(9;22)
                                    BCR/ABL
                                                       p210 NEG dyploid FALSE
## 01010
               FALSE
                                                       <NA> POS dyploid FALSE
                      simple alt.
                                        NEG
## 03002
                  NA
                              <NA>
                                    BCR/ABL
                                                       p190 NEG dyploid FALSE
## 04006
               FALSE
                           t(4;11) ALL1/AF4
                                                       <NA> NEG dyploid FALSE
## 04007
               FALSE
                           del(6q)
                                        NEG
                                                       <NA> NEG dyploid FALSE
                                                       <NA> NEG hyperd. FALSE
## 04008
               FALSE complex alt.
                                        NEG
##
         relapse transplant
                                            f.u date.last.seen
## 01005
           FALSE
                        TRUE BMT / DEATH IN CR
                                                           <NA>
## 01010
            TRUE
                       FALSE
                                                     8/28/2000
                                            REL
## 03002
            TRUE
                      FALSE
                                            REL
                                                    10/15/1999
                                                     1/23/1998
## 04006
            TRUE
                      FALSE
                                            REL
## 04007
            TRUE
                      FALSE
                                            REL
                                                     11/4/1997
## 04008
            TRUE
                      FALSE
                                            REL
                                                    12/15/1997
summary(pdata$sex)
      F
##
           M NA's
##
     42
          83
                3
summary(pdata$cyto.normal)
                               NA's
##
             FALSE
                       TRUE
      Mode
                                 35
## logical
                69
                         24
Remind yourselves about various ways to subset and access columns of a data.frame
pdata[1:5, 3:4]
##
         sex age
## 01005
           М
              53
## 01010
           M 19
## 03002
              52
           F
## 04006
           М
              38
## 04007
           М
              57
pdata[1:5, ]
          cod diagnosis sex age BT remission CR
                                                    date.cr t.4.11. t.9.22.
## 01005 1005 5/21/1997
                              53 B2
                                            CR CR 8/6/1997
                                                               FALSE
                                                                        TRUE
                           Μ
## 01010 1010 3/29/2000
                                                               FALSE
                                                                       FALSE
                           Μ
                              19 B2
                                            CR CR 6/27/2000
                           F
## 03002 3002 6/24/1998
                              52 B4
                                            CR CR 8/17/1998
                                                                  NA
                                                                          NA
## 04006 4006 7/17/1997
                           M
                              38 B1
                                            CR CR 9/8/1997
                                                                TRUE
                                                                       FALSE
## 04007 4007 7/22/1997
                           M
                              57 B2
                                            CR CR 9/17/1997
                                                               FALSE
                                                                       FALSE
                            citog mol.biol fusion.protein mdr
##
         cyto.normal
                                                                  kinet
                                                                          ccr
```

```
## 01005
              FALSE
                        t(9;22) BCR/ABL
                                                   p210 NEG dyploid FALSE
## 01010
                                                   <NA> POS dyploid FALSE
              FALSE simple alt.
                                     NEG
                           <NA> BCR/ABL
## 03002
                 NA
                                                   p190 NEG dyploid FALSE
## 04006
                        t(4;11) ALL1/AF4
              FALSE
                                                   <NA> NEG dyploid FALSE
                        del(6q)
## 04007
              FALSE
                                     NEG
                                                   <NA> NEG dyploid FALSE
                                         f.u date.last.seen
##
        relapse transplant
## 01005
                      TRUE BMT / DEATH IN CR
          FALSE
                                                       <NA>
                                                  8/28/2000
## 01010
           TRUE
                     FALSE
                                         REL
## 03002
           TRUE
                     FALSE
                                         REL
                                                 10/15/1999
## 04006
           TRUE
                     FALSE
                                         REL
                                                  1/23/1998
## 04007
           TRUE
                     FALSE
                                          REL
                                                   11/4/1997
head(pdata[, 3:5])
         sex age BT
##
## 01005
         M 53 B2
## 01010
         M 19 B2
## 03002
         F 52 B4
## 04006
          M 38 B1
## 04007
         M 57 B2
## 04008
         M 17 B1
tail(pdata[, 3:5], 3)
         sex age BT
## 65003
           M 30 T3
## 83001
           M 29 T2
## LAL4 <NA> NA T
head(pdata$age)
## [1] 53 19 52 38 57 17
head(pdata$sex)
## [1] M M F M M M
## Levels: F M
head(pdata[pdata$age > 21,])
##
          cod diagnosis sex age BT remission CR
                                                 date.cr t.4.11. t.9.22.
## 01005 1005 5/21/1997
                         M 53 B2
                                    CR CR 8/6/1997
                                                                    TRUE
## 03002 3002 6/24/1998
                                         CR CR 8/17/1998
                         F 52 B4
                                                              NΑ
                                                                      NA
## 04006 4006 7/17/1997
                        M 38 B1
                                         CR CR 9/8/1997
                                                            TRUE
                                                                   FALSE
## 04007 4007 7/22/1997 M 57 B2
                                         CR CR 9/17/1997
                                                           FALSE
                                                                   FALSE
## 08001 8001 1/15/1997 M 40 B2
                                         CR CR 3/26/1997
                                                           FALSE
                                                                   FALSE
## 08011 8011 8/21/1998
                        M 33 B3
                                         CR CR 10/8/1998
                                                           FALSE
                                                                   FALSE
                           citog mol.biol fusion.protein mdr
##
         cyto.normal
                                                               kinet
## 01005
              FALSE
                         t(9;22) BCR/ABL
                                                 p210 NEG dyploid FALSE
## 03002
                            <NA> BCR/ABL
                                                   p190 NEG dyploid FALSE
                 NA
## 04006
              FALSE
                         t(4;11) ALL1/AF4
                                                    <NA> NEG dyploid FALSE
## 04007
              FALSE
                         del(6q)
                                      NEG
                                                    <NA> NEG dyploid FALSE
## 08001
              FALSE
                                                   p190 NEG
                        del(p15)
                                  BCR/ABL
                                                                <NA> FALSE
## 08011
              FALSE del(p15/p16)
                                  BCR/ABL
                                               p190/p210 NEG dyploid FALSE
##
        relapse transplant
                                         f.u date.last.seen
## 01005
          FALSE
                      TRUE BMT / DEATH IN CR
                                                        <NA>
## 03002
           TRUE
                     FALSE
                                         REL
                                                 10/15/1999
## 04006
           TRUE
                     FALSE
                                         REL
                                                 1/23/1998
```

```
## 04007 TRUE FALSE REL 11/4/1997
## 08001 TRUE FALSE REL 7/11/1997
## 08011 FALSE TRUE BMT / DEATH IN CR <NA>
```

It seems from below that there are 17 females over 40 in the data set. However, some individuals have NA for the age and / or sex, and these NA values propagate through some computations. Use table() to summarize the number of females over 40, and the number of samples for which this classification cannot be determined. When R encounters an NA value in a subscript index, it introduces an NA into the result. Observe this (rows of NA values introduced into the result) when subsetting using [ versus using the subset() function.

```
idx <- pdata$sex == "F" & pdata$age > 40
table(idx, useNA="ifany")
## idx
## FALSE
          TRUE
                 <NA>
##
     108
             17
                     3
dim(pdata[idx,])
                              # WARNING: 'NA' rows introduced
## [1] 20 21
tail(pdata[idx,])
##
                 diagnosis
                             sex
                                  age
                                        BT remission
                                                                        CR
            cod
## 49006 49006
                 8/12/1998
                               F
                                   43
                                        B2
                                                                        CR.
                                                   CR
                                F
   57001 57001
                 1/29/1997
                                   53
                                        ВЗ
                                                 <NA>
                                                       DEATH IN INDUCTION
                                                  REF
   62001 62001 11/11/1997
                               F
                                   50
                                        В4
                                                                       REF
  NA.1
           <NA>
                       <NA>
                            <NA>
                                   NA
                                      <NA>
                                                 <NA>
                                                                      <NA>
   02020
                               F
##
           2020
                 3/23/2000
                                   48
                                        T2
                                                 <NA> DEATH IN INDUCTION
## NA.2
           <NA>
                       <NA> <NA>
                                   NA
                                      <NA>
                                                 <NA>
                                                                      <NA>
             date.cr t.4.11. t.9.22.
##
                                       cyto.normal
                                                             citog mol.biol
## 49006 11/19/1998
                           NA
                                    NA
                                                               <NA>
                                                                     BCR/ABL
                                                 NA
                        FALSE
## 57001
                                 FALSE
                                               TRUE
                <NA>
                                                            normal
                                                                          NEG
## 62001
                        FALSE
                                  TRUE
                                                                     BCR/ABL
                <NA>
                                              FALSE
                                                    t(9;22)+other
## NA.1
                <NA>
                           NA
                                    NA
                                                 NA
                                                                         <NA>
                                                               <NA>
                        FALSE
                                 FALSE
                                              FALSE
## 02020
                <NA>
                                                      complex alt.
                                                                         NEG
## NA.2
                <NA>
                           NA
                                    NA
                                                 NA
                                                               <NA>
                                                                         <NA>
##
                           mdr
                                           ccr relapse transplant
         fusion.protein
                                  kinet
                                                                     f.u
## 49006
                           NEG dyploid FALSE
                                                  TRUE
                                                                     REL
                     p210
                                                             FALSE
## 57001
                     <NA>
                           NEG hyperd.
                                            NΑ
                                                     NA
                                                                 NA
                                                                    <NA>
## 62001
                     <NA>
                           NEG hyperd.
                                            NA
                                                     NA
                                                                 NA
                                                                    <NA>
                     <NA>
## NA.1
                          <NA>
                                                                    <NA>
                                   <NA>
                                            NA
                                                     NΑ
                                                                 NΑ
## 02020
                     <NA>
                           NEG
                               dyploid
                                            NA
                                                     NA
                                                                 NA
                                                                    <NA>
## NA.2
                     <NA>
                          <NA>
                                   <NA>
                                            NA
                                                     NA
                                                                 NA <NA>
##
         date.last.seen
## 49006
               4/26/1999
## 57001
                     <NA>
## 62001
                     <NA>
## NA.1
                     <NA>
## 02020
                     <NA>
## NA.2
                     <NA>
dim(subset(pdata, idx))
                              # BETTER: no NA rows
## [1] 17 21
dim(subset(pdata, (sex == "F") & (age > 40))) # alternative
```

#### ## [1] 17 21

##

#### tail(subset(pdata,idx))

```
diagnosis sex age BT remission
## 28032 28032
                 9/26/1998
                              F
                                 52 B1
                                               CR
                                                                    CR 10/30/1998
## 30001 30001
                 1/16/1997
                              F
                                 54 B3
                                             <NA> DEATH IN INDUCTION
                                                                             <NA>
## 49006 49006
                8/12/1998
                              F
                                 43 B2
                                               CR
                                                                    CR 11/19/1998
## 57001 57001
                 1/29/1997
                                 53 B3
                                             <NA> DEATH IN INDUCTION
                                                                             <NA>
## 62001 62001 11/11/1997
                                              REF
                                                                             <NA>
                              F
                                 50 B4
                                                                   R.F.F
  02020
          2020
                3/23/2000
                              F
                                 48 T2
                                             <NA> DEATH IN INDUCTION
                                                                             <NA>
##
         t.4.11. t.9.22. cyto.normal
                                                citog mol.biol fusion.protein
## 28032
             TRUE
                    FALSE
                                 FALSE
                                              t(4;11) ALL1/AF4
## 30001
           FALSE
                     TRUE
                                                       BCR/ABL
                                 FALSE t(9;22)+other
                                                                           p190
## 49006
               NA
                                    NA
                                                 <NA>
                                                        BCR/ABL
                       NA
                                                                           p210
## 57001
                                  TRUE
                                               normal
           FALSE
                    FALSE
                                                            NEG
                                                                           < NA >
## 62001
           FALSE
                     TRUE
                                 FALSE t(9;22)+other
                                                        BCR/ABL
                                                                           <NA>
## 02020
           FALSE
                    FALSE
                                 FALSE
                                         complex alt.
                                                            NEG
                                                                           <NA>
##
         {\tt mdr}
               kinet
                        ccr relapse transplant
                                                  f.u date.last.seen
                                           FALSE
                                                  CCR
## 28032 NEG dyploid
                       TRUE
                               FALSE
                                                            5/16/2002
## 30001 NEG hyperd.
                          NA
                                              NA <NA>
                                                                  <NA>
                                  NA
## 49006 NEG dyploid FALSE
                                TRUE
                                           FALSE
                                                  REL
                                                            4/26/1999
## 57001 NEG hyperd.
                         NA
                                  NA
                                              NA <NA>
                                                                  < NA >
## 62001 NEG hyperd.
                          NA
                                  NA
                                              NA <NA>
                                                                  <NA>
## 02020 NEG dyploid
                                              NA <NA>
                                                                  <NA>
                         NΑ
                                  NΑ
## robust `[`: exclude NA values
dim(pdata[idx & !is.na(idx),])
```

CR

date.cr

#### ## [1] 17 21

Use the mol.biol column to subset the data to contain just individuals with 'BCR/ABL' or 'NEG', e.g.,

```
bcrabl <- subset(pdata, mol.biol %in% c("BCR/ABL", "NEG"))</pre>
```

The mol.biol column is a factor, and retains all levels even after subsetting. It is sometimes convenient to retain factor levels, but in our case we use droplevels() to removed unused levels

```
bcrabl$mol.biol <- droplevels(bcrabl$mol.biol)</pre>
```

The BT column is a factor describing B- and T-cell subtypes

#### levels(bcrabl\$BT)

```
"B1" "B2" "B3" "B4" "T" "T1" "T2" "T3" "T4"
```

How might one collapse B1, B2, ... to a single type B, and likewise for T1, T2, ..., so there are only two subtypes, B and T? One strategy is to replace two-letter level (e.g., B1) with the single-letter level (e.g., B). Do this using substring() to select the first letter of level, and update the previous levels with the new value using levels<-.

```
table(bcrabl$BT)
```

```
##
##
   B B1 B2 B3 B4
                  T T1 T2 T3 T4
   4 9 35 22 9 5 1 15 9 2
levels(bcrabl$BT) <- substring(levels(bcrabl$BT), 1, 1)</pre>
table(bcrabl$BT)
```

```
##
## B T
## 79 32
```

Use aggregate() to count the number of samples with B- and T-cell types in each of the BCR/ABL and NEG groups

```
aggregate(rownames(bcrabl) ~ BT + mol.biol, bcrabl, length)
```

```
## BT mol.biol rownames(bcrabl)
## 1 B BCR/ABL 37
## 2 B NEG 42
## 3 T NEG 32
```

Use aggregate() to calculate the average age of males and females in the BCR/ABL and NEG treatment groups.

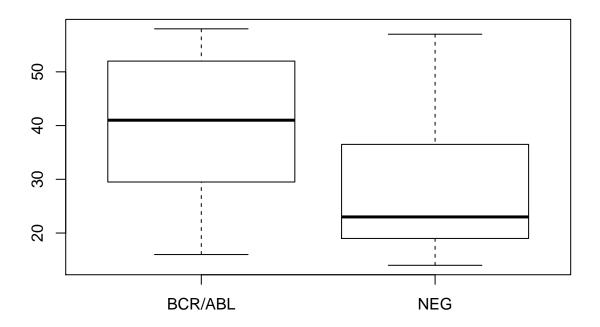
```
aggregate(age ~ mol.biol + sex, bcrabl, mean)
```

```
##    mol.biol sex        age
## 1    BCR/ABL        F     39.93750
## 2        NEG        F     30.42105
## 3    BCR/ABL        M     40.50000
## 4        NEG        M     27.21154
```

Use t.test() to compare the age of individuals in the BCR/ABL versus NEG groups; visualize the results using boxplot(). In both cases, use the formula interface. Consult the help page ?t.test and re-do the test assuming that variance of ages in the two groups is identical. What parts of the test output change?

```
t.test(age ~ mol.biol, bcrabl)
```

```
##
   Welch Two Sample t-test
##
##
## data: age by mol.biol
## t = 4.8172, df = 68.529, p-value = 8.401e-06
## alternative hypothesis: true difference in means is not equal to 0
## 95 percent confidence interval:
    7.13507 17.22408
##
## sample estimates:
## mean in group BCR/ABL
                             mean in group NEG
                40.25000
                                      28.07042
boxplot(age ~ mol.biol, bcrabl)
```



## 12 Exploration and simple univariate measures

```
path <- file.choose() # look for BRFSS-subset.csv

stopifnot(file.exists(path))
brfss <- read.csv(path)</pre>
```

#### 12.1 Clean data

R read Year as an integer value, but it's really a factor

```
brfss$Year <- factor(brfss$Year)</pre>
```

#### 12.2 Weight in 1990 vs. 2010 Females

Create a subset of the data

##

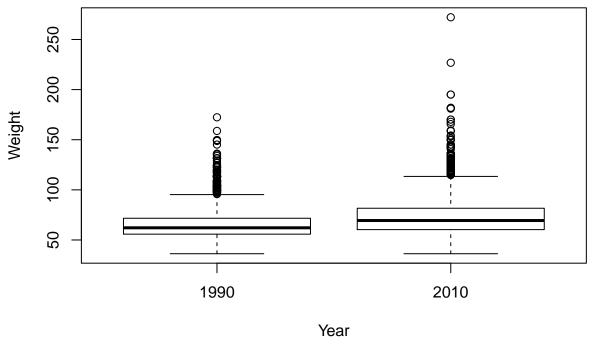
```
brfssFemale <- brfss[brfss$Sex == "Female",]
summary(brfssFemale)</pre>
```

```
##
                                            Sex
         Age
                          Weight
                                                            Height
           :18.00
                           : 36.29
                                        Female: 12039
                                                               :105.0
##
    Min.
                     Min.
                                                        Min.
##
    1st Qu.:37.00
                     1st Qu.: 57.61
                                        Male :
                                                        1st Qu.:157.5
    Median :52.00
                     Median: 65.77
                                                        Median :163.0
##
    Mean
           :51.92
                            : 69.05
                                                                :163.3
                     Mean
                                                        {\tt Mean}
                     3rd Qu.: 77.11
##
    3rd Qu.:67.00
                                                        3rd Qu.:168.0
##
    Max.
            :99.00
                     Max.
                             :272.16
                                                        Max.
                                                                :200.7
                     NA's
                             :560
                                                        NA's
##
    NA's
            :103
                                                                :140
##
      Year
##
    1990:5718
    2010:6321
##
```

```
##
##
##
##
```

#### Visualize

```
plot(Weight ~ Year, brfssFemale)
```



#### Statistical test

```
t.test(Weight ~ Year, brfssFemale)
```

```
##
## Welch Two Sample t-test
##
## data: Weight by Year
## t = -27.133, df = 11079, p-value < 2.2e-16
## alternative hypothesis: true difference in means is not equal to 0
## 95 percent confidence interval:
## -8.723607 -7.548102
## sample estimates:
## mean in group 1990 mean in group 2010
## 64.81838 72.95424</pre>
```

#### 12.3 Weight and height in 2010 Males

Create a subset of the data

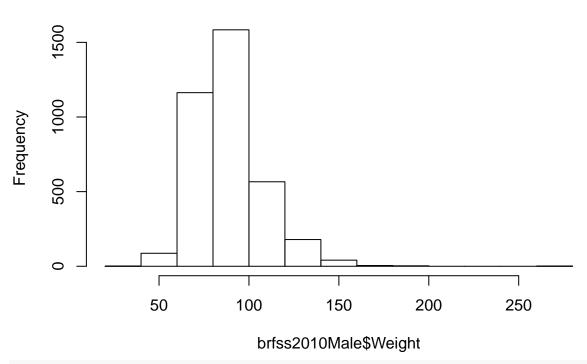
```
brfss2010Male <- subset(brfss, Year == 2010 & Sex == "Male")
summary(brfss2010Male)
         Age
                        Weight
                                         Sex
                                                        Height
                                                                    Year
## Min.
          :18.00
                          : 36.29
                                               0
                                                                  1990:
                    Min.
                                     Female:
                                                   Min.
                                                           :135
```

```
1st Qu.:45.00
                    1st Qu.: 77.11
                                      Male :3679
                                                    1st Qu.:173
                                                                   2010:3679
##
    Median :57.00
                    Median : 86.18
                                                    Median:178
           :56.25
                    Mean
                           : 88.85
                                                    Mean
                                                            :178
##
    3rd Qu.:68.00
                    3rd Qu.: 99.79
                                                    3rd Qu.:183
    Max.
           :99.00
                            :278.96
                                                    Max.
                                                            :218
##
                    Max.
    NA's
                    NA's
                                                    NA's
##
           :30
                            :49
                                                            :31
```

Visualize the relationship

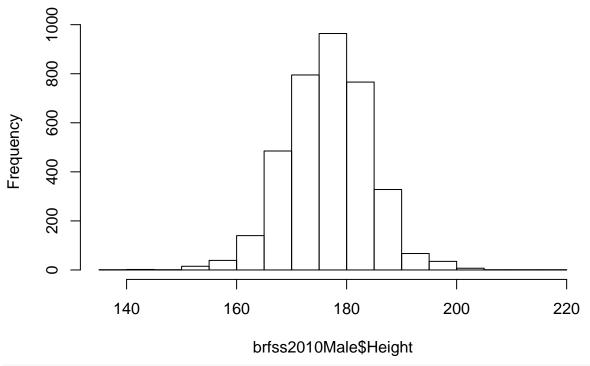
hist(brfss2010Male\$Weight)

# Histogram of brfss2010Male\$Weight

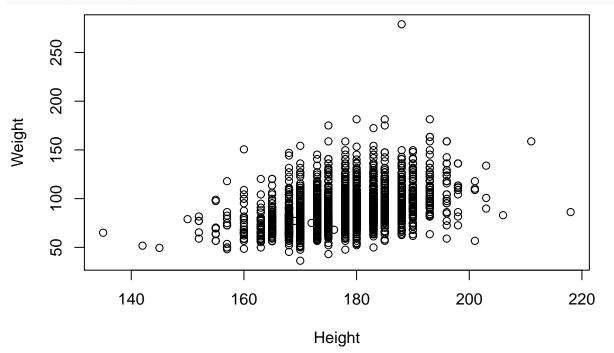


hist(brfss2010Male\$Height)

# Histogram of brfss2010Male\$Height



plot(Weight ~ Height, brfss2010Male)



Fit a linear model (regression)

```
fit <- lm(Weight ~ Height, brfss2010Male)
fit</pre>
```

##

## Call:

```
## lm(formula = Weight ~ Height, data = brfss2010Male)
##
## Coefficients:
  (Intercept)
##
                      Height
      -86.8747
                      0.9873
Summarize as ANOVA table
anova(fit)
## Analysis of Variance Table
##
## Response: Weight
##
               Df Sum Sq Mean Sq F value
                                               Pr(>F)
                1 197664 197664
                                     693.8 < 2.2e-16 ***
## Height
## Residuals 3617 1030484
                               285
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
Plot points, superpose fitted regression line; where am I?
plot(Weight ~ Height, brfss2010Male)
abline(fit, col="blue", lwd=2)
points(180, 88, col="red", cex=4, pch=20)
                                                         0
     250
     200
Weight
     150
                                                                             0
                                  0
                                                                      0
     100
                                                                                   0
                                                                        0
                140
                                 160
                                                  180
                                                                  200
                                                                                   220
                                             Height
Class and available 'methods'
class(fit)
                            # 'noun'
methods(class=class(fit))
                            # 'verb'
Diagnostics
```

plot(fit)
?plot.lm

### 13 Multivariate analysis

This is a classic microarray experiment. Microarrays consist of 'probesets' that interogate genes for their level of expression. In the experiment we're looking at, there are 12625 probesets measured on each of the 128 samples. The raw expression levels estimated by microarray assays require considerable pre-processing, the data we'll work with has been pre-processed.

#### 13.1 Input and setup

Start by finding the expression data file on disk.

```
path <- file.choose() # look for ALL-expression.csv
stopifnot(file.exists(path))</pre>
```

The data is stored in 'comma-separate value' format, with each probeset occupying a line, and the expression value for each sample in that probeset separated by a comma. Input the data using read.csv(). There are three challenges:

- 1. The row names are present in the first column of the data. Tell R this by adding the argument row.names=1 to read.csv().
- 2. By default, R checks that column names do not look like numbers, but our column names do look like numbers. Use the argument check.colnames=FALSE to over-ride R's default.
- 3. read.csv() returns a data.frame. We could use a data.frame to work with our data, but really it is a matrix() the columns are of the same type and measure the same thing. Use as.matrix() to coerce the data.frame we input to a matrix.

```
exprs <- read.csv(path, row.names=1, check.names=FALSE)
exprs <- as.matrix(exprs)</pre>
class(exprs)
## [1] "matrix"
dim(exprs)
## [1] 12625
               128
exprs[1:6, 1:10]
##
                01005
                          01010
                                    03002
                                             04006
                                                      04007
                                                                 04008
## 1000_at
             7.597323
                       7.479445 7.567593 7.384684 7.905312
                                                             7.065914
## 1001 at
             5.046194
                      4.932537 4.799294 4.922627 4.844565 5.147762
## 1002 f at 3.900466
                       4.208155 3.886169 4.206798 3.416923
                                                             3.945869
## 1003 s at 5.903856
                       6.169024 5.860459 6.116890 5.687997
## 1004 at
             5.925260 5.912780 5.893209 6.170245 5.615210 5.923487
## 1005 at
             8.570990 10.428299 9.616713 9.937155 9.983809 10.063484
##
                 04010
                           04016
                                    06002
                                               08001
## 1000 at
              7.474537
                        7.536119 7.183331
                                            7.735545
                        5.016132 5.288943
## 1001_at
              5.122518
                                            4.633217
## 1002_f_at
              4.150506
                        3.576360 3.900935
                                            3.630190
## 1003_s_at
                        5.665991 5.842326
              6.292713
                                            5.875375
## 1004_at
              6.046607 5.738218 5.994515
                                            5.748350
## 1005_at
             10.662059 11.269115 8.812869 10.165159
range(exprs)
```

## [1] 1.984919 14.126571

We'll make use of the data describing the samples

```
# look for ALL-phenoData.csv
path <- file.choose()</pre>
stopifnot(file.exists(path))
pdata <- read.csv(path, row.names=1)</pre>
class(pdata)
## [1] "data.frame"
dim(pdata)
## [1] 128
head(pdata)
##
          cod diagnosis sex age BT remission CR
                                                    date.cr t.4.11. t.9.22.
## 01005 1005 5/21/1997
                           Μ
                              53 B2
                                            CR CR
                                                   8/6/1997
                                                               FALSE
                                                                        TRUE
## 01010 1010 3/29/2000
                              19 B2
                                            CR CR 6/27/2000
                                                               FALSE
                                                                       FALSE
                           Μ
## 03002 3002 6/24/1998
                           F
                              52 B4
                                            CR CR 8/17/1998
                                                                  NA
                                                                          NA
## 04006 4006 7/17/1997
                                            CR CR 9/8/1997
                                                                TRUE
                                                                       FALSE
                              38 B1
                           М
## 04007 4007 7/22/1997
                           М
                              57 B2
                                            CR CR 9/17/1997
                                                               FALSE
                                                                       FALSE
                                                               FALSE
## 04008 4008 7/30/1997
                             17 B1
                                            CR CR 9/27/1997
                                                                       FALSE
##
         cyto.normal
                             citog mol.biol fusion.protein mdr
                                                                   kinet
                                                                            ccr
## 01005
                           t(9;22)
                                    BCR/ABL
                                                       p210 NEG dyploid FALSE
               FALSE
## 01010
               FALSE simple alt.
                                        NEG
                                                       <NA> POS dyploid FALSE
## 03002
                  NA
                              <NA> BCR/ABL
                                                       p190 NEG dyploid FALSE
## 04006
               FALSE
                           t(4;11) ALL1/AF4
                                                       <NA> NEG dyploid FALSE
## 04007
               FALSE
                           del(6q)
                                        NEG
                                                       <NA> NEG dyploid FALSE
## 04008
               FALSE complex alt.
                                        NEG
                                                       <NA> NEG hyperd. FALSE
##
         relapse transplant
                                            f.u date.last.seen
## 01005
           FALSE
                        TRUE BMT / DEATH IN CR
                                                           <NA>
## 01010
                       FALSE
            TRUE
                                            REL
                                                     8/28/2000
## 03002
            TRUE
                       FALSE
                                            REL
                                                    10/15/1999
## 04006
            TRUE
                       FALSE
                                            REL
                                                     1/23/1998
                                                     11/4/1997
## 04007
            TRUE
                       FALSE
                                            REL
            TRUE
                      FALSE
## 04008
                                            REL
                                                    12/15/1997
```

Some of the results below involve plots, and it's convenient to choose pretty and functional colors. We use the RColorBrewer package; see colorbrewer.org

```
library(RColorBrewer) ## not available? install package via RStudio
highlight <- brewer.pal(3, "Set2")[1:2]</pre>
```

'highlight' is a vector of length 2, light and dark green.

For more options see ?RColorBrewer and to view the predefined palettes display.brewer.all()

#### 13.2 Cleaning

We'll add a column to pdata, derived from the BT column, to indicate whether the sample is B-cell or T-cell ALL.

```
pdata$BorT <- factor(substr(pdata$BT, 1, 1))
```

Microarray expression data is usually represented as a matrix of genes as rows and samples as columns. Statisticians usually think of their data as samples as rows, features as columns. So we'll transpose the expression values

```
exprs <- t(exprs)</pre>
```

Confirm that the pdata rows correspond to the exprs rows.

stopifnot(identical(rownames(pdata), rownames(exprs)))

### 13.3 Unsupervised machine learning – multi-dimensional scaling

Reduce high-dimensional data to lower dimension for visualization.

Calculate distance between samples (requires that the expression matrix be transposed).

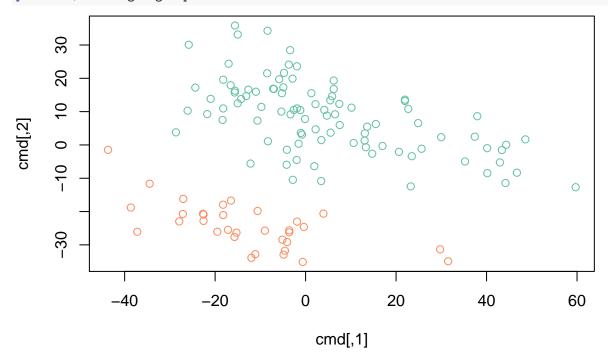
```
d <- dist(exprs)</pre>
```

Use the cmdscale() function to summarize the distance matrix into two points in two dimensions.

```
cmd <- cmdscale(d)</pre>
```

Visualize the result, coloring points by B- or T-cell status

plot(cmd, col=highlight[pdata\$BorT])



# 14 Using R in real life

#### 14.1 Organizing work

Usually, work is organized into a directory with:

- A folder containing R scripts (scripts/BRFSS-visualize.R)
- 'External' data like the csv files that we've been working with, usually in a separate folder (extdata/BRFSS-subset.csv)

- (sometimes) R objects written to disk using saveRDS() (.rds files) that represent final results or intermediate 'checkpoints' (extdata/ALL-cleaned.rds). Read the data into an R session using readRDS().
- Use setwd() to navigate to folder containing scripts/, extdata/ folder
- Source an entire script with source("scripts/BRFSS-visualization.R").

R can also save the state of the current session (prompt when choosing to quit() R), and to view and save the history() of the the current session; I do not find these to be helpful in my own work flows.

#### 14.2 R Packages

All the functionality we have been using comes from packages that are automatically loaded when R starts. Loaded packages are on the search() path.

```
search()
```

Additional packages may be *installed* in R's libraries. Use 'installed.packages() or the RStudio interface to see installed packages. To use these packages, it is necessary to attach them to the search path, e.g., for survival analysis

```
library("survival")
```

There are many thousands of R packages, and not all of them are installed in a single installation. Important repositories are

- CRAN: https://cran.r-project.org/
- Bioconductor: https://bioconductor.org/packages

Packages can be discovered in various ways, including CRAN Task Views and the Bioconductor web and Bioconductor support sites.

To install a package, use install.packages() or, for *Bioconductor* packages, instructions on the package landing page, e.g., for GenomicRanges. Here we install the ggplot2 package.

```
install.packages("ggplot2", repos="https://cran.r-project.org")
```

A package needs to be installed once, and then can be used in any R session.

# 15 Graphics and Visualization

Load the BRFSS-subset.csv data

```
path <- "BRFSS-subset.csv" # or file.choose()
brfss <- read.csv(path)</pre>
```

Clean it by coercing Year to factor

```
brfss$Year <- factor(brfss$Year)
```

#### 15.1 Base R Graphics

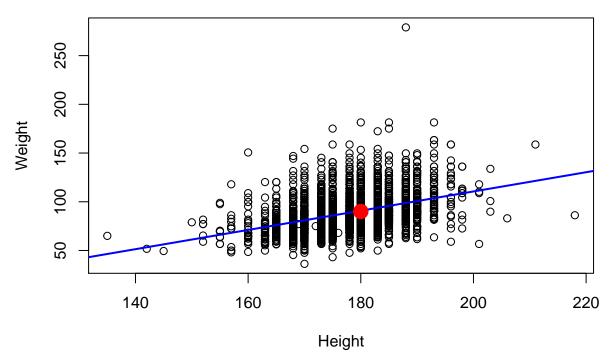
Useful for quick exploration during a normal work flow.

- Main functions: plot(), hist(), boxplot(), ...
- Graphical parameters see ?par, but often provided as arguments to plot(), etc.
- $\bullet$  Construct complicated plots by layering information, e.g., points, regression line, annotation.

```
brfss2010Male <- subset(brfss, (Year == 2010) & (Sex == "Male"))
fit <- lm(Weight ~ Height, brfss2010Male)

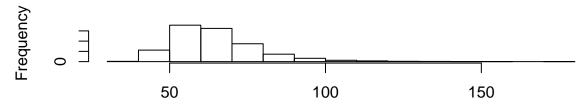
plot(Weight ~ Height, brfss2010Male, main="2010, Males")
abline(fit, lwd=2, col="blue")
points(180, 90, pch=20, cex=3, col="red")</pre>
```

## **2010, Males**



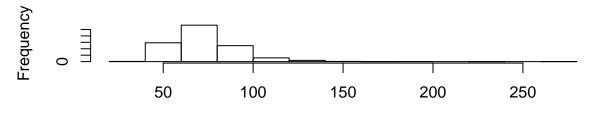
• Approach to complicated graphics: create a grid of panels (e.g., par(mfrows=c(1, 2)), populate with plots, restore original layout.





brfssFemale[brfssFemale\$Year == 1990, "Weight"]

# Female, 2010



brfssFemale[brfssFemale\$Year == 2010, "Weight"]

par(opar) # restore original layout

### 15.2 What makes for a good graphical display?

- Common scales for comparison
- Efficient use of space
- Careful color choice qualitative, gradient, divergent schemes; color blind aware; ...
- Emphasis on data rather than labels
- Convey statistical uncertainty

## 15.3 Grammar of Graphics: ggplot2

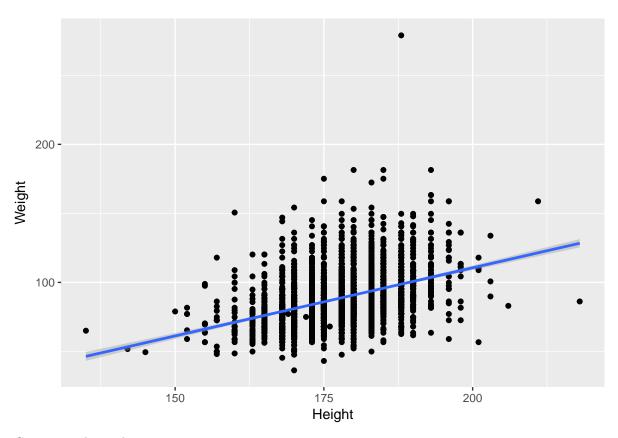
#### library(ggplot2)

• http://docs.ggplot2.org

'Grammar of graphics'

- Specify data and 'aesthetics' (aes()) to be plotted
- Add layers (geom\_\*()) of information

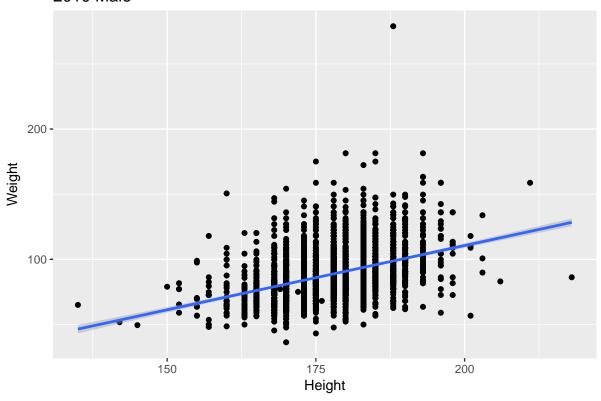
```
ggplot(brfss2010Male, aes(x=Height, y=Weight)) +
   geom_point() +
   geom_smooth(method="lm")
```



• Capture a plot and augment it

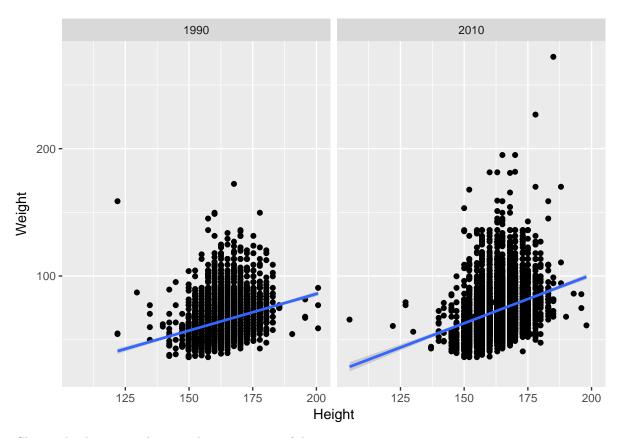
```
plt <- ggplot(brfss2010Male, aes(x=Height, y=Weight)) +
    geom_point() +
    geom_smooth(method="lm")
plt + labs(title = "2010 Male")</pre>
```

## 2010 Male



• Use facet\_\*() for layouts

```
ggplot(brfssFemale, aes(x=Height, y=Weight)) +
    geom_point() + geom_smooth(method="lm") +
    facet_grid(. ~ Year)
```



• Choose display to emphasize relevant aspects of data

```
ggplot(brfssFemale, aes(Weight, fill=Year)) +
   geom_density(alpha=.2)
```

